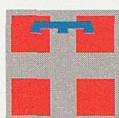
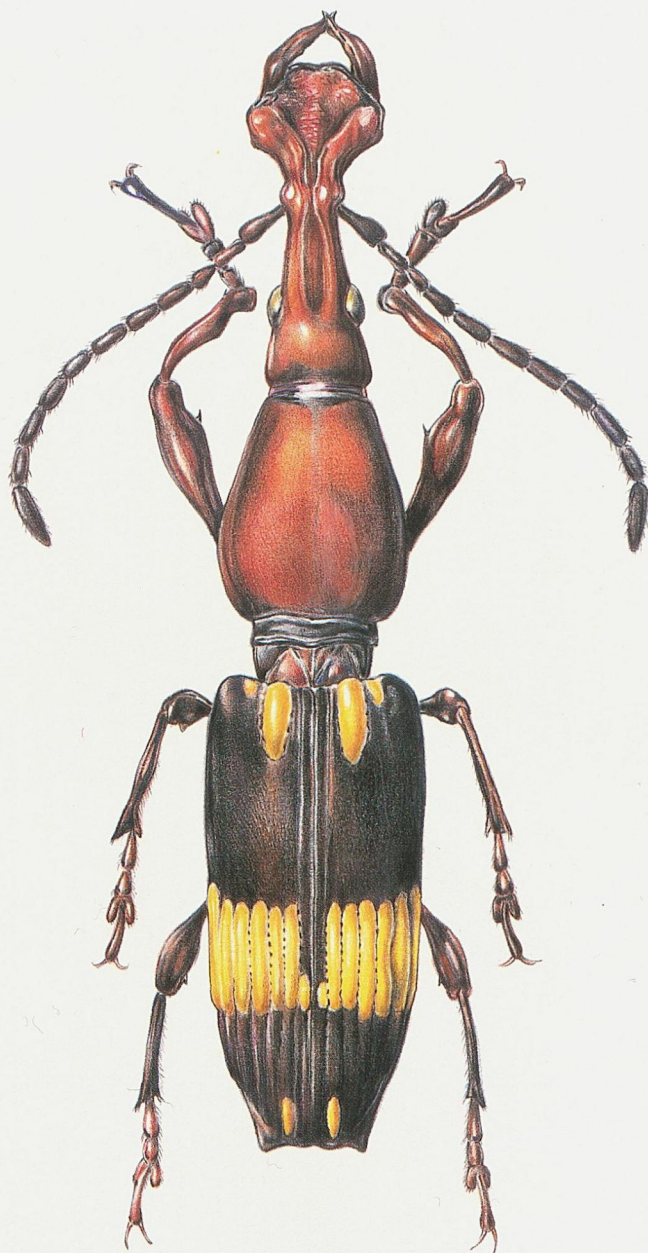


Alessandra SFORZI - Luca BARTOLOZZI
Editors

Brentidae of the world (Coleoptera, Curculionoidea)



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Museo Regionale di Scienze Naturali

MONOGRAFIE XXXIX

2004

The family Brentidae treated here includes the following sub-families and tribes: Brentinae, Cyphagoginae, Pholidochlamydinae, Taphroderinae, Trachelizinae, Ulocerinae (Brentidae *sensu Auctorum*), plus Antliarhininae, Cyladinae, Eurhynchinae, and Microcerini.

This book, which updates the last checklist published by Richard Kleine in 1938, includes notes on the morphology, biology, ecology, ethology and zoo-geography of the Brentidae, as well as an historical review and a complete bibliography. All the described taxa have been catalogued.

At present, the Brentidae *sensu Auctorum* (Brentinae, Cyphagoginae, Pholidochlamydinae, Taphroderinae, Trachelizinae, Ulocerinae) comprise 293 genera and 1690 species, for the most part from the circumtropics. The Antliarhininae are all African, with 2 genera and 7 species and are exclusively associated with cycads of the genus *Encephalartos*. The Cyladinae consist of the single circumtropical genus *Cylas*, with 24 species. The Eurhynchinae have 3 genera with 30 species from Australia and New Guinea. Finally, the Microcerini comprise 3 genera with 67 species of sub-Saharan distribution.

Alessandra Sforzi was born in Pistoia (Italy) on 1 October 1966. She began studying Brentidae in 1991, after graduating from the University of Florence with a degree in Biological Sciences and now has a contract as Researcher in the Entomology Department of the Florence University Natural History Museum (Zoology Section). She has participated in various scientific expeditions in Africa, Asia and South America, and has worked on the Brentidae collections in the main European Museums. She has published many papers dealing with the taxonomy and faunistics of this family and has described several new taxa.

Luca Bartolozzi was born in Florence (Italy) on 17 October 1954. He is the Curator of the Entomology Department of the Florence University Natural History Museum (Zoology Section) and a specialist of Coleoptera Brentidae and Lucanidae. The author of over 100 scientific papers describing many new taxa, he has collected in Africa, Asia, Oceania and South America, and has worked on the Brentidae collections in the main European Museums.

The chapters on Antliarhininae and Eurhynchinae were written by Rolf Oberprieler, Research Scientist at CSIRO Entomology, Canberra, Australia and a specialist of weevil systematics.

The chapter on Cyladinae was written by Miguel Angel Alonso-Zarazaga, Entomologist at the Museo Nacional de Ciencias Naturales in Madrid, Spain and a specialist of weevil systematics and the taxonomy of Apionidae and Nanophyidae.

The chapter on Microcerini was written by Schalk vdM Louw, Professor at the Department of Zoology & Entomology of the University of the Free State, Bloemfontein, South Africa and a specialist of weevil systematics and taxonomy, and insect-plant interaction.

Front cover:

Estenorhynus sp. from Ecuador (drawing by Cristina Girard)

Back cover:

Nemorhynus myrmecophagus (Herbst) (plate from Labram & Imhoff, 1838)

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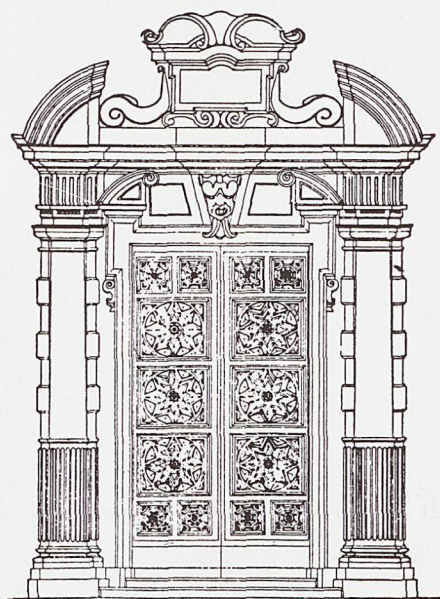
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MONOGRAFIE XXXIX

Brentidae of the world (Coleoptera, Curculionoidea)

Alessandra SFORZI and Luca BARTOLOZZI
Editors





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*This book is dedicated to all those
who have dealt with this
fascinating family
of Coleoptera, so rich in strange
and various forms*

“To conclude, it cannot too often be recommended and enjoined, that men’s diligence in investigating and amassing natural history be henceforward entirely changed, and turned into the direction opposite to that now in use. For hitherto men have used great and indeed over-curious diligence in observing the variety of things, and explaining the exact specific differences of animals, herbs, and fossils; most of which are rather sports of nature than of any serious use towards science. Such things indeed serve to delight, and sometimes even give help in practice; but for getting insight into nature they are of little service or none. Men’s labour therefore should be turned to the investigation and observation of the resemblances and analogies of things, as well in wholes as in parts. For these it is that detect the unity of nature, and lay a foundation for the constitution of sciences.”

Francis Bacon, 1620, *Novum Organum*, II, 27

"To conclude, it cannot too often be recommended and expected that men's diligence in investigating and examining natural history be transformed entirely changed, and turned into the direction opposite to that now in use. For hitherto men have used given and indeed over-certain diligence in observing the variety of things, and explaining the exact specific differences of animals, herbs, and fossils; most of which are rather sports of nature than of any serious use towards science. Such things indeed serve to delight, and sometimes even give help in practice; but for getting insight into nature they are of little service or none. Men's labour therefore should be turned to the investigation and observation of the resemblances and analogies of things, as well in wholes as in parts. For there it is that even the unity of nature, and lay a foundation for the constitution of sciences."

Francis Bacon, 1620, *Novum Organum*, II, 21

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Individual contributions may be cited thus:

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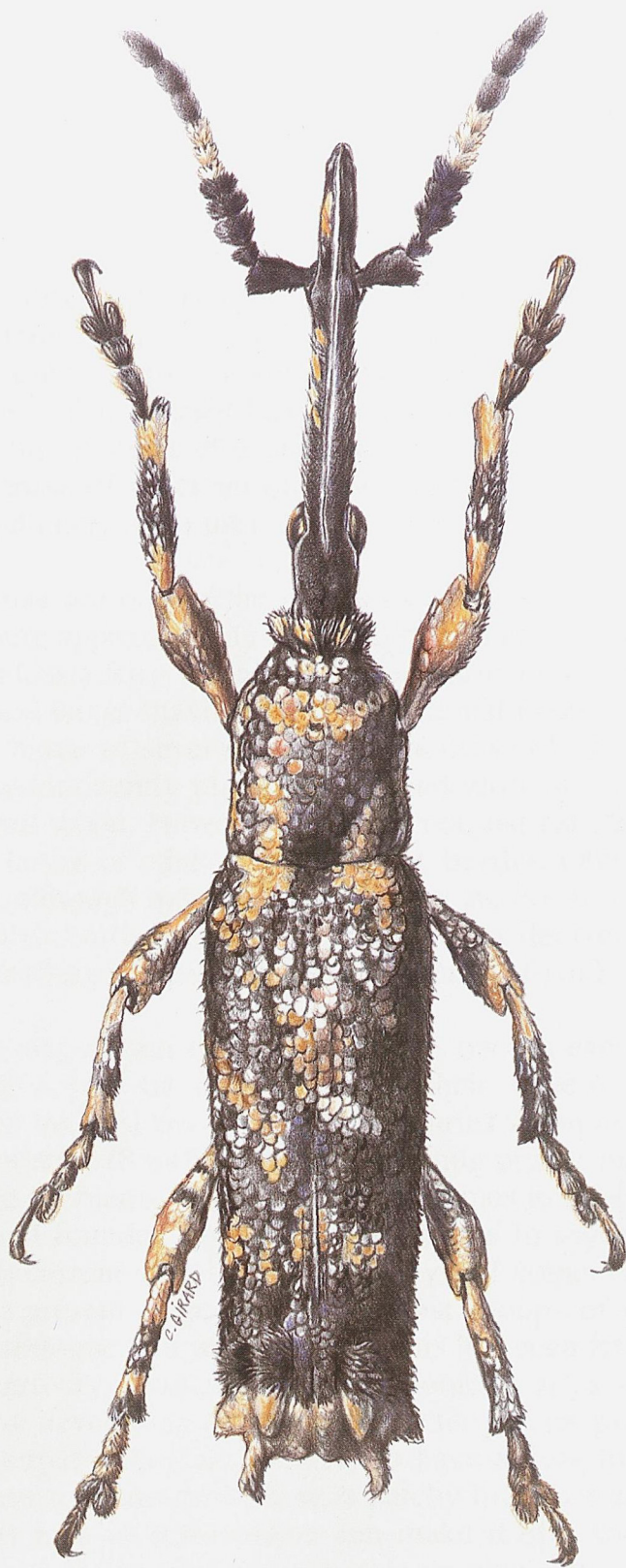


Plate I - *Pholidochlamys madagascariensis* Lacordaire, Madagascar (drawing by Cristina Girard).

FOREWORD

This book started life as a catalogue, a type of product generally considered by taxonomists and others as both extraordinarily useful and monumentally dull. However, in its gestation it has grown, and now is nothing less than a natural history of an extraordinary and diverse family of insects, together with the essential element of a checklist of all the subfamilies, genera and species and a mass of other information of great interest and importance to taxonomists and many other users.

The Brentidae are one of the families of weevils, the huge insect superfamily containing approximately one-fifth of all beetles. My first introduction to them was in Costa Rica when a fallen tree turned out, once we had reduced it to sawdust and wood shavings in the traditional manner of coleopterists, to have been the home of several hundred Brentidae of three different species. Weevils are predominantly plant-feeding, and most of the Brentidae develop as larvae in dead wood. However, some do not just eat the wood, but instead prey upon the larvae of other wood-dwelling beetles. Others live on or in living plants and, although most of the Brentidae are creatures of the moist tropics, one group recently added to the family is a denizen of arid lands and rather than tunnelling in trees species are generally found under stones.

Catalogues may appear dull (although this one, as even a slight glance inside it will tell, is very far from dull!), but their value is without doubt very great indeed. In the past two and a half centuries taxonomists have described somewhere around 1.78 million species of living organisms, and are currently facing up to the problems inherent in giving names to the other 4-100 million. Even in the well-founded taxonomic institutions in industrialized countries, discovering the current state of the taxonomy and nomenclature of a group of organisms can present huge problems. Most groups of organisms do have some sort of catalogue, but in many cases this has been left behind by the tide of taxonomic activity. Workers striving to come to grips with insects such as the Brentidae in developing countries face a far greater problem. Unlike their northern counterparts they are unlikely to have access to even the old catalogues, and have to make do with such patchy literature as they can lay their hands on. This lack of information can make it effectively impossible for them to take part in the global taxonomic community, and can also lead to their countries being unable to determine what species are found within their borders. Given that obligations of countries under the Convention on Biologi-

cal Diversity include developing inventories of their biodiversity, this barrier to taxonomic activity can be of considerable significance. In the case of the Brentidae, the last catalogue at species level was published in 1938, and certainly did not include all the subfamilies and tribes now considered part of this family. There have been some regional publications, but nothing approaching the completeness of the present volume. This treatment cannot be viewed as simply a publication for taxonomists to mull over in dusty corners, but as a tool in the global armoury of conservation and sustainable use.

Right across the natural world there is a considerable effort underway to make biodiversity information globally available via the internet. This programme is extremely laudable, and deserves every support. However, although funding is being directed at this process to some extent, much less support is being given to efforts to populate this global database – to compiling the catalogues and checklists on which it will be based. There are a number of possible reasons for this, both from the point of view of funding bodies, many of which direct their support at ‘cutting-edge’ work but not at information gathering at the more scholarly end of the research spectrum, and from scientists, who see few papers in high-profile journals coming out of cataloguing. The work required is less glamorous and, arguably, more difficult than some other areas of research and technology development. The needs for catalogues are clear, but the capabilities, scholarship and the energy to actually compile them are far from common.

This catalogue has been compiled by a group of people with all of the necessary background, skills and energy. The editors have authored the section on the Brentidae (as the family has been considered in the past) themselves but, not content with that, have enlisted Miguel Alonso-Zarazaga, Rolf Oberprieler and Schalk Louw to cover the family-groups that have been added on to the Brentidae in recent years – the Cyladinae, Antliarhininae, Eurhynchinae and Microcerini. This area of the great superfamily of weevils has been the focus of a great deal of systematic study in the past ten years, and what looked a few years ago to be a well-defined group of primarily xylophagous beetles is now expanded to include weevils tunnelling in the seeds of cycads, feeding in sweet potatoes, forming galls, and ground-dwelling in arid regions. Very often the changes made by taxonomists as they attempt to move classifications closer to a phylogenetically-accurate arrangement are made available to the wider biological community only through scattered publications in a variety of academic journals. This catalogue is an excellent model of a taxonomic output required to provide timely overviews for the wider community and to help people understand the results of the work that has taken place.

In addition to the high-level needs mentioned above, one of the gaps in weevil publications has been of treatments that give a good background to the

group. Instead of these some regional or national publications have been the only resources for people who just want to know more about this fascinating group, since observations that have been made are scattered through the literature. This volume for the first time brings together information about Brentidae on larval and adult biology, as well as zoogeographic information and morphology. This really will be an invaluable resource for students, researchers and amateurs alike.

This volume has been compiled with enthusiasm, scholarship and intelligence; it will be used for many years as a source of information in many different lands and for many different purposes. It will also highlight and encourage research into a fascinating and important group of beetles. Taxonomy has a poor image in some quarters. Books like this serve to dispel that erroneous perception.

Christopher H. C. Lyal
The Natural History Museum
London, September 2003

PREFACE

For several years we have been interested in the taxonomy and faunistics of Coleoptera Brentidae. Over this period, we realised the necessity of updating the last checklist, published as many as 66 years ago by Kleine in the *Genera Insectorum*, to include all the new taxa described since and consider the changes in the taxonomy and systematics of the family that have occurred.

So, five years ago we set out on our "adventure" blissfully unaware of the demanding job ahead. As we proceeded, we realised the extent and nature of the data we had to verify and it became indispensable to examine the type material deposited in the most important museums in Europe and consult old and often rare works. This was in great part made possible through the financial support from the European Union's IHP (Improving Human Potential) Programme (1998-2002).

At first, we thought we would just provide a simple checklist, but we soon realised the benefits of including a general section on the family offering an updated picture of our knowledge on its morphology, biology, ethology, ecology and zoo-geography.

For classification purposes we decided to follow the excellent "World catalogue of families and genera of Curculionoidae" (Alonso-Zarazaga & Lyal, 1999, 2002). Consequently, we treat the Brentidae *sensu Auctorum* (Brentinae, Cyphagoginae, Pholidochlamyinae, Taphroderinae, Trachelizinae, Ulocerinae), whilst Miguel Alonso-Zarazaga and Rolf Oberprieler cover the Cyladinae and Anthliarhininae respectively. We then include two more chapters, one on the Eurhynchinae (by Rolf Oberprieler) and one on the Microcerini (by Schalk Louw); these two groups in fact are considered by some authors as belonging to the brentids, although their phylogenetic position still awaits critical scrutiny.

At present, the Brentidae *sensu Auctorum* (Brentinae, Cyphagoginae, Pholidochlamyinae, Taphroderinae, Trachelizinae, Ulocerinae) comprise 293 genera and 1690 species, for the most part of circumtropical distribution. The Anthliarhininae are exclusively African, with 2 genera and 7 species; they are entirely associated with cycads of the genus *Encephalartos*. The Cyladinae consist of a single circumtropical genus, with 24 species. The Eurhynchinae

have 3 genera with 30 species that live in Australia and New Guinea. Finally, the Microcerini comprise 3 genera with 67 species of sub-Saharan distribution.

We hope that our work will prove to be not only a useful tool for specialists, but also that young entomologists will find it a stimulus for studying this fascinating family of beetles.

The Editors
Alessandra Sforzi
Luca Bartolozzi

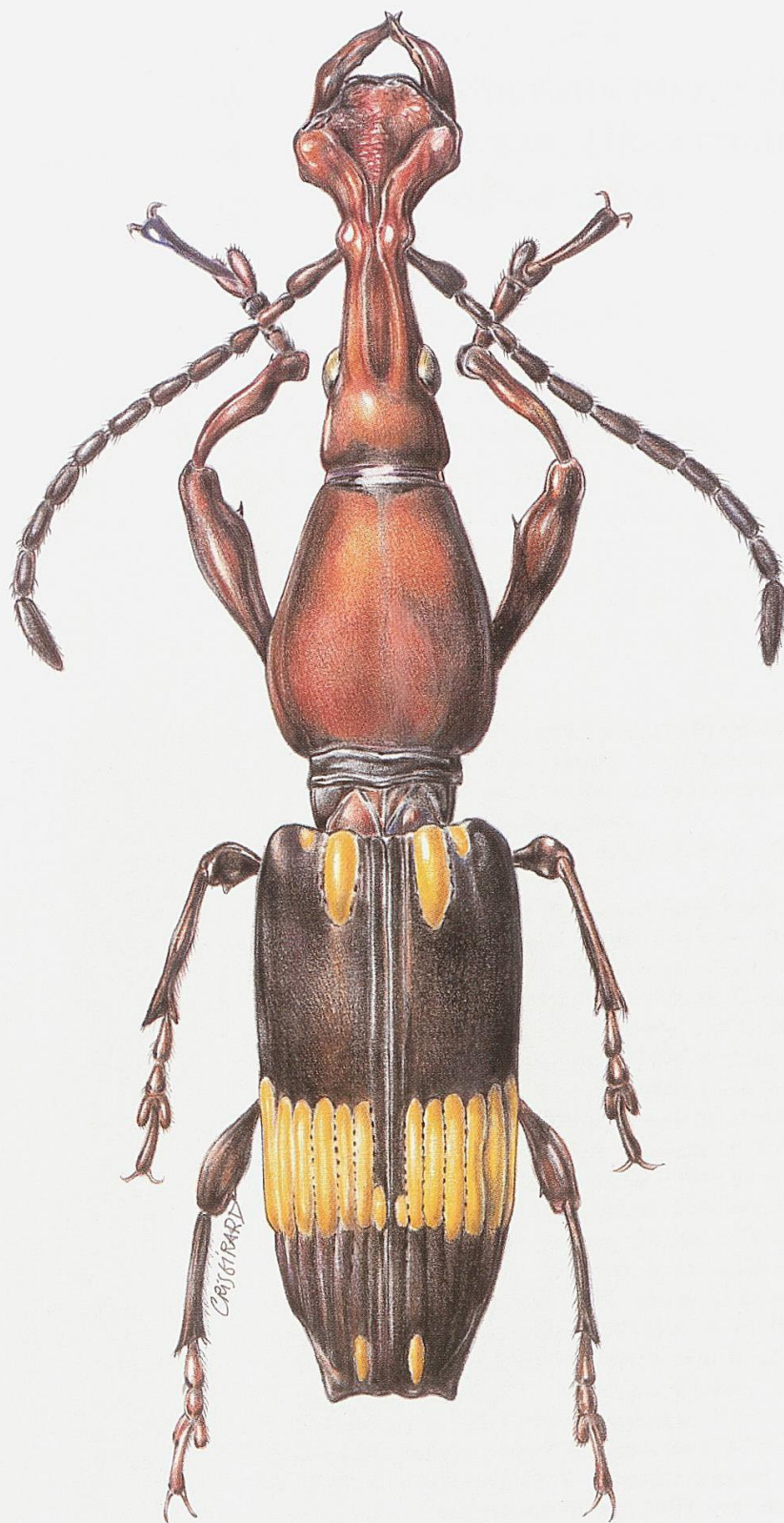


Plate II - *Estenorhinus* sp., Ecuador (drawing by Cristina Girard).

Brentidae Billberg, 1820 (Brentinae, Cyphagoginae, Pholidochlamydinae, Taphroderinae, Trachelizinae, Ulocerinae) (Coleoptera, Curculionoidea)

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ABSTRACT

This publication lists 293 genera with 1690 valid species of Brentidae (Brentinae, Cyphagoginae, Pholidochlamydinae, Taphroderinae, Trachelizinae, Ulocerinae). Their morphology, biology, ecology, ethology and zoo-geography, as well as an historical review, are summarized, and all the described species are catalogued. A complete bibliography is also given.

The new taxon *Ithystenus bicolor* Sforzi & Bartolozzi **n. sp.** is established to replace *I. bicolor* Kleine, 1919 (non Guérin-Méneville, 1831).

The following two generic new synonymies are proposed: *Homales* Kolbe, 1883 **syn. n.** of *Pia-zocnemis* Lacordaire, 1866; *Tinoteramocerus* Kleine, 1927 **syn. n.** of *Belorhynchus* Berthold, 1827.

The following 25 new synonymies are proposed: *Baryrhynchus lineicollis* f. *kleinei* Damoiseau, 1972 **syn. n.** of *Baryrhynchus (Eupsalomimus) lineicollis* Power, 1879; *Bolbocephalus mechowii* var. *peregrinus* Kolbe, 1916 **syn. n.** of *Bolbocranius mechowii* (Kolbe, 1883); *Brenthus metallicus* Boheman, 1840 **syn. n.** of *Amerismus cavicaudatus* (Chevrolat, 1839); *Brentus armillatus* var. *erythromerus* Kirsch, 1868 **syn. n.** of *Brentus armillatus* Kirsch, 1868; *Calorychodes decens* Kleine, 1925 **syn. n.** of *Calorychodes tuberculatus* (Senna, 1894); *Cerobates debilis* var. *sulcata* Kolbe, 1897 **syn. n.** of *Cerobates* (s. s.) *debilis* J. Thomson, 1858; *Claeoderes kirschi* Soares & Scivittaro, 1977 **syn. n.** of *Claeoderes radulirostris mexicanus* Boheman, 1840; *Curculio druryanus* Pallas, 1781 **syn. n.** of *Arrenodes minutus* Drury, 1773; *Diurus grootaerti* Damoiseau, 1989 **syn. n.** of *Ceocephalus philippinus* (Senna, 1911); *Heteroblysmia condensa* Kleine, 1926 **syn. n.** of *Ectocemus cinnamomi* (Herbst, 1783); *Higonius abruptus* Kleine, 1935 **syn. n.** of *Taphrocomister singularis* Senna, 1895; *Mygaleicus vittipennis* var. *nitida* Calabresi, 1920 **syn. n.** of *Gynandrorhynchus vittipennis* (Fåhræus, 1871); *Pericordus australis* Damoiseau, 1967 **syn. n.** of *Pericordus freyi* Damoiseau, 1966; *Prophthalmus potens* var. *amethystina* Kleine, 1916 **syn. n.** of *Prophthalmus potens potens* Lacordaire, 1866; *Prophthalmus tricolor* f. *celebensis* Kleine, 1916 **syn. n.** of *Prophthalmus tricolor* Power, 1878; *Prophthalmus tricolor* f. *ceramensis* Kleine, 1916 **syn. n.** of *Prophthalmus tricolor* Power, 1878; *Prophthalmus tricolor* f. *philippinensis* Kleine, 1916 **syn. n.** of *Prophthalmus tricolor* Power, 1878; *Schizotrachelus bakeri* f. *concolor* Kleine, 1922 **syn. n.** of *Tracheloschizus bakeri* Kleine, 1922; *Stereobates chiriquensis* Sharp, 1895 **syn. n.** of *Stereobates pedator* Sharp, 1895; *Stereobates gracilis* Sharp, 1895 **syn. n.** of *Stereobates pedator* Sharp, 1895; *Stereodermus carinatus* var. *tenuis* Sharp, 1895 **syn. n.** of *Stereodermus carinatus* Sharp, 1895; *Tinoteramocerus enormis* Kleine, 1927 **syn. n.** of *Belorhynchus nasutus* (Fabricius, 1781); *Trachelizus hirtellus* Sharp, 1895 **syn. n.** of *Paratrachelizus aureopilosus* (Senna, 1890); *Tychaeus*

curvidens var. *signatus* Sharp, 1895 **syn. n.** of *Nemorhinus myrmecophaga* (Herbst, 1797); *Zemioses camerunus* var. *nigricollis* Kolbe, 1916 **syn. n.** of *Zemioses porcatus* Pascoe, 1862.

The following **new combinations** are established: *Achrionota celebica* (Senna, 1897) **comb. n.**; *Belorhynchus stratiorrhinoides* (Kleine, 1942) **comb. n.**; *Dentisebus malgasicus* (Senna, 1895) **comb. n.**; *Hypomiolispis amoena* (Perroud, 1853) **comb. n.**; *H. rufovittata* (Perroud, 1853) **comb. n.**; *Kolbrentus metallicus* (Kolbe, 1883) **comb. n.**; *Metatrachelus unisulcatus* (Damoiseau, 1966) **comb. n.**; *Microsebus biakianus* (Damoiseau, 1989) **comb. n.**; *M. fijianus* (Damoiseau, 1989) **comb. n.**; *Microtrachelizus rectestriatus* (Fairmaire, 1897) **comb. n.**; *Neacratus argentinus* (Haedo Rossi, 1954) **comb. n.**; *N. brevicostatus* (Kleine, 1922) **comb. n.**; *N. clavipes* (Boheman, 1840) **comb. n.**; *N. crinitus* (Kleine, 1922) **comb. n.**; *N. deplanatus* (Sharp, 1895) **comb. n.**; *N. dolosus* (Kleine, 1927) **comb. n.**; *N. erectus* (Kleine, 1927) **comb. n.**; *N. famulus* (Boheman, 1840) **comb. n.**; *N. fasciatus* (Kleine, 1922) **comb. n.**; *N. femoratus* (Sharp, 1895) **comb. n.**; *N. fulgidus* (Kleine, 1928) **comb. n.**; *N. guatemalensis* (Senna, 1893) **comb. n.**; *N. longiceps* (Perroud, 1853) **comb. n.**; *N. muticus* (Kleine, 1922) **comb. n.**; *N. nodosus* (Kleine, 1937) **comb. n.**; *N. obtusus* (Lund, 1800) **comb. n.**; *N. piceus* (Perroud, 1853) **comb. n.**; *N. puncticeps* (Sharp, 1895) **comb. n.**; *N. punctulatus* (Boheman, 1840) **comb. n.**; *N. scotti* (Kleine, 1923) **comb. n.**; *N. tonsus* (Kleine, 1928) **comb. n.**; *Nemocephalus avarus* (Kleine, 1927) **comb. n.**; *N. enodis* (Kleine, 1927) **comb. n.**; *N. moderatus* (Kleine, 1922) **comb. n.**; *Orphanobrentus emarginatus* (Chevrolat, 1839) **comb. n.**; *Protoproctus minor* (K. E. Schedl, 1961) **comb. n.**; *Schizotrachelus javanicus* (Perroud, 1853) **comb. n.**; *Sennaiella devisa* (Kleine, 1926) **comb. n.**; *Tracheloschizus alienus* (Kleine, 1922) **comb. n.**; *T. bakeri* (Kleine, 1922) **comb. n.**; *T. birmanicus* (Senna, 1892) **comb. n.**; *T. brunneus* (Kleine, 1922) **comb. n.**; *T. cameratus* (Lacordaire, 1866) **comb. n.**; *T. castaneicolor* (Kleine, 1933) **comb. n.**; *T. consideratus* (Kleine, 1926) **comb. n.**; *T. consimilis* (Kleine, 1923) **comb. n.**; *T. corpulentus* (Kleine, 1922) **comb. n.**; *T. dichrous* (Lacordaire, 1866) **comb. n.**; *T. fortis* (Kleine, 1935) **comb. n.**; *T. gemellus* (Kleine, 1925) **comb. n.**; *T. generalis* (Kleine, 1925) **comb. n.**; *T. imbricellus* (Kleine, 1925) **comb. n.**; *T. imitator* (Kleine, 1925) **comb. n.**; *T. inconstans* (Kleine, 1922) **comb. n.**; *T. interruptus* (Senna, 1898) **comb. n.**; *T. intrusus* (Senna, 1898) **comb. n.**; *T. marginatus* (Senna, 1898) **comb. n.**; *T. metallicus* (Senna, 1897) **comb. n.**; *T. nanus* (Senna, 1894) **comb. n.**; *T. puncticollis* (Boheman, 1840) **comb. n.**; *T. quaesitus* (Kleine, 1936) **comb. n.**; *T. salomonensis* (Kleine, 1933) **comb. n.**; *T. scrobicollis* (Gyllenhal, 1833) **comb. n.**; *T. sculptiventris* (Senna, 1892) **comb. n.**; *T. suspensus* (Kleine, 1943) **comb. n.**; *T. timoriensis* (Senna, 1898) **comb. n.**; *T. viridescens* (Shaw, 1953) **comb. n.**; *T. vitalisi* (Calabresi, 1921) **comb. n.**; *Usambioproctus collarti* (Damoiseau, 1963) **comb. n.**; *U. compressicornis* (Kleine, 1916) **comb. n.**; *U. congoensis* (De Muizon, 1955) **comb. n.**; *U. conjunctus* (Kleine, 1916) **comb. n.**; *U. decellei* (Damoiseau, 1967) **comb. n.**; *U. dissimilis* (Kleine, 1916) **comb. n.**; *U. gracilis* (Kleine, 1916) **comb. n.**; *U. intermedius* (Damoiseau, 1961) **comb. n.**; *U. muizoni* (Damoiseau, 1963) **comb. n.**; *U. pierloti* (Damoiseau, 1967) **comb. n.**; *U. planistriatus* (Damoiseau, 1961) **comb. n.**; *U. rufus* (Kolbe, 1916) **comb. n.**; *U. tempestivus* (Kolbe, 1916) **comb. n.**; *U. vestigialis* (Kolbe, 1916) **comb. n.**

The following taxa are resurrected: *Belopherus* Schoenherr, 1833 **stat. res.**; *Brentus linearis* Olivier, 1790 **stat. res.**; *Brentus turbatus* Boheman, 1840 **stat. res.**; *Cormopus filum* Kolbe, 1916 **stat. res.**; *Megalosebus fallaciosus* Kolbe, 1916 **stat. res.**; *Podozemius comparabilis* (Kolbe, 1916) **stat. res.**; *Pseudomygaleicus somereni* De Muizon, 1960 **stat. res.**; *Schizoadidactus napaeus* (Kolbe, 1916) **stat. res.**; *Symmorphocerus monticola* Schoenherr, 1847 **stat. res.**

The genus *Belorhynchus* Berthold, 1827 is transferred from Arrhenodini Lacordaire, 1866 to Ithystenini Lacordaire, 1866 (**new placement**).

The type series of 410 previously described taxa have been examined to establish their identities, and lectotypes are designated for the following taxa: *Abrentodes auratus* Sharp, 1895; *A. pusillus* Sharp, 1895; *A. robustus* Sharp, 1895; *A. sulcipennis* Sharp, 1895; *Achrionota bilineata* Pascoe, 1872; *Acramorhophocephalus costipennis* Calabresi, 1920; *Acratus apicalis* Sharp, 1895; *A. armatus* Senna, 1897; *A. chontalensis* Sharp, 1895; *A. disjunctus* Kleine, 1927; *A. expressus* Kleine, 1927; *A. exquisitus* Kleine, 1927; *A. extrarius* Kleine, 1927; *A. fidelis* Kleine, 1927; *A. filum* Sharp, 1895; *Agrioblepis hova* Kleine, 1921; *Allaeometrus breviceps* Senna, 1903; *A. deformis* Kleine, 1925; *Amerismus ardens* Kleine, 1923; *A. felix* Kleine, 1925; *Amorphocephalus calvei* Power, 1879; *Ananesiotes insularis* Kleine, 1922; *A. kleinei* Calabresi, 1920; *Anepsiotes schenklingi* Kleine,

1917; *Apterorhynchus albatrus* Kleine, 1922; *Araiorrhynchus fausti* Senna, 1898; *A. ophthalmicus* Calabresi, 1920; *Arrhenodes approximans* F. Walker, 1859; *A. concolor* Sharp, 1895; *A. confrater* Kleine, 1944; *A. designatus* Boheman, 1840; *A. forficatus* J. Thomson, 1858; *A. funebris* Sharp, 1895; *A. gentilis* J. Thomson, 1858; *A. latirostris* Gyllenhal, 1833; *A. pygmaeus* Gyllenhal, 1833; *Aulacoderes immotus* Chevrolat, 1839; *Autosebus setosellus* Kolbe, 1916; *Baryrhynchus anthracinus* Kleine, 1916; *B. merocephalus* Kleine, 1916; *B. rudis* Senna, 1893; *B. speciosissimus* Kleine, 1916; *B. umbraticus* Kleine, 1916; *B. deyrollei* Power, 1879; *B. ochraceus* Kleine, 1916; *B. poweri* Roelofs, 1879; *B. rugosicollis* Power, 1879; *B. eximus* Kleine, 1927; *Bolbocranius bicolor* Senna, 1898; *B. mandibularis* Kolbe, 1897; *B. opacus* Kolbe, 1916; *B. parvulus* De Muizon, 1955; *B. unicolor* De Muizon, 1960; *Brentus clavipes* Boheman, 1840; *B. decollatus* Chevrolat, 1839; *B. planicaudatus* Chevrolat, 1839; *B. pugionatus* Chevrolat, 1839; *B. chiriquensis* Sharp, 1895; *B. clavipes* Sharp, 1895; *B. consentaneus* Perroud, 1853; *B. efferratus* Kleine, 1927; *B. finitimus* Kleine, 1927; *B. guatemalensis* Sharp, 1895; *B. laticornis* Sharp, 1895; *B. multilineatus* Sharp, 1895; *B. quadrilineatus* Sharp, 1895; *B. rufescens* Sharp, 1895; *B. sculptipennis* Sharp, 1895; *B. unidentatus* Perroud, 1853; *B. variegatus* Sharp, 1895; *Cacopsalis rudis* Sharp, 1895; *Caenorhynchodes rubrosignatus* Kleine, 1921; *C. versicolor* Kleine, 1921; *C. fasciatus* Kleine, 1921; *C. insulanus* Kleine, 1921; *Callipareius feae* Senna, 1892; *C. foveatus* Senna, 1894; *Calodromus cyrtotrachelus* J. Thomson, 1857; *C. mellyi* ssp. *sarawakensis* Heller, 1910; *Calorychodes decens* Kleine, 1925; *Carcinopisthius felschei* Kolbe, 1892; *C. loriae* Senna & Calabresi, 1919; *Catablysmia stoermeri* Kleine, 1925; *Cediocera longicornis* Pascoe, 1887; *C. tristis* Senna, 1892; *Centrophorus aeneolus* Kolbe, 1883; *C. compressipes* Chevrolat, 1839; *C. curvirostris* Chevrolat, 1839; *C. emarginatus* Chevrolat, 1839; *C. laevicollis* J. Thomson, 1858; *C. rectirostris* Kolbe, 1883; *C. validirostris* Kolbe, 1883; *Ceocephalus cavus* Walker, 1859; *C. fraterculus* Kolbe, 1883; *C. internatus* Pascoe, 1872; *C. opacus* Chevrolat, 1839; *Cephalobarus pumillus* Montrouzier, 1855; *Cerobates birmanicus* Senna, 1894; *C. canaliculatus* var. *carinensis* Senna, 1892; *C. collectivus* Kleine, 1925; *C. concisus* Kleine, 1925; *C. sennae* Calabresi, 1920; *C. sexsulcatus* var. *glaberrima* Senna, 1892; *C. tristriatus* ssp. *elytralis* Senna, 1897; *C. tristriatus* ssp. *longirostris* Senna, 1898; *Ceunonus minutus* Kleine, 1922; *Chalybdicus reverens* Kleine, 1928; *Claeoderes guatemalensis* Sharp, 1895; *C. mexicanus* var. *tristis* Senna, 1890; *Coptorhynchus francoisi* Desbrochers des Loges, 1892; *Cordus latirostris* Power, 1879; *Cormopus distinctus* Kleine, 1916; *C. diversitarsis* Kleine, 1916; *C. diversus* Kleine, 1926; *Cyphagogus appendiculatus* Fairmaire, 1889; *C. bipunctatus* Senna, 1897; *C. buccatus* Kleine, 1916; *C. costipennis* Fairmaire, 1889; *C. delicatus* f. *tristriatus* Kleine, 1916; *C. modiglianii* Senna, 1893; *C. perrieri* Fairmaire, 1899; *C. signipes* Lewis, 1883; *C. silvanus* Senna, 1903; *C. simulator* Senna 1903; *Cyriodontus evanidus* Kleine, 1927; *C. exactus* Kleine, 1927; *Dactylobarus carbonarius* Kleine, 1916; *D. fraterculus* Kleine, 1916; *Diplohoplizes armatus* Kleine, 1916; *Diurus celsus* Kleine, 1923; *D. completus* Kleine, 1923; *D. deruptus* Kleine, 1923; *D. philippinicus* Senna, 1911; *D. poultoni* Senna, 1911; *Dysmorphorhynchus amabilis* Kleine, 1916; *Ectocemus granulirostris* Gestro, 1876; *E. madrasanus* Kleine, 1931; *E. pterygorhinus* Gestro, 1876; *E. wallacei* Pascoe, 1862; *Elytracantha cerbera* Kleine, 1919; *E. contraria* Kleine, 1926; *Episphales championi* Sharp, 1895; *E. elegans* Sharp, 1895; *E. facilis* Sharp, 1895; *E. similis* Sharp, 1895; *Estenorhinus evidens* Kleine, 1927; *E. guttatus* Sharp, 1895; *Eubactus fuscoanthinus* Fairmaire, 1881; *Eupsalis coracina* Kolbe, 1883; *E. kolbei* Kleine, 1917; *E. lecontei* Power, 1878; *E. parviornata* Kleine, 1917; *E. promissus* Pascoe, 1872; *E. sallei* Power, 1878; *E. testacea* Kleine, 1917; *E. tuberculata* Senna, 1894; *Euschizus alatus* Kleine, 1922; *Eutrachelus morio* Kleine, 1936; *Glaucocephalus robustus* Kleine, 1916; *G. seminitens* Kleine, 1916; *Gyalostoma elegans* Kleine, 1916; *Hemiorhynchodes compactus* Kleine, 1931; *H. curvus* Kleine, 1925; *H. dissonus* Kleine, 1925; *Hemipsalis crassus* Sharp, 1895; *Hephebocerus mexicanus* Sharp, 1895; *Heterobrenthus distans* Sharp, 1895; *Heteroplites celebicus* Senna, 1897; *Higonius nudus* Kleine, 1920; *Holotrachelus comparabilis* Kleine, 1923; *Homales glaber* Kolbe, 1883; *Homoeolizus salvini* Sharp, 1895; *Homophylus castaneus* Kleine, 1920; *Hoplopisthius celebensis* Kolbe, 1892; *H. javanus* Kolbe, 1892; *H. kolbei* Senna, 1893; *Hormocerus annulipes* Schaufuss, 1887; *Hypomiolispa colorata* Kleine, 1925; *H. conjugalis* Kleine, 1925; *H. exigua* Kleine, 1918; *H. ferox* Kleine, 1926; *H. incerta* Kleine, 1937; *Hyposphales factus* Kleine, 1927; *Ionthocerus angulaticeps* Senna, 1898; *I. asiaticus* Kleine, 1920; *I. nigripes* Lewis, 1883; *Isomorphus trifasciatus* Kleine, 1916; *Ithystenus adoptivus* Kleine,

1919; *I. alatus* Kleine, 1919; *I. appendiculatus* Kleine, 1919; *I. caudatus* Kleine, 1919; *I. cultellatus* Kleine, 1919; *I. cupreus* Kleine, 1943; *I. decorus* Kleine, 1919; *I. densepunctatus* Kleine, 1919; *I. franklini* Kleine, 1925; *I. franklinmuelleri* Kleine, 1925; *I. impar* Kleine, 1943; *I. leveri* Kleine, 1939; *I. perlongus* Kleine, 1919; *I. sabulosus* Kleine, 1919; *I. similis* Kleine, 1919; *I. spinosus* Kleine, 1919; *Mesoderes aridus* Kleine, 1923; *M. sexnotatus* Senna, 1898; *Metatrachelizus artificialis* Kleine, 1923; *M. congruens* Kleine, 1925; *M. constans* Kleine, 1926; *M. ferox* Kleine, 1944; *Metatrachelus hospes* Kleine, 1926; *Microsebus kolbei* Kleine, 1916; *M. lorae* Senna, 1894; *Microtrachelizus aethiopicus* Calabresi, 1920; *M. beneficus* Kleine, 1925; *M. reductus* Kleine, 1941; *M. silvicola* Senna, 1903; *M. targionii* Senna, 1893; *Miolispa enganica* Senna, 1893; *M. exarata* var. *humeralis* Senna, 1903; *M. exarata* var. *sumatrana* Senna, 1903; *M. fausti* Senna, 1895; *M. fulgida* Kleine, 1925; *M. fundata* Kleine, 1925; *M. funebris* Kleine, 1925; *M. javanica* Senna, 1892; *M. lineata* Senna, 1898; *M. metallica* Senna, 1892; *M. metallica* var. *impressa* Senna, 1903; *M. nupta* Senna, 1892; *M. pasteurii* Senna, 1898; *M. pygmaea* Senna, 1895; *M. salomonensis* Senna, 1894; *M. semivelata* Schaufuss, 1885; *M. siporana* Senna, 1898; *M. sponsa* Kleine, 1918; *M. trachelizoides* Senna, 1895; *Miolispoides sculpturatus* Calabresi, 1921; *M. somereni* De Muizon, 1960; *M. tarsalis* Kleine, 1920; *M. vittipennis* var. *nitida* Calabresi, 1920; *Nematocephalus guatemalensis* Senna, 1893; *Nemobrenthus aeneipennis* Sharp, 1895; *N. expletus* Kleine, 1927; *Nemocephalinus dubitabilis* Kleine, 1927; *Nemocephalus dolosus* Kleine, 1927; *N. femoratus* Sharp, 1895; *N. gounellei* Kleine, 1944; *N. piceus* Perroud, 1853; *N. planicollis* F. Walker, 1859; *N. puncticeps* Sharp, 1895; *N. scotti* Kleine, 1923; *N. spinirostris* F. Walker, 1859; *Opisthenoplus commodus* Kleine, 1923; *Orychodes andrewsi* Gahan, 1900; *O. cinnamomi* var. *tuberculatus* Senna, 1903; *O. femoratus* Schaufuss, 1885; *O. insignis* Lewis, 1883; *O. pictus* Pascoe, 1862; *Ozodecterus forficulatus* Chevrolat, 1839; *Ozodecterus rugicollis* Chevrolat, 1839; *Paramorphocephalus binotatus* Calabresi, 1922; *Parorychodes cereus* Kleine, 1925; *P. comes* Kleine, 1921; *Paryphobrenthus beringei* Kolbe, 1897; *Perisymphocerus gracilis* Kleine, 1919; *Phobetromimus exiguus* Kleine, 1916; *P. simulans* Kleine, 1916; *Phobetrum paradoxum* Kleine, 1916; *Phocylides collaris* Pascoe, 1872; *Pithoderes corporatus* Kleine, 1924; *Prodeutor fruhstorferi* Senna, 1897; *P. laminatus* Pascoe, 1862; *Prophthalmus assimilis* Kleine, 1916; *P. bourgeoisii* Power, 1878; *P. brevis* Power, 1878; *P. heikertingeri* Kleine, 1916; *P. mutabilis* Senna, 1893; *P. mutabilis* var. *mentaweicus* Senna, 1898; *P. planipennis* Pascoe, 1872; *P. potens* Lacordaire, 1866; *P. pugnator* Power, 1878; *P. ruficornis* Senna, 1903; *P. tricolor* Power, 1878; *P. versicolor* Senna, 1892; *P. wichmanni* Kleine, 1916; *Proteramocerus enervatus* Kleine, 1927; *P. fidus* Kleine, 1927; *Protosebus piceus* Kleine, 1922; *Protusambius praecursor* Kolbe, 1916; *Pseudanchisteus neglectus* Kleine, 1922; *Pseudophocylides clarus* Kleine, 1923; *P. insularis* Kleine, 1920; *Pseudorychodes crassus* Senna, 1894; *P. damnosus* Kleine, 1925; *P. helleri* Senna, 1899; *P. honoris* Kleine, 1926; *P. tenuirostris* Senna, 1894; *Pyrestema kuntzeni* Kleine, 1922; *Raphidorrhynchus sexvittatus* Senna, 1894; *R. variabilis* Senna, 1894; *R. vicinus* Senna, 1894; *Raphidorrhynchus auricollis* Sharp, 1895; *R. chiriquensis* Sharp, 1895; *R. excellens* Kleine, 1927; *R. longulus* Sharp, 1895; *R. obscurus* Sharp, 1895; *R. sennai* Sharp, 1895; *Raphidorrhynchus consanguineus* Senna, 1895; *R. rothschildi* Senna, 1895; *Rhynchoneus belti* Sharp, 1895; *Rhyticephalus aulaconotus* Chevrolat, 1839; *R. brevicornis* Chevrolat, 1839; *Rhytidopterus dilucidus* Kleine, 1924; *R. variabilis* Kleine, 1916; *Schizoadidactus tibialis* Kleine, 1916; *S. alienus* Kleine, 1922; *S. altilis* Kleine, 1922; *S. birmanicus* Senna, 1892; *S. cameratus* Lacordaire, 1866; *S. carinirostris* Senna, 1892; *S. celebicus* Senna, 1899; *S. consanguineus* Senna, 1892; *S. consideratus* Kleine, 1926; *S. dichrous* Lacordaire, 1866; *S. gemellus* Kleine, 1925; *S. intermedius* Senna, 1892; *S. metallicus* Senna, 1897; *S. nanus* Senna, 1894; *S. suspensus* Kleine, 1943; *Spatherrhinus curiosus* Kleine, 1926; *S. longiceps* Kolbe, 1888; *Stereobates chiriquensis* Sharp, 1895; *S. gracilis* Sharp, 1895; *Stereodermus barbirostris* Sharp, 1895; *S. breviceps* Sharp, 1895; *S. calvus* Sharp, 1895; *S. carinatus* Sharp, 1895; *S. chontalensis* Sharp, 1895; *S. dentipennis* Sharp, 1895; *S. fessus* Kleine, 1927; *S. filum* Sharp, 1895; *S. gestroi* Senna, 1893; *S. infidus* Senna, 1898; *Stereodermus longiceps* Sharp, 1895; *S. puncticollis* Sharp, 1895; *S. sansibaricus* Kolbe, 1897; *S. zunilensis* Sharp, 1895; *Stratiorrhina concors* Kleine, 1923; *S. major* Calabresi, 1921; *S. pulla* Kleine, 1936; *Stroggylosternum nitens* Kleine, 1922; *Subdysmorphorhynchus carinatus* Kleine, 1917; *Suborychodes intermedius* Kleine, 1917; *Syngenithysten nigrinitens* Kleine, 1919; *Symphocerus beloni* Power, 1879; *S. cardoni* Senna, 1894; *S. consequens* Kleine, 1926; *S. lottoi*

De Muizon, 1960; *Synorychodes glaber* Kleine, 1937; *Taphroderes apicalis* Sharp, 1895; *T. bel-tianus* Sharp, 1895; *T. mexicanus* Sharp, 1895; *T. nevermanni* Kleine, 1927; *T. oscillator* Sharp, 1895; *T. rectus* Sharp, 1895; *T. ventralis* Sharp, 1895; *Taphroderomimus disjunctus* Kleine, 1927; *Ter-amocerus belti* Sharp, 1895; *Tetanocephalus nigronitens* Kleine, 1916; *Trachelizus advena* Sharp, 1895; *T. alarius* Kleine, 1922; *T. arduus* Sharp, 1895; *T. cognatus* Sharp, 1895; *T. dispar* Sharp, 1895; *T. elevatus* Sharp, 1895; *T. filiformis* Sharp, 1895; *T. fracticornis* Sharp, 1895; *T. frontalis* Sharp, 1895; *T. hirtellus* Sharp, 1895; *T. lineatus* Sharp, 1895; *T. modestus* Senna, 1892; *T. nigri-cornis* Sharp, 1895; *T. occlusus* Sharp, 1895; *T. oscillator* Sharp, 1895; *T. prolixus* Sharp, 1895; *T. robustus* Sharp, 1895; *T. serratus* Sharp, 1895; *T. sternalis* Sharp, 1895; *T. vasseleti* Boheman, 1840; *Tulotus maculipennis* Senna, 1894; *Tychaeus curvidens* var. *signatus* Sharp, 1895; *Ubanus aeneus* Senna, 1895; *Ulocerus cinereus* Senna, 1896; *U. dalmani* Senna, 1896; *U. laticornis* Sharp, 1895; *U. longicornis* Senna, 1896; *U. luridus* Senna, 1896; *U. mexicanus* Sharp, 1895; *U. nigro-lineatus* Senna, 1896; *U. severini* Senna, 1896; *Uropterus gestroi* Senna, 1894; *Usambius conradti* Kolbe, 1892; *U. conradti* var. *laevis* Senna, 1898; *Xestocoryphus gracilis* Kleine, 1916; *Zemioses camerunus* var. *bicolor* Kolbe, 1916; *Z. camerunus* var. *nigricollis* Kolbe, 1916; *Z. celtis* Lewis, 1883; *Z. crassicollis* Fairmaire, 1899; *Z. laetus* Senna, 1892; *Z. striolatus* Fairmaire, 1897.

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INTRODUCTION

The first ever checklist published on the Brentidae *sensu Auctorum* (Brentinae, Cyphagoginae, Pholidochlamydinae, Taphroderinae, Trachelizinae, Ulocerinae) was by Schönfeldt in *Genera Insectorum* (1908), comprising 111 genera and 624 species; this was followed by a second in *Coleopterorum Catalogus* (Schönfeldt, 1910a) where the number of species rose to 735. Later, Kleine (1927a) listed 1279 species in the *Coleopterum Catalogus* and finally he again listed 274 genera and 1356 species in *Genera Insectorum* (1938d).

Since 1938, a great many works on brentids have been published, including some important monographies, but they were limited to specific geographical areas, e. g. "Faune des Brenthides d'Afrique" by De Muizon (1960), "Brentidos argentinos" by Haedo Rossi (1961), "Monographie des Coléoptères Brentidae du Continent africain" by Damoiseau (1967a) and "Australian Weevils" by Zimmerman (1994).

In 1999, Alonso-Zarazaga & Lyal published "A world catalogue of families and genera of Curculionoidea", in which the systematics of subfamilies, tribes and genera of brentids was revised and updated. In this volume we follow the classification of Alonso-Zarazaga & Lyal (1999) and the modifications and updates furnished by the authors themselves (Alonso-Zarazaga & Lyal, 2002), with the suppression of the Belorhynchini tribe and the resurrection of the Tychaeini tribe.

This work lists 293 genera with 1690 valid species. We also thought it would be helpful to give an update on our knowledge of brentid morphology, biology, ecology, ethology and zoo-geography as well as an historical review offering information on the most important specialists that have dedicated themselves to the family. We also furnish a complete bibliography on all the papers dealing with brentids.

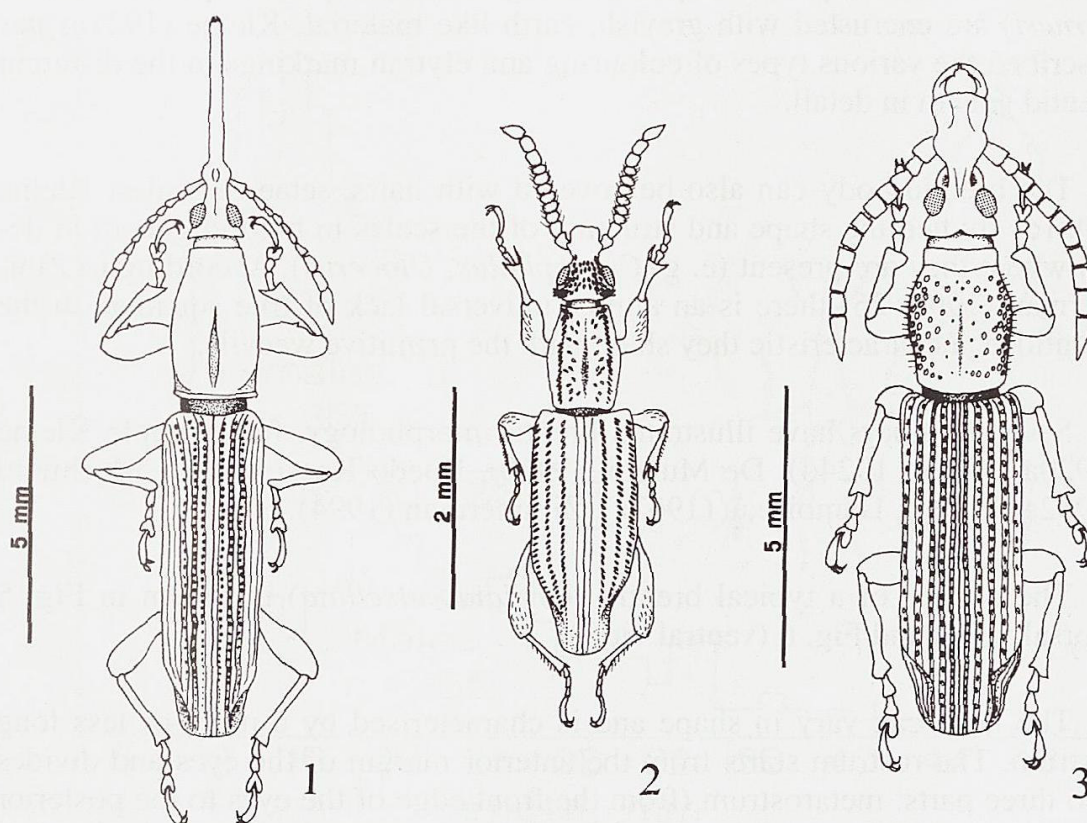
Also included are four Appendices. In Appendix 1 we list all the known brentid host plants; in Appendix 2 the species of wood borer beetles (Platypodidae, Scolytidae, Cerambycidae) in whose tunnels Brentidae may lay eggs; in Appendix 3 the known brentid parasites (Acarina); in Appendix 4 the host ant species for myrmecophilous brentids.

MORPHOLOGY

Adults

According to Kuschel (1995) the characters of the Brentinae (= Brentidae *sensu Auctorum*) are as follows: antennal club not or hardly wider than funicle; tibial spurs present; eye completely glazed over with a crystalline coating; scutellum not visible (visible only in the New Zealand species *Lasiorhynchus barbicornis*); claw segment distinctly longer than segments 2 and 3 combined; claws divergent; fore coxa considerably closer to hind than front margin of prothorax; female tergite 8 denticulate at apex.

In general, the brentid body is long and slender with a rostrum that can be filiform, short and blunt or very wide and strong (Figs 1-3). Size varies from a few millimetres to 70-80 millimetres, as in *Eutrachelus temminckii* and *Lasiorhynchus barbicornis*. Individual variability within the same species can be remarkable. For example, in *Brentus anchorago* the largest individuals can be up to five times as large as the smallest (Sharp, 1936; Johnson, 1982).



Figs 1-3 - Different brentid body shapes. Fig. 1: *Gynandrorhynchus vittipennis* (Fåhræus); Fig. 2: *Autosebus kolbei* De Muizon; Fig. 3: *Afrocordus vicarius* (Kleine).

Johnson (1982) studied a population of about 1000 *B. anchorago* specimens on a dead trunk of *Bursera simaruba* L. in Costa Rica. The females ranged in length from 8 to 27 mm and the males from 9 to 49 mm, but some males reached 52 mm (Johnson, 1983b). Weight variation was even greater. The heaviest female was 22.4 times as heavy as the lightest female, and the heaviest male in the population was 26.5 times as heavy as the lightest male (Fig. 4). Johnson (1982) suggested that most of the variation in body size is environmental in origin rather than genetic. The environmental factors could be: larval density, wood moisture levels (affecting the growth of the fungal food of the larvae and the toughness of the wood), growth on non optimal host tree species, and parasites of the larvae.

Brentids are usually dark in colour, from reddish brown to black. Sometimes the pronotum can be a different colour from the elytra (*Bolbocranius bicolor*, *Ithystenus bicolor*, *Phocylides bicolor*). The elytra can bear a pattern formed of coloured areas arranged along the striae. The arrangement of the pattern can vary to different degree within the same species and in general markings are yellow or reddish in colour. Some species are opaque, others shiny and yet others a brilliant metallic colour (e. g. *Ozodecerus*, *Proteramocerus*, *Teramocerus*); some species (e. g. *Rhinopteryx foveipennis* and *R. wernerii*) are encrusted with greyish, earth-like material. Kleine (1921a) has described the various types of colouring and elytran markings in the different brentid genera in detail.

The brentid body can also be covered with hairs, setae or scales. Kleine (1921c) studied the shape and structure of the scales in brentid genera in detail where they are present (e. g. *Ceocephalus*, *Ulocerus*). According to Zimmerman (1994: 25) there is an almost universal lack of true squamae in the Brentidae, a characteristic they share with the primitive weevils.

Several authors have illustrated brentid morphology, for example Kleine (1920a, 1920e, 1924d), De Muizon (1960), Haedo Rossi (1961), Morimoto (1962a, 1976b), Damoiseau (1967a), Zimmerman (1994).

The outline of a typical brentid (*Orfilaia vulsellata*) is shown in Fig. 5 (dorsal view) and Fig. 6 (ventral view).

The head can vary in shape and is characterised by a more or less long rostrum. The rostrum starts from the anterior margin of the eyes and divides into three parts: metarostrum (from the front edge of the eyes to the posterior edge of the scrobes or of the mesorostral plate), mesorostrum (at the level of insertion of the antennae, sometimes with a mesorostral plate), prorostrum (from the anterior margin of the mesorostral plate or the cavity where the ant-

ennae are inserted as far as the mandibles). In almost all families and tribes the rostrum is long or very long, whilst in many Cyphagogini the rostrum is extremely short. In many of the tribes the rostrum is characterised by varying degrees of sexual dimorphism. In females the rostrum is generally shorter, cylindrical and filiform. In the Ithystenini the rostrum in males can be filiform and several times longer than the head itself (Fig. 71). Many species of the genus *Brentus* also have the rostrum far longer than the head. In some species of the genera *Prophthalmus*, *Baryrhynchus* and *Eupeithes*, for example, the rostrum in males is very wide and strong (Fig. 72). Usually the rostrum is straight and curved downwards, but in some cases it can turn slightly upwards (*Gynandrorhynchus audax*; *G. corneolus*, *Agriorrhynchus*). Sometimes the rostrum in males can be saw-toothed laterally (*Anepsiotes*, *Belorhynchus stratiorrhinoides*, *Megateras*, *Stratiorrhina*, *Teraticorhynchus*, *Toxobrentus*) (Fig. 11) carry a spiny protuberance pointing upwards (*Hopliterrhynchus*) (Fig. 17) or downwards (*Ankleineella*, *Heterorrhynchus*) (Fig. 79). Sometimes teeth occur beneath the head (males of *Mesoderes*, *Microsebus*, *Podozemius*, and *Rhynchoneus*) (Figs 15, 16). The anterior edge of the prorostrum can be straight, concave (e. g. *Adidactus*) or jutting forwards between the mandibles

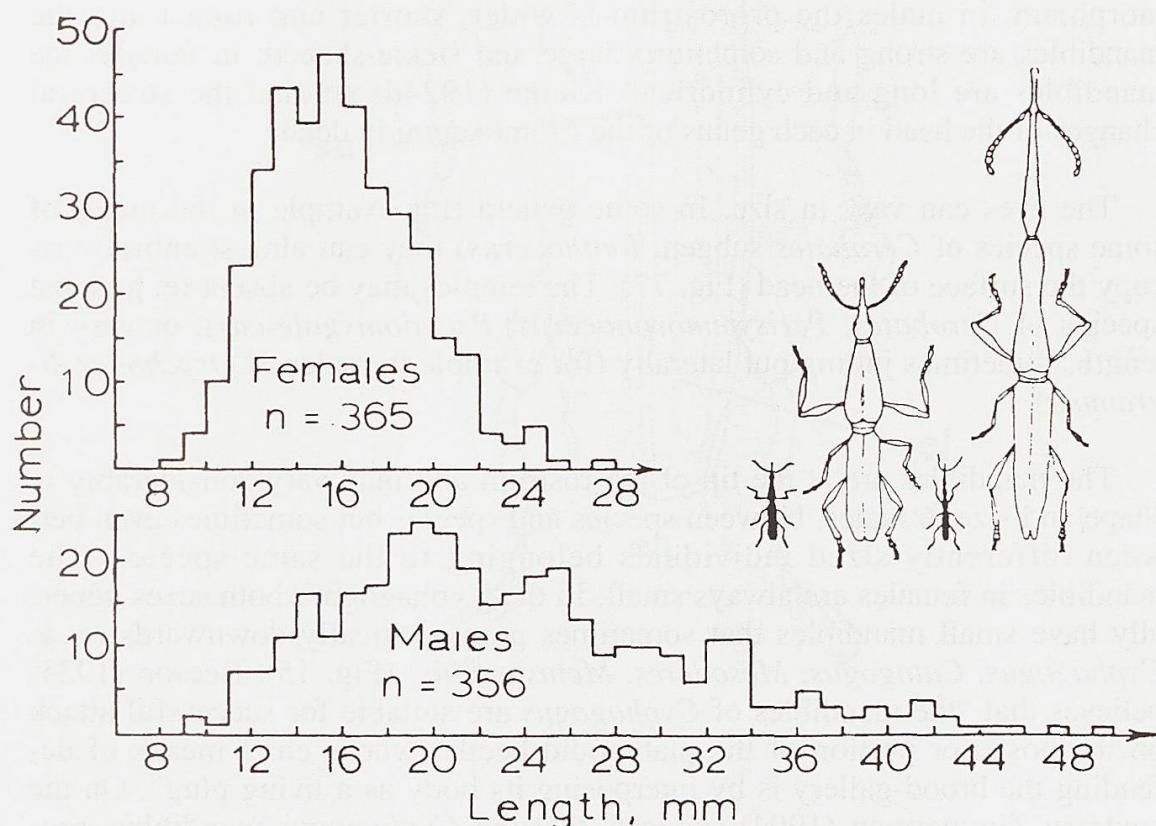


Fig. 4 - Lognormal distribution of body lengths in an aggregation of 365 female and 356 male *Brentus anchorago* L. from Costa Rica (from Johnson, 1982).

(*Bolbocranius delhaisei*, *B. mechowii*, *B. rostralis*) (Fig. 8). The rostrum surface can be smooth, grooved or punctated to varying degree.

The most profound structural modifications to the head and rostrum are found in the Eremoxenini (Figs 7, 13, 14), with the occurrence of sulci, apophyses, carinae, grooves and pubescent areas. These modifications are ascribable to the particular biology of the Eremoxenini, most of which are associated with ants. Lateral apophyses also occur in the genus *Orfilaia* (Fig. 9) belonging to the Arrhenodini tribe. The head may be smooth, punctated or covered with scales. The vertex can be flat, raised, grooved or keeled and project more or less backwards on the occipital bulb. Grooves of varying length and depth can occur behind the eyes, as can gland tuberosities underneath; the ventral surface of the head and rostrum may be sulcated, as in the genus *Cordus* (Fig. 10). Secretions from the cephalic glands flow down these sulci to the apex of the rostrum, where they become available to the ants (Zimmerman, 1994: 55). The metarostrum can be extensively deformed, keeled (e. g. *Symmorphocerus*) or deeply concave and bear brushes of stiff setae (e. g. *Amorphocephala*, *Ankleineella*). The mesorostrum generally consists of a mesorostral plate (Figs 7, 14) of varying width and shape, to which the apophyses may be attached. The shape of the prorostrum is subject to pronounced sexual dimorphism: in males the prorostrum is wider, shorter and robust and the mandibles are strong and sometimes large and sickle-shaped; in females the mandibles are long and cylindrical. Kleine (1924d) studied the structural changes of the head in each genus of the Eremoxenini in detail.

The eyes can vary in size. In some genera (for example in the males of some species of *Cerobates* subgen. *Ionthocerus*) they can almost entirely occupy the surface of the head (Fig. 77). The temples may be absent (e. g. some species of *Cerobates*, *Perisymphocerus*, *Pseudomygaleicus*), or vary in length, sometimes jutting out laterally (for example in males of *Orychodes digramma*).

The mandibles are at the tip of the rostrum and may vary considerably in shape and size in males, between species and species but sometimes even between differently sized individuals belonging to the same species. The mandibles in females are always small. In the Cyphagogini, both sexes generally have small mandibles that sometimes point vertically downwards (e. g. *Cyphagogus*, *Catagogus*, *Mesoderes*, *Metusambius*) (Fig. 15). Beeson (1925) believes that "the mandibles of *Cyphagogus* are suitable for successful attack on the posterior portion of the platypodid beetle, whose chief means of defending the brood-gallery is by interposing its body as a living plug". On the contrary Zimmerman (1994) suggests that the *Cyphagogus* mandibles, projecting downwards and resembling tusks, could act as hooks to assist the bren-tid to pull itself along the narrow tunnels of its host.

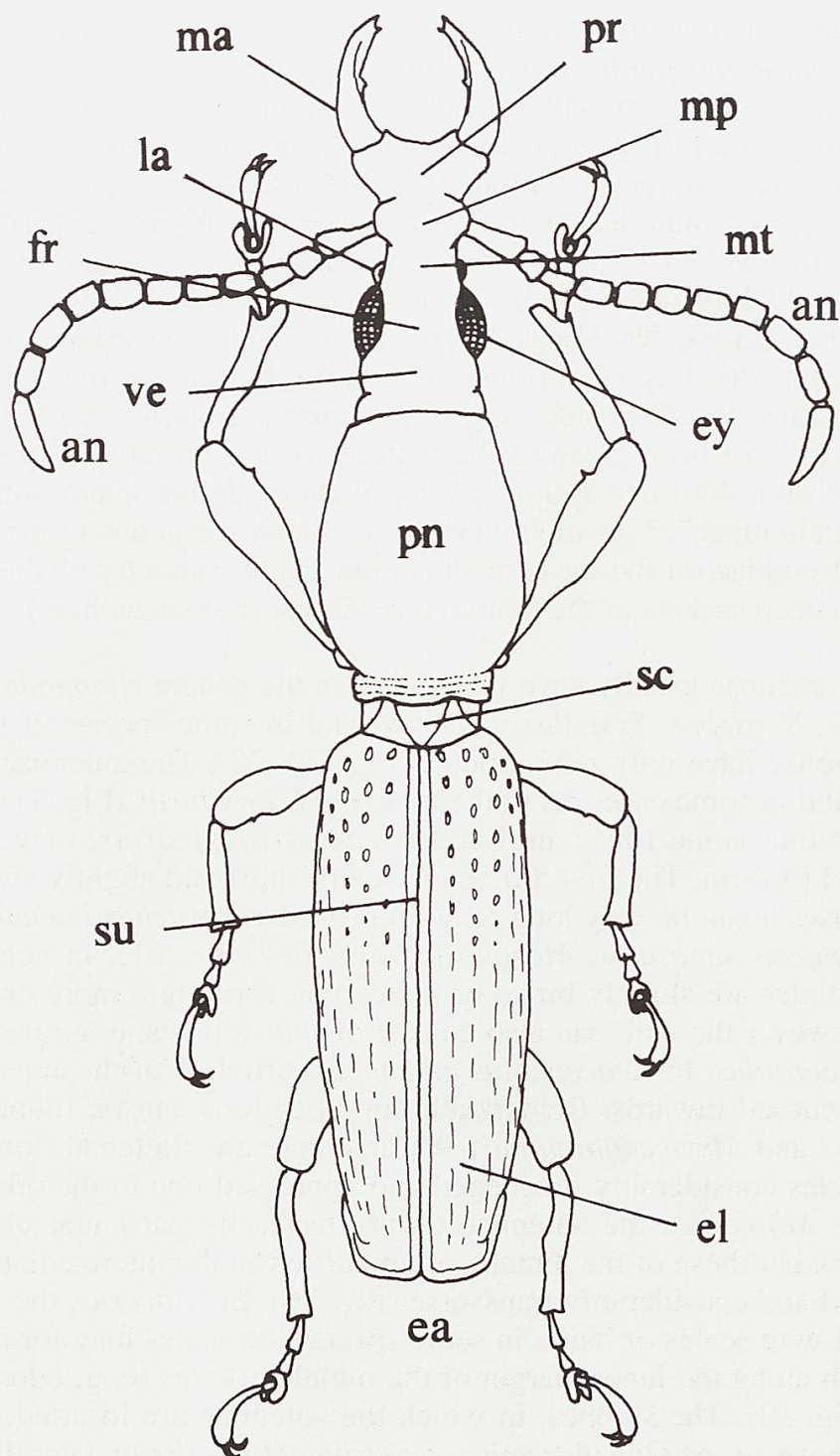


Fig. 5 - *Orfilaia vulsellata* (Gyllenhal), male, outline of dorsal view. Abbreviations: an: antenna; ea: elytral apex; el: elytron; ey: eye; fr: frons; la: lateral apophysis; ma: mandible; mp: mesorostral plate; mt: metarostrum; pn: pronotum; pr: prorostrum; sc: scutellum; su: suture; ve: vertex.

Sickle-shaped mandibles, that define a wide space internally, occur in the genera *Amorphocephala*, *Estenorhinus*, *Eupeithes*, *Orfilaia*, *Prophthalmus*, etc. (Figs 7, 9). The mandibles in *Bolbocranius bicolor* and *B. unicolor* are long and slender (Fig. 12). In *Bolbocranius distortus* and *B. csikii* the mandibles are asymmetric and the left mandible is remarkably curved and far more developed than the right (Fig. 73). The mouth parts are arranged inside the rostrum. Olivier (1807), Westwood (1849), Lacordaire (1866), Kempers (1932) and Morimoto (1962a, 1976b) have all studied the mouth parts of various brentid species; Haedo Rossi (1961: 23) has described and illustrated in detail the structure of various mouth parts in *Brentus lineicollis* from Argentina (Figs 26, 27). According to Zimmerman (1994: 30) "little attention appears to have been given to the features of the buccal cavity since Lacordaire's (1866: 400) discussion"...such features define major subdivisions of the Curculionidae". Morimoto (1976b) included the genus *Cylas* Schoenherr, in the Brentidae on the basis of the apomorphic character of the labial palpi borne in deep sockets of the labium (see Alonso-Zarazaga, h. o.).

The antennae usually have 11 articles. In the genera *Higonodes*, *Myrmecobrenthus*, *Neosebus*, *Systellus*, *Ulocerus* and in some species of *Ceocephalus* the antennae have only nine articles (Figs 20, 22). The antennae can vary in length and in some cases, as in the males of *Eterodiurus* (Fig. 81) and various Arrhenodini, some Ithystenini and *Ulocerus longicornis*, they can be very long and filiform. The first article is usually short and slightly swollen, but in other cases it can be very long (e. g. in *Gynandrorhynchus bocandei*) or spiny (*Allacompsus separator*, *Megateras horribile*) (Fig. 21). In general the last three articles are slightly broad or pubescent, forming a more or less distinct club, however the club can also consist of four articles, e. g. in some species of *Bolbocranius*. In *Ceocephalus antennatus* article 5 of the antennae is more or less curved inwards. Remarkable modifications can be found in *Pausso-brenthus* and *Myrmecobrenthus*: the antennae are flattened dorso-ventrally, the articles considerably broadened and appressed one to the other (Fig. 20). In male *Allagogus*, the antennae do not have any particular characteristics (Fig. 75), but those of the females are modified as the intermediate articles are flattened and considerably transverse (Fig. 76). In *Ulocerus*, the antennae are covered with scales or hairs; in some species the scales may form an elongated brush along the inner margin of the middle articles (e. g. *Ulocerus lacerosus*) (Fig. 22). The scrobes, in which the antennae are inserted, can be seen from above (e. g. *Opisthozemius*, *Sebasius*) (Fig. 18) or laterally (e. g. *Microsebus*) (Fig. 19). In the Eremoxenini, the mesorostral plate covers the scrobes.

The prothorax can vary in shape (barrel-shaped, pyriform, heart-shaped, cylindrical, globular). In *Brentus anchorago* the prothorax in the male is long, narrowing in the middle (Fig. 66). In general, the sides are rounded, but the

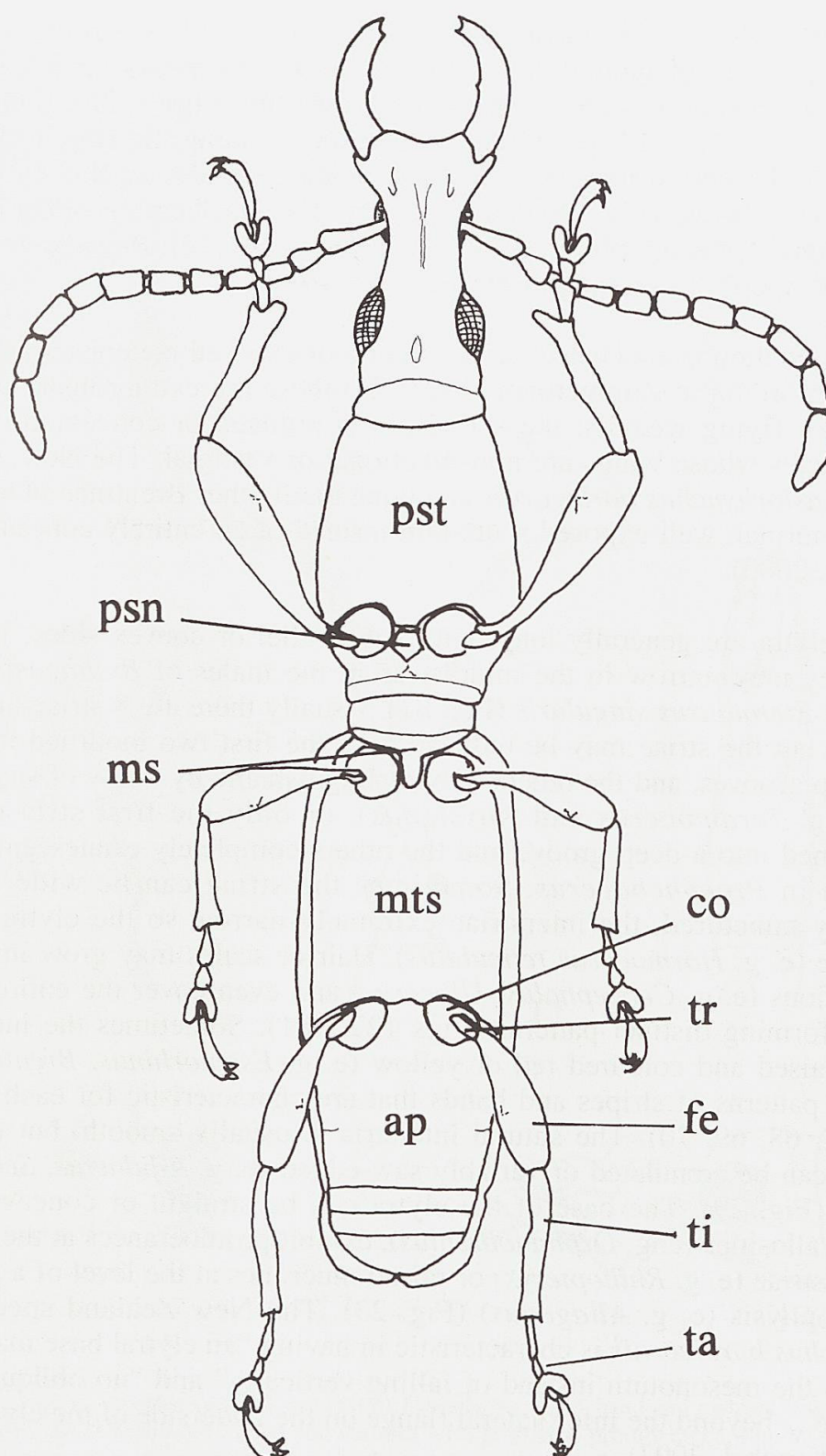


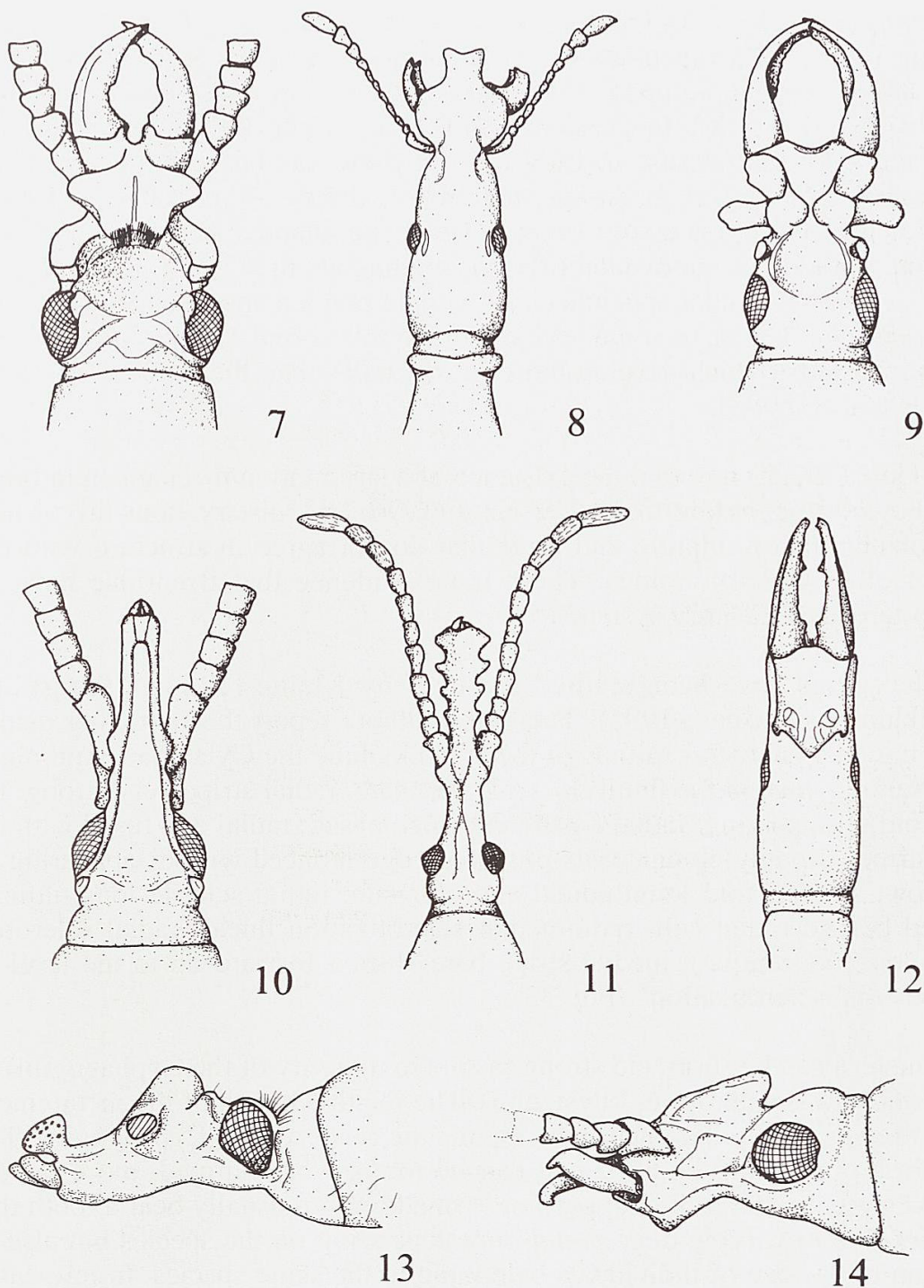
Fig. 6 - *Orfilaia vulsellata* (Gyllenhal), male, outline of ventral view. Abbreviations: ap: abdominal plate (fused ventrites I + II); co: coxa; fe: femur; ms: mesosternum; mts: metasternum; psn: prosternellum; pst: prosternum; ta: tarsus; ti: tibia; tr: trochanter.

males of *Belopherus* and *Thaumastopsis* have a more or less cornered process (Fig. 24). In the Cyphagogini the anterior portion of the prothorax is generally compressed laterally to accommodate the profemurs (Figs 2, 25). Sometimes the narrow portion of the pronotum can be keeled along the edge to varying degree. The pronotum may be smooth or sulcate, convex or flat. In *Cyphagogus* and *Cormopus* (Fig. 25) there is an anterior raised carina whilst in *Allagogus* a long, spiny apophysis points backwards (Fig. 23). *Rhynchoneus belti* has a large apophysis on the prosternum (Fig. 16).

The scutellum is concealed and the width of exposed metepisternum is reduced. According to Zimmerman (1994: 26) these are extraordinary characteristics for flying weevils: the scutellum is reduced or concealed only on those species whose wings are non-functional or vestigial. The New Zealand species *Lasiorrhynchus barbicornis* is unique to all other Brentinae in having a distinct, normal, well-exposed scutellum instead of an entirely concealed one (Kuschel, 2003).

The elytra are generally long, with subparallel or convex sides. In some cases they may narrow in the middle, as in the males of *Bulbogaster* (Fig. 175) and *Eterodiurus singularis* (Fig. 81). Usually there are 8 striae and 9 interstriae, but the striae may be reduced with the first two modified into narrow, deep grooves, and the others only distinguishable by a row of superficial pits (e. g. *Teramocerus* and *Ithystenus*), or only the first stria may be transformed into a deep groove and the others completely evanescent, as for example in *Proephebocerus*. Sometimes the striae can be wide and remarkably punctured, the interstriae extremely narrow so the elytra appear reticulate (e. g. *Hormocerus reticulatus*). Hair or scales may grow inside the punctuations (e. g. *Ceocephalus*, *Ulocerus*) and even cover the entire elytral surface forming distinct patterns (Figs 122, 181). Sometimes the interstriae can be raised and coloured red or yellow (e. g. *Estenorhinus*, *Brentus*, etc.) forming patterns of stripes and bands that are characteristic for each species (Figs 67, 68, 69, 70). The sutural interstria is usually smooth, but in some cases it can be crenulated or variably saw-edged (e. g. *Adidactus*, *Schizoadidactus*) (Fig. 29). The base of the elytra can be straight or concave, carry omeral callosities (e. g. *Orphanobrentus*), exhibit protuberances at the level of the interstriae (e. g. *Rhinopteryx*) or raised mucrones at the level of a prothoracic apophysis (e. g. *Allagogus*) (Fig. 23). The New Zealand species *Lasiorrhynchus barbicornis* is characteristic in having “an elytral base that slopes down to the mesonotum instead of falling vertically” and “no oblique carina or flange ... beyond the inferolateral flange on the underside of the elytral declivity” (Kuschel, 2003).

Declivity can be more or less steep and marked; some *Ulocerus* have tufts of raised scales (Fig. 28). Usually the elytra are rounded at the tip, or angular, bearing an extension at the level of the suture (e. g. *Hoplopisthius*,



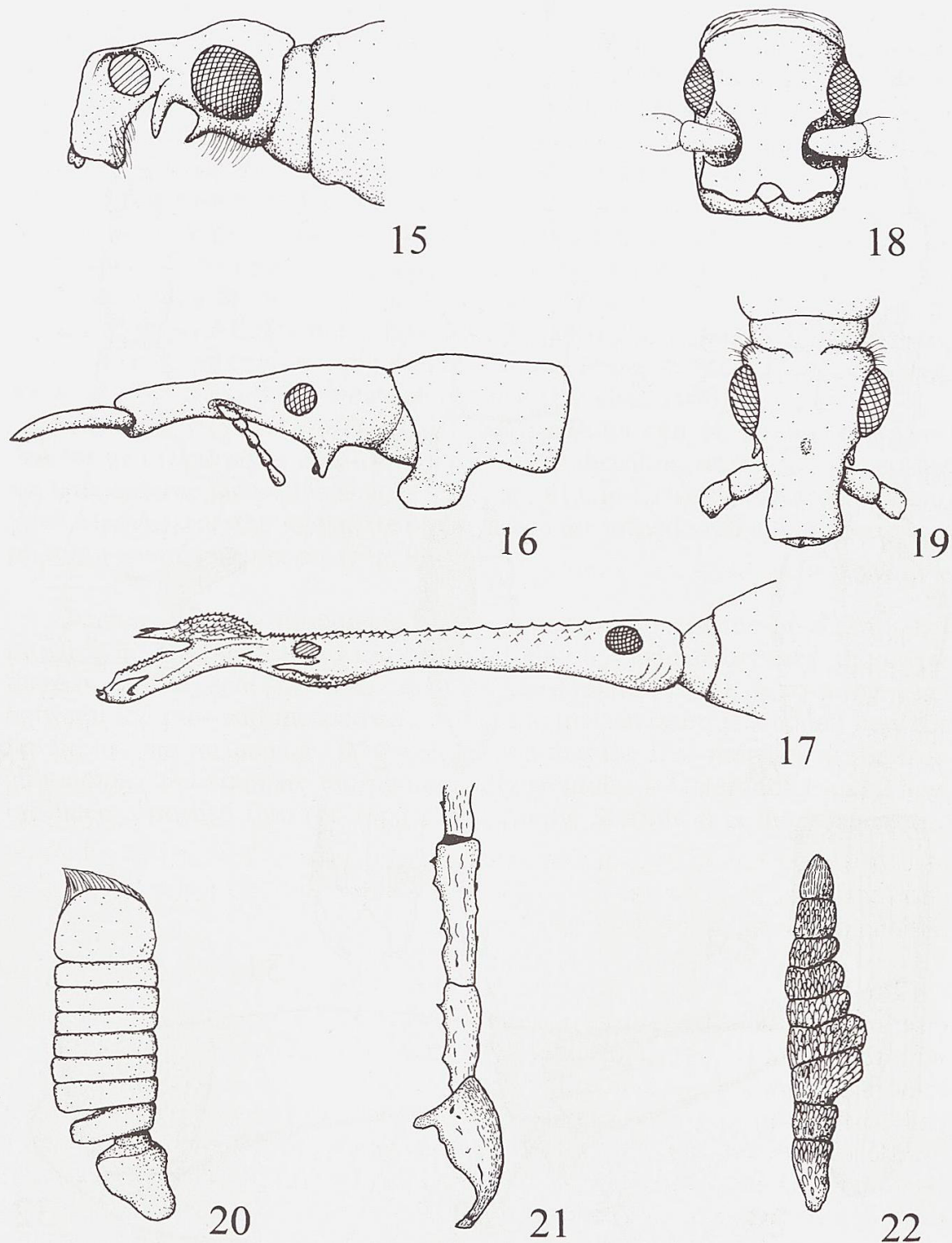
Figs 7-14 - Fig. 7: *Amorphocephala excantator* Damoiseau, male, head and rostrum; Fig. 8: *Bolbocranius delhaisei* (Damoiseau), head and rostrum with prorostral protuberance; Fig. 9: *Orfilaia nigra* Damoiseau, male, head and rostrum with lateral apophysis; Fig. 10: *Cordus demarzi* Damoiseau, sulci on the rostral ventral surface; Fig. 11: *Stratiorrhina tarsalis* Sforzi & Bartolozzi, male, head and rostrum; Fig. 12: *Bolbocranius bicolor* Senna, head and rostrum; Fig. 13: *Eremoxenus al-girus* Bartolozzi, male, lateral view of head and rostrum; Fig. 14: *Amorphocephala aethiopica* Bartolozzi, male, lateral view of head and rostrum.

Opisthozemius) (Figs 30, 168) or sunken in the middle (e. g. *Entomopisthius*). The tip may even be spatulate (e. g. *Prodeutor laminatus*, *Storeosomus riisii* (Fig. 84) or carry more or less long tails that can vary considerably in shape (Figs 83, 85, 174, 188). In *Carcinopisthius* (Fig. 31) the tails are short and robust and curved inwards, so they resemble the caudal appendices of the Dermaptera. According to Zimmerman (1994) these remarkable modifications of the elytral apices in *Carcinopisthius* are adapted as "shovels" to enable the brentids to remove debris from the tunnels they occupy in wood. In some genera the caudal appendices are simple pointed spurs (e. g. *Elytracantha*) (Fig. 32), whilst in some species of *Ithystenus* and *Ceocephalus* for example, the tails, which develop more in the males than the females, are long and slender (Fig. 83).

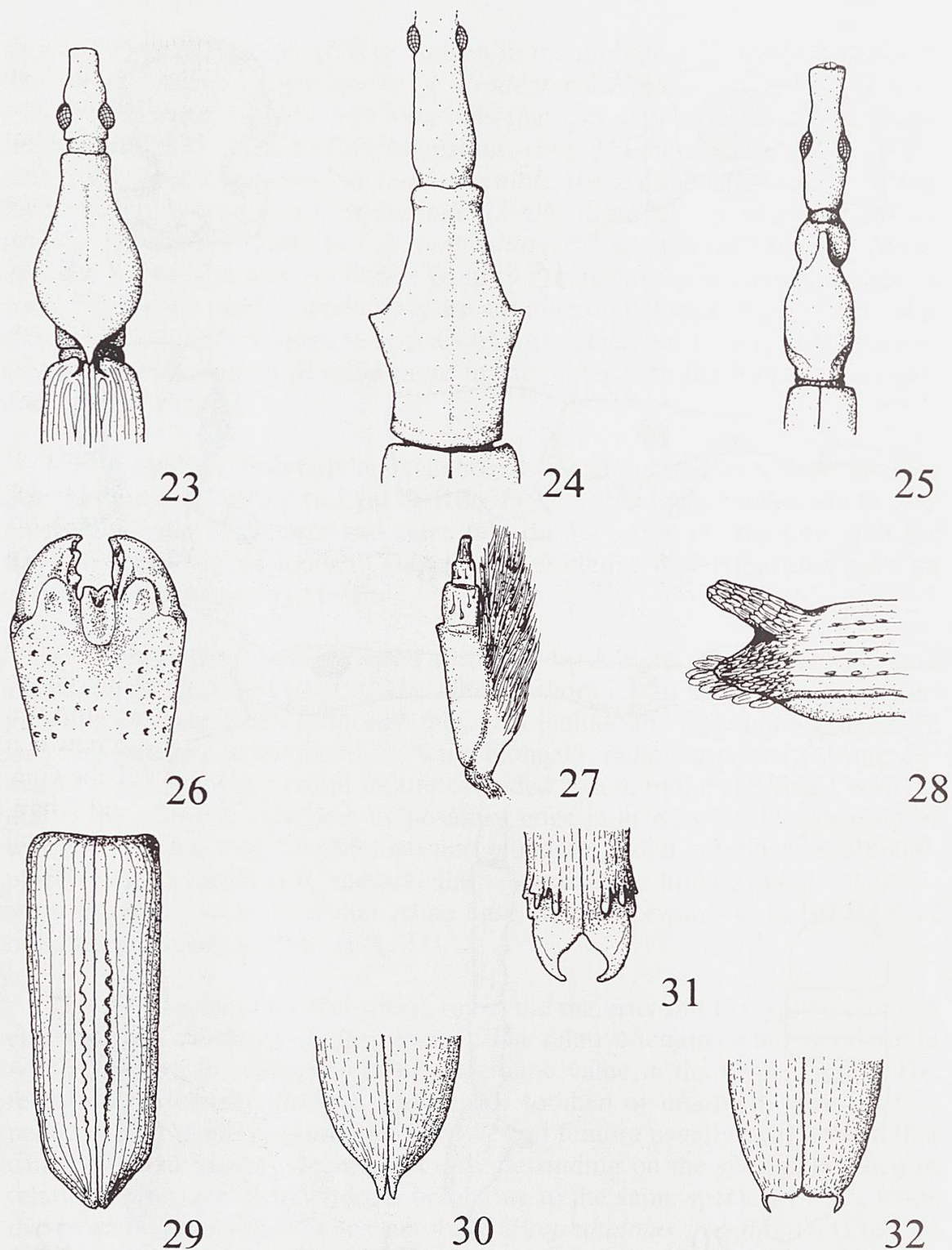
Kleine (1920a) reported the existence of a stridulation mechanism in brentids, however according to Lyal & King (1996) "his observations are of heterogeneous microsculpture and setae that do not agree in structure with the file of other Curculionoidea. There is no evidence that Brentidae have an elytro-tergal stridulatory system".

The wings have been studied in detail by Kleine (1920e, 1926g) and Zherikhin & Gratshev (1995). The latter authors report the following distinguishing characteristics (although they also include the Cyladinae, Apioninae and Nanophyinae in the family): "wing elongate; radial stripe very strong; radial fold disappearing; radial fissure extended basad; radial cell fused with radial stripe, strongly elongate; its posterior edge reduced with disappearing of window; stigmal fold longitudinal and elongate; radial sclerite longitudinal, placed behind radial cell; radiomedial sclerotization linear; radial sclerotization lost (not unique); medial stripe base shifted forward up to the level of radiomedial sclerotization" (Fig. 33).

The legs can be short and strong (as in the majority of the Cyphagogini) or else long and slender (e. g. *Ithystenini*). The relative length of the metafemora with respect to the abdomen is of systematic value in the Cyphagogini. The femora are normally clubbed and can be toothed or unarmed, more or less pedunculated in the proximal portion. Armed femora usually bear a tooth that can develop to varying degree, not only depending on the species but also in relation to the size of individuals belonging to the same species. In rare cases there can be two (Fig. 35) or even three (*Prophthalmus tridentatus*) (Fig. 36) femoral teeth. In the males of *Ithystenomorphus femoralis*, the posterior femora exhibit a square shaped swelling on the lower portion (Fig. 37). In the Cyphagogini, the anterior femora are flattened laterally so they can fit into the appropriate cavities in the prothorax (see Zimmerman, 1994: 135, Fig. 157). In *Myrmecobrenthus* and *Paussobrenthus* both the femora and the tibiae of the three pairs of legs are flattened. The tibiae are normally straight and end in



Figs 15-22 - Fig. 15: *Mesoderes guttatus* (Kleine), male, teeth on the ventral part of head and rostrum; Fig. 16: *Rhynchoneus belti* Sharp, ventral cephalic and prothoracic protuberances; Fig. 17: *Hopliterrhynchus emmae* Senna, dorsal protuberance on rostrum; Fig. 18: *Opisthozemius vittatus* Kolbe, scrobes visible from front; Fig. 19: *Microsebus adelphus* Kolbe, scrobes not visible from front; Fig. 20: *Myrmecobrenthus wasmanni* Kleine, antenna; Fig. 21: *Allacompsus separator* Kleine, first antennal segments; Fig. 22: *Ulocerus lacerosus* Schoenherr, antenna.



Figs 23-32 - Fig. 23: *Allagogus brunneus* Gahan, posterior prothoracic apophysis; Fig. 24: *Thaumastopsis gemmaria* Kleine, thorax; Fig. 25: *Cyphagogus signipes* Lewis, pronotal carina; Fig. 26: *Brentus lineicollis* Boheman, dorsal view of the apex of pronotum; Fig. 27: *B. lineicollis*, lateral view of maxilla; Fig. 28: *Ulocerus laticornis* Sharp, hair tufts on elytral declivity; Fig. 29: *Adidactus striolatus* (Fairmaire), saw-toothed elytral suture; Fig. 30: *Opisthozemius appendiculatus* Kolbe, pointed elytral apex; Fig. 31: *Carcinopisthius lorae* Senna & Calabresi, caudal appendages; Fig. 32: *Elytracantha contraria* Kleine, caudal appendages.

one or two spurs. The genus *Cerobates* retains the primitive tibial spur formula 2:2:2 (Zimmerman, 1994). In the Stereodermini, the protibiae have a deep hollow on the internal margin that serves as a grooming organ for the antennae (Fig. 34); in other Cyphagogini this organ is sometimes a mere tooth or small carina. According to Kuschel (2003) the New Zealand species *Lasiorynchus barbicornis* have the “fore legs with a broad, densely hairy grooming device between outer tooth and spur”. In brentids the tarsi are normally rather short, but in some cases the first article can be elongated. In *Carcinopisthius* “the first tarsite of the protarsus is serrate beneath to form a kind of rake which is used in cleaning the burrows” (Zimmerman, 1994). The third article can be entire or bilobate. All Brentidae have simple tarsal claws. *Lasiorynchus barbicornis* has no protruding apical lobe on the ventral surface of the claw segment, and the claws have one or more setae (Kuschel, 2003).

Extraordinary morphological modifications can be found in several species of *Calodromus* and *Cormopus*, where the tibiae as well as the posterior tarsi assume incredible shapes (Figs 38, 91). In *Ceragogus appendiculatus* from Madagascar, the metatibiae of the males are armed with a long, backward pointing spiniform process (Fig. 93).

The fore legs are articulated on the lower portion of the prothorax. The prosternum lies anterior to the procoxal cavities, whilst between the coxae there is a pentagonal structure called the prosternellum. The mesosternum lies between the pro- and mesocoxae, whilst the metasternum is situated between the meso- and metacoxae. It is well known that the five ventrites of the Curculionoidea abdomen are morphologically sternites 3-7, sternites 1 and 2 having been absorbed into the hind coxal cavity. Sternite 8 is the next ventral

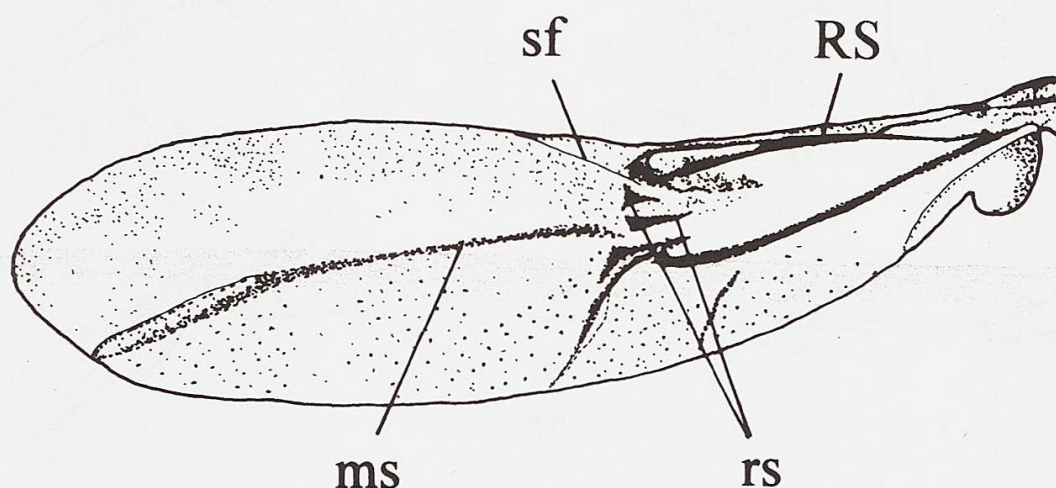
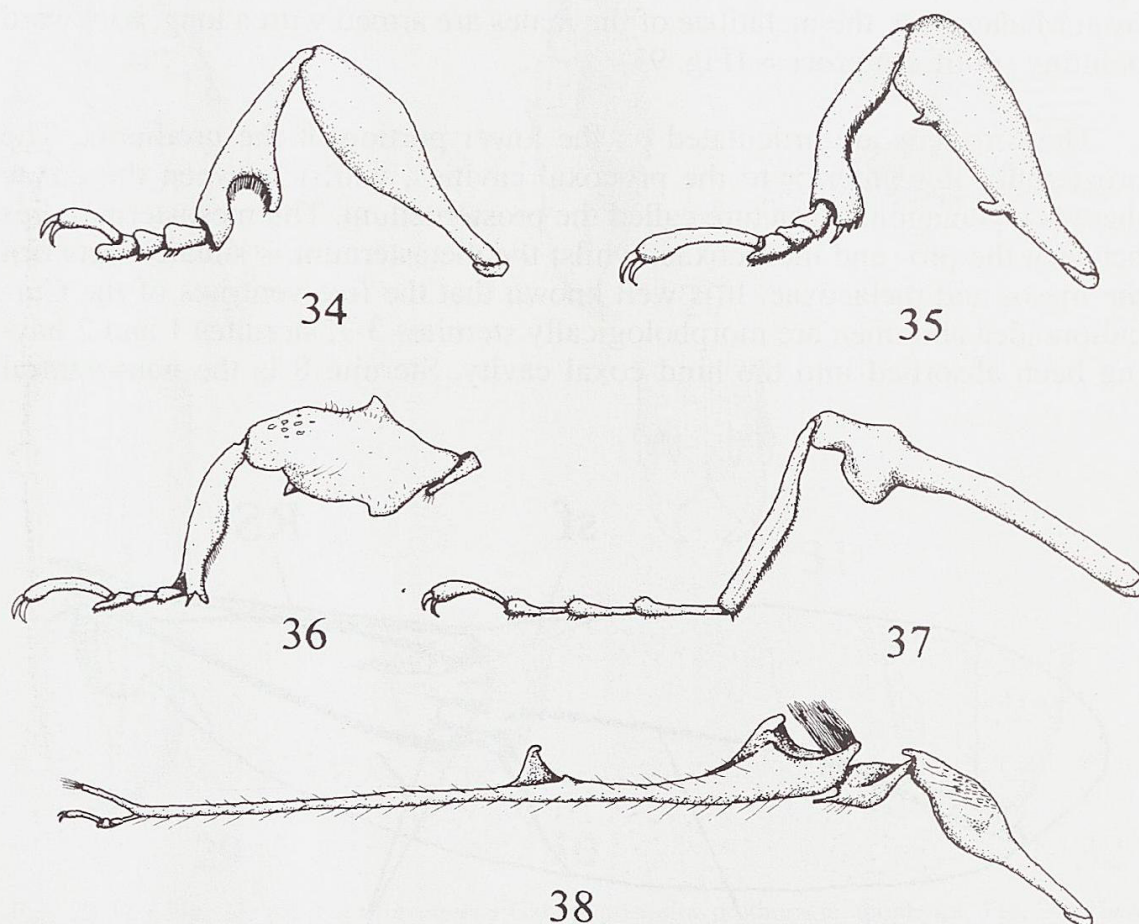


Fig. 33 - *Cyphagogus samoanus* Kleine, wing. Abbreviations: ms: medial stripe; rs: radial sclerite; RS: radial stripe; sf: stigmal fold (from Zherikhin & Gratshev, 1995, modified).

sclerite after ventrite 5 and is concealed within the genital chamber (Thompson, 1992). In brentids, ventrites 1-2 are much longer than the others and are fused together, the last three ventrites are short and free in the female, whilst ventrites 4-5 are fused in the male. Ventrites 1-2, fused together, give rise to the abdominal plate. Thompson (1992) studied the morphology of several anatomical parts of the Curculionoidea: for brentids in particular he studied and illustrated venter and sternite 8 of *Cyphagogus westwoodii*, *Brentus anchorago* and *Pseudoceocephalus depressus* and the tegmina of *Brentus anchorago*.

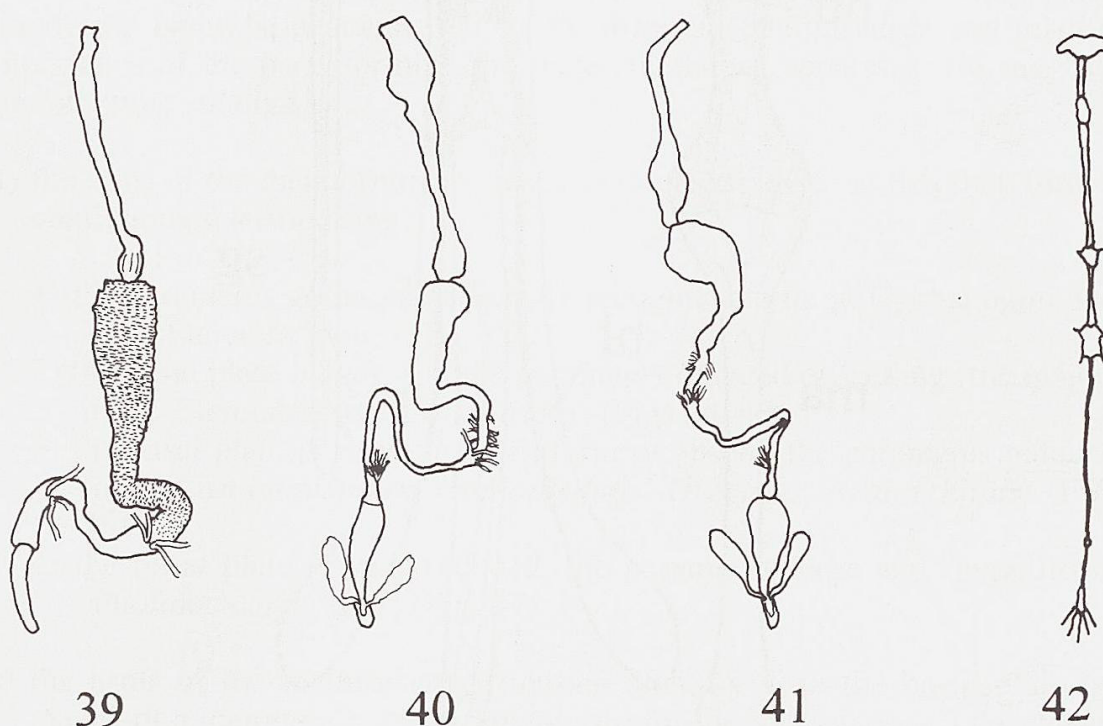
Kasap & Crowson (1977) studied the alimentary canal in *Arrenodes minutus* and *Cerobates (Ionthocerus) ophthalmicus* [sub *Cerobates australasiae*]. Calder (1989) examined the digestive tracts of *Allaeometrus breviceps* (Fig. 39), *Baryrhynchus (Eupsalomimus) lineicollis* (Fig. 40), *Ectocemus decem-*



Figs 34-38 - Fig. 34: *Cerobates sulcatus* Boheman, fore leg; Fig. 35: *Brentus vulneratus* Gyllenhal, male, hind leg; Fig. 36: *Prophthalmus tridentatus* (Lund), male, hind leg; Fig. 37: *Ithystenomorphus femoralis* Kleine, male, hind leg; Fig. 38: *Calodromus mellyi* Guérin-Méneville & Gory, male, hind leg with first tarsomere extremely modified.

maculatus (Fig. 41), *Ithystenus hollandiae*, *Trachelizus bisulcatus*, and *Tracheloschizus dichrous*. All these species, except *A. breviceps*, "have an elongate fore gut which is slightly longer than the mid gut and occupies 23-36 % of the length of the alimentary canal. However in *A. breviceps* the fore gut is very much shorter than the mid gut ... The mid gut is usually shorter than the hind gut except in *A. breviceps* where the mid gut is slightly longer than the hind gut. ... There are 4 cryptonephric Malpighian tubules usually inserted anteriorly in 1 group, except for *A. breviceps* which has the tubules arranged in 2 closely sited groups of 2" (Calder, 1989: 1210).

The central nervous system of *Orfilaia* has been studied by Kasap & Crowson (1977); Calder (1989) examined *Allaeometrus breviceps*, *Trachelizus bisulcatus* (Fig. 42), *Baryrhynchus* (*Eupsalomimus*) *lineicollis* and *Ectocemus decemmaculatus*. "All these species have a discrete cerebrum, sub-



Figs 39-42 - Fig. 39: *Allaeometrus breviceps* Senna, alimentary canal; Fig. 40: *Baryrhynchus* (*Eupsalomimus*) *lineicollis* (Power), alimentary canal; Fig. 41: *Ectocemus decemmaculatus* (Montrouzier), alimentary canal; Fig. 42: *Trachelizus bisulcatus* (Lund), central nervous system (from Calder, 1989, modified).

oesophageal ganglion and prothoracic ganglion, a fused meso-metathorac ganglion and two discrete abdominal ganglia. The abdominal connectives are extraordinarily elongate between the thoracic and first free abdominal ganglion and are separate in *Allaeometrus breviceps*. However the connectives are apparently fused in *Trachelizus bisulcatus*, *Baryrhynchus* (*Eupsalomimus*) *lineicollis* and *Ectocemus decemmaculatus*" (Calder, 1989: 1227).

Kasap & Crowson (1977) studied the abdominal skeletal structure and the abdominal musculature; Zimmerman (1994: 28) contributed some remarks.

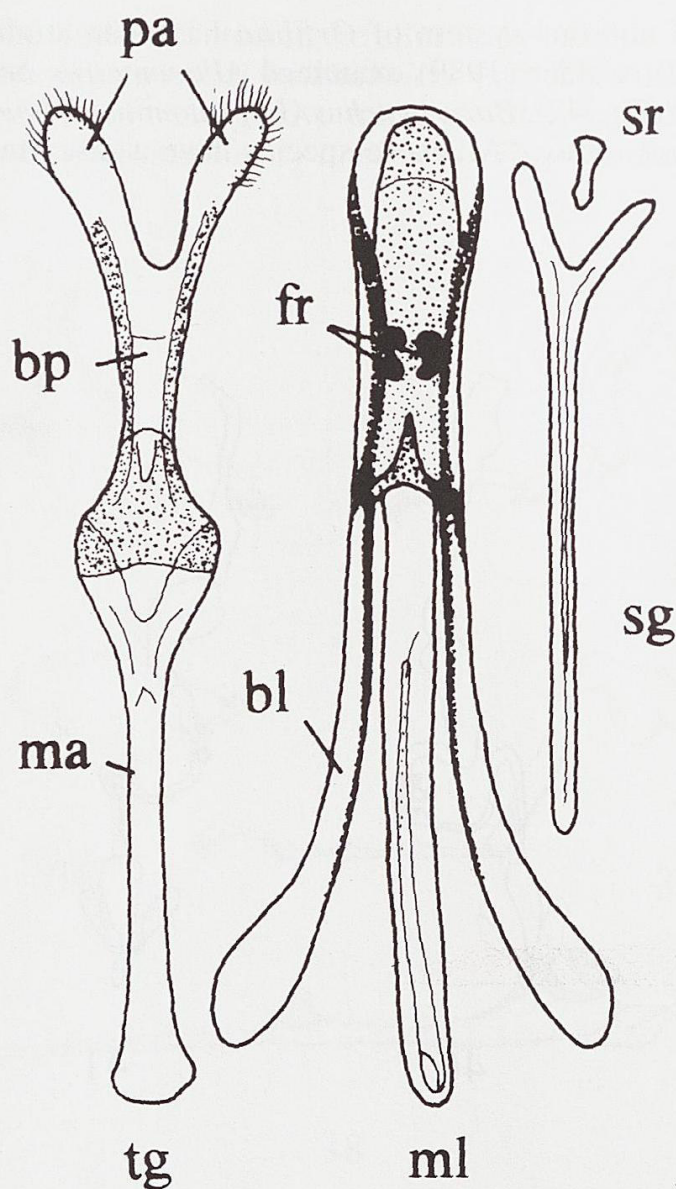


Fig. 43 - *Allagogus brunneus* Gahan, male genitalia. Abbreviations: bl, basal lamina; bp, basal plate; fr, frena; ma, manubrium; ml, median lobe; pa, parameres; sg, spiculum gastrale; sr, spiculum relictum; tg, tegmen.

The male copulatory apparatus (Fig. 43) consists of a Y shaped spiculum gastrale (sternite 9), a small sclerified part called spiculum relictum (an apodeme of sternite 8), a tegmen, formed of a ventral manubrium and a basal dorsal plate that carries the parameres at its tip, and a median lobe (aedeagus). The parameres can be well developed, more or less reduced or even absent. The manubrium and basal plate can articulate between each other or be fused together to form a ring in which the median lobe moves freely; the latter is a long, straight or curved tube with a dorso-ventral, more or less chitinised, bi-laminar structure that holds the internal sac. The basal portion of the median lobe lengthens into two long basal laminae that serve for the insertion of the muscles. The internal sac is a membranous structure that ends in an ostium and is equipped with two small chitinised reniform plates called frena. Sometimes the internal wall may have sclerified raised areas or spiny or dentiform processes (Pseudoceocephalinae). At the proximal tip of the internal sac, in correspondence to the testis and seminal duct, there is a more or less developed flagellum (Damoiseau, 1967a: 16). According to Zimmerman (1994: 26) the occurrence of the frena is noteworthy stable from genus to genus and tribe to tribe.

According to Damoiseau (1963b: 195) several groups can be distinguished among the brentids, characterised by the different conformation and relative importance of the parts forming the male copulatory apparatus. He suggests the following subdivision:

- A) the arms of the manubrium are fused to the basal plate, so that they form a continuous sclerified ring.
 - 1) the parameres are large and easily recognizable (e. g. Cyphagogini and some Stereodermini) (Fig. 43)
 - 2) the basal plate is long and the parameres reduced or lacking (the majority of Stereodermini, *Callipareius*) (Figs 44, 45)
 - 3) the basal plate is very small, filiform or absent, the parameres reduced (e. g. *Carcinopisthius*, *Hoplopisthius*, *Higonius*, *Araiorrhinus*) (Fig. 46)
 - 4) the basal plate is very reduced, the parameres large and flagelliform (Pholidochlamydinae) (Fig. 47)
- B) the arms of the manubrium articulate dorsally with the basal plate by means of a membrane; the parameres are for the most part well developed and cover the median lobe dorsally and laterally (Brentinae, Ulocerinae, Taphroderinae) (Fig. 48)
- C) the arms of the manubrium are fused to expansions of the basal plate, the sclerified ring is interrupted in the tergal region (*Nemocephalus*, *Acratus*, etc.) (Fig. 49).

We observed that the genera *Stratiorrhina* (Arrhenodini) (see Sforzi & Bartolozzi, 1993) and *Rhinopteryx* (Pseudoceocephalini) (see Orbach, Bartolozzi & Sforzi, 1995) fall within group B described by Damoiseau, whilst the genus *Ceocephalus* (Ithystenini) (see Sforzi & Bartolozzi, 1997) falls within group C.

The female reproductive apparatus (Fig. 50) consists of tergite 8, triangular or semicircular in shape, toothed or non-toothed on the external margin, the dorsal surface equipped with a pair of glands or "tergal pouches" (Kuschel, 1987), and by sternite 8 that is shaped like an oar. According to Kuschel (2003), *Lasiorrhynchus barbicornis* is unique to all other Brentinae in having "four intersegmental glandular pouches instead of two or none".

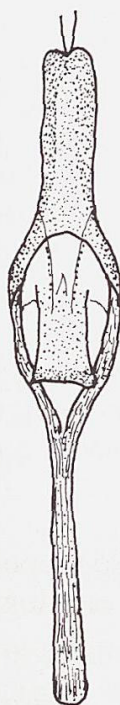
The coxites are drawn out into a pair of long ventral baculi, with small stili at the tip (Brentinae, Pseudoceocephalinae, Ithysteninae) or on the side (Cyphagoginae). A membranous vagina without reinforcement or sclerotization, a bursa copulatrix and a common oviduct are present; the spermatheca is linked to the bursa copulatrix by a small circumvolved tube of variable length and is equipped with an accessory gland.

Morimoto (1962a: 362) illustrated the bursa copulatrix, the spermatheca and its gland in *Baryrhynchus poweri*. In this species the capsule is sickle-shaped, the gland is small and distally distended, and the duct is very long and convoluted.

Kasap & Crowson (1977) illustrated the soft male reproductive organs in *Arrenodes minutus* and *Cerobates (Ionthocerus) ophthalmicus* and the female reproductive organs in *Arrenodes minutus*. The male *A. minutus* has "a pair of bilobed testis, a pair of tubular accessory glands, a pair of round and large vesiculae seminales and a short and simple ejaculatory duct. Each testis lobe is multifollicular and enclosed in a separate membranous sheath. The vasa efferentia are greatly enlarged. In this species, the accessory glands open to the vasa efferentia, thus presenting an unusual situation as they open to the vesiculae seminales in other Curculionoidea. The vesiculae seminales are soft and membranous-looking whereas they are usually heavily muscled in the others. ... The female reproductive organs consist of two ovaries each of two ovarioles, short oviducts, a globular and very distinct bursa copulatrix and a long vagina. The spermathecal duct is long and convoluted." According to Calder (1990: 457) the presence of a pair of single tubular accessory glands in males should be checked again in the species, as an accessory gland was not present in any of the six species of Brentidae he examined. He studied the morphology of the soft parts of the male and female reproductive systems in the Curculionoidea. He illustrated the reproductive systems for the following brentid species: *Allaeometrus breviceps* (Figs 51, 54), *Arrenodes minutus*, *Baryrhynchus (Eupsalomimus) lineicollis*; (Figs 52, 55), *Ectocemus decem-*



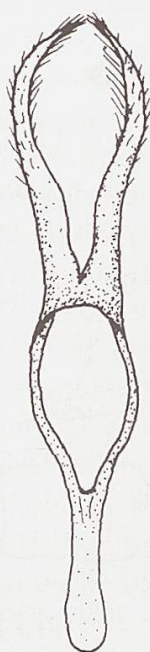
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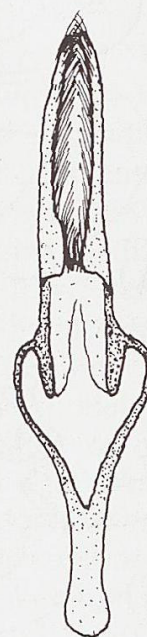
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Figs 44-49 - Tegmen. Fig. 44: *Cerobates (Cerobates) sulcatus sulcatus* Boheman; Fig. 45: *Cerobates (Cerobates) dilutus* Kleine; Fig. 46: *Carcinopisthius doriae* (Senna); Fig. 47: *Pholidochlamys madagascariensis* Lacordaire; Fig. 48: *Bolbocranius delhaisei* (Damoiseau); Fig. 49: *Acratus tarsatus* (Gyllenhal).

maculatus, *Trachelizus bisulcatus* (Figs 53, 56), *Tracheloschizus dichrous*.

The characters of the brentidae family are as follows: Male - "testis bilobed; anterior section of vasa deferentia greatly enlarged; accessory glands absent; seminal vesicle multilobed or single-lobed"; female - "two ovarioles per ovary; ovarioles usually with terminal filaments; spermatheca and spermathecal gland well-developed; spermathecal duct and gland inserted on spermatheca separately; spermathecal duct entering vagina-bursa copulatrix ventrally" (Calder, 1990: 476).

Howden (1995) studied the structures related to oviposition in the Curculionoidea, examining the species *Arrenodes minutus* and mentioning *Brenatus anchorago* and *Claeoderes radulirostris mexicanus* [sub *Claeoderes guatemalensis*] in the brentids.

Johnson (1983a) and Howden (1995) report observations on the use of the dentate tergite 8 (see the chapter on ethology). The dentate tergite 8 of the

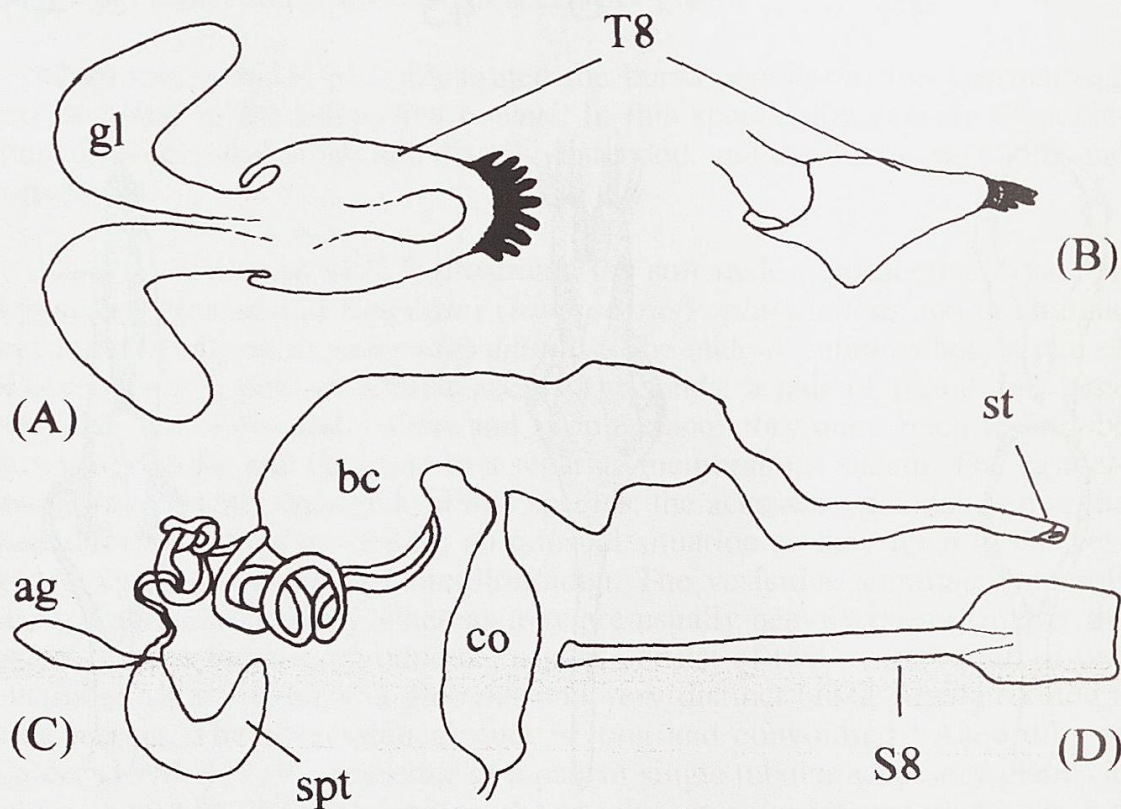


Fig. 50 - *Arrenodes minutus* (Drury), female reproductive structure: (A) tergite 8, ventral view; (B) tergite 8, lateral view; (C) reproductive tract, lateral view; (D) sternite 8. Abbreviations: ag, accessory gland; bc, bursa copulatrix; co, common oviduct; gl, gland; spt, spermatheca; st, stylus; S8, sternite 8; T8, tergite 8 (from Howden, 1995, modified).

brentid is associated with the method of oviposition, in particular with the spreading of the patch material over the hole. The ovipositor of *Brentus anchorago* was illustrated by Tanner (1927); Morimoto (1962a) illustrated those of *Baryrhynchus poweri*, and Kasap & Crowson (1977) those of *Arrenodes minutus*.

Virkki studied the chromosomes of *Brentus anchorago*, *B. mexicanus*, *Neacratus femoratus*, *Proepisphales elegans*, and *Trachelizus* sp. The karyotype $10 + X_y$ is present in all species. The chromosome relations resemble those of Curculionids. In *B. anchorago* the y chromosome is particularly large; in *P. elegans* the sex bivalent is associated with some autosome bivalents by means of the nucleolus.

Larvae

Compared with the number of adults described in the Brentidae, very few larvae have been studied. The larvae are white, slender, fleshy and have an exserted hypognathous head. Small, soft and few segmented thoracic legs are often present (Oberprieler & Louw, 1985), but the larvae of the Cyphagogini are peculiar in lacking any trace of leg segmentation (Gardner, 1935; May, 1993).

According to May (1994: 439) the brentid larval characters are: "Body elongate, curved, truncate posteriorly; abdominal segments with 3-4 folds. Head free; postoccipital condyles absent; frontal suture reaching mandibular membrane; endocarinal line absent; ocelli absent; antenna 1-segmented, a slender cone on a basal cushion, enclosed within frontal plate. Mandibles bifid (or unidentate if eroded) with sharp incisor edge and small median tooth; mola undeveloped. Labrum having 2 or 3 pairs of primary setae; 1 or 3 sensilli; tormae short, strong, convergent. Epipharyngeal lining with 0-3 anterolateral setae. Hypopharyngeal bracon clear or pigmented. Tentorial bridge sometimes laterally subdivided. Maxilla lacking a palpiger; palpi 2-segmented; mala conical, without a lacinial spine. Mentum free laterally. Labial palpi 1 or 2-segmented. Postlabium with posterior pair of postlabial setae as wide apart as median pair. Pronotum strongly transverse. Mesothorax with a dorsolateral triangular area, behind the postdorsal setae, usually rugose, often pigmented. Spiracles variable in shape, with or without airtubes, lateral on abdominal segment VIII. Legs 2-segmented or absent. Alimentary canal (only *Lasiiorhynchus* examined) with proventriculus expanded; lacking mycetomes around cardiac valve; gastric caeca absent; with 4 Malpighian tubules; cryptonephridium symmetrical; rectal bracon a membraneous ring". May assembled these characters from the New Zealand species *Lasiiorhynchus barbicornis*, the Australian species *Allaeometrus breviceps* and the genera *Cyphagogus* and *Cerobates*.

Harris (1838) was the first to describe a brentid larva, which he attributed to the species *Arrenodes minutus*. Chapuis & Candèze (1853) briefly re-described the larva of *A. minutus* (from Harris), but expressed doubts about its correct identification. Riley (1874) verified that the specimen Harris described was actually a Tenebrionidae larva and described and illustrated the true *A. minutus* larva and pupa for the first time. Packard (1881), Felt (1905) and Sharp (1936) resumed Riley's original descriptions and figures. Böving & Craighead (1931) again described and illustrated the *A. minutus* larva. Sanborne (1981) illustrated the first instar larva of *A. minutus* (Fig. 57).

Motschulsky (1856) described the larva of *Estenorhinus* sp. (sub *Arrhenodes*) from Venezuela, although Sharp (1936: 295) did not consider it to be a brentid.

The larva and pupa of *Lasiorhynchus barbicornis* from New Zealand were first described by Broun (1881). Muir (1913) and Hudson (1934) both reconsidered the description; Muir illustrated the larva (Fig. 58) and pupa. In the male pupa (Fig. 63) the antennae, stemming from near the tip of the rostrum, flank each side of the head, and as the head and rostrum are deflexed, they lie on the ventral surface of the thorax. In the female (Fig. 64) the antennae, rising from about mid rostrum, encircle the thorax.

On the basis of Riley's illustrations Lameere (1903) treated the systematic position of the brentids in the Curculionoidea, but according to Heller (1904) he was probably misled by the imprecision of the drawings.

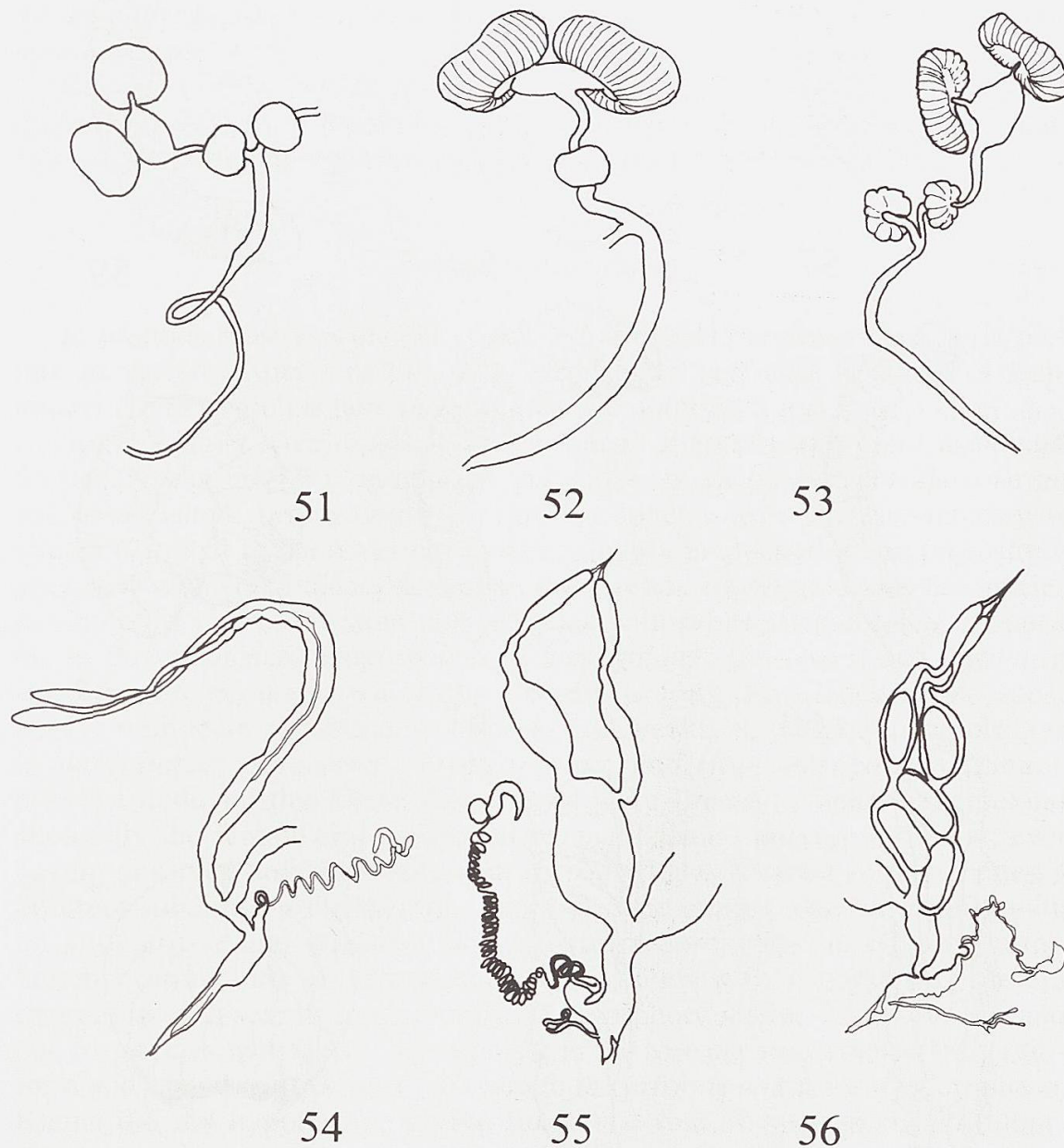
Heller (1904) describes the larva of the Brazilian species *Brentus lineicollis* that exhibits three pairs of 2-jointed legs, furnishing an historical excursus on the brentid larvae described to that date.

Gardner (1925) described the larva of *Cerobates* (s. s.) *tristriatus* (drawings that De Muizon and Zimmerman re-used in 1960 and 1994 respectively), *C.* (s. s.) *sexsulcatus* and *Cyphagogus corporaali*, and in 1935 he published a paper entitled "Immature stages of Indian Coleoptera". In this paper he gave the key to the subfamilies and some genera and species occurring in India, as well as describing and illustrating in detail the larvae of the following species: *Callipareius* (s. s.) *bistriatus* [sub *Opisthenoxys bistriatus*], *Callipareius* (s. s.) *foveatus* [sub *Pittodes mediocris*], *Cyphagogus incisus* [sub *Cyphagogus obconiceps*], *Cerobates* (*Ionthocerus*) *carinensis* (Fig. 59), *Trachelizus bisulcatus* (Fig. 60), *Higonius cilo*, *Carcinopisthius oberthueri*, *Anepsiotes commendabilis*, *Ectocemus cinnamomi*, *Prophthalmus heikertingeri*, *Orfilaia truncata* [sub *Eupsalis truncata*], *Schizotrachelus cavus* [sub *Opisthenoplus cavus*].

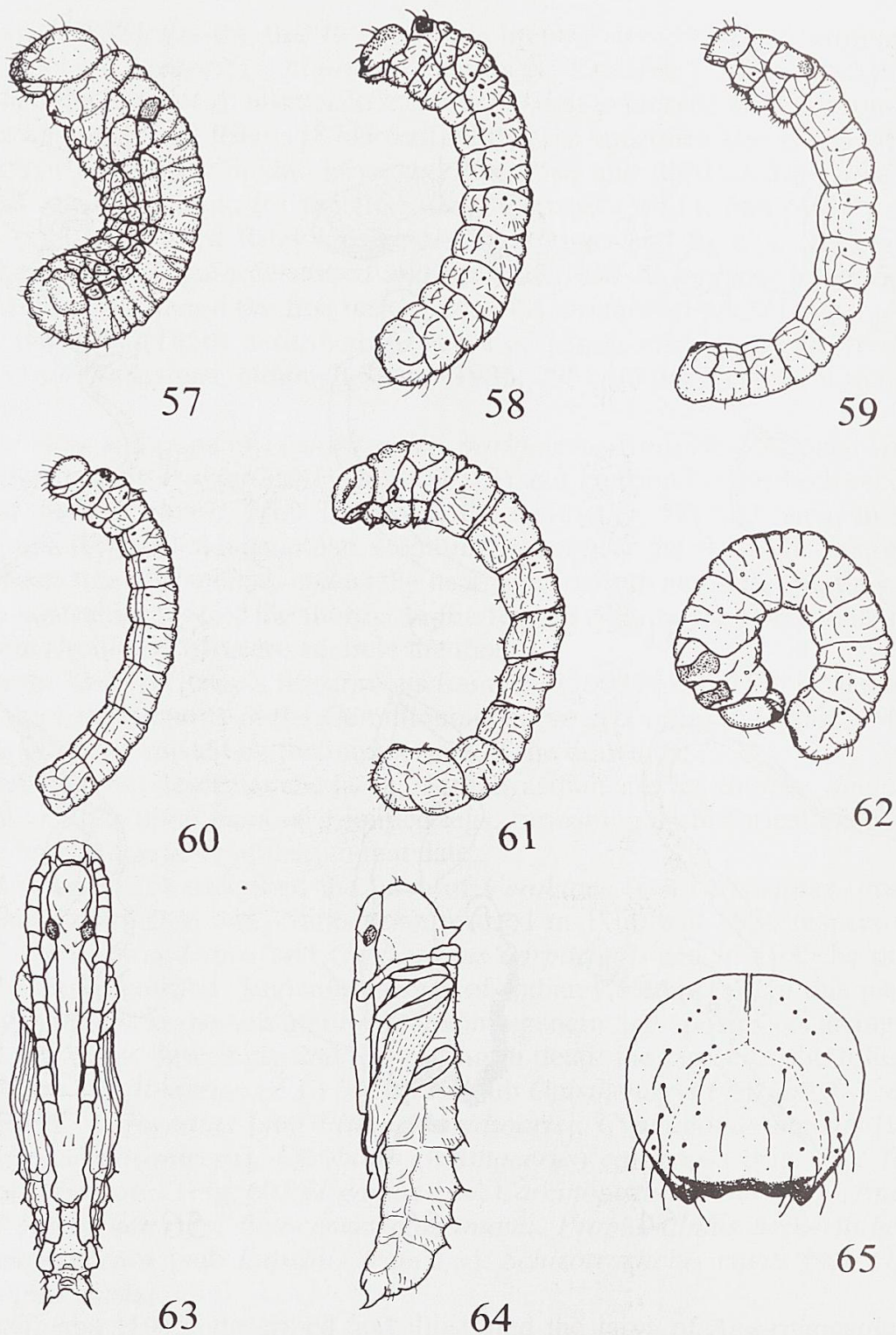
Hoffmann (1945) described and illustrated the larva of *Amorphocephala coronata* (Fig. 61).

Haedo Rossi (1961) illustrated the larva and pupa of *Brentus lineicollis*; Morimoto (1962a) furnished a key to distinguish brentid larvae from the larvae of other Japanese Curculionoidea.

Sanborne (1981) performed a comparison between the larvae of *Arrenodes*



Figs 51-56 - Male reproductive organs. Fig. 51: *Allaeometrus breviceps* Senna; Fig. 52: *Baryrhynchus (Eupsalomimus) lineicollis* Power; Fig. 53: *Trachelizus bisulcatus* (Lund). Female reproductive organs. Fig. 54: *Allaeometrus breviceps*; Fig. 55: *Baryrhynchus (Eupsalomimus) lineicollis*; Fig. 56: *Trachelizus bisulcatus* (from Calder, 1990, modified).



Figs 57-65 - Fig. 57: *Arrenodes minutus*, first instar larva; Fig. 58: *Lasiorhynchus barbicornis*, larva; Fig. 59: *Cerobates (Ionthocerus) carinensis*, larva; Fig. 60: *Trachelizus bisulcatus*, larva; Fig. 61: *Amorphocephala coronata*, larva; Fig. 62: *Allaeometrus breviceps*, larva; Fig. 63: *Lasiorhynchus barbicornis*, male, pupa; Fig. 64: *Lasiorhynchus barbicornis*, female, pupa; Fig. 65: *Allaeometrus breviceps*, head (drawings from: Muir, 1913; Gardner, 1935; Hoffmann, 1945; Sanborne, 1981; May, 1993; 1994; modified).

minutus, *Ithycerus noveboracensis* (Forster, 1771) (Ithyceridae) and *Antliarhinus* [sub *Antliarhis*] sp. (Antliarhininae).

May (1994: 441) provided a key for the identification of the larvae of some Australasian brentids and illustrated the larvae and pupae of *Allaeometrus breviceps* (Figs 62, 65) and *Cerobates (Ionthocerus) ophthalmicus*.

Kuschel (1995) and Oberprieler (2000) employed several larval characters (generally taken from the above mentioned literature) for their cladistic analysis of Brentidae phylogenesis (see the chapter on systematics).

Sexual dimorphism

In brentids most species exhibit marked sexual dimorphism. A general picture of various aspects of this dimorphism can be found in Sforzi & Bartolozzi (1998) which is here re-examined and amplified.

Some authors have reported on the sexual dimorphism present in brentid beetles. Senna (1892b) studied the variability of shape in the male rostrum and established three categories: one for species in which the rostrum is almost identical in the male and female, slightly broadened at the prorostrum apex, and with small mandibles (as in *Brentus* and *Ithystenus*); one for species in which the rostrum is stout and wrinkled with robust mandibles at the apex (as in *Baryrhynchus*, *Prophthalmus*, *Estenorhinus*, *Arrenodes*, and *Orfilaia*); and one for species in which the rostrum is long, broadened at the apex, always with teeth or tubercles or spines and weakly developed mandibles (as in *Belopherus*, *Ectocemus*, *Stratiorrhina*, and *Hopliterrhynchus*). Senna (1892b) deduced that these differences were linked to behaviour (sexual choice by the female and protection from predators) without, however, ever having observed any such behaviour directly. Later Kleine (1931b) created a different subdivision for brentids, again in three groups: one for species with no apparent sexual dimorphism in rostrum conformation (Calodromini, Stereodermini); one for myrmecophilous species with a markedly different rostrum in each sex, strongly modified (Amorphocephalini, *Mesitogenus*); and one for species with sexual dimorphism in the rostrum and females with a fili-form and cylindrical rostrum with which to perforate wood for ovodeposition. Kleine did not hypothesize on the functional role of rostrum sexual dimorphism in males.

De Muizon (1960) and Damoiseau (1967a) reviewed some of the secondary sexual characters encountered in African brentids; Zimmerman (1994) synthesized this argument for Australian species.

In some brentids sexual dimorphism is absent or very slight, such as in most Cyphagoginae and Trachelizinae, but it is usually not only obvious but occasionally even striking, affecting various parts of the body: the head



Fig. 66 - *Brentus anchorago* (L.), male (on left), female (on right) (photo by S. Bambi).



Fig. 67 - *Nemorhinus myrmecophaga* (Herbst), male (on left), female (on right) (photo by S. Bambi).

(rostrum, mandibles, antennae), pronotum, elytra, legs (femora, tibiae, tarsi), and underside (abdominal plate, last abdominal segment).

The rostrum is usually cylindrical and filiform in the females; in the males it is usually longer (e. g. *Brentus*, *Nemorhinus*) (Figs 66, 67) and can be either very dilated apically (e. g. *Estenorhinus*, *Gyalostoma*) (Figs 68, 69), rough (*Ectocemus decemmaculatus*) (Fig. 70) or saw-toothed laterally (*Stratiorrhina*) (Fig. 11), with a medial upright protuberance (*Hopliterhynchus*) (Fig. 17) or markedly pubescent beneath (*Lasiorrhynchus barbicor-*



Fig. 68 - *Estenorhinus* spp., male (on left), female (on right) (photo by S. Bambi).

nis) (Fig. 71). In the Eremoxenini the rostrum is generally highly modified in both sexes, but in males it is usually more robust and the mandibles are more developed; numerous examples are illustrated in Zimmerman (1994) (Figs 7, 10, 13, 14).

With regard to the genus *Diurus* (now *Ceocephalus*), Senna (1911: 32) points out that the relative lengths of the pro - and metarostrum of females and males are inversely proportional. In species where the male metarostrum is much longer than the prorostrum, the females have a much longer prorostrum



Fig. 69 - *Gyalostoma elegans* Kleine, male (on left), female (on right) (photo by S. Bambi).

than metarostrum, where the male metarostrum is shorter than the prorostrum the opposite occurs in the females.

De Muizon (1955a: 465) shows how the anterior margin of the rostrum can be more or less hollowed in both sexes even in those species with modest sexual dimorphism, e. g. *Adidactus striolatus*.

Damoiseau (1989b: 93) points out that in the female of *Tulotus maculipennis*, the only example among the Trachelizini, the prorostrum widens at the tip as in males; as there is no other secondary sexual characteristic, the two sexes can only be distinguish by the copulatory apparatus.

The male mandibles can be long and falcate or very robust (e. g. *Estenorhinus*, Fig. 68; *Gyalostoma*, Fig. 69; *Prophthalmus*, Fig. 72). In male *Bolbocranius csikii* (Fig. 73) and *B. distortus* the left mandible is extremely



Fig. 70 - *Ectocemus decemmaculatus* (Montrouzier), male (on left), female (on right) (photo by S. Bambi).

long, flat and sinuate, the right one very small; in the female the mandibles are small and normal in shape (Fig. 74) (Arrow, 1940). The use of such an unusual structure is unknown.

The antennae are sometimes much longer in the males, as in *Cerobates* subgen. *Ionthocerus*, *Ceocephalus forcipatus*, *Eterodiurus singularis* (Sforzi & Bartolozzi, 1993) (Fig. 81). In *Allagogus* the antennal articles are of normal shape in the males (Fig. 75) but enlarged and modified in the females (Fig. 76); it is one of the rare instances in which the female, and not the male, presents strong structural modifications. De Muizon (1960: 20) points out that in some African *Pseudoceocephalini* the rostrum is similar in both sexes, but in males the antennae are inserted further from the base and therefore the pro-rostrum is shorter than in females. Senna (1911) remarks that the number of



Fig. 71 - *Lasiorynchus barbicornis* (F.), male, rostrum (above), female, rostrum (below) (photo by S. Bambi).

articles of the antenna can vary from nine to eleven within the genus *Diurus* [now *Ceocephalus*], depending on the species but not on the sex as occurs in other Curculionoidea.

In the males of *Cerobates* subgen. *Ionthocerus* the eyes cover almost the entire head (Fig. 77), whilst in the females they are smaller, lateral, with a large interocular distance (Fig. 78).

In some species of the genera *Carcinopisthius* and *Hoplopisthius* the head of the females bears a large pubescent rhomboid or elongated area (Fig. 82), whilst the metarostrum in males is smooth.

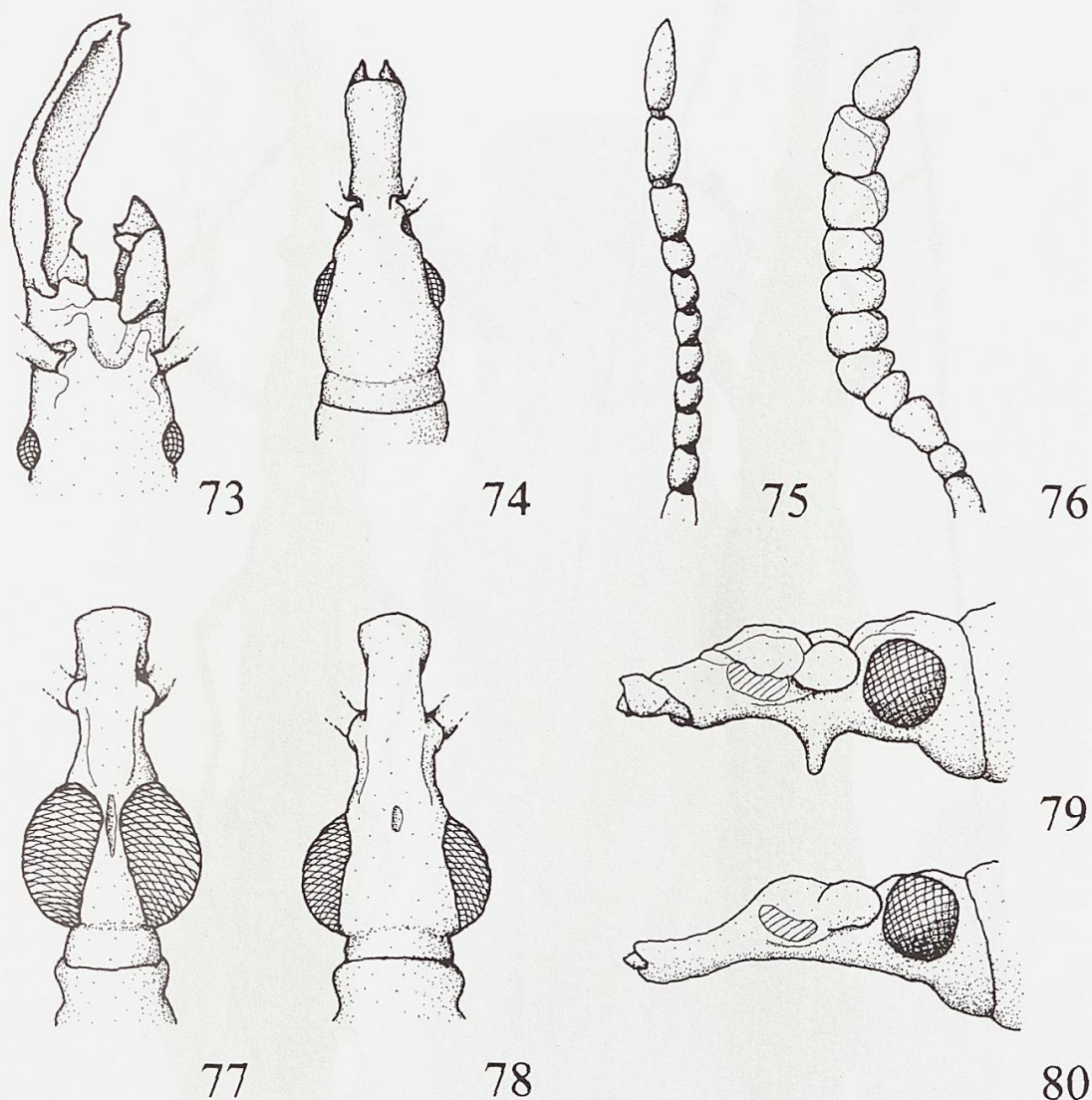


Fig. 72 - *Prophthalmus planipennis* Pascoe, male (on left), female (on right) (photo by S. Bambi).

De Muizon (1960: 20) remarks that in some species of African *Cerobates* the males exhibit a triangle of golden appressed hairs on the lower part of the head, which is absent in the females.

In some species of Cyphagogini the males may bear strong and pointed teeth on the ventral part of the head and rostrum (e. g. various species of *Mesoderes*) (Fig. 15).

In *Ankleineella*, the males have a strong ventral medial tooth on the metarostrum, absent in the females (Zimmerman, 1994) (Figs 79, 80). There is also a strong ventral process in the males of *Rhynchoneus belti*, but



Figs 73-80 - Fig. 73: *Bolbocranius csikii* (Bolkay), male, head, rostrum and mandibles; Fig. 74: *B. csikii*, female, head, rostrum and mandibles; Fig. 75: *Allagogus brunneus* Gahan, male, antenna; Fig. 76: *A. brunneus* Gahan, female, antenna; Fig. 77: *Cerobates (Ionthocerus) conradti* Senna, male, head; Fig. 78: *C. (Ionthocerus) conradti*, female, head; Fig. 79: *Ankleineella australis* (Lacordaire), male, lateral view of head; Fig. 80: *A. australis*, female, lateral view of head.



Fig. 81 - *Eterodiurus singularis* Senna, male (on left), female (on right) (photo by S. Bambi).

this is situated under the head and not under the rostrum (Fig. 16).

The pronotum is long, slender and narrowed in the middle in male *Brentus anchorago*, shorter and trapezoidal in the females (Fig. 66).

In some species, the male elytra have long caudal appendages, which can be short or absent in the females (e. g. *Ceocephalus forcipatus*, *Prodeutor*



Fig. 82 - *Carcinopisthius oberthueri* (Senna), female, head and rostrum (photo by L. Bartolozzi & A. Sforzi).

laminatus) (Figs 83, 84). In the male of *Eterodiurus singularis* the elytra are strongly narrowed in the middle, slightly so in the female (Fig. 81). In *Carcinopisthius* caudal appendages are found in both sexes but they differ in shape (Arrow, 1940) (Figs 85, 86). Miyatake (1956) points out that the tip of the elytra also differ in shape even though there are no appendices in the males or females of *Higonius cilo* (Figs 87, 88).

The legs of the males often have more or less stout spines on the femora, and sometimes also on the protibiae (*Orychodes digramma*, *Spatherrhinus*; Figs 89, 90). In *Cormopus* the posterior legs are strongly modified, and differ



Fig. 83 - *Ceocephalus forcipatus* (Westwood), female (on left), male (on right), caudal appendages (photo by S. Bambi).



Fig. 84 - *Prodeutor laminatus* Pascoe, male (on left), female (on right) (photo by S. Bambi).

in both sexes (Figs 91, 92). In *Ithystenomorphus femoralis* the male metafemora are markedly swollen and differ in shape from those of the females (Fig. 37). In *Ceragogus appendiculatus* the male metatibia exhibit a long, strong, backward pointing spiniform process, almost as long as the tarsi (Figs 93, 94).

In *Hoplopisthius trichemerus* the males bear long hairs under their pro-femora, whilst the females have a thick brush of much shorter hairs.

In *Rhynchoneus belti* the males have a very particular raised process on the lower part of prothorax that is not found in other species of brentids (Fig. 16).

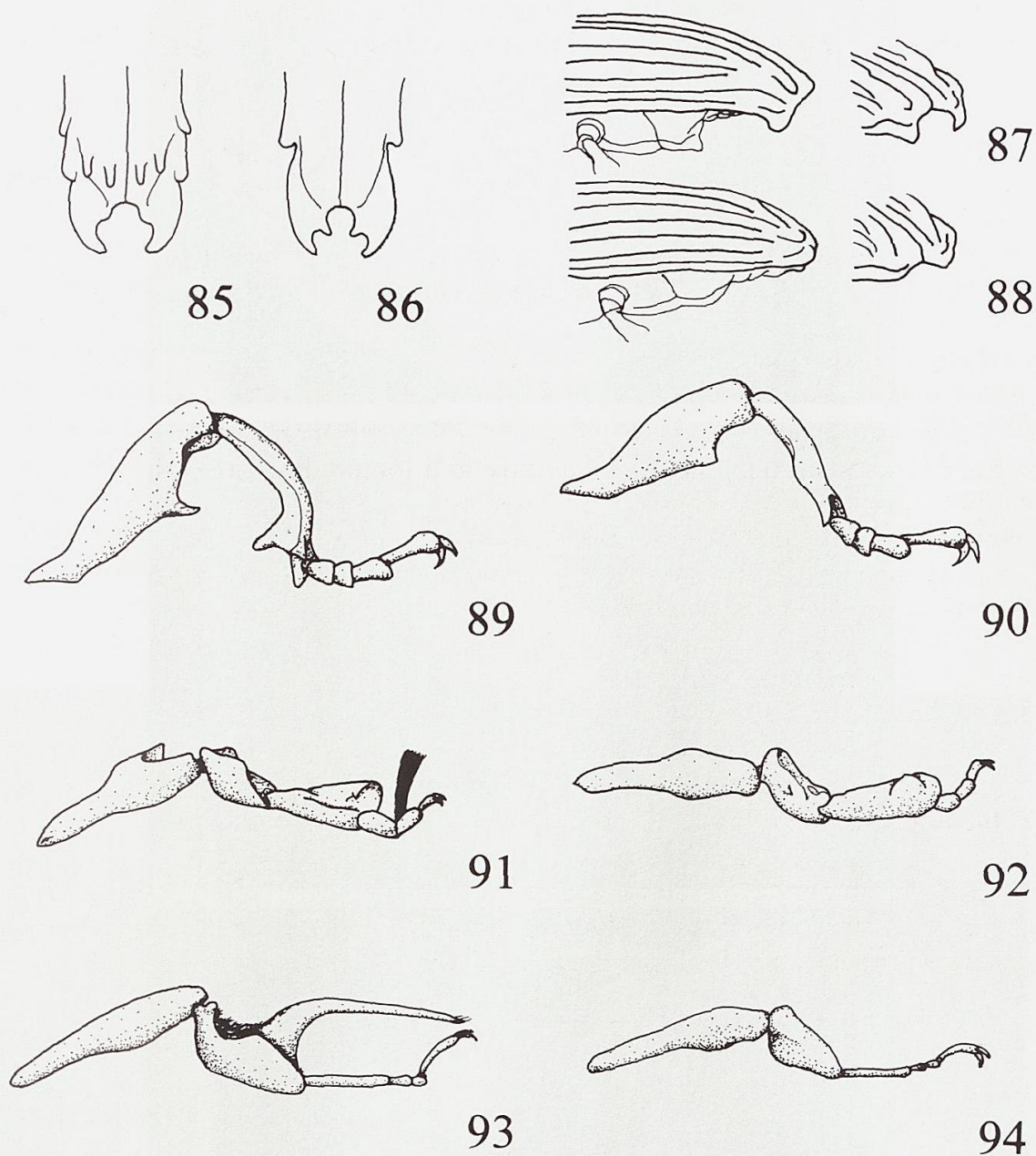
The male underside usually has a more or less concave abdominal plate, flat in the females, and a differently shaped last abdominal segment.

For instance, in male *Pseudoceocephalus depressus* the last abdominal segment bears two deep hollows laterally to a longitudinal medial carina and the posterior margin is concave; in females the same segment is flat with a transverse sulcus and the posterior margin is convex. It is interesting that in the other species of the same genus, *Pseudoceocephalus formosus*, the last abdominal segment has two hollows in both sexes (Damoiseau, 1967a).

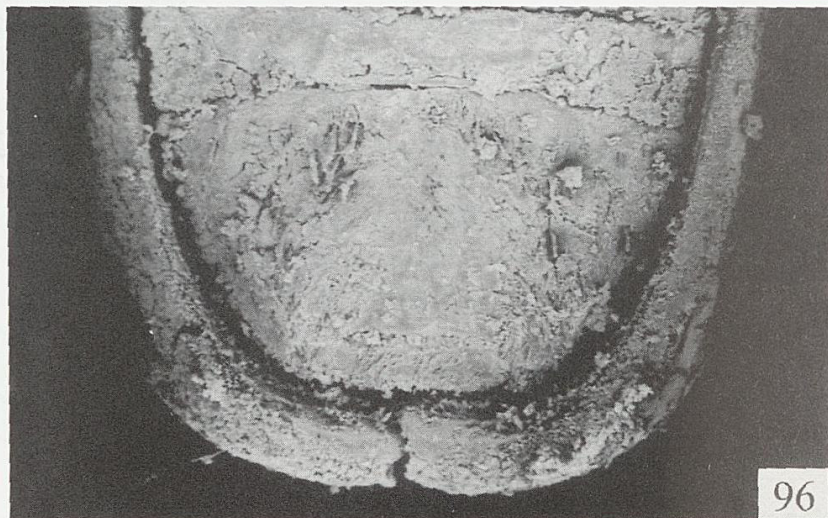
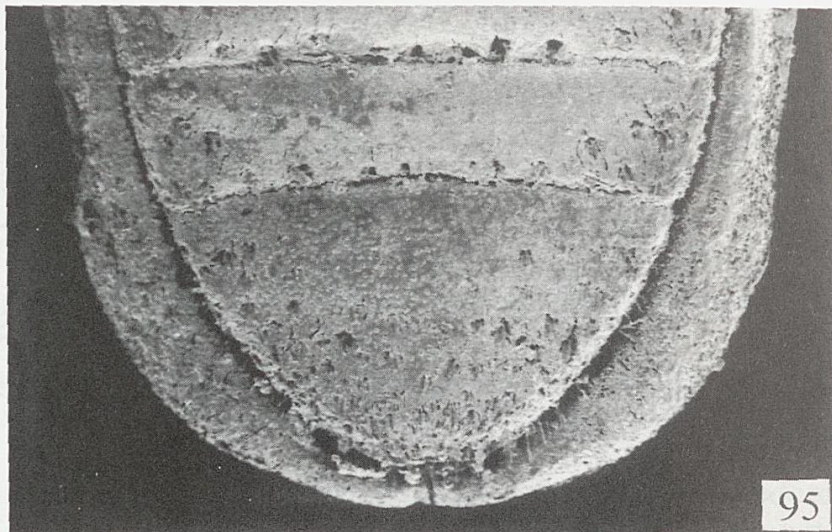
In *Paleoceocephalus nitidissimus* males, the apical sternite has 3 foveolae arranged in a triangle, whilst in the females the last segment is simply punctuated (Damoiseau, 1989b).

In *Rhinopteryx foveipennis*, the last abdominal segment is flat and convex at apex in the female, whilst it is hollow in the middle and posteriorly truncate in the male (Figs 95-97).

Both the function of the structures linked to sexual dimorphism and the use of such structures in understanding the phylogenetic relationships among genera and species are far from clear.



Figs 85-94 - Fig. 85: *Carcinopisthius loriae* Senna & Calabresi, male, caudal appendages; Fig. 86: *C. loriae*, female, caudal appendages; Fig. 87: *Higonius cilo* Lewis, male, tip of the elytra; Fig. 88: *H. cilo*, female, tip of the elytra; Fig. 89: *Orychodes digramma* (Boisduval), male, fore leg; Fig. 90: *O. digramma*, female, fore leg; Fig. 91: *Cormopus penicillifer* Kolbe, male, hind leg; Fig. 92: *C. penicillifer*, female, hind leg; Fig. 93: *Ceragogus appendiculatus* (Fairmaire), male, hind leg; Fig. 94: *C. appendiculatus*, female, hind leg.



Figs 95-97 - Fig. 95: *Rhinopteryx foveipennis* (J. Thomson), female, last abdominal segment; Figs 96-97: *R. foveipennis*, male, last abdominal segment.

ECOLOGY AND BIOLOGY

Brentids mostly live in rain forests, but some genera and species also occur in temperate, arid or sub-desert climates. The vast majority of forest-related species are xylophagous, whilst in xeric habitats they are mostly myrmecophilous. Brentids inhabit not only primary forest but also secondary and degraded forest.

Over the last few years, we have been on several expeditions to various parts of the world looking for brentids and have had the opportunity of observing the different life habitats of these beetles. In South America we collected several species of *Brentus* and *Estenorhinus* under the barks of felled trees in the Yasuni National Park, Ecuador (Fig. 98). This area is a lowland rain forest on the Amazonian side of the Andes, near the river Rio Napo. Again in Ecuador, but in an Andean montane forest, near Rio Hollin (Fig. 99) we observed other species of *Brentus* under the barks of fallen trees and individuals of *Estenorhinus* on the leaves, whilst in highland secondary forest at Union del Toachi (Fig. 100) we found *Brentus approximatus*, *B. caudatus*, and various species of *Estenorhinus* and *Stereodermus* under tree barks.



Fig. 98 - Ecuador: rainforest in the Yasuni National Park (photo by A. Sforzi).



Fig. 99 - Ecuador: Andes mountain forest, near Rio Hollin (photo by L. Bartolozzi).



Fig. 100 - Ecuador: highland secondary forest, near Union del Toachi (photo by L. Bartolozzi).

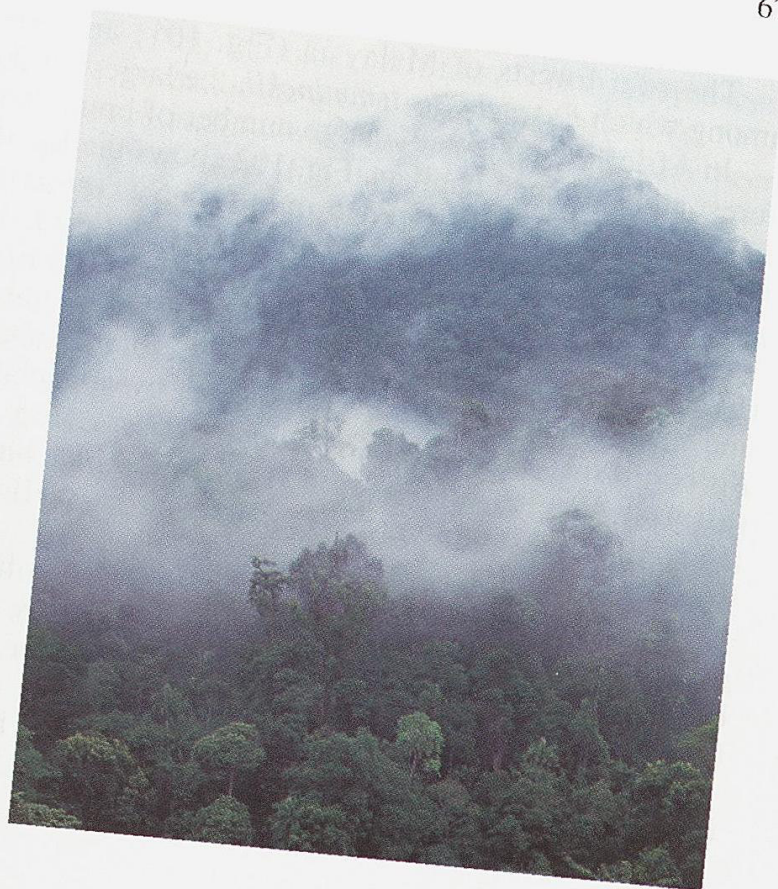


Fig. 101 - Malaysia: rainforest in the surroundings of Cameron Highland (photo by S. Bambi).

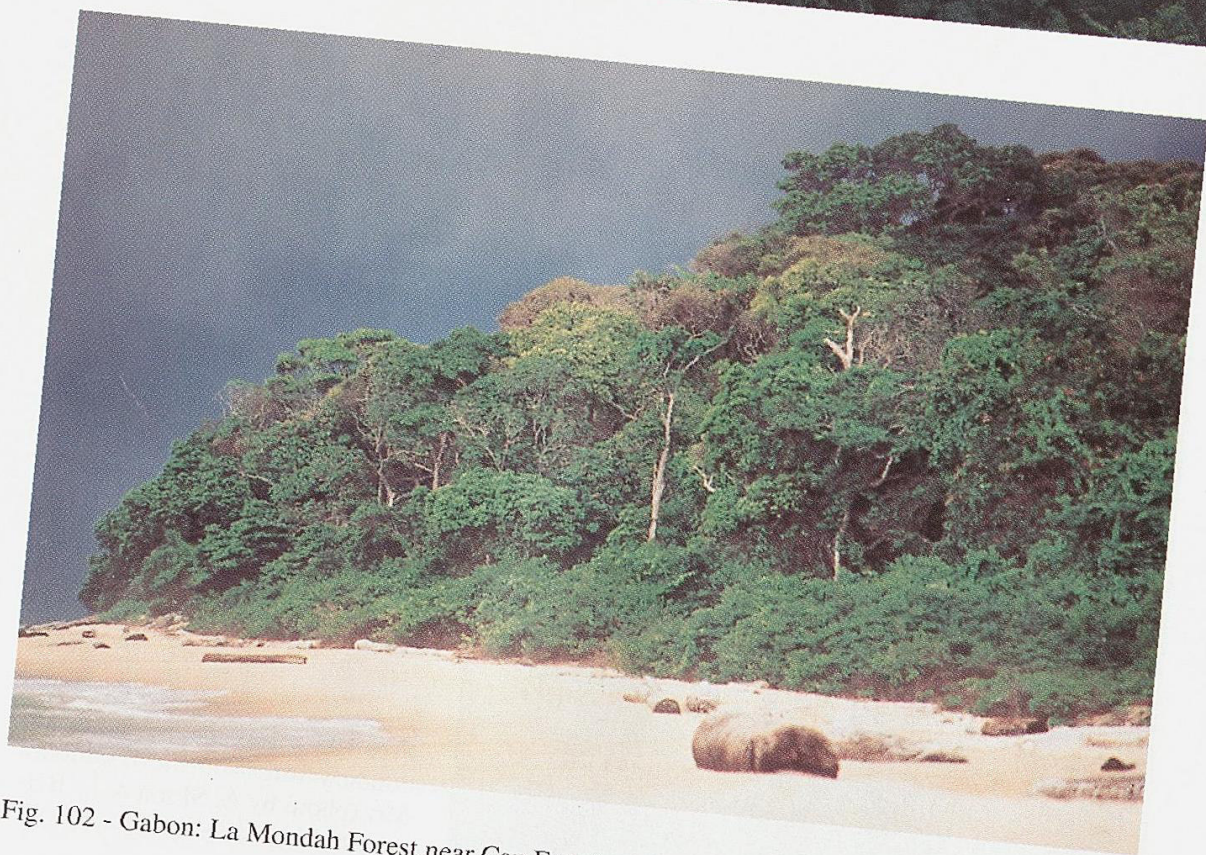


Fig. 102 - Gabon: La Mondah Forest near Cap Esterias (photo by L. Bartolozzi).

The rain forests of Malaysia (Fig. 101) are extremely rich in species, among which *Eutrachelus temminckii*, the largest of the brentids.

In Africa we collected a large number of brentids from both the equatorial forest and savannah habitats. Fig. 102 shows the La Mondah costal rain forest in Gabon, where *Debora bocandei*, *Synsebasius roubaudi*, *S. seminittens*, *Usambioproctus conjunctus*, *Cerobates* (s. s.) *conveniens*, *C.* (s. s.) *punctulatus*, and *Storeosomus riisii* all live (Bartolozzi & Sforzi, 1997). In the Usambara Mountains secondary forest habitat at Amani, Tanzania (Fig. 103) we found several species of *Cerobates* under the bark of fallen trees and managed to collect several species by artificial light at night, including *Bolbocranius csikii* and *Orfilaia vulsellata* (Fig. 104). We also collected this last species in Ethiopia from under the bark of a large fallen tree (Fig. 105) in secondary forest habitat as well as from thorny scrub in Botswana (Fig. 106).

We collected *Cormopus penicillifer* (Cyphagogini) and *Micramorphocephalus* sp. *prope frater* (Eremoxenini) by artificial light from a low bushland rocky habitat at Dakhata Valley, near Harar in Eastern Ethiopia (Fig. 107).

Various species of Eremoxenini can be found in the semi-arid thorny scrub of Kenya (Fig. 108) and Somalia (Fig. 109) (see Bartolozzi, 1986, 1991; Bar-



Fig. 103 - Tanzania: Amani, secondary forest on the Usambara Mts (photo by A. Sforzi & L. Bartolozzi).

tolozzi & Osella, 1990b; Sforzi, 1992; Bartolozzi & Sforzi, 1994). Eremoxeniini of the genus *Cordus* (Fig. 110) inhabit the arid areas of Australia (Fig. 111), while *Amorphocephala excantator* and *Symmorphocerus piochardi* have been collected by night in xeric habitats of Israel (Fig. 112).

The majority of adult brentids can be found under the barks of dead or partially decomposed trees and in some cases on flowers, leaves and blossoms. Other species can be seen by day walking over the surface of felled tree trunks in search of Platypodidae and Scolytidea tunnels where they often lay their eggs. Many species of Brentidae seem to be active only by night and are often attracted by artificial light.

In general, it can be said that little is known of brentid biology, although a few papers have been published. However, these mainly concern a limited number of species or particular geographical areas, for example the work by Schedl (1961a) on the biology of brentids in the Congo. Kleine (1921f, 1921i, 1923g, 1924d, 1927c, 1931b, 1931c, 1931d, 1931e, 1933a, 1933b, 1933c, 1933d, 1933e) published a number of papers on the biology of brentids, but usually only cited observations taken from previous papers, for example by Beeson (1925, 1941) on Brentids from India.

Brentids can be divided into three "biological groups" (Kleine, 1926j, 1931b, 1938d; Beeson, 1941; Haedo Rossi, 1961; Zimmerman, 1994):

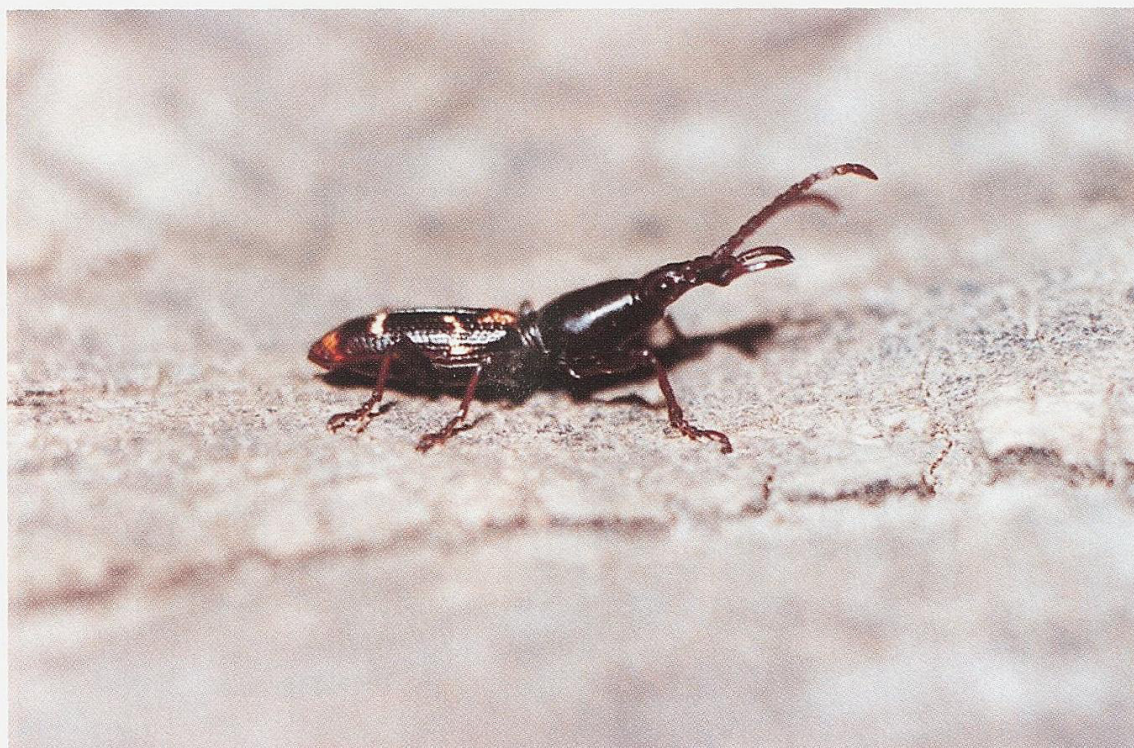


Fig. 104 - *Orfilaia vulsellata* (Gyllenhal) (Botswana) (photo by A. Sforzi & L. Bartolozzi).



Fig. 105 - Ethiopia: one of the Authors looking for Brentidae under the bark of a fallen tree near Bedele (photo by L. Bartolozzi).



Fig. 106 - SE-Botswana: thorny scrub near the Limpopo River (photo by A. Sforzi & L. Bartolozzi).

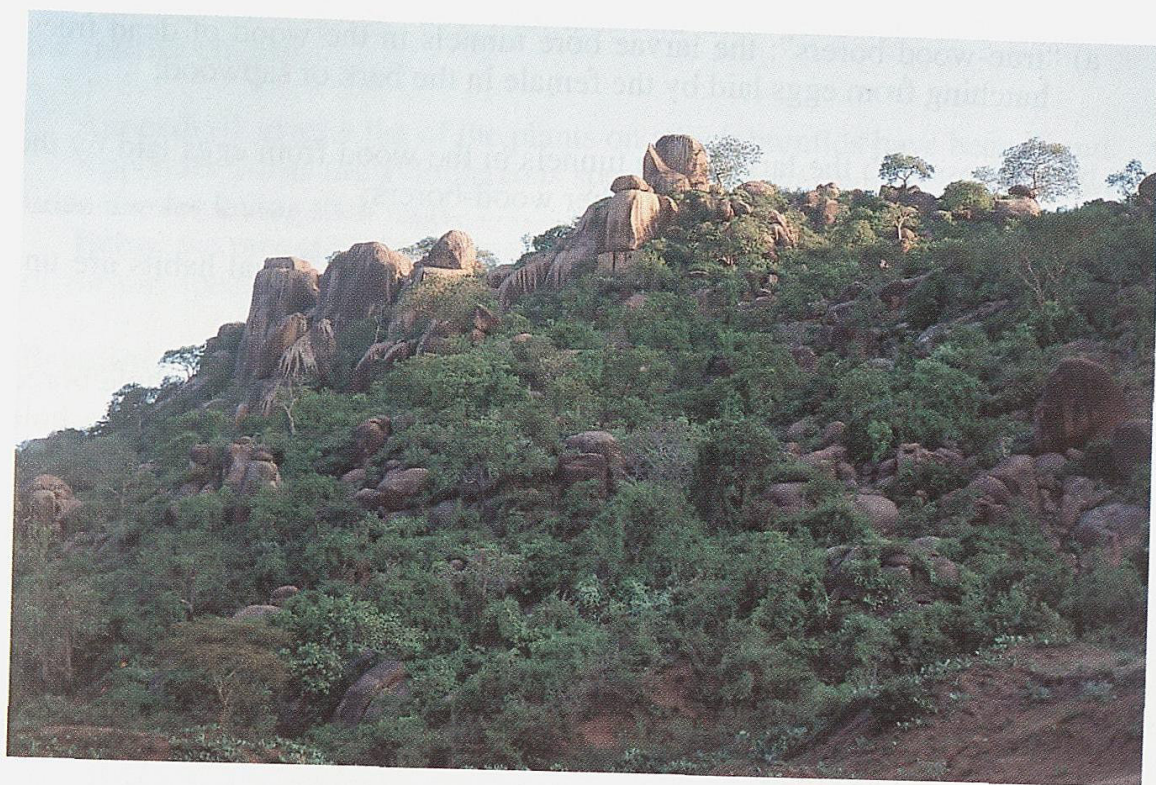


Fig. 107 - Ethiopia: Dakhata Valley near Harar (photo by A. Sforzi & L. Bartolozzi).

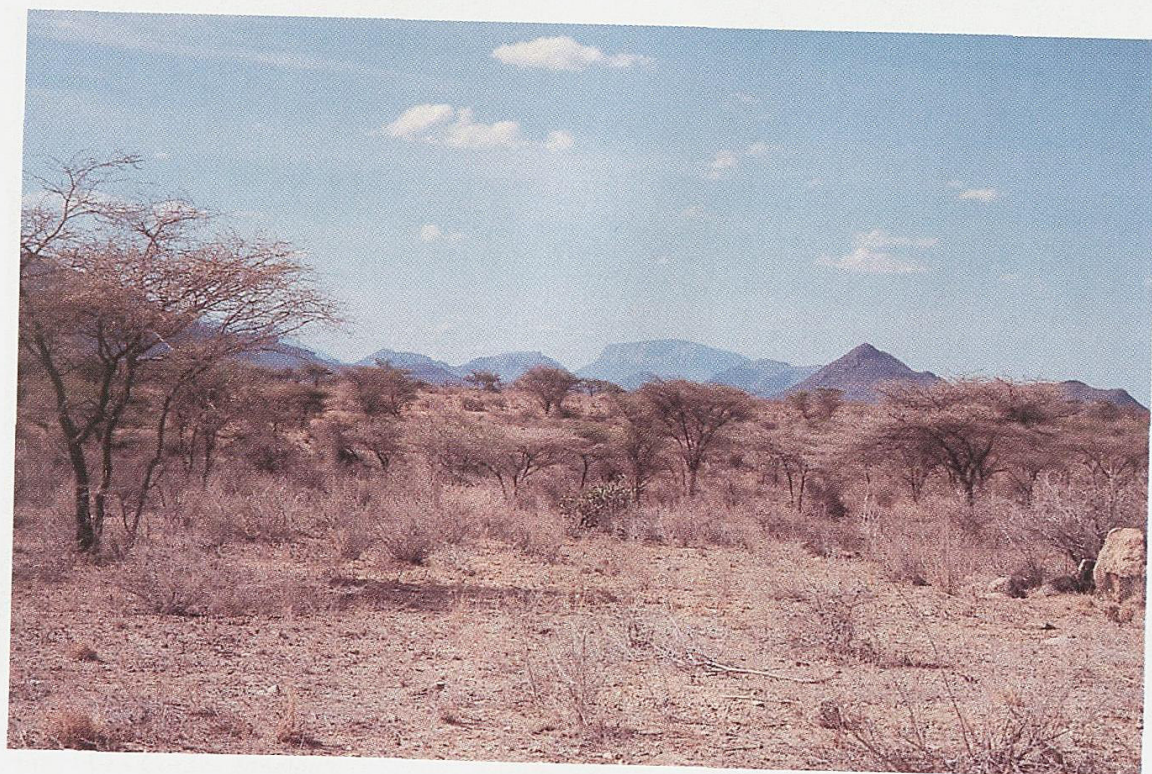


Fig. 108 - Kenya: dry scrub near Samburu (photo by A. Sforzi & L. Bartolozzi).

- a) "true-wood-borers": the larvae bore tunnels in the wood of dead trees, hatching from eggs laid by the female in the bark or sapwood;
- b) "aggressors": the larvae bore tunnels in the wood from eggs laid by the female inside the tunnels of other wood-borers;
- c) "myrmecophiles": the beetles live with ants, their larval habits are unknown.

"True-wood-borers" lay their eggs in sap-wood, sometimes in a hole bored by the female, and often protected by the sawdust produced when the hole was originally excavated. The larvae dig tunnels of various length from the site where the egg was laid and feed on sap or saprophytic fungi growing in the gallery. Pupation usually occurs in widened cells lying along the tunnel axis. The larvae emerge at the external opening of the tunnel on the trunk surface over the site of oviposition.

"Aggressors" exploit tunnels for their oviposition that have already been excavated by other wood-borers, such as the Platypodidae, Scolytidae and Cerambycidae. Several authors report that the galleries are used after the host has been ejected or killed.

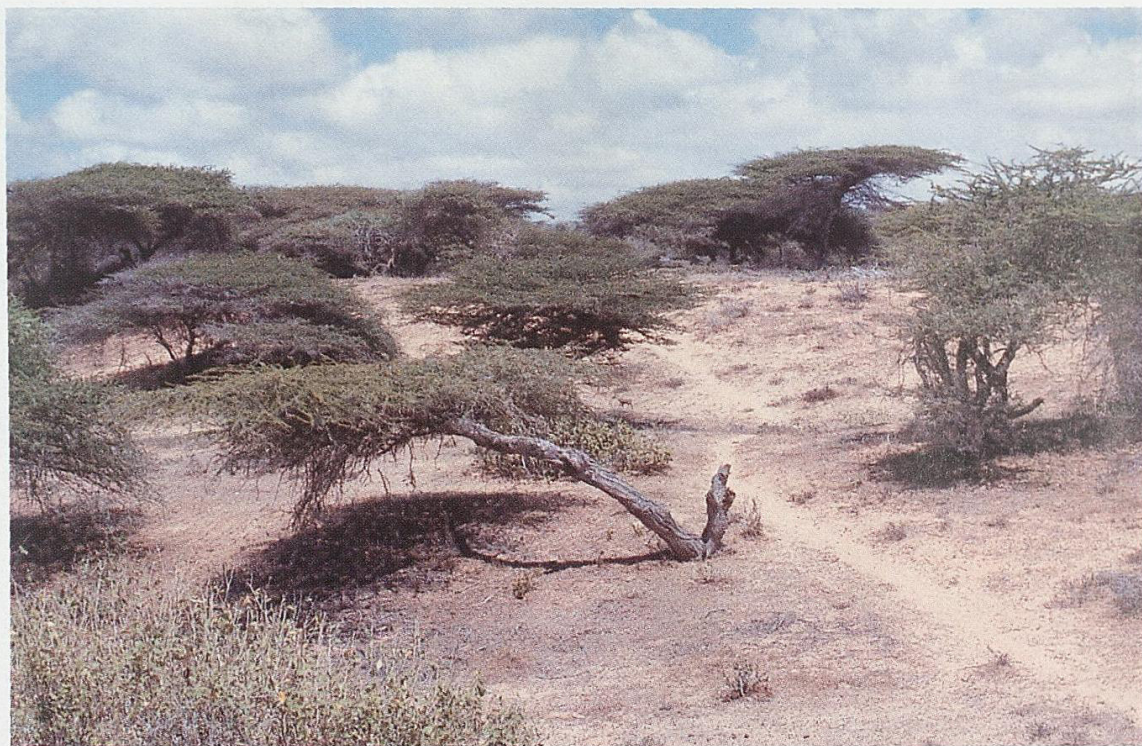


Fig. 109 - Somalia: dry scrub near Gesira (photo by L. Bartolozzi).

The “myrmecophiles” live with ants and their behaviour is discussed in the relative chapter.

Appendix 1 gives a list of the plants on which brentids have been found.

Appendix 2 lists the xylophagous species of beetles whose galleries Brentidae use for laying their eggs.

Below we provide information available from the literature on the biology of various species of Brentids, grouped by tribes.

Brentini

Haedo Rossi (1959b) described a *Brentus vulneratus* gallery in the trunk of *Balfourodendron riedelianum* Engl. in Argentina. The gallery, reaching a total of 6.5 cm in length with a maximum diameter of 4.5 mm, was filled in the initial part, 1.7 cm long, with tiny pieces of sawdust. The nymphal chamber, 2.4 cm long, was situated at the end of the first portion of the gallery and was followed by another 2.4 cm long tract.

Johnson (1983b) studied *Brentus anchorago* in Costa Rica. The adult brentids were frequently found in large numbers at densities of up to four hundred per square meter. The female bores a small hole in the bare wood of



Fig. 110 - *Cordus ganglbaueri* Senna (Karijini Nat. Park, Pilbara, Western Australia) (photo by R. Oberprieler).

a decaying tree with her mandibles, working her rostrum into the wood to the depth of the antennal insertion. In the hole, she deposits an elliptical, yolk egg 1-2 mm long, depending on the size of the female. The larva chews a gallery below the cambium, where it presumably feeds on sap or fungal mycelia. The separate larval galleries are crowded close to one another but are not thought to come into contact. The larval period may last a year. As adults, the beetles can survive for more than one breeding season. During the dry season the adults live by feeding on sap or by visiting flowers. They insert their rostra into flowers of *Cordia curassavica* (Jacq.) Roem & Schult., *C. inermis* (Mill.) IM Johnst. (Boraginaceae), *Casearia corymbosa* Kunth (Flacourtiaceae) and *Calycophyllum candidissimum* (Vahl.) DC. (Rubiaceae).

Johnson (1983a) investigated *Claeoderes radulirostris mexicanus* [sub *C. bivittata*] in Panama on a dying *Quararibea asterolepis* Pittier tree. The female chews a hole for 30-60 minutes until her rostrum is buried as far as the antennal insertion. Periodically she withdraws her rostrum, lifts her head, and expels sawdust from her jaws. To oviposit, she turns around and locates the drilled hole by tapping with her rear end and hind legs and remains still for 70 seconds to 3 minutes laying a single egg in the hole. A female may drill and oviposit three times in succession.



Fig. 111 - Australia: Karijini National Park, Pilbara, Western Australia (photo by R. Oberprieler).

Arrhenodini

The first observations on the biology of the North American species *Arrhenodes minutus* (Fig. 113) come from Riley (1874) and Packard (1881, 1890). Sanborne (1983) completed far more detailed observations on the biology and ethology of this species (see also the chapter on ethology). He studied a population of about 300 adult individuals on freshly cut red oaks near Ottawa in Canada. Adult *A. minutus* are usually found from May to September (Buchanan, 1960; Solomon, 1995). There are two periods of oviposition, from mid-June to late July and from early to mid-September (Sanborne, 1983). Females chew cylindrical holes into the sapwood and lay single eggs. Riley (1874) reports that the female takes about one day to prepare the hole in the wood prior to oviposition, whilst according to Buchanan (1960) and Sanborne (1983) it takes one or two hours to dig an egg hole. During this operation the male stands on guard. The eggs are round, less than 1 mm in diameter, initially translucent but gradually turning opaque. Their incubation requires from a few days to 3 weeks, depending on the temperature. The newly hatched larva (Fig. 57) bores directly into the wood. The tunnels are 0.2 to



Fig. 112 - Israel: Lower Galilee, Alonim Hills (photo by E. Orbach).

3 mm in diameter which at first are sufficiently wide for the larvae to move along them, but they are eventually widened. Full grown larvae are 12 to 24 mm long. The tunnels almost reach the opposite side of the tree, then make a sharp U-turn and go back across the wood grain toward the entrance. The larvae keep their tunnels clean by pushing the frass outside. Evidence for larvae consists of little piles of frass that accumulate around the egg holes. According to Kleine (1927c), *A. minutus* feeds on wood fungi. Pupation occurs near the gallery exit and the adults emerge through circular holes near the egg site. The life cycle is 2 to 3 years (Solomon, 1995), sometimes 4 (Buchanan, 1960).

Stebbing (1914) and Beeson (1925) report that pupae of *Prophthalmus tridentatus* were found in fresh, moist *Pterospermum acerifolium* Willd. sapwood in India. "The chambers were elongate and parallel to the long axis of the tree, and about half an inch [= 12.5 mm] deep in the sapwood."

Eremoxenini

Species of this tribe live with ants (see chapter on ethology), but their larval habits are unknown.



Fig. 113 - *Arrenodes minutus* (Drury) (U.S.A.) (photo by S. Marshall).

Cyphagogini

The first to carry out observations on the brentids of this tribe in the field was Lewis (1883) in Japan. He noticed that some species of Cyphagogini, e. g. *Eterozemus celtis* and *Cyphagogus signipes* entered Platypodidae galleries excavated in tree trunks, using their cylindrical body and exploiting their ability to put their pro-femora in the excavations of the fore part of the thorax when inside the tunnels, tucking their meso-femora into the space between the thorax and the elytra, and stretching out their hind legs behind them. Sharp (1936: 296) reports the observations Lewis made and concludes that these species of Cyphagogini "are predaceous and enter the burrows of wood-boring insects to search for larvae as prey".

Beeson (1925, 1941) carried out observations in India on the biology of *C. corporaali*, *C. cyrtotrachelus* [sub *C. buccatus*], *C. tabacicola* and *C. westwoodii*. All these species deposit their eggs in Platypodidae galleries. Beeson (1925) wrote that *Cyphagogus westwoodii* and *C. tabacicola* "were observed in the entrance tunnels of the *Crossotarsus* and of other platypodids, sometimes near the partially destroyed bodies of the brood-parents. As there was no other evident reason for the death of the platypodid, it was concluded that the brentids were predaceous". Beeson also reports that the larvae of *C. westwoodii* and *C. incisus* [sub *C. obconiceps*] both dig galleries grouped in clusters or whorls originating from the horizontal tunnels of *Crossotarsus saundersi* Chapuis. The larval galleries are curved and short, no longer than five times the length of the brentid, and terminate in pupal cells closed by plugs of sawdust and fine fibres.

According to Beeson (1925, 1941) *C. corporaali* galleries (Fig. 114) originate from the wall of the main horizontal *Platypus solidus* Walker gallery, from where they curve outwards and vertically upwards or downwards for a distance of up to 2.5 cm. The terminal portion forms a pupal chamber, where the insects lie with their heads directed towards the *Platypus* gallery, through which successful individuals make their escape. The prepupal portion of the gallery is packed with fine wood dust.

Kleine (1923g, 1931a, 1933f) claims that *Cyphagogus* are predators and can kill platypodids in order to take over their galleries. Haedo Rossi (1961), writing to report observations by Beeson (1925, 1941), categorically confirms that adult *Cyphagogus* kill Platypodidae females to take over their galleries and that the larvae of these brentids feed on Platypodidae larvae. Oberprieler & Louw (1985) as well as Damoiseau (1967a; 1989a) also hypothesise that adult *Cyphagogus* can prey on Scolytidae and Platypodidae.

According to Zimmerman (1994), *Cyphagogus* species "may extirpate the original owners of the tunnels, and then lay their eggs in the host tunnels from which their own larvae bore short tunnels outwards. The repeated reports in literature from other lands that these brentids are cannibalistic are, I believe, erroneous"... "More probably they only eject ambrosia beetles from their nests".

Browne (1961) wrote that "whether, as has frequently been suggested, *Cyphagogus* kills and devours the rightful occupants of the nest has not been proved, but it evidently at least ejects the guardian male from the entrance, and its presence, if only by its interference with the normal organisation of the nest, appears usually to result in the destruction of the ambrosia beetles and their brood. In Malaysia, these species are probably an important factor in control; in a large colony of ambrosia beetles, only a few nests are generally usurped, but the whole of the broods in these is destroyed."

Beeson (1941) also reports that *Calodromus mellyi* occurs in the galleries of Scolytidae in Assam and it is a "predaceous species like *Cyphagogus*".

Schedl (1961a) observed that *Allagogus brunneus* deposited its eggs in a 6 cm long larval gallery excavated by the Platypodidae *Doliopygus subditivus* Schedl in a large branch of *Tetrapleura tetraptera* (Thonn.) Taub. The galleries excavated by *Allagogus* larvae spread out radially with respect to the Platypodidae galleries and generally branch out in groups of four in an almost symmetrical pattern (Fig. 117). At each egg-laying site along the Platypodidae gallery were two to three groups of radial galleries. The brentid pupated in the larval gallery with its head directed towards the Platypodidae tunnel. The larva is xylomicetophagous.

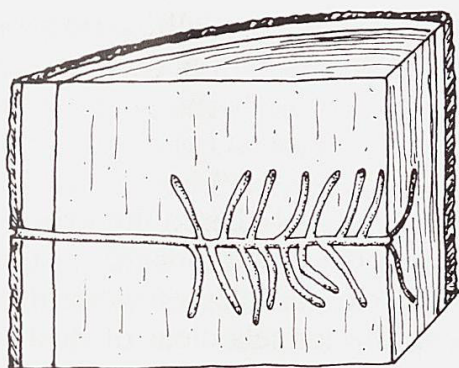
The African species *Nannobrenthus compressithorax*, *Cormopus distinctus* and *C. penicillifer* lay their eggs in a Platypodidae gallery at a depth of between one or two centimetres. The larval galleries branch out radially in all directions, starting from the host tunnel (Schedl, 1961a). Mayné & Donis (1962) report that in the Congo they observed an individual of *Cormopus filum* devouring a Scolytidae belonging to the genus *Doliopygus* inside its gallery.

Löyttyniemi & Löyttyniemi (1987) write that *Cormopus diversus* from Zambia "can fly throughout the year, the flight pattern being similar to that of their principal hosts, the platypodids. The long flight period and rearing results indicate that at least in some Calodromini [= Cyphagogini] species overlapping generations are produced annually and that development takes place during the dry and cool season as well as the hot, wet season. Also in this respect their biology is similar to that of the ambrosia beetles."

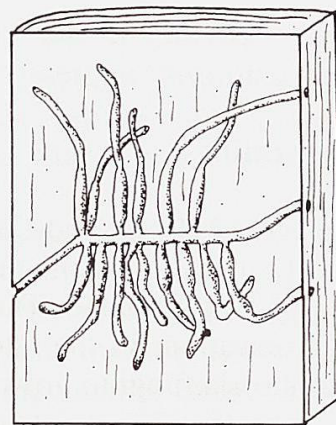
Schedl (1961a) observed larval galleries of *Oncodemerus sennai* [sub *O. costipennis*] in a *Doliopygus ghesquierei* Schedl tunnel, excavated in *Myrianthus arboreus* P. Beauv wood (Fig. 115). The larval galleries varied between 1.5 and 3.5 centimetres in length.

Kleine (1921f) reports examining the excrements of *Opysthenoxis ochraceus* and finding the remains of herbaceous plants, thus deducing that some species of Cyphagogini can be phytophagous.

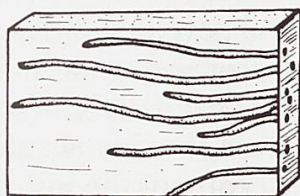
Stebbing (1914) reports finding individuals of a species belonging to the Cyphagogini [that he mistakenly indicated as *Zemioses* sp.] in India on a *Shorea robusta* Gaertn. f. trunk. These brentids dig circular tunnels through



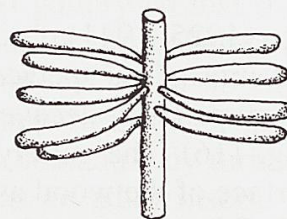
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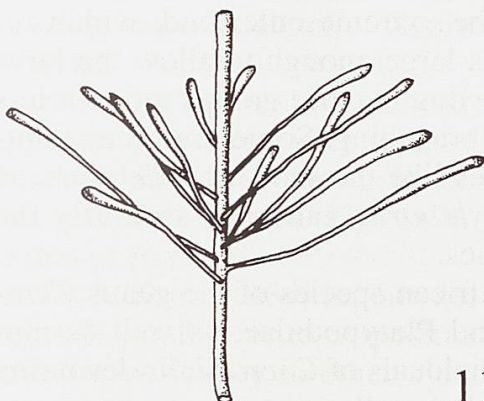
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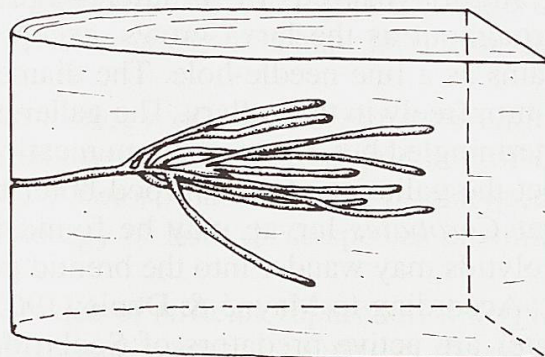
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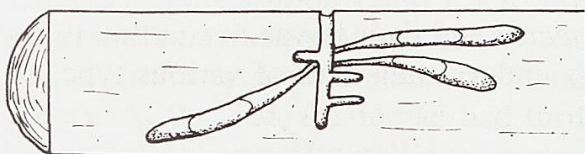
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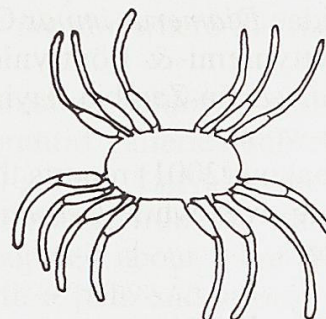
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Figs 114-121 - Larval tunnels of Brentidae. Fig. 114: *Cyphagogus corporaali* Kleine; Fig. 115: *Oncodemerus costipennis* Senna; Fig. 116: *Cerobates tristriatus* (Lund); Fig. 117: *Allagogus brunneus* Gahan; Fig. 118: *Bolbocranius csikii* (Bolkay); Fig. 119: *Bolbocranius bicolor* Senna; Fig. 120: *Taphroderes sahlbergi* (Sharp); Fig. 121: *Microtrachelizus beneficus* Kleine.

the bark where they lay their eggs. It would thus seem that not all Cyphagogiini are predators or "aggressors", and that some are "true wood-borers".

Hoplopisthiini

Beeson (1941) studied *Carcinopisthius oberthueri* in India: in this species the larval tunnels occur in clusters connected with the brood-tunnels of large species of Platypodidae. They produce very coarse particles of wood-dust during excavation; in the later stages they keep the tunnels clear of dust to allow ambrosia fungi to grow.

Stereodermini

Beeson (1925, 1941) observed egg laying and larval development in *Cerobates tristriatus*. This species lays eggs singly on the surface of the sapwood of trees with partially decayed bark; the larva bores a radial gallery in the soft wood (Fig. 116). The gallery increases in diameter from the fine needle-hole on the surface of the wood as it proceeds towards the centre of the log. It never exceeds 7.5 cm in length and its course on the horizontal plane is straight or slightly curved. In its later stages, the gallery is widened uniformly throughout as the larva grows, except at the extreme outer end, which remains as a fine needle-hole. The diameter is large enough to allow the larva to turn freely in the gallery. The galleries are dug in close groups more or less intermingled but are non-communicating or branching. Sometimes they intersect the galleries of other wood-borer beetles like the scolytid *Xyleborus*, so that *Cerobates* larvae may be found in *Xyleborus* galleries, similarly the scolytids may wander into the brentid galleries.

According to Mayné & Donis (1962), African species of the genus *Cerobates* are active predators of Scolytidae and Platypodidae, as well as mycetophagous. These authors report seeing individuals of *C. credibilis* devouring Scolytidae *Diamerus impar* Chap. larvae in their galleries.

Löyttyniemi & Löyttyniemi (1987) observed *Cerobates (Ionthocerus) usambaricus* in Zambia, laying eggs on bark and the larvae boring deep into the sap-wood.

Kabakov (2001) reports that *Ionthocerus* and their larvae live in thin twigs of young trees while *Cerobates* occurs under rotten bark of various types of dead trees.

Taphroderinae

According to Sharp (1895), South American Taphroderini "are probably all predaceous insects: their peculiar structure allows the legs to be packed close to the body when the insects enter a burrow in wood in search of its prey".

Kleine (1929b, 1938d) reports that *Taphroderopsis oscillator* and *Taphroderes nevermanni* live in galleries of the Scolytidae *Xyleborus confusus* Eichh. and underlines the similarities in the biology between Taphroderini and Cyphagogini. Damoiseau (1967a: 439) claims that Taphroderini prey on Scolytidae and Platypodidae larvae.

Schedl (1961a) observed the biology of *Bolbocranius csikii* [sub *Anisognathus distortus*] in the Congo. He reports that this brentid takes over the galleries of a large number of different species of Platypodidae and Scolytidae. The choice of host gallery depends on the size of the brentid. Small sized individuals tend to prefer galleries excavated by the smaller Scolytidae whilst larger individuals must of necessity find galleries that have been excavated by larger wood-borers. The female lays a series of eggs starting at about two centimetres deep, moving downwards in the host gallery so that the last egg laid is the furthest from the entrance hole. The larvae dig galleries that reach several centimetres in length and slope at an angle of about 45° away from the Platypodidae tunnel (Fig. 118). The larvae pupate in the galleries; there are no pupal cells and the insect pupates with its head already turned towards the Platypodidae gallery, where it will emerge as an adult insect. There are no traces of either the hosts or their larvae in galleries taken over by the brentids. Mayné & Donis (1962) observed *Bolbocranius csikii* in the Congo devouring large numbers of Scolytidae *Diamerus impar* Chap. larvae in their tunnels.

Schedl (1961a) also described the biology of *Bolbocranius mechowii*, apparently the exclusive guest of tunnels belonging to Platypodidae of the *Trachyostus* genus, and of *Bolbocranius bicolor*, that oviposits in platypodid and scolytid galleries (Fig. 119). The biology of both these species is similar to that of *B. csikii*.

De Muizon (1960) reports seeing females of African Taphroderini digging holes in wood prior to oviposition. This in part seems to contradict the use of galleries belonging to other xylophagous species or predation. Mayné & Donis (1962) also observed a pair of *Bolbocranius bicolor* where the male was intent in digging a gallery in wood.

In Brazil, Mecke *et al.* (2001) observed *Taphroderes sahlbergi* galleries in *Araucaria angustifolia* (Bertol.) Kuntze. The brentid galleries started from within a *Corthylus praealtus* Schedl (Scolytidae) gallery. Larval tunnels were 3.5-4.5 cm long and increased in diameter with distance from the start of the gallery. At the end, the larvae had turned and pupated about 2 cm from the *Corthylus* gallery. The galleries were filled with a pale and strongly compressed frass. The emerging imagines passed through the scolytid gallery to leave the trunk (Fig. 120).

Trachelizini

Zimmerman (1994) writes that he can offer nothing regarding the hosts or

habits of this tribe; most species, however, appear to be generalised borers of dead wood.

Senna (1895c) mentioned some individuals of *Hypomiolisa exarata* were collected in Western Java on flowers.

According to Beeson (1941) *Trachelizus bisulcatus* is a "wood borer making long cylindrical tunnels more or less horizontal (to the axis of the standing tree) and radial or concentric with the zones of growth; the tunnels contain loosely packed coarse wood-dust which is subsequently cleared out and the walls of the tunnels discolour or blacken indicating growth of mould or food-fungus. The life-cycle is annual [in India], beginning emergence in April and appearing irregularly until October. Beetles shelter for the cold season under loose bark".

Kabakov (2001) reports that in Vietnam *Trachelizus bisulcatus* "occurs in various types of forest and in cutting areas accumulating under the bark of dead trees of various species in groups of 10 to 12 beetles per sq. decimetre, often in combination with other species of Brentidae (*Cerobates*, *Higonius*)".

Ithystenini

Wallace (1874: 478) wrote that females of *Ithystenus angustatus* on the Aru islands bore deep holes in the bark of dead trees with their pointed rostrum and then deposit their eggs in these holes, which are as deep as the rostrum as far as the eyes.

Hudson (1934), Meads (1976) and May (1993) report several observations on the New Zealand species *Lasiornychus barbicornis*. The female brentid prepares her egg-laying site by chewing a cavity in the bark of various species of dying or dead trees (see Appendix 1) and freshly felled logs. The female bores a 0.5 mm wide hole through the bark by tilting her rostrum at about 45° to the body axis and twisting her head from side to side. After boring for about 0.3 mm, she extracts her rostrum and ejects the debris collected inside the rostrum and between the mandibles. The female continues this procedure until she has made a hole 3-4 mm deep, then she reverses her position and lays the egg in the hole (Meads, 1976). The larvae bore radially into the solid wood.

The diameter of the tunnel is enlarged as the larva grows, except in the short initial section which remains narrow. Ambrosia fungi grow in the larval tunnel. The gut contents of sub-mature larvae contained fungal material including coelomycete spores (*Seimatosporium*, *Monochaetia*) but no wood fragments (May, 1993). Pupation occurs in a cell along the tunnel axis near the exit, that has been widened but not lengthened through the original entry tunnel in the bark. In the pupae (Figs 63, 64) the head and rostrum are deflexed and they lie on the ventral surface of the thorax. It is left for the emergent adults, when hardened, to chew through the final section. Some of them per-

ish at this time with their rostra protruding, having apparently failed to make sufficient space to clear their rather wide shoulders through the tunnel (Broun, 1881; May, 1993). The pupation cell is only slightly larger than the pupa in circumference. Since the adults are always found in the pupal cell with their rostra normally pointing forwards, it can be assumed that the head and rostrum are straightened out immediately after emergence when the tissues are still soft, otherwise this operation would be impossible once the tissues become chitinised (Muir, 1913). The adults usually stay on tree trunks near the edge of the forest and on rare occasions quite a number may be found in close association.

Zimmerman (1994) mentions an individual of *Mesetia amoena* found in Australia on *Leptospermum* sp. blossoms.

Microtrachelizini

Beeson (1925, 1941) observed the reproductive biology of *Microtrachelizus beneficus* in India. This species exploits the prepupal tunnels of *Hoplocerambyx spinicornis* Newm. (Cerambycidae) for its oviposition sites (Fig. 121). The brentid enters the bark on the fallen tree by means of the ejection-hole of the Cerambycidae and while the mature larvae of *H. spinicornis* are hibernating, they penetrate the tunnel leading from the sapwood to the pupal chamber in the heart-wood. Individual, but abundant, eggs are laid on the tunnel wall. Once the larvae hatch, they excavate tunnels from 2.5 to 5 cm in length that curve outwards in a whorl from the longhorn beetle tunnel. Presumably they feed on sap and saprophytic fungi, as the larvae evidently move freely to-and-fro and the abandoned tunnels are found to contain mainly mildewed excrements with very little wood-dust. Pupation takes place in a slightly enlarged chamber at the base of the larval gallery; the beetle emerges by enlarging the hole of the larval gallery and escapes into the *Hoplocerambyx* tunnel.

Pseudoceocephalini

According to Wilson (1971) the Pseudoceocephalini, together with Cyphagogini and Taphroderini, are specialised cleptoparasites of ambrosia scolytoid nests, but literature holds no evidence of such behaviour in the Pseudoceocephalini.

De Muizon (1955b) observed the species *Pithoderes gestri* in Gabon laying eggs on fallen logs of Bilinga trees (*Sarcocephalus diderrichii* Wildem.). The female always takes up a position on the side of the trunk, standing vertically and holding her head upwards. She searches for bark-free areas and begins digging a hole with her rostrum, directed upwards at an angle of about 45°. Excavation lasts about 15 minutes and about every 30 seconds the female extracts her rostrum to eliminate any sawdust and clean it with her antennae.

The hole is as long as the female rostrum as far as the antennal insertion. The female never bores into areas protected with bark because her rostrum would not be long enough to reach the sapwood. Subsequently, the female turns round by 180° and lays a single egg in the hole, then with the tip of her abdomen she covers the hole with the accumulated sawdust so that it is invisible. The female then moves on and begins the whole process over again in another place. De Muizon also observed that *P. gestri*, apparently rare in virgin forests, is extremely common in areas under felling, since the female finds plenty of suitable sites for oviposition on many of the debarked trunks. De Muizon (1955b) reports seeing the same egg-laying technique in females of *Storeosomus riisii*, *Heterothesis elegans* and *Orphanobrenthus picipes*.

Löyttyniemi & Löyttyniemi (1987) observed *Storeosomus riisii* in Zambia laying eggs on bark and the larvae boring deep into the sapwood.

According to Kabakov (2001), *Hormocerus reticulatus* in Vietnam “inhabits various types of tropical forest, occurring mostly on fallen and burned trees. Aggregations of beetles occur under thick bark in rotten cambium with specific odour of beer in association with Passalidae, Carabidae and other Brentidae (*Baryrhynchus*, *Parorychodes*, *Orychodes*). Females of *H. reticulatus* were found crawling over the surface of the bark of *Ficus* stumps and putting their rostrum into the burrows of Platypodidae and Scolytidae, apparently to eat their larvae”.

Mimicry

Shelford (1902) quoted some Cerambycidae mimicking Brentidae in Borneo. Some species of the longicorn subfamily Hippopsinae mimic certain brentid species. *Alibora* sp. mimics *Baryrhynchus dehiscens* (Gyllenhal) in the colour of the elytra. Moreover, the three basal articles of the Cerambycidae are covered with long, close hairs. The insect holds the articles close together so that it imitates the shape of a Brentid rostrum. *Aegoprepis insignis* (Pascoe) imitates the brentid *Ceocephalus silvanus* in colour and the shape of the elytral tip. Shelford reports that “the mimic has the ground-colour of the prothorax and elytra black, and their dorsal surfaces are covered with tufts of a fine pale ochreous pubescence; these represent very well the scales of the brentid, and a very similar mottled appearance is thus produced in both species by totally different means”..... “Both model and mimic were taken on a fallen log close together”. *Ectatosia moorei* (Pascoe) mimics the female of *Ceocephalus* (Fig. 122) in the same manner as *Aegoprepis insignis*. Other Cerambycidae species mentioned by Shelford (1902) are *Dymascus porosus* (Pascoe) that mimics *Ceocephalus forcipatus* and *Stegenus dactylon* (Pascoe) (subfamily Agniinae) that mimics *Ceocephalus silvanus* in colour and shape of the elytra.



Fig. 122 - *Ceocephalus furcillatus* Guérin-Ménéville (Brentidae) (on left) and *Ectotasia moorei* (Pascoe) (Cerambycidae) (on right) (photo by P. Magrini).

Predation

Johnson (1983b) reports cases of predation of *B. anchorago* in Costa Rica by lizards of the genus *Anolis* and woodpeckers of the genus *Phloeoceastes*. Vitt (1993) mentions the occurrence of brentids in the diet of an Amazonian population of lizards of the genus *Tropidurus*. Vitt (2003, personal communication) ascertained the presence of brentids in the diet of other species of lizards: *Anolis fuscoauratus*, *Plica plica*, and *Tropidurus oreadicus*, all from the Amazonian rainforest, and *Anolis lemurinus* (Nicaraguan rainforest).

In response to predator attacks, *B. anchorago* defends itself by letting itself drop from the trunk and entering into thanatosis for a minute or two. Sanborne (1983) reports that *Arrenodes minutus* also draws in its legs when disturbed and falls to the floor, where it feigns death for a few moments.

Some species of *Ceocephalus* bear extremely long and slender caudal appendices. When the insect is threatened by a predator, it freezes holding its antennae out in front. In this way it is difficult to distinguish the caudal appendices from the antennae so the predator is probably tricked and cannot distinguish the true position of the head.

Sanborne (1983) observed ants attacking adult individuals of *Arrenodes minutus*: "brentids were feeding at sap flows and were attacked by numerous ants. The ants bit and pulled at the beetles but were unable to dislodge or injure them. The brentids responded by drawing in their legs and antennae but made no attempt to resist or flee. It was obvious that the ants' mandibles were no match for the heavily chitinised exoskeletons of the beetles" ... "It was likely that the ants were competing with the brentids for the food source and were not predators of these beetles."

Parasites

Brentids are often infested by mites. Appendix 3 lists the brentids on which mites have been found, with the names of their respective parasites (Vitzthum, 1921a, 1921b).

Schedl (1961a: Fig. 13) illustrates the case of individuals of *Bolbocranius csikii* in the Congo on which unidentified mites had settled on the internal side of the large left mandible of the males and between the first articles of the female antennae.

B. anchorago is often attacked by huge numbers of mites. Johnson (1983b) reports that these brentids tried to scrape the mites off with their forelegs, but any mites dislodged by this method found little difficulty in clambering back onto their hosts.

Zimmerman (1994) observed fungi of the Laboulbeniaceae family growing on an unidentified brentid from Lord Howe Island (Australia). According to Prof. W. Rossi (University of L'Aquila, Italy), a specialist in Laboulbeniaceae, there is no other mention of this type of fungus on Brentidae.

ETHOLOGY

Not many studies exist on Brentid ethology. Wallace (1874) commented on the behaviour of *Ithystenus angustatus* on the Aru islands (Indonesia). Observations on the behaviour of the North American species *Arrenodes minutus* have been carried out by Riley (1874), Leconte & Horn (1876, 1883), Blatchley & Leng (1916), Buchanan (1960), and Sanborne (1983). Meads (1976) and Hamilton (1979) observed the endemic New Zealand species *Lasiorrhynchus barbicornis*.

Schedl (1960a) studied the biology and behaviour of some species of brentids in the Congo.

Concerning the Central and South American species, Johnson (1983a) and Howden (1995) studied the reproductive behaviour of *Claeoderes radulirostris mexicanus* [sub *C. bivittatus* and sub *C. guatemalensis* respectively] in Panama and Guatemala. Johnson (1982, 1983b) and Johnson & Hubbell (1984) investigated that of *Brentus anchorago* in Costa Rica and Panama, and Garcia (1989) that of *Estenorhinus* sp. *prope dispar* in Costa Rica.

Grandi (1951), Tassi (1965), Torossian (1965, 1966), Le Masne & Torossian (1965), Ponel (1981) and García-París & Olmos (1992) have reported their observations on *Amorphocephala coronata* in Italy, Spain, and France. The behaviour of this species is described in detail in the paragraph on myrmecophily.

Here we give a summary of the behaviours observed in the above mentioned brentid species; further information on reproductive behaviour can be found in the chapter on their biology.

Wallace (1874: 478), writing on the behaviour of *Ithystenus angustatus* on the Aru islands, comments: "Those curious little beetles, the Brentidae, were very abundant in Aru. The females have a pointed rostrum, with which they bore deep holes in the bark of dead trees, often burying the rostrum up to the eyes, and in these holes deposit their eggs." ... "I once saw two males fighting together; each had a fore-leg laid across the neck of the other, and the rostrum bent quite in an attitude of defiance, and looking most ridiculous. Another time, two were fighting for a female, who stood close by busy at her boring. They pushed at each other with their rostra, and clawed and thumped, apparently in the greatest rage, although their coats of mail must have saved both from injury. The small one, however, soon ran away, acknowledging himself vanquished. In most Coleoptera the female is larger than the male, and it is therefore interesting, as bearing on the question of sexual selection, that in this case, as in the stag-beetles where the males fight together, they should be not only better armed, but also much larger than the females." Darwin also

mentions this behaviour in his book "The descent of man and selection in relation to sex" (1893).

The males of *Arrenodes minutus* have a wide rostrum and robust mandibles (Price, 1975). They are highly aggressive towards each other and dominance is based on size and strength (Riley, 1874; Leconte & Horn, 1876, 1883; Blatchley & Leng, 1916). Aggression takes the form of stereotyped combats, with no evidence of injury to the opponents. The females are not known to fight. Usually, large males grab smaller males with their mandibles and throw them off the trunk (Sanborne, 1983), otherwise the vanquished male will fly away (Leconte & Horn, 1876, 1883). Sanborne (1983) also reported courtship and mating behaviour: "There is no obvious courtship in this species. Males determined the gender of another beetle by passing their rapidly fluttering antennae up and down the other beetle's head and body. Upon recognizing females, males became noticeably more excited, and their antennae fluttered even more rapidly. Attempts to mount the female usually followed recognition." ... "Successful males forced the female's head up, causing her to stand still, and copulation followed. The chase often had to be repeated several times before the female stopped resisting. If a female were completely uncooperative, the male eventually gave up and wandered away." ... "Successful copulation occurred only between individuals of similar size. If the size difference was too great, copulation was physically impossible. However, size was not a factor in whether or not an attempt to copulate was made." Sanborne (1983) observed a female of *A. minutus* laying a single egg in a hole, then the female "... plugged the hole by pressing the tip of the abdomen on the wood surface and drawing it back and forth across the opening. The plug seemed to be a mixture of wood frass and a secreted sticky substance". However, according to Buchanan (1960) "no protective cover or fluid was placed over the egg".

Johnson (1983a) and Howden (1995) observed ovipositing females of *Claeoderes radulirostris mexicanus*. Both confirmed the observations of Sanborne (1983): the distal margin of tergite 8 is similarly dentate in the females of *A. minutus* (Fig. 50) and *C. radulirostris mexicanus*. The tip of the abdomen is used to scrape the surface of the log in a similar manner to that described for *A. minutus*. After oviposition, the female *C. radulirostris mexicanus* rocks by bending and straightening her forelegs, repeatedly moving the tip of the abdomen back and forth from the hole to positions further back. As the female rocks out, the dentate tergite 8 is everted and used as a rake, to which bits of sawdust and other debris adhere. As she rocks in, the material is added to the hole.

According to Riley (1874) and Holland (1883), the male of *A. minutus* occasionally helps the female in extracting her rostrum. Riley (1874) describes the process as follows: "... this he does by stationing himself at a right angle with her body, and by pressing his heavy pro-sternum against the tip of her abdomen; her stout forelegs serving as a fulcrum, and long body as a lever."

Sanborne (1983) does not report this behaviour but writes that: "When a male came upon an unpaired female drilling an egg hole, he initiated the usual recognition procedure and positioned himself at right angles to her, with his head resting on her thorax or elytra".

Buchanan (1960) reported that "females disturbed while digging holes did not return to finish them". This observation contradicts Sanborne (1983). On six occasions this author observed females, previously disturbed whilst drilling, successfully relocate their unfinished egg holes up to one hour later. "They systematically walked up and down and around the general area where the hole was located with their rostra pressed lightly on the substrate. Once relocated, the hole was completed. It was probably a combination of visual and olfactory clues which led them back to the holes."

When a male on guard is disturbed by a male intruder, a fight may break out between similarly sized individuals. If they differ considerably in size, the smaller individual leaves (Sanborne, 1983). This author also reports two cases of territorial defence by *A. minutus* males. A 'territory' considered worth defending is the wide circle that the male defines round the female by walking in a circular fashion during guarding behaviour. Defending a good hiding place against intruding males is another form of territorial behaviour, although one or more female can be accepted inside the retreats. Sanborne (1983) also describes grooming movements in *A. minutus* in detail.

Meads (1976) reported that in *Lasiorrhynchus barbicornis* "fighting occurs frequently when an extra male or males finds a copulating pair. An attacking male first uses his downwardly curved mandibles and long rostrum with a raking action across the back of the copulating male in order to dislodge him. If this is not successful both contestants endeavour, rather awkwardly, to secure any part of their opponent's leg with their mandibles. If this difficult manoeuvre is accomplished and a firm hold is achieved, the defeated male immediately becomes submissive and is lifted bodily off the tree and dropped. Some beetles have legs missing (mainly tibia or tarsi) and this may be attributed to such fights. Smaller males tend to retreat quickly when challenged and do not fight. The defeated males spend long periods hiding in crevices, holes or under loose bark." ... "The function of the long rostrum of the male is not so obvious. The mandibles are large and well developed, though they are not used in any special way for feeding. The fighting behaviour of males shows one way in which the rostrum is used to good effect. Small males, with correspondingly short rostrums, are unable to compete successfully with the larger males for females, but occasionally mate when larger males are distracted by fighting". Hamilton (1979) also contributes some remarks on the behaviour of *L. barbicornis*. May (1993) reports that the "oviposition sites of *L. barbicornis* are prepared by the female weevil chewing in the bark of dying or suppressed trees and in freshly felled logs". Mating male and female *L. barbicornis* are illustrated in Fig. 123.

Schedl (1961a) reports some observations on the reproductive behaviour of *Allagopus brunneus* in the Congo. When the female of the species finds a Platypodidae gallery, she enters just far enough for the tip of her abdomen to remain outside. The male mates with her by keeping the hind end of his body lifted and bent at an acute angle to the fore end, in this way his head leans against the trunk (Fig. 124). Scivittaro (1975), observing the mating in the Brazilian species *Taphroderopsis curvidens*, also notes that the male mated with the female whilst the latter was partially inside a tunnel belonging to an xylophagous species, and that only her abdomen was protruding.

In the Congo, Schedl (1961a) also noted adult individuals of *Bolbocranius csikii* rummaging around by day on the trunks of fallen trees searching for suitable tunnels. The males of the species used their left mandible, asymmetrically far more developed than the right (Fig. 73), to force the Platypodidae or Scolytidae out of their tunnels. Schedl also observed a *B. csikii* male wandering around a series of holes of galleries made by a platypodid of the genus *Trachyostus*, and then entering one of them head first. After a few moments the male exited backwards, followed by a female of the same species walking in the same fashion. The female immediately entered the hole again, then the male turned round 180° and followed her, walking backwards. According to Schedl, this behaviour could be a courtship ritual preceding copulation.

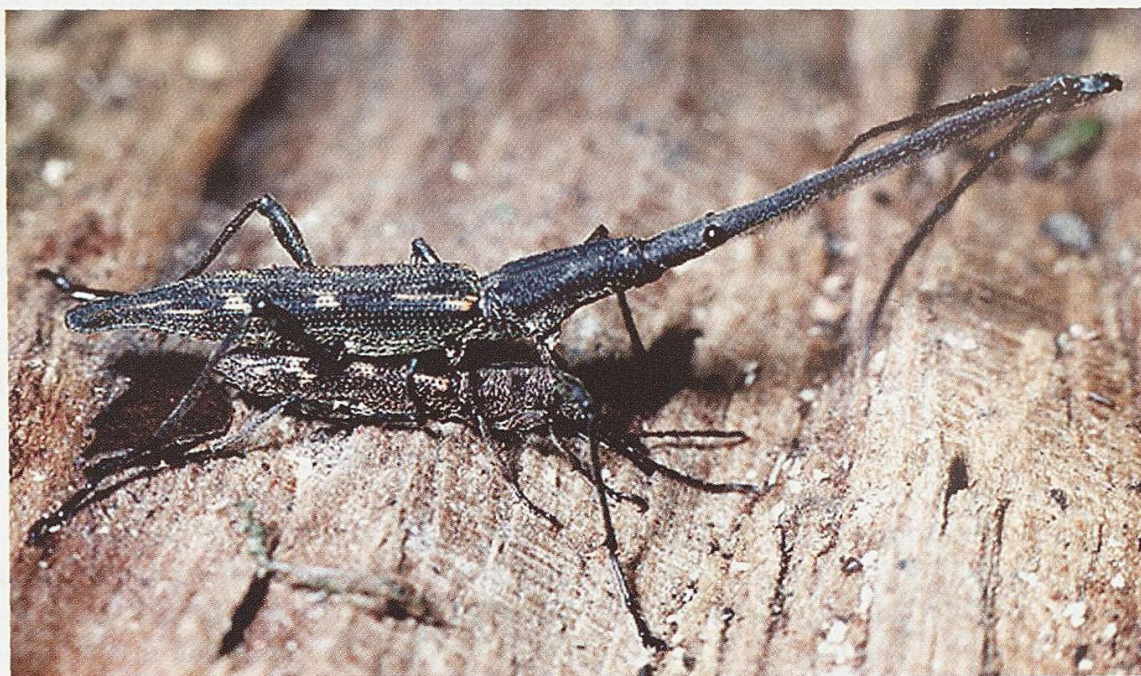


Fig. 123 - *Lasiorrhynchus barbicornis* (F.) (New Zealand), mating male and female (photo by M. Meads).

Johnson (1983a) studied the reproductive behaviour of *Claeoderes radulirostris mexicanus* [sub *C. bivittata*] in Panama. She observed male and female aggressive behaviour, guarding and mating. "Males fight to get access to females. Initially they may intermingle, jerk, or lash their antennae. Attacks involve nipping, kicking, poking with the snout, or putting the snout under the rival's body and jerking upwards three times/second. A male may also interfere with copulation by thrusting his snout between a mating pair and pushing. An attacked male may flee, or reciprocate in kind. Two males of approximately the same length may engage in a more stylized contest in which they align themselves, side by side, 1-10 mm apart, facing opposite directions. On the side of the rival a male taps his antenna and hind leg 4-5 times/sec, and when the opponent does likewise, the males fence legs against antenna at either end." ... "Such an appendage-lashing contest may permit rivals to assess one another's relative size".

Female fights usually happen between females prior to drilling or just after oviposition, when the egg-layer may attack another female drilling her hole. "The encounter may involve only an intention movement, or the instigator may push, poke, or swat a drilling female with her snout, or pry her out of her hole by sticking the snout under her abdomen and lifting." ... "Reciprocated aggression may result in a fight lasting 6 minutes or more in which the combatants kick, face one another and swivel their heads and forebodies, or thrust

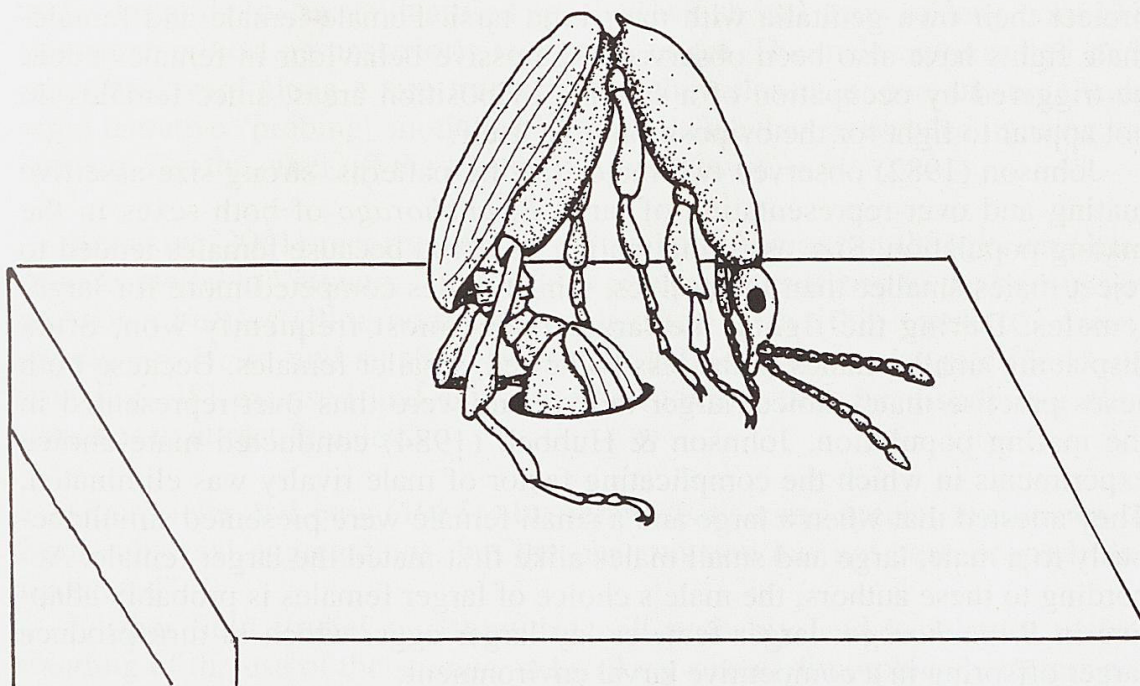


Fig. 124 - *Allagopus brunneus* Gahan, mating male and female (from Schedl, 1961a, modified).

the snout under the other and lift suddenly; females of the same length may also stack themselves head-to-tail and sweep their snouts over the tip of their opponent's abdomen. Fights end when a female leaves or is flipped from the tree" (Johnson, 1983a).

Male guarding behaviour consists in keeping his rostrum or body over a drilling or egg-laying female. If a rival male or another female approaches, he responds aggressively. The male may mate with the female several times during the drilling process if he is on guard. Sometimes a female may be guarded by two males, if one is large and the other small. Copulation lasts about a minute. Oviposition usually occurs less than a minute after copulation is complete.

Johnson (1982, 1983b) mentions that the adults of *Brentus anchorago* live under loose bark and in holes made in the sapwood of fallen branches and logs of various trees (see Appendix 1). Mating and oviposition take place by day on the exposed surface, provided the wood is not wet with rain or heated by direct rays of the noon sun. The male wanders about on the surface of the log. When he finds a drilling female, he lays his rostrum across her and guards her, copulating for about one minute at intervals, until the oviposition hole is ready and the female begins laying. Fights between males for mating with the female have been observed. The males employ their rostrum as a lever to throw the rival off the trunk. If the size difference between rival males is considerable, the larger individual always gets the better in the fight. When two similarly-sized males fight, they stack themselves head-to-tail and nip at one another's genitalia with their tiny mandibles, at the same time trying to protect their own genitalia with their hind tarsi. Female-female and female-male fights have also been observed. Aggressive behaviour in females could be triggered by occupation of a suitable oviposition areas, since females do not appear to fight for the oviposition hole itself.

Johnson (1982) observed two major mating patterns: strong size-assertive mating and over-representation of large *B. anchorago* of both sexes in the mating population. Size assorting mating occurred because females tended to reject males smaller than themselves, whilst males competed more for larger females. During the fights, the larger males most frequently won, often displacing smaller males to the less contested, smaller females. Because both sexes practise mate choice, larger individuals were thus over-represented in the mating population. Johnson & Hubbell (1984) conducted mate-choice experiments in which the complicating factor of male rivalry was eliminated. They attested that when a large and a small female were presented simultaneously to a male, large and small males alike first mated the larger female. According to these authors, the male's choice of larger females is probably adaptive in *B. anchorago*: larger females lay larger eggs, which in turn produce larger offspring in a competitive larval environment.

The main differences between the behaviours of *B. anchorago* and *C. radulirostris mexicanus* are that in *B. anchorago*, the bigger the male, the

more he mates (Johnson, 1982), whilst in *C. radulirostris mexicanus* mating males are significantly more likely to be either large or small, the middle-sized males mating the least (Johnson, 1983a). The disproportionate mating success of small males may be explained in part by the tendency of smaller individuals to wait partially hidden under a drilling female, emerging for copulation when the larger males are not guarding the female.

Garcia (1989) observed courtship and copulation behaviour in *Estenorhinus* (quoted as *Arrhenodes*) sp. *prope dispar* in Costa Rica. "When the male encountered the female he began to tap her body gently with his antennae. He gradually shifted position until he reached her antennae and then the drumming became faster and softer. While continuing to tap her antennae, the male climbed onto the female and with his mandibles he grasped her at the suture just behind her head. When the female moved, the male accelerated the soft drumming on her antennae. Since the male was shorter than the female, in order to copulate he had to open his mandibles (thereby releasing her) and move slightly backwards." ... "Before copulation, the male rubbed the female's copulatory opening with his aedeagus from side to side like a pendulum. After copulation, which lasted at least 15 minutes, the male remained on top of the female, rubbing her in the same way for several more minutes."

Eberhard (1979: 254) quotes a personal communication from E. Sleeper, who "observed what appeared to be an aggressive interaction between two males brentids [of an unidentified species] which had pairs of long horn-like structures projecting posteriorly from their elytra. The males approached each other backward along a tree trunk, and as they closed on one another, made rapid tentative "probing" motions backwards. One then succeeded in using his horns to flip the other off the tree with a quick twist."

Kabakov (2001) reports a rather particular behaviour in the species *Calodromus mellyi* in Vietnam: "...the males occupy the entrance to the burrows of ambrosia beetles (Platypodidae), most often in the thin trunks of *Liquidambar* or *Ficus*, and sit there with their extraordinarily elongate hind tarsi [see Fig. 38] exposed and scissors-like moving, thereby dispensing pheromones to attract females".

Oberprieler & Louw (1985) also report that "some species are known to form subsocial aggregations, but the phenomenon has not been properly investigated".

Further field studies on Brentidae will probably lead to a better understanding of the use of the strange and striking sexual dimorphic characters occurring in this family.

Myrmecophily

All references to myrmecophily in the family refer to species distributed in the Palearctic, Ethiopian, Oriental and Australian Regions; there are no known cases for the Nearctic and Neotropical realms.

Kleine (1921i; 1924d) wrote two works on myrmecophily in the brentids. In one (1924d) he listed all the myrmecophilous genera of Brentidae and examined in detail all their morphological modifications. The genera considered belong to the Eremoxenini tribe, even if in the previous paper (1921i) Kleine also included the species *Orfilaia vulsellata* (Arrhenodini) and *Rhinopteryx foveipennis* as well as *Orphanobrentus laevicollis* (Pseudocecephalini) among the myrmecophiles. Two years later Kleine (1923g) wrote that these three species were not true myrmecophiles, but had been found on occasions close to ant nests. However, Schedl (1975a) reports the occurrence of *Orfilaia reichei* in a nest of *Camponotus compressus* ssp. *fellah* Dalla Torre in Israel. It has therefore to be ascertained if some species of *Orfilaia* are true myrmecophiles.

The main morphological modifications in the Eremoxenini are evident at the level of the rostrum, head and antennae, with the occurrence of hollows, trichobothria, glands, grooves etc. (see the chapter on morphology).



Fig. 125 - *Amorphacephala coronata* (Germar) (Italy, Tuscany Region) (photo by S. Bambi).

According to Schedl (1961a) all the Eremoxenini are exclusively myrmecophiles, but Löyttyniemi & Löyttyniemi (1987) throw some doubt on this affirmation, since in Zambia they observed *Symmorphocerus frontalis* "...reared from logs".

Appendix 4 gives those Brentids whose respective species of ant hosts are reported in literature. The ants belong to four subfamilies: Dolichoderinae, Dorylinae, Formicinae and Myrmicinae. The Brentid that has been studied most is undoubtedly the Palaearctic species *Amorphocephala coronata*; it has been found together with ants of the genera *Camponotus* Mayr, *Formica* L., *Crematogaster* Lund and *Pheidole* Westwood. The citation of the genus *Lasius* F. (C. Azam, 1884; J. Azam, 1893) has been doubted by some authors (Kleine, 1921i; Schedl, 1970), who retain the genus should be *Camponotus*.

Schedl (1970; 1975a) has investigated the circum-Mediterranean species, whilst Zimmerman (1994) treated the Australian species; his paper also includes observations by Lea (1898; 1905; 1910b; 1912) and Walker (1905). Lea (1912: 77) wrote: "In October 1910, in examining some nests of *Iridomyrmex nitidus* Mayr at Glenfield (New South Wales), I saw thousands of specimens of this beetle [*Cordus hospes*]. In parts of the nest they were clustered so thickly together that from a space, a square inch in extent, several dozens could have been taken. The ants moved freely about them without in any way interfering with them." Zimmerman (1994) wrote that *Cordus*, *Ankleineella* and *Kleinella* species in Australia are associate with ants, but the nature of the association and their biology remain unknown.

Some observations on the ethology of *Amorphocephala coronata* (Fig. 125) were carried out by C. Azam (1884) and Ponel (1981) in France, by Grandi (1951) and Tassi (1965) in Italy and by Kleine (1916g) and García-París & Olmos (1992) in Spain. In depth studies on the behaviour of this species with ants of the genus *Camponotus* have been performed by Torossian (1965, 1966) and Le Masne & Torossian (1965). C. Azam (1884) found *A. coronata* to be very common in the Var region of France, living in nests of *Lasius niger* (L.) (see previous observations on the correct identification of the species) under stones in the woods. The Author reports he collected as many as 500 specimens in just one hour and never found isolated individuals under the stones but always two to six individuals together. Azam also observed that the ants busied themselves to recuperate any Brentids that had wandered away from the nest and brought them back, both when the beetles were intent on leaving after they had been disturbed and when the Author displaced them himself nearby. Tassi (1965) also reports an ant belonging to the *Camponotus* genus carrying a live individual of *A. coronata* in its mandibles. He mentioned that he collected approximately 200 specimens of *A. coronata* in just one *Camponotus* nest, near Pomezia (Latium, Central Italy).

Torossian (1965; 1966) and Le Masne & Torossian (1965) observed how

individuals of *Amorphocephala* introduced into a colony of ants of the genus *Camponotus* were first attacked by the workers and reacted by keeping absolutely still. During this phase, any ants that came into contact with the pubescence located in the cephalic hollow of *Amorphocephala* stopped their attack and began to lick the secretion from the cephalic pubescence. The hostility the ants show towards the brentids can last for several days, after which the beetle is accepted by the colony and begins food exchange by mouth with the ants. Wilson (1971) and Hölldobler & Wilson (1990) have reported the results of these studies, emphasising some examples from *A. coronata* that "...receive food from some of the host workers and regurgitate part of it back to other host workers". This is the first reported example that could be construed as altruistic behaviour on the part of the symbiont beetles. Quantitative investigations are needed, however, to see how much of the received food the beetles share with the other host ants. It could very well be that *Amorphocephala* return only very small portions of the food to the colony and this "pseudoaltruistic" behaviour is one other mechanism used by symbionts to become fully integrated into the host colony. Torossian (1966) reports various experiences of adoption of *A. coronata* by ants which do not belong to the genus *Camponotus*, the usual host of this myrmecophile. He experimented with ants of the genera *Formica*, *Tapinoma* Foerster and *Myrmica* Latreille. After introduction into the nest, the brentid was attacked by the ants, but after several hours it was completely ignored. It gained no benefit from any attention such as licking, neither did the ants provide it with food; it was merely tolerated as a guest. Torossian (1966: 47) concludes by hypothesising that these beetles can enter many ant nests of French fauna at no risk and this probably allows the beetle to search for the colony of its usual host without running into danger.

Lea (1898; 1905; 1910b) reported that *Cordus hospes* was "taken in the nests of some termites and native bees" but Zimmerman (1994: 106) writes that "I do not have irrefutable evidence that the species occurs in association with termites and bees." De Muizon (1960) reported that *Afrocordus asper* was found with termites under barks. Kistner (1969) states that some species of Brentidae are termitophiles; Löyttyniemi & Löyttyniemi (1987) write that "some brentids in miombo woodland may also be termitophiles ... though the evidence of this relationship is insufficient". Kabakov (2001) reports a male of *Paussobrenthus bakeri* in nest of the bee *Apis cerana* F. in North Vietnam.

GEOGRAPHICAL DISTRIBUTION

Brentidae are mainly tropical insects. According to Damoiseau (1967a) and Osella *et al.* (1998), only 25 species occur north of 30° latitude; actually there are more than 40 (3 species in the Nearctic Region, 39 in the Palaearctic Region).

Kleine (1922c; 1926a; 1938d) considered the geographical distribution of the family in great detail. In the many papers he has dedicated to various genera, Kleine has always discussed their geographical distribution, similarly Damoiseau (1989a) has examined the problems regarding the distribution of species pertaining to the genus *Cyphagogus*. Damoiseau (1967a) and Osella *et al.* (1998) examined the distribution of the African species, whilst Zimmerman (1994) discussed the distribution of the species in the Australian Region.

The distribution of brentids throughout the different Zoogeographical Regions is shown in Figs 126-131 and summed up in Fig. 132 and Table 1. The Oriental Region has the highest number of species and genera (646 e 120 respectively); a high number of species also occurs in the Neotropical and Ethiopian Regions. The poorest region as far as Brentid taxa are concerned is the Nearctic, with a mere 7 species of 6 different genera, mostly localised in the South. The Palaearctic Region (including Taiwan) is remarkably richer than North America, with 85 species belonging to 31 genera.

Table 2 gives the species and genera of the various subfamilies and tribes divided according to Zoogeographical Regions. It is clear that the tribe Brentini is almost exclusively neotropical, with two species (*Brentus anchorago* and *B. mexicanus*) reaching the southern part of the Nearctic Region (California and Florida) and one (*Brentus cylindrus*) very probably introduced into Polynesia. The Arrhenodini is a tribe rich in genera and species distributed in

Tab. 1 - Number of genera and species of Brentidae present in each Zoogeographical Region.

Zoogeographical Regions	genera	species
Palaearctic	31	85
Ethiopian	109	363
Oriental	120	646
Nearctic	6	7
Neotropical	57	419
Australian	65	276

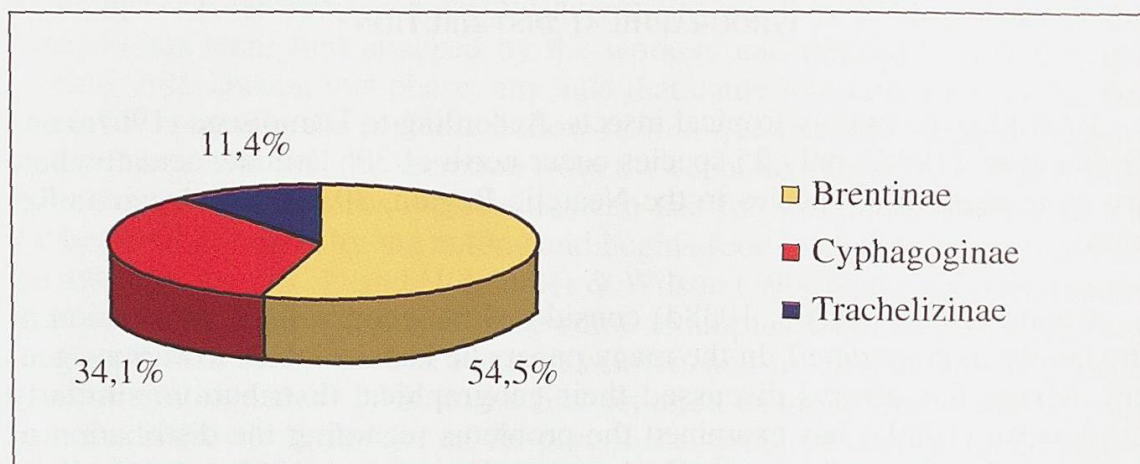


Fig. 126 - Percentages of the brentid subfamilies in the Palaearctic Region.

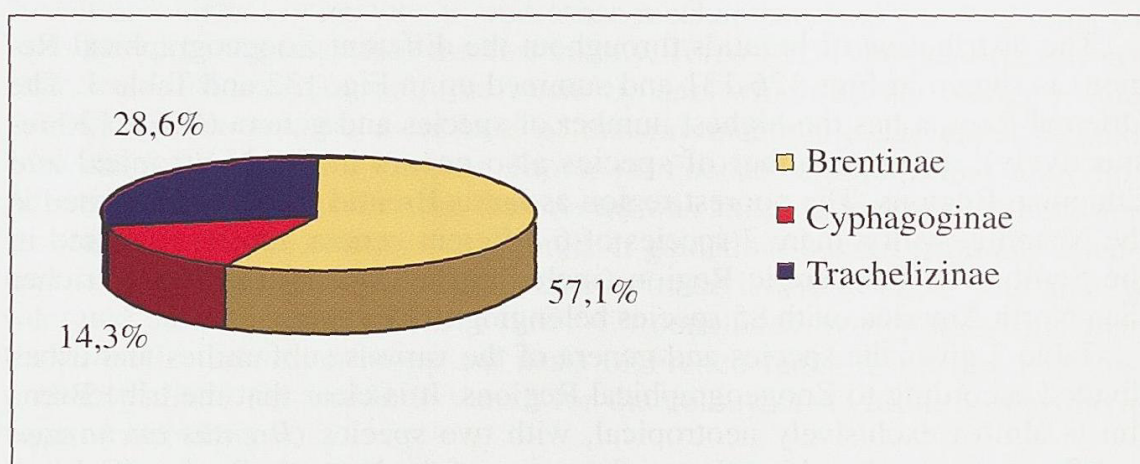


Fig. 127 - Percentages of the brentid subfamilies in the Nearctic Region.

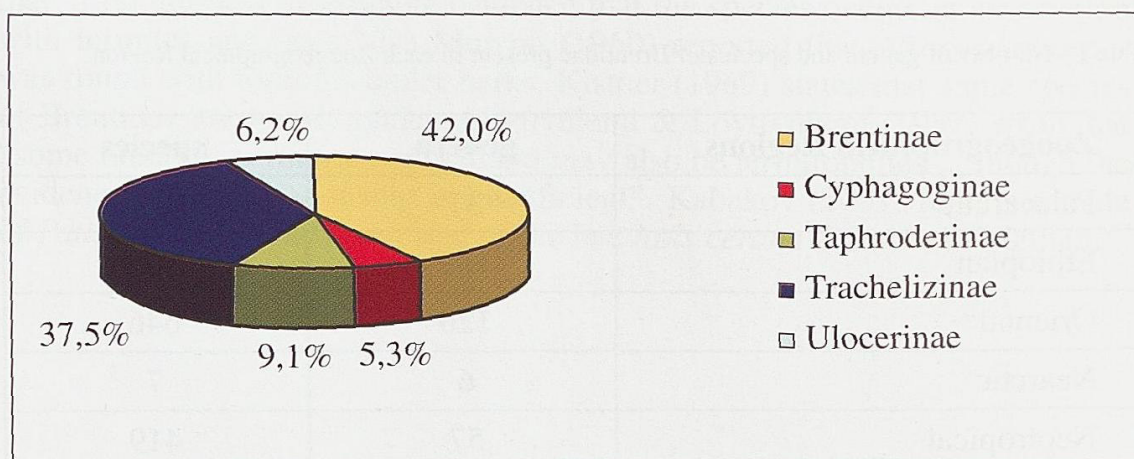


Fig. 128 - Percentages of the brentid subfamilies in the Neotropical Region.

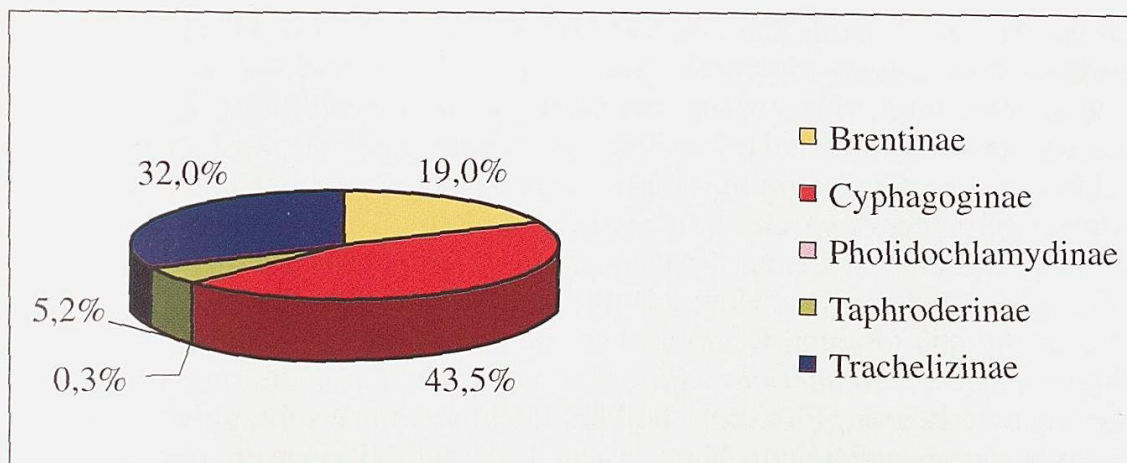


Fig. 129 - Percentages of the brentid subfamilies in the Ethiopian Region.

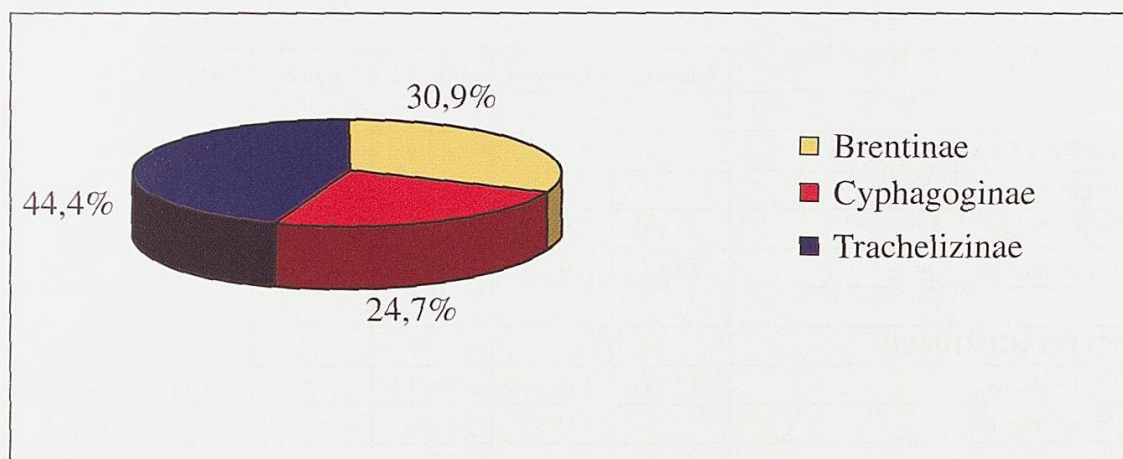


Fig. 130 - Percentages of the brentid subfamilies in the Oriental Region.

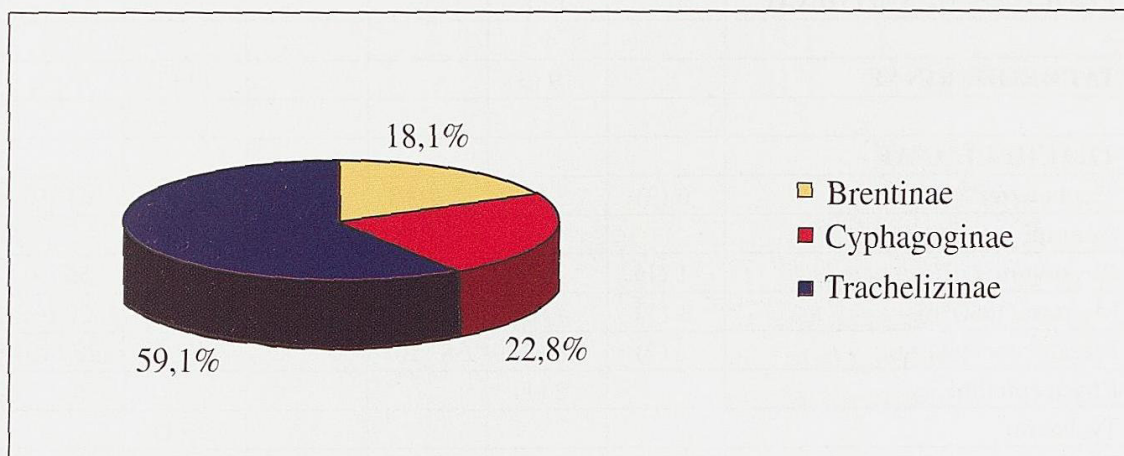


Fig. 131 - Percentages of the brentid subfamilies in the Australian Region.

all the Regions, mainly the Oriental and Neotropical. This tribe includes the northernmost species (*Arrenodes minutus*) that has reached as far as Canada. The Eremoxenini tribe, whose members are myrmecophilous, occurs in all the Regions except for the Nearctic and Neotropical. In the Cyphagoginae subfamily, the Cyphagogini tribe is distributed throughout the Old World, with a contingent of species in the Australian Region and a single species, *Sebasius brasiliensis*, in the Neotropics. The Atopobrentini are exclusively African and Oriental; the Hoplopisthiini are a small tribe that lives in the east Palaearctic and Oriental Regions. The Stereodermini are pantropical, with a single species, *Stereodermus exilis*, that reaches as far as the southern part of the Nearctic Region (Florida). The Pholidochlamydinae subfamily consists of a single species endemic to Madagascar. The Taphroderinae are only found in

Tab. 2 - Number of genera and species (in brackets) of the Brentidae tribes present in each Zoogeographical Region.

	Palearctic	Ethiopian	Oriental	Nearctic	Neotropical	Australian
BRENTINAE						
Brentini				2 (1)	38 (2)	1 (1)
Arrhenodini	35 (9)	20 (3)	172 (40)	2 (2)	137 (25)	20 (10)
Eremoxenini	13 (5)	49 (10)	28 (8)			31 (4)
CYPHAGOGINAE						
Cyphagogini	17 (7)	118 (44)	109 (23)		1 (1)	44 (13)
Atopobrentini		15 (2)	1 (1)			
Hoplopisthiini	2 (2)		5 (3)			7 (1)
Stereodermini		25 (3)	45 (3)	1 (1)	23 (5)	12 (2)
PHOLIDOCHLAMYDINAE		1 (1)				
TAPHRODERINAE		19 (3)			43 (5)	
TRACHELIZINAE						
Trachelizini	6 (3)	2 (1)	154 (13)	2 (2)	41 (4)	61 (9)
Acratini					115 (12)	
Ithystenini	1 (1)	7 (3)	46 (6)		3 (2)	55 (9)
Microtrachelizini	1 (1)	7 (2)	49 (7)			20 (6)
Pseudocoecephalini	2 (2)	98 (35)	38 (10)			27 (10)
Rhyticephalini		2 (1)				
Tychaeini					1 (1)	
ULOCERINAE					26 (1)	

Africa and the Neotropical Region. The Trachelizinae subfamily comprises seven tribes. The Trachelizini tribe occurs in all the Zoogeographical Regions, mainly in the Oriental. The Acratini and Tychaeini occur exclusively in Central and South America. The Ithystenini tribe is widespread, except in the Nearctic Region and with only one species in the Palaearctic Region. The Microtrachelizini and Pseudoceocephalini are not found in the New World, but they do occur in the other Regions, including the Australian one. The Rhyticephalini are a small tribe endemic to Madagascar. Finally, the Ulocerinae subfamily is exclusive to the Neotropical Region.

Cases of accidental importation of Brentid species in several parts of the world are not unusual. Senna (1893e) described new species that have arrived in Europe from the East with dried tobacco; Bartolozzi (1988a) cites several species of imported African Brentids in the port of Venice that arrived with exotic timber; Zimmerman (1994: 152) quotes "a specimen of the widely distributed *Opisthenoxys ochraceus* ... found in Indonesian timber in quarantine in Adelaide". According to Zimmerman (1994: 19) the three brentid species established in Tasmania are introduced species probably carried by man. Zimmerman (1994: 18) also mentions the case of the species on the Marquesas Islands that Kleine described as *Brentus efferatus* but is actually the South American species *Brentus cylindrus*, introduced accidentally by man to these isolated islands.

We can signal the case of the Nearctic species *Arrenodes minutus*, a species that damages timber and for this reason has been included in the list of harmful species by the European Union under Council Directive 2000/29/EC of 8th May, 2000 for "Protective measures against the introduction into the Community of organisms harmful to plants or plant products and against their spread within the Community".

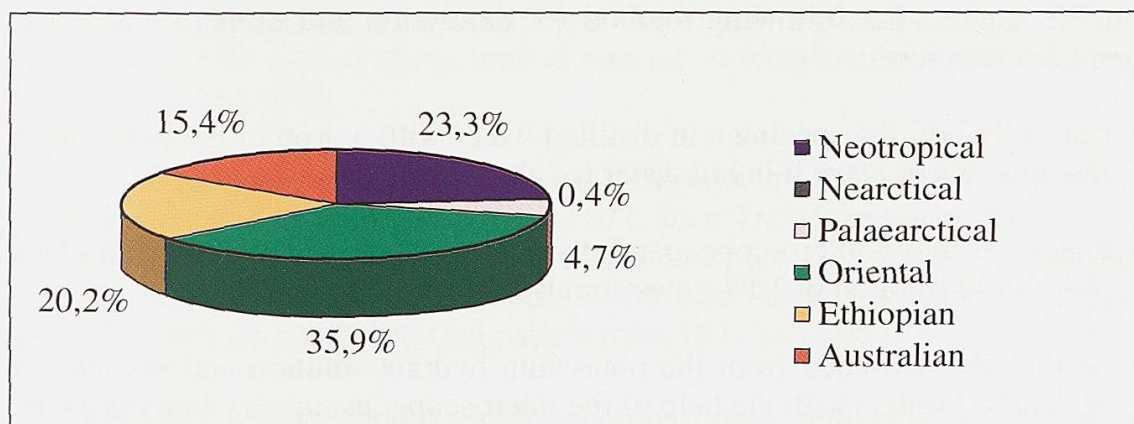


Fig. 132 - Percentages of the brentid species in the different Zoogeographical Regions.

COLLECTING AND PREPARATION METHODS

Brentids can usually be collected during the day under the barks of trees or from rotting and fallen branches. Some genera (e. g. *Cerobates*) prefer the barks still partly attached to the sapwood, whilst others (e. g. *Brentus*) are found under loose bark (Johnson, 1982, 1983b). Some Cyphagogini occur on the wood of trees attacked by Scolytidae and Platypodidae. In tropical countries, brentids are more abundant during the rainy season. According to De Muizon (1960), in Africa it is possible to collect large numbers of specimens from parts of the forest that the local people have burnt for new plantations: the bark of the burnt trunks is in part lifted away and the fermenting lymph attracts many species of *Cerobates* and *Microtrachelizus*. In general, brentid species are not strictly associated with just one plant species and may be found on several trees (see Appendix 1). Brentids can also be found resting on leaves and flowers. Myrmecophilous species occur under stones or wood where there are ant nests.

Collecting specimens is most successful at night with a bulb, as they are attracted by light.

Brentids should be prepared by pinning or, if they are very small, by glueing them onto the tip of a triangular label; the underneath of the body should be free for examination for identification purposes.

Correct preparation of the reproductive apparatus is extremely important, as its examination is often necessary for identification. Kleine (1917b) wrote a paper on the preparation of copulatory organs, but the method he suggests is not altogether satisfactory, nor his drawings of the aedeagi. Damoiseau prepared the genitalic structures by flattening them on slides, but this method causes considerable deformation of the parameres and other structures. Zimmerman (1994) also writes that "I have seen some of his [Damoiseau's] slide mounts and have found them rendered worthless by deterioration of the mountant".

We suggest the following method for extraction and preparation of the genitalic structures:

- Carefully boil the specimen in distilled water with a drop of detergent for a few minutes or leave it in hot water for about one hour;
- under the stereo-microscope, carefully detach all of the abdomen and place it in a cold solution of 10% potassium hydrate for 12-24 hours;
- remove the abdomen from the potassium hydrate solution and immerse it in distilled water; with the help of the microscope, using very fine tweezers, isolate and clean the copulatory apparatus taking care not to remove the various membranous structures pertaining to the chitinous parts;

- mount the apparatus in a drop of DMHF (Dimethyl Hydantoin Formaldehyde resin) on a label to be placed on the same pin holding the specimen.

DMHF is a perfectly transparent, water-soluble product. This means it is possible to examine the structure without deforming it and to move the apparatus if required as the medium can be made fluid again by adding a drop of distilled water. If the surface of the DMFH inclusion crinkles during the drying, it is sufficient to wet the surface with alcohol to flat it and to make it perfectly transparent (M. Alonso-Zarazaga, 2003, personal communication).

HISTORICAL REVIEW

The first author to describe two species of brentids (*Curculio anchorago* and *C. dispar*) was Carl von Linné (1758). In 1787, Johann Christian Fabricius transferred the Linnaean species to the new genus *Brentus*, together with several new species he had himself described. Subsequently several authors, including Niels Tønder Lund (1800) and Antoine Guillaume Olivier (1807), studied the group, then still considered as a tribe of the Curculionidae, and described several taxa. In his work "Entomologie, ou Histoire Naturelle des Insectes" (1808), Olivier illustrated several species of brentids with beautiful colour plates (Figs 133, 134). It was Gustav Johann Billberg (1820) who was responsible for assigning the status of family and who united the species of the genera *Episus*, *Brentus*, *Cylas*, *Antliarhis* and *Apius* under the "Brentithides" group.

Charles Henri Boheman and Leonhard Gyllenhal described a large number of new species in papers published by Carl Johann Schoenherr (1833, 1840), dividing the genera into twelve tribes. David Labram and Ludwig Imhoff (1838; 1841) illustrated many brentid species in wonderfully detailed colour plates (Figs 135-138).

In his monumental work in nine volumes on Coleopteran genera, published in Paris between 1854 and 1871, Jean Théodore Lacordaire treated the Curculionoidae in Volume VII (1866) and thus also the Brentidae, describing several new genera and species. Gustave Power also worked in Paris and published several interesting papers from 1878 to 1880.

David Sharp, the famous English entomologist, published an important monograph on Central American brentids (1895a) in which he described and illustrated many new species (Figs 139, 140). Von Schönfeldt, a German, was

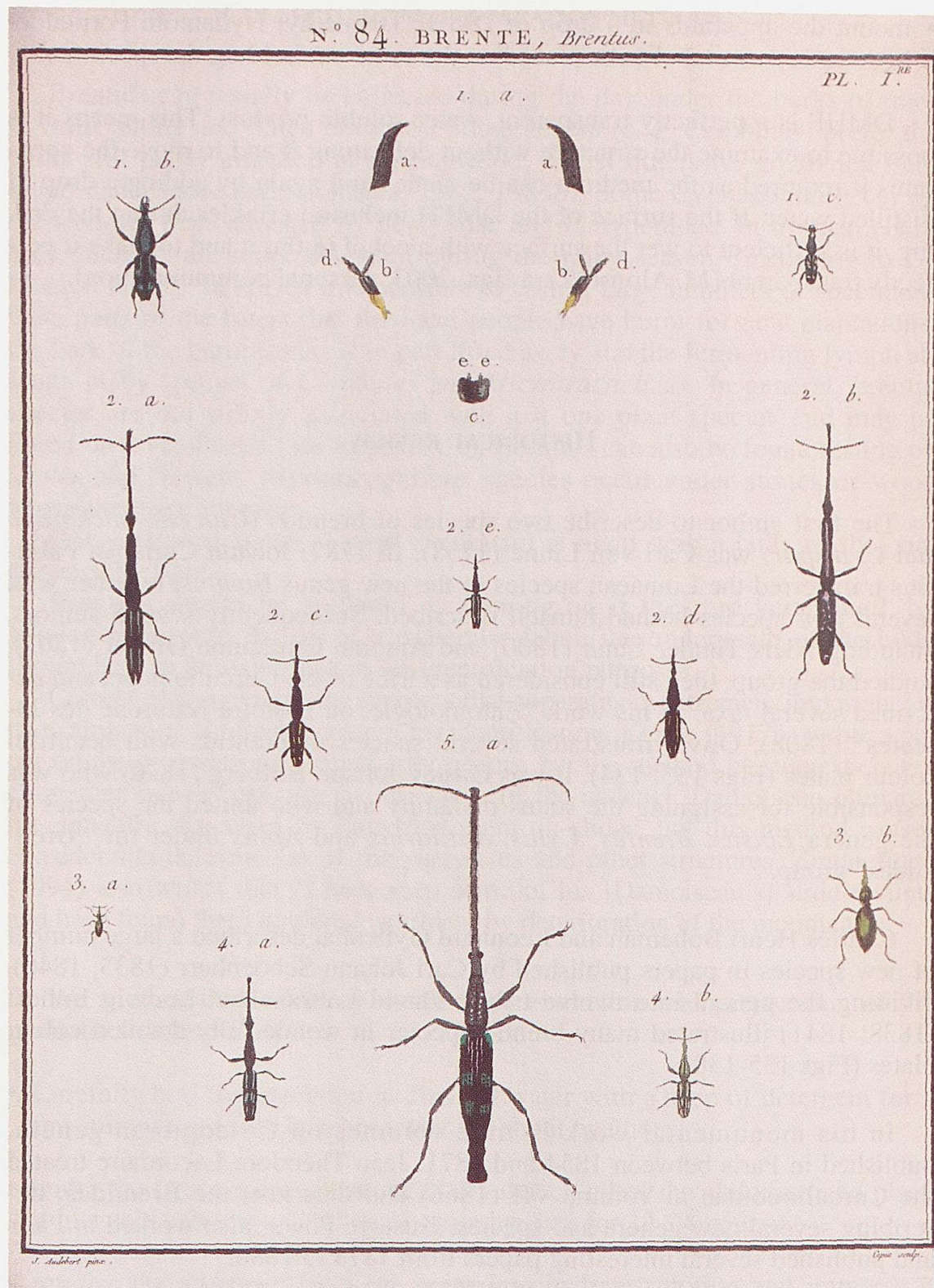


Fig. 133 - Plate I of Brentidae from Olivier's "Entomologie, ou Histoire Naturelle des Insectes" (1808).

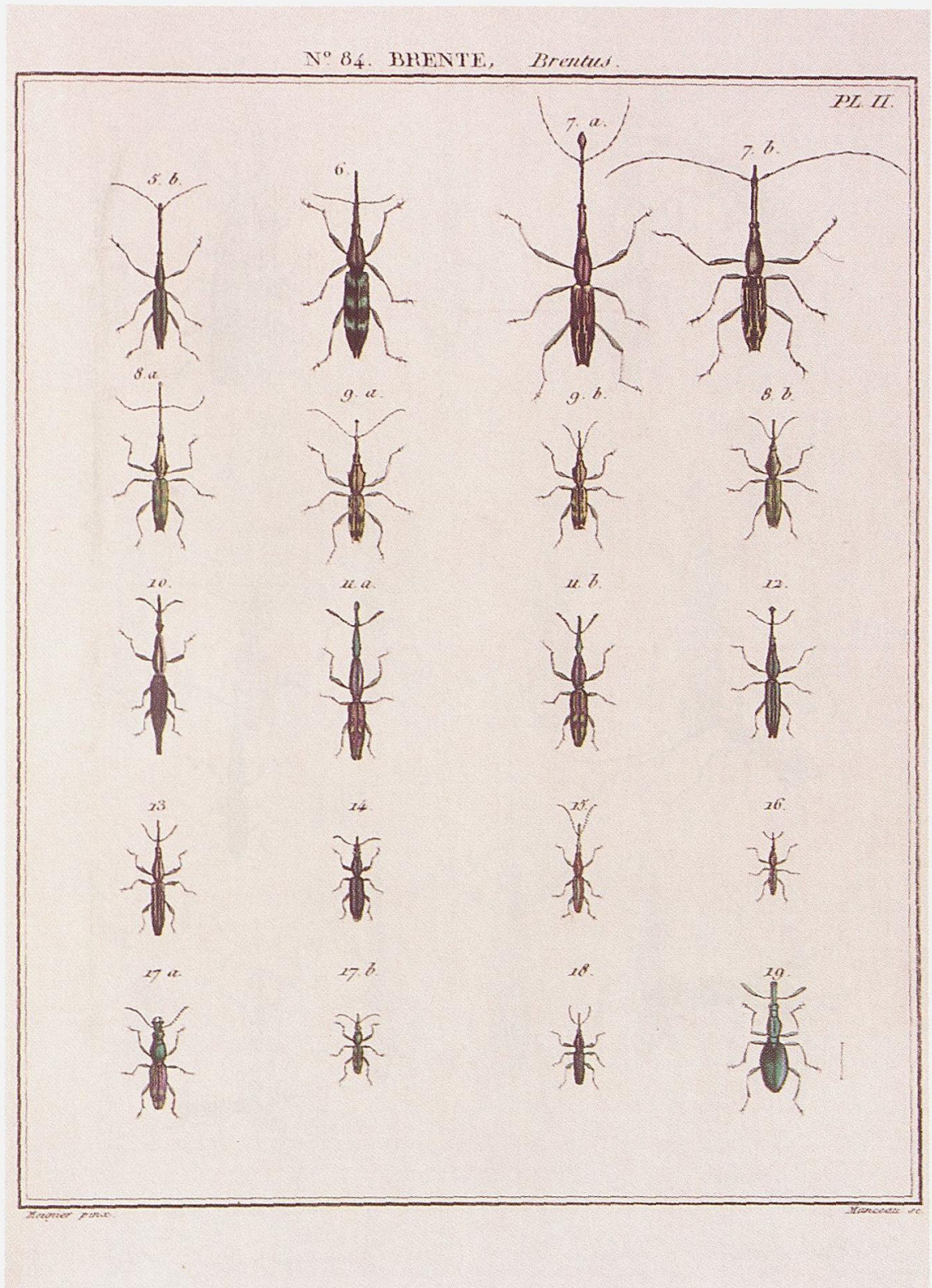


Fig. 134 - Plate II of Brentidae from Olivier's "Entomologie, ou Histoire Naturelle des Insectes" (1808).



Fig. 135



Fig. 136



Fig. 137

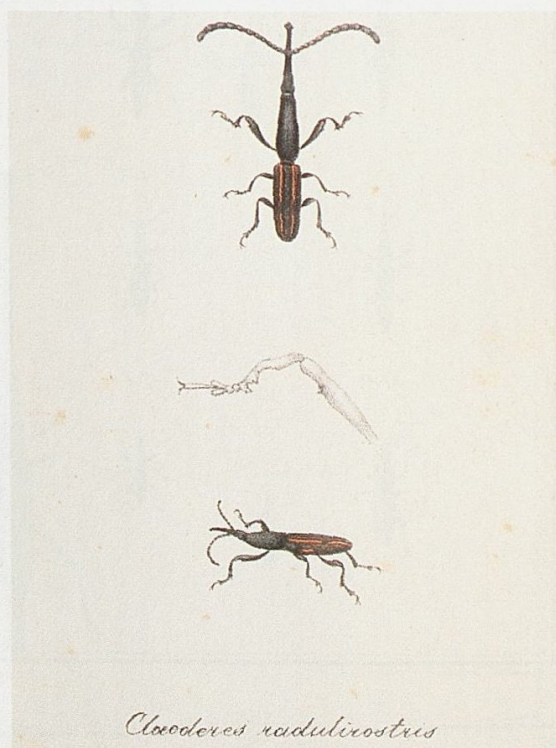


Fig. 138

Figs. 135-138 - Plates from Labram & Imhoff (1838; 1841). Fig. 135: *Estenorhinus faldermanni* (Gyllenhal); Fig. 136: *Stereodermus pygmaeus* (Gyllenhal); Fig. 137: *Nemorhinus myrmecophaga* (Herbst); Fig. 138: *Claeoderes radulirostris radulirostris* Boheman.

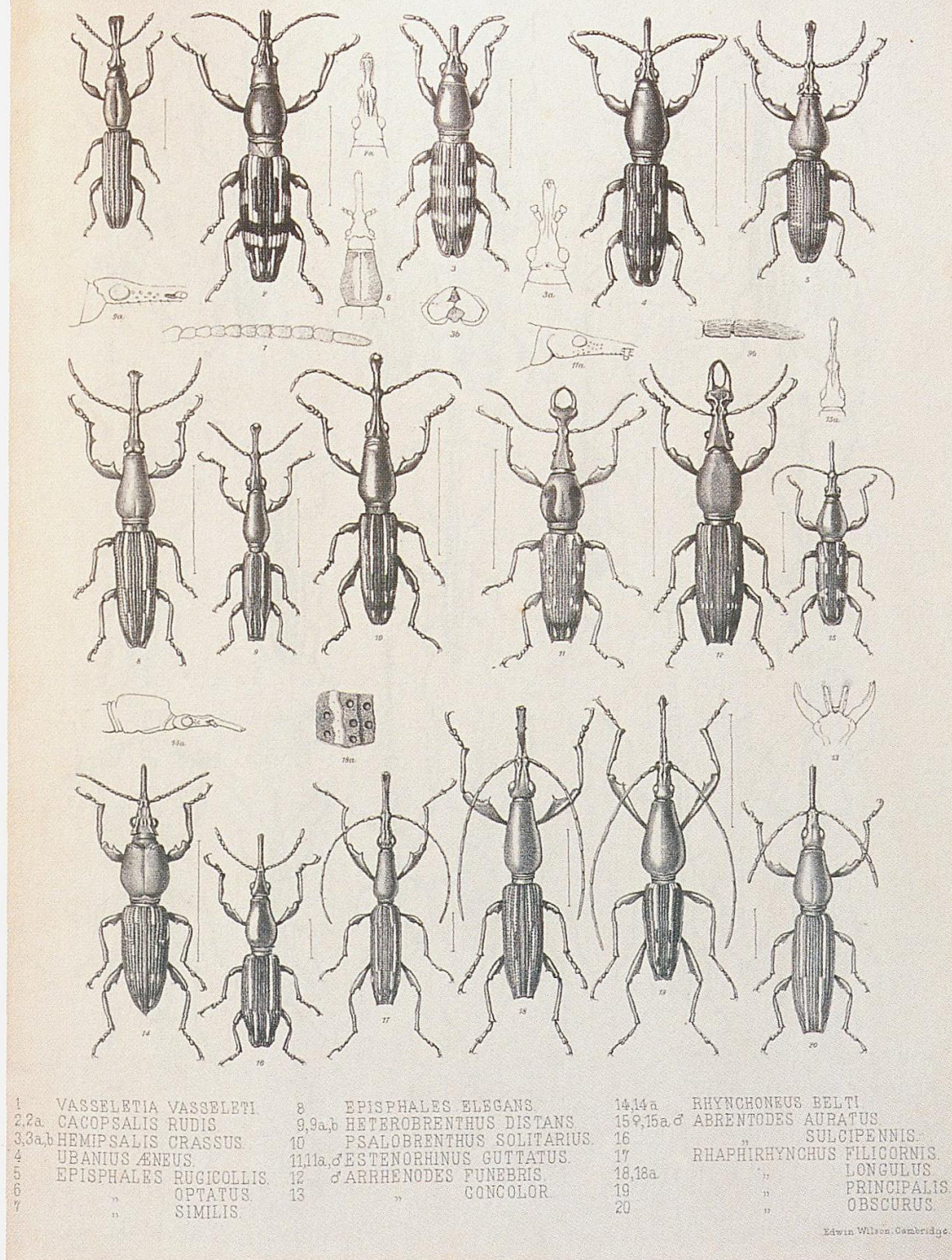
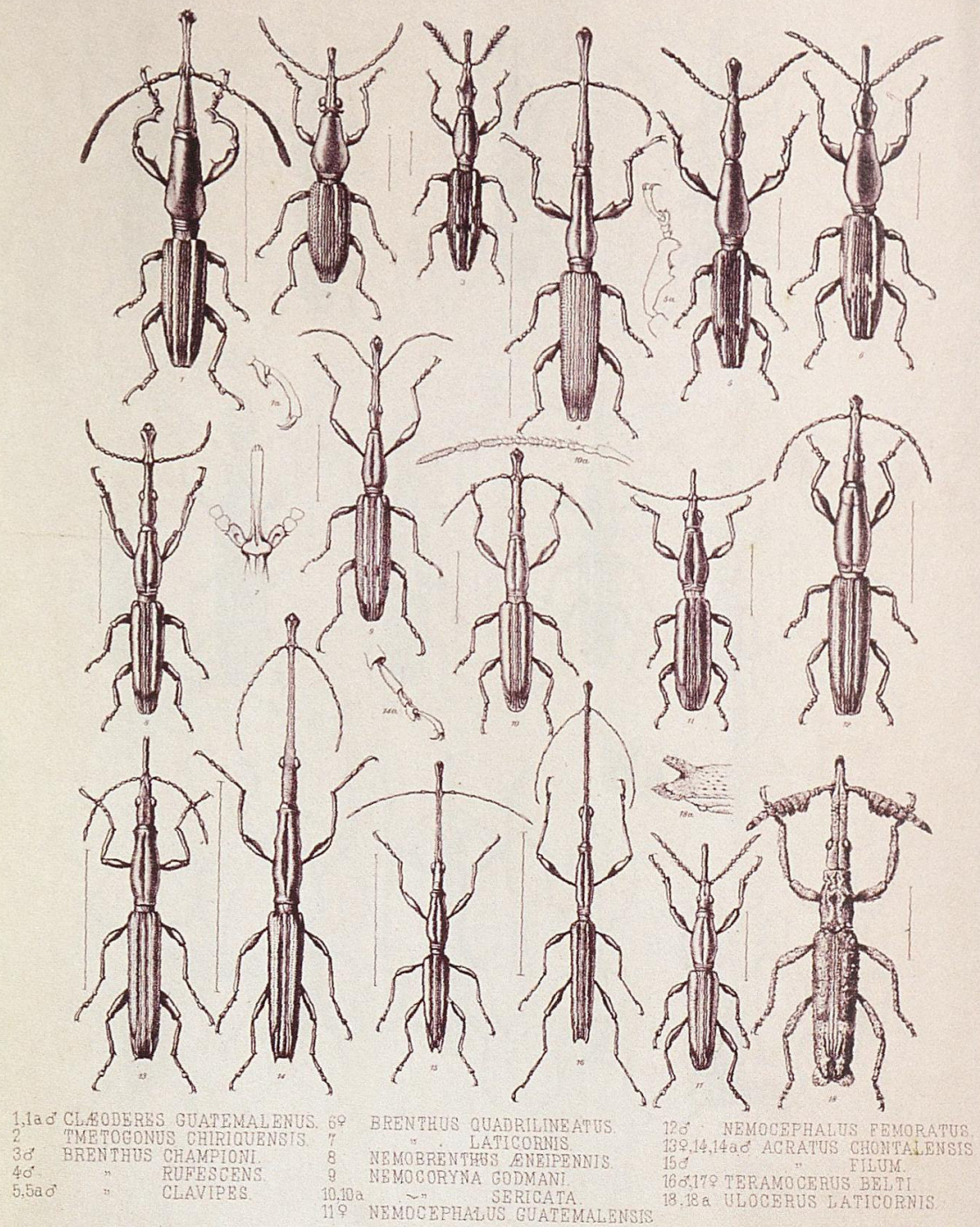
*Biol. Centr. Am.**Coleoptera. Vol. IV. Pt. 6. Tab. 2.*

Fig. 139 - Plate II from Sharp's "Biologia Centrali-Americana" (1895a).



Edwin Wilson Cambridge.

Fig. 140 - Plate III from Sharp's "Biologia Centrali-Americana" (1895a).

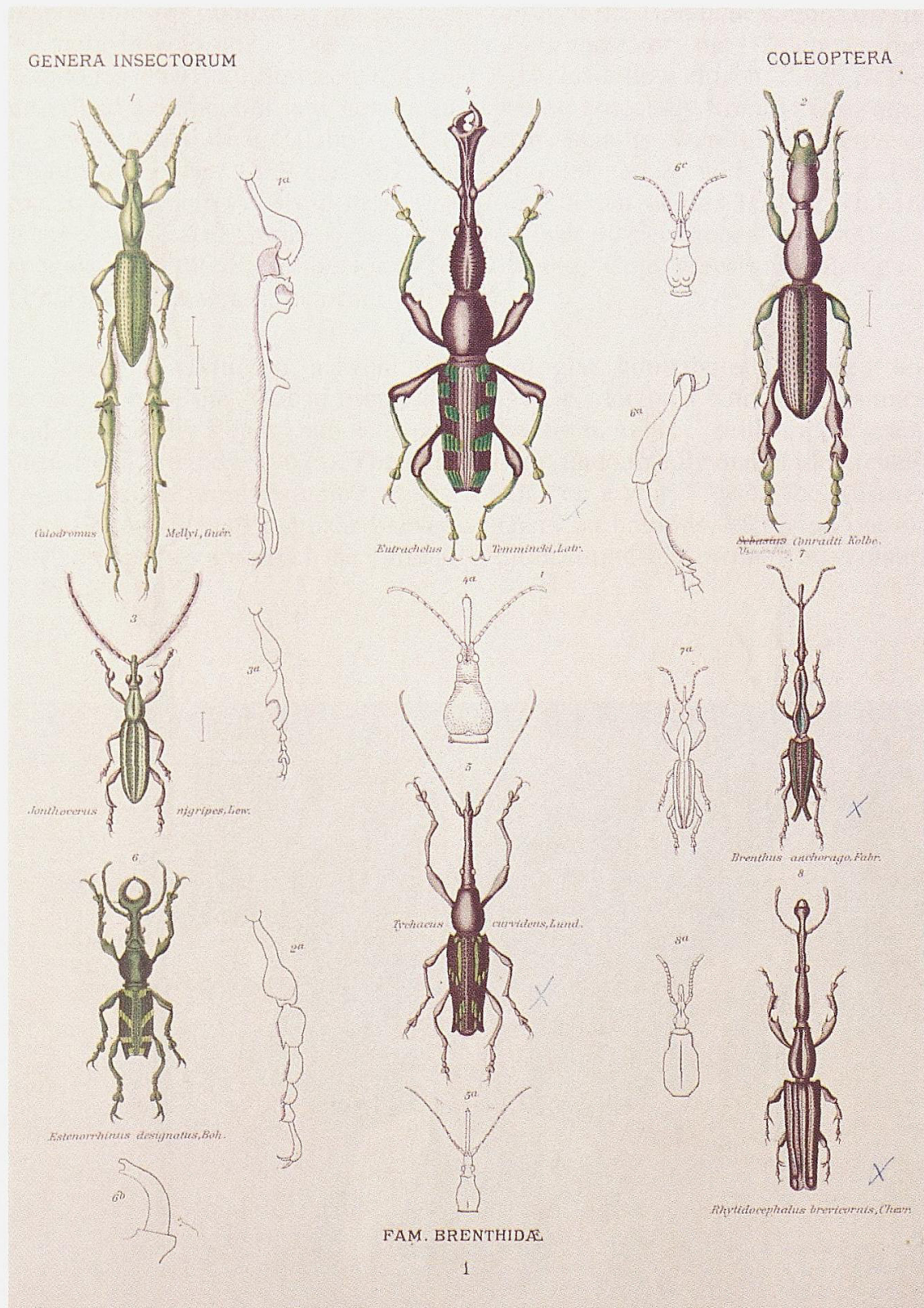


Fig. 141 - Plate I from von Schönfeldt's "Genera Insectorum" (1908).

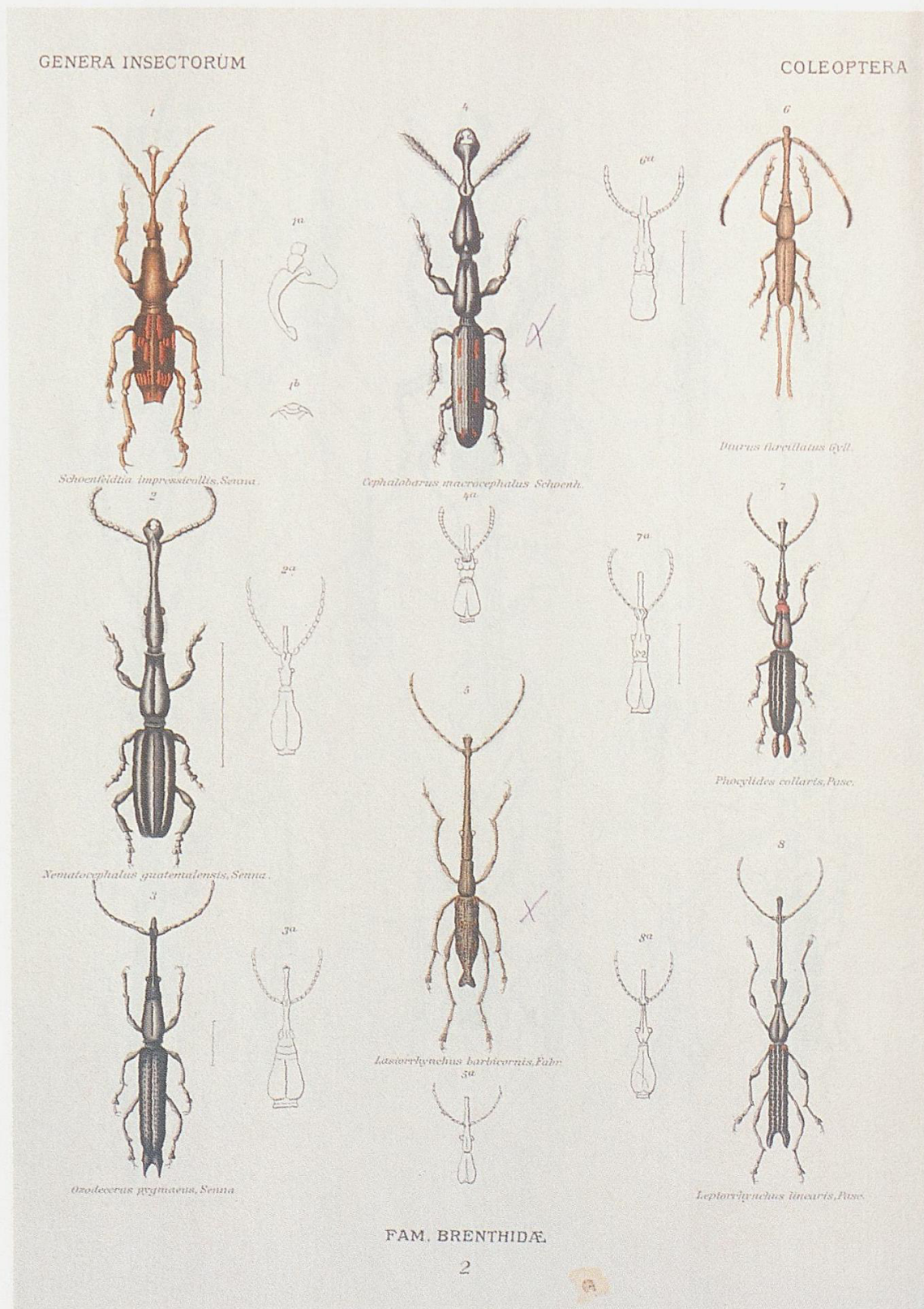


Fig. 142 - Plate II from von Schönfeldt's "Genera Insectorum" (1908).

responsible for publishing the first checklists of the Brentidae family for the "Genera Insectorum" (1908) (Figs 141, 142) and later the "Coleopterorum Catalogus" (1910a). Between 1883 and 1916, his fellow citizen, Julius Kolbe, described a number of new genera and species, especially from Africa. Angelo Senna and Enrica Calabresi, of Italian nationality, were highly active in Florence between 1889 and 1922, enriching the family with new taxa. Between 1914 and 1948 the most prolific specialist of this family, Richard Kleine, worked in Germany and described over 90 new genera and 500 new species. He edited the checklists for both the "Coleopterorum Catalogus" (1927) and "Genera Insectorum" (1938d).

Joseph De Muizon, a French entomologist, contributed an important monograph on the African brentids (1960). De Muizon's work was resumed and enlarged by Roger Damoiseau, from Belgium, in his excellent monograph on African brentids (1967a). The latter author, undoubtedly one of the greatest experts of the family, studied its systematics on a world level. The Austrian Karl Eduard Schedl published several papers on brentids, including one on African species in which he furnished an accurate list of their host plants (1961a).



Fig. 143 - Eylon Orbach, Alessandra Sforzi, Luca Bartolozzi (Florence, 2002) (photo by S. Bambi).

The South American fauna has recently been studied by Sydney Mello Dias, Eugênio Izecksohn, Carlos R. Meyer, Adelmo Scivittaro and Benedicto Abilio Monteiro Soares, all of Brazilian nationality, as well as by José Haedo Rossi, from Argentina. Interesting papers on the brentid ethology of some South American species were made by the American Leslie Johnson.

The brentids of Oceania have been revised by Elwood C. Zimmerman (1994), an Australian, who also published photographs of a large number of species (1991).

At present, as well as the authors of this work, Eylon Orbach, (Fig. 143) from Israel, Antoine Mantilleri, from France, Oleg N. Kabakov from Russia, Adelmo Scivittaro and Marcia Aparecida Sanches, from Brasil, and Jacques Goossens, from Belgium, are all involved in Brentidae (*sensu Auctorum*) taxonomy and faunistics. The systematic position of the family within the Curculionidae and its phylogenesis have been studied by Richard Thompson (1992) from England, Guillermo Kuschel (1995), from New Zealand, Juan Morrone (1998, 2000), from Mexico, Rolf Oberprieler (2000) from Australia, Adriana Marvaldi (2000) from Argentina and Marek Wanat (2001) from Poland. An up-to-date checklist of brentid genera has been published by Miguel Alonso-Zarazaga and Chris Lyal (1999, 2002) in the volume "A World catalogue of families and genera of Curculionoidea".

We believe that the following biographical information on the most important specialists to study Brentids over the last century will interest our readers. Other information on a number of authors can be found in Zimmerman (1993).

David Sharp (15th October 1840, Towcester, England – 27th August 1922, Brockenhurst, England) (Fig. 144) was a doctor, but left his medical practice in 1883 to dedicate himself to entomology. In 1890 he became curator of the insect collections at the Cambridge University Museum of Zoology where he worked until he retired in 1909 (Zimmerman, 1993). He was one of the most famous entomologists and published over 250 papers. In five papers devoted to Brentidae he described 10 genera and 88 new species. His most important work on brentids is undoubtedly that on the Central American species published in "Biologia Centrali Americana" (1895a). He was a Fellow of the Linnean Society, Zoological Society, Royal Society and Entomological Society of London, serving the latter as Vice-President and Secretary. His Coleopteran collection was donated to the Natural History Museum of London (Harvey *et al.*, 1996). Further information on the life and works of Sharp can be found in Zimmerman (1993).

Hermann Julius Kolbe (2nd June 1855, Halle in Westfalen, Germany – 26th November 1939, Berlin, Germany) (Fig. 145) graduated in zoology at Münster. From 1878 to 1882 he taught at the Oeding Gymnasium in Westfalen and in 1882 became Assistant of the Entomology Section at the Museum für Naturkunde der Humboldt-Universität in Berlin. From 1890 to 1921 he was Curator of the Coleopteran and Neuropteran collections there. He was a specialist in Scarabaeidae and Brentidae but also studied practically all the principal Coleopteran groups. The types for the species he described are held in the Berlin Museum where he worked (Uhlig & Jaeger, 1995). He published over 110 papers on African fauna, including his particularly important work “Die Käfer Deutsch-Ost-Afrikas” (1897). In his 12 papers dedicated to brentids, he described 21 new genera and 56 new species, considering only those then still valid.

One of the most important specialists of the Brentidae family was undoubtedly the Italian Angelo Senna (1st April 1866, Milan, Italy – 15th March 1952, Florence, Italy) (Fig. 146). He was Professor of Zoology at the University of Florence (Colosi, 1952; Baccetti, 1986; Conci & Poggi, 1996). He did not limit his studies to insects, but was also interested in Chiroptera and Crustacea, publishing over a hundred papers, 43 of which were dedicated to the study of brentids. He described 32 new genera and 249 new species and varieties, most of which are still valid. Among his most important works are the revisions of the genera *Hoplopisthius* (1893c), *Raphidorhynchus* [= *Raphirhynchus*] (1894d), *Cerobates* (1896a), *Ulocerus* (1896c) and *Diurus* [= *Ceocephalus*] (1911). Damoiseau (1967a: 8) considered Senna as... “le plus remarquable des systématiciens qui se soient occupés de Brentidae”. His rich collection is presently deposited in the Zoological Section ‘La Specola’ of the Natural History Museum of the University of Florence and the type material contained therein is listed in Bartolozzi *et al.* (1985; 2002).

Senna’s pupil was Enrica Calabresi (10th November 1891, Ferrara, Italy – 18th January 1944, Florence, Italy) (Fig. 147). She held the post of Assistant Professor of Zoology at the University of Florence and was responsible for herpetology and Cnidaria as well as insects. She published 9 papers on brentids in which she described 7 new genera and 34 new species. Some of her most important works are the revisions of the Hoplopistiini tribe (Senna & Calabresi, 1919), the Belopherinae [= Arrhenodini] sub-family (Calabresi, 1920b, 1921c) and the genus *Stratiorrhina* (Calabresi, 1921b). Her sad personal life had long been forgotten until recently brought to light again (Poggesi & Sforzi, 2001). Enrica Calabresi was Jewish and in 1938, because of the enforcement of racial laws by the Fascist government in Italy, she was forced to resign from her position at the University. In January, 1944, she was arrested by the Nazis to be deported to a concentration camp; to avoid such a fate she committed suicide by taking poison. Most of the types of the species she

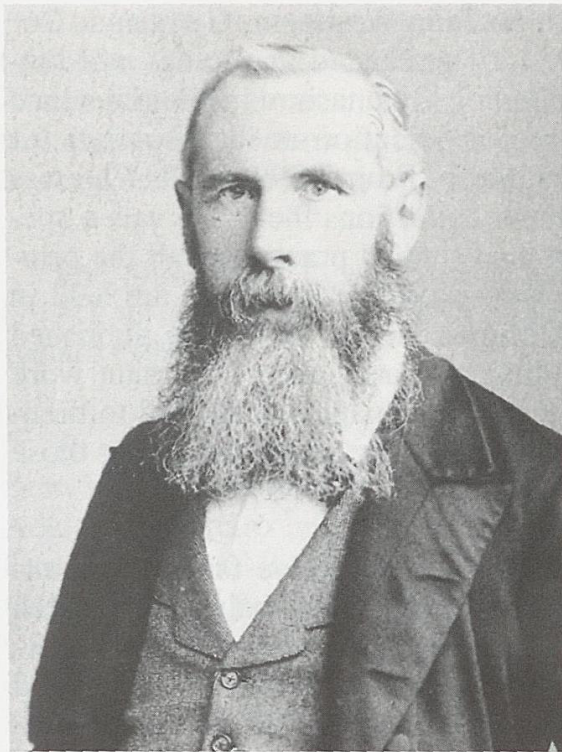


Fig. 144



Fig. 145



Fig. 146



Fig. 147

Figs 144-147 - Fig. 144: David Sharp; Fig. 145: Hermann Julius Kolbe; Fig. 146: Angelo Senna; Fig. 147: Enrica Calabresi.

described are deposited in the Zoological Section 'La Specola' of the Natural History Museum of the University of Florence (Bartolozzi *et al.*, 1985; 2002).

Richard Kleine (4th July 1874, Magdeburg, Germany – 10th April 1948, Halle a. S., Germany) (Fig. 148) was one of the greatest specialists of the family, of which he described as many as 91 new genera and 534 new species. His father was a carpenter and his mother an expert horticulturist. He took an interest in plants and insects at an early age, stimulated by his mother, but was forced by his father to work as a printer, first at Magdeburg and then, from 1899 to 1910, at Halle a. S. (Sachtleben, 1949). Whilst he was living in this city, he took the opportunity of attending evening classes at University and a naturalist group. During this period he also began to publish a good number of scientific papers. In 1911 he decided to leave his job as a printer to take the position of Assistant at the Phytopathological Station of the Agricultural Office in Saxony Province. In 1912 he moved to Stettin, near the Agricultural Office in Pomeran Province, where he stayed until 31st December 1937 when he asked to retire. From 1938 he collaborated with the Stettin Museum of Natural History. On 10th March 1945, after the city was evacuated, he was obliged to leave Stettin with his wife and move first to Zettemin and then Pinnow. This put an end to his entomological activity, but on 10th April 1946 he returned to Halle Agricultural Station as a collaborator. After losing all his possessions after the evacuation of Stettin, in March 1947 he suffered another loss, the death of his wife. He died of tuberculosis on 10th April, 1948, leaving as many as 417 scientific papers on several Insect orders. His international fame as a coleopterist is due to his studies on the systematics of the Brentidae and Lycidae. Some of his most important works on brentids include the *Coleopterorum Catalogus*, Pars 89 (1927a), the *Bestimmungstabellen der Brentidae* (1927-1933) and the *Genera Insectorum*, Fasc. 207 (1938d). His research into brentids not only concerned systematics, but also their biology, zoogeography and faunistics. According to Damoiseau (1967a: 356) and Zimmerman (1993: 601) Kleine's collection was destroyed when the Stettin Museum was bombed during World War II, but actually a good part of the types of the Kleine collection is now in the Warsaw Museum. In spite of the enormous contribution Kleine gave to our knowledge of brentid taxonomy, mention should be made (as several authors such as Damoiseau and Zimmerman have pointed out) of the number of errors that occur in the material he studied. In the words of Zimmerman (1993: 601), "he left to posterity a great burden".

Joseph De Muizon (29th December 1890 – 23rd October 1958, Paris, France) (Fig. 149) was an Agricultural Engineer. He participated in numerous expeditions to Africa and began to take an interest in brentids under the guide of Prof. Jeannel from the Museum of Paris. He dedicated himself almost exclusively to African brentids and in 1960 his monograph "Faune des Bren-



Fig. 148

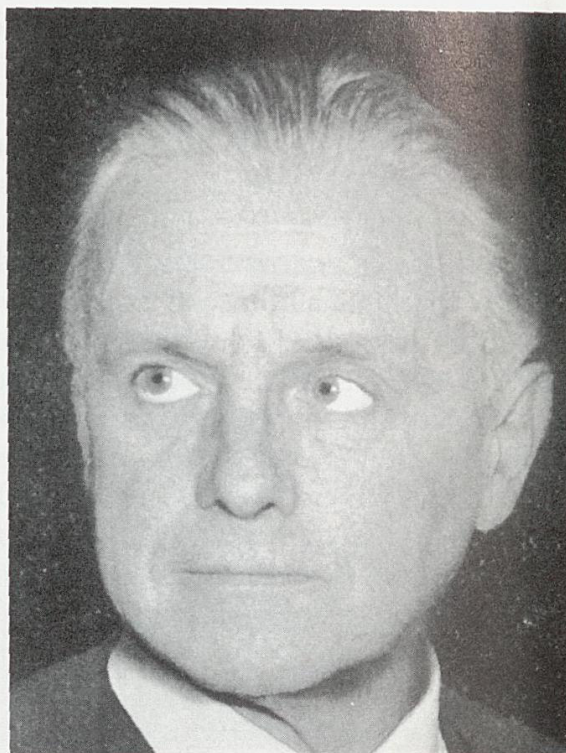


Fig. 149



Fig. 150

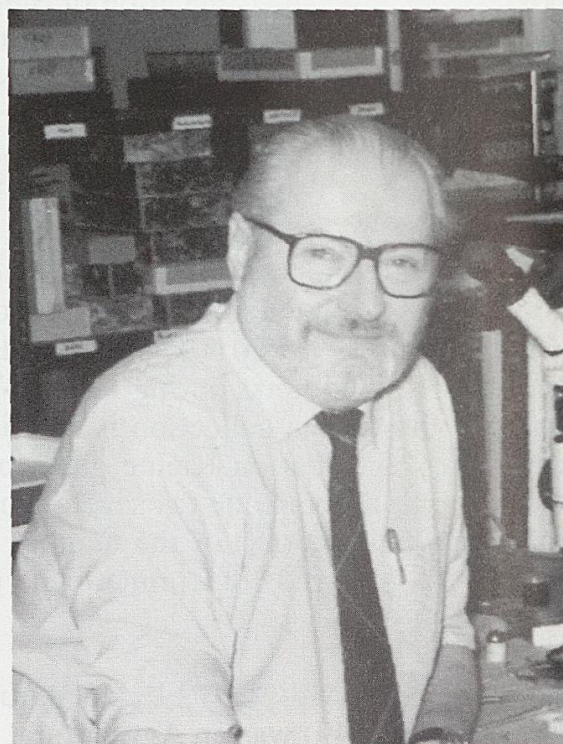


Fig. 151

Figs 148-151 - Fig. 148: Richard Kleine; Fig. 149: Joseph De Muizon; Fig. 150: Karl Eduard Schedl; Fig. 151: Roger Damoiseau.

thides d'Afrique" was published posthumously. He described several new genera and species in six studies on brentids.

Karl Eduard Schedl (17th January 1898 – 18th May 1979, Lienz, Austria) (Fig. 150) graduated in Engineering in 1921 and began taking an interest in entomology, particularly the fight against the Coleoptera Scolytidae. From 1926 to 1932 he carried out his activity as entomologist in Canada, at the Entomological Branch of the Department of Agriculture in Chatham (Ontario). In 1932 he returned to Vienna. He taught at the University and worked in several forestry research institutes in Germany and Austria. In 1952 he led a scientific expedition to the Congo and in 1953 to Madagascar. He assembled an important collection of Scolytidae, Platypodidae and Brentidae (Fischer, 1979). He wrote 413 scientific publications, six of which concerning brentids, especially African species.

One of the most important specialists in brentids was undoubtedly Roger Damoiseau (13th September 1929, Namur, Belgium – 3rd April 1984, Bruxelles, Belgium) (Fig. 151). He was a Forestry Agricultural Engineer. From 1955 to 1961 he worked in the Belgian Congo as a forestry entomologist; then after Congo gained independence, he returned to Belgium where he was employed at the Institut Royal des Sciences Naturelles de Belgique in Bruxelles

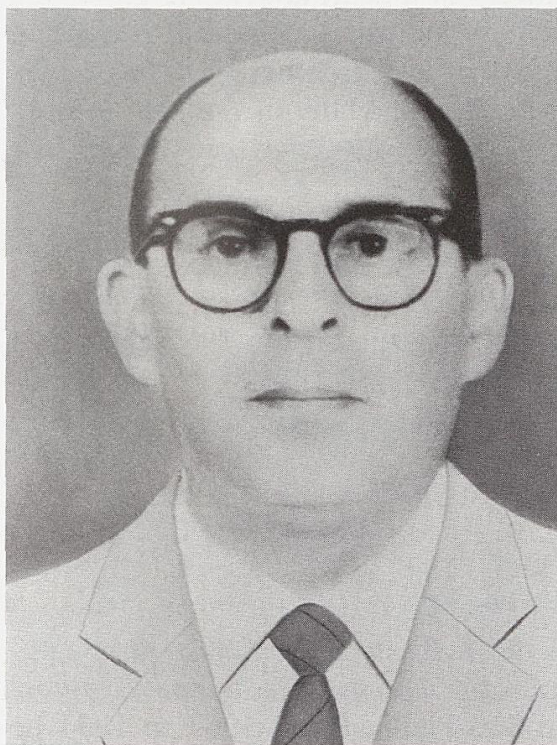


Fig. 152

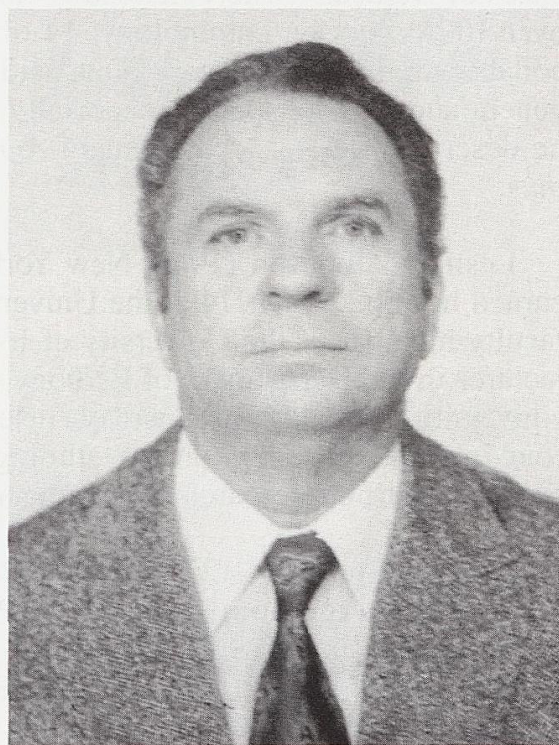


Fig. 153

Figs 152-153 - Fig. 152: Benedicto Abilio Monteiro Soares; Fig. 153: Adelmo Scivittaro.

until his untimely death (Staner, 1984). He described 27 new genera and 225 new species of brentids. He was also interested in Scolytidae. His most important works on the Brentidae include his revision of the genera *Calodromus*, *Cyphagogus*, *Eterozemus*, *Neoceocephalus*, *Amorphocephala* and *Cordus*.

One of his fundamental works is his monograph on African brentids (1967a) in which Damoiseau revised and enlarged previous studies by De Muizon (1960). Three important contributions were published posthumously (Damoiseau, 1987, 1989a, 1989b) by entomologists of the Institut Royal des Sciences Naturelles de Belgique, based on a collection of notes they found. Damoiseau was undoubtedly one of the greatest specialists in brentids and his works are all extremely accurate, richly illustrated and show particular detail to the study of the reproductive organs. His collection is held in the Institut Royal des Sciences Naturelles de Belgique in Bruxelles.

Benedicto Abilio Monteiro Soares (12th September 1914 – 12th December 1985, Botucatu, Brazil) (Fig. 152) was an important specialist in South American brentids. He was Professor of Biology from 1941 to 1950 at the Department of Zoology of the Secretaria da Agricultura do Estado de São Paulo (now the Museu de Zoologia da Universidade de São Paulo). From 1950 to 1966 he held the chair for Zoology at the Escola Nacional de Agronomia, Rio de Janeiro. In 1966 he moved to the faculty of Medical and Biological Sciences of Botucatu, where he died prematurely of a heart attack. He published over 100 papers on entomology, 14 of which dedicated to brentids (some in collaboration with Scivittaro, Dias and Meyer), in which he described 2 new genera and 39 new species, especially from Brazil. Many of the new species he described belong to the genera *Acratus*, *Proteramocerus* and *Nemobrenthus*.

Leslie K. Johnson (1946, New York - 16th May 1997, Princeton, U.S.A.) earned her Ph.D. in 1974 at the University of Berkeley, California, and was a faculty member of the University of Iowa for 13 years. From 1989 she was a lecturer in the Department of Ecology and Evolutionary Biology at Princeton University. She was an expert on insect behavioural ecology and published four interesting papers on the ethology of some South American brentids, based on her field research in Panama and Costa Rica in the 1980's.

Adelmo Scivittaro (27th May 1932, Salto, São Paulo, Brazil) (Fig. 153) graduated in Agronomy in 1958 at Piracicaba and from 1959 to 1966 he worked at the Instituto Biológico and at the Instituto Agrônômico da Secretaria de Estado da Agricultura de São Paulo and Diretoria de Ensino Agrícola da Secretaria de Estado da Educação de São Paulo. From 1967 to 1991 he taught at the Department of Zoology of the Faculty of Medical and Biological Sciences at Botucatu of the Universidade Estadual Paulista (UNESP). Until now he published 12 papers on the systematics of South American brentids,

some together with Soares. His most important works include the revision of the neotropical Taphroderinae (1975). He described a total of 3 new genera and 27 new species.

Between 1948 and 1961, the Argentinian José Haedo Rossi wrote a number of works on South American brentids, including an interesting monograph on the brentids of Argentina (1961). Unfortunately we have not been able to obtain a photograph or biographical information from this specialist, in spite of our repeated requests.

To assist attribution and identification of old museum material, we have reproduced some of the original labels we found in the handwriting of Gustave Power (Fig. 154), Hermann Julius Kolbe (Fig. 155), Angelo Senna (Fig. 156) and Richard Kleine (Fig. 157).

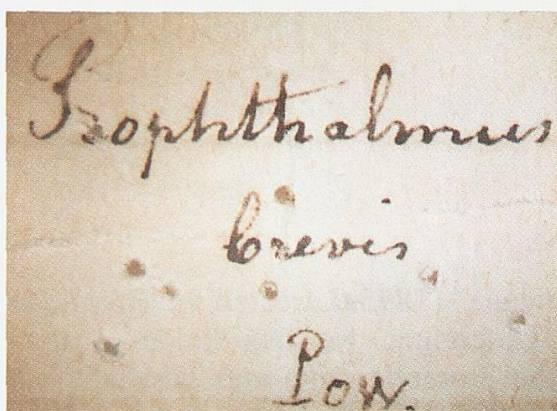


Fig. 154

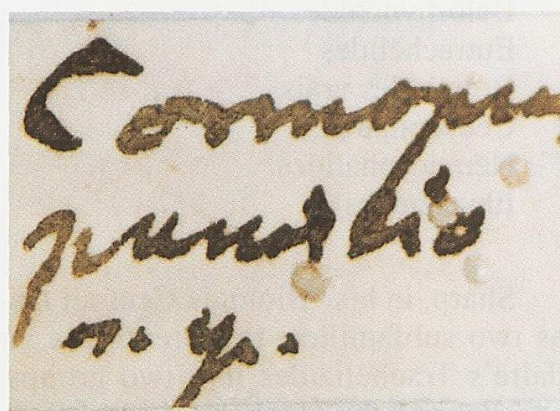


Fig. 155

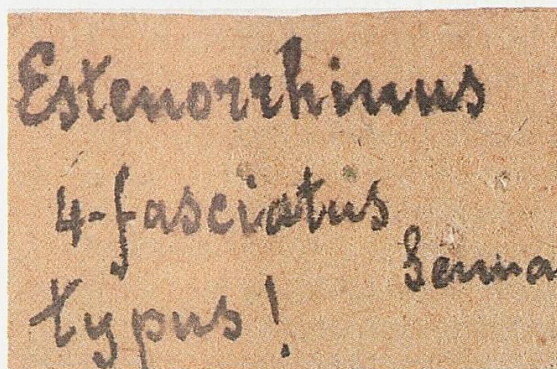


Fig. 156

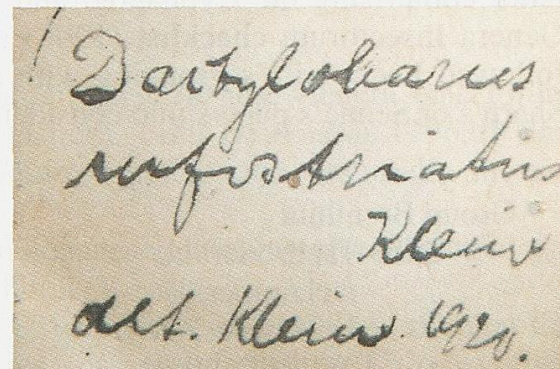


Fig. 157

Figs 154-157 - Handwritten labels. Fig. 154: Gustave Power; Fig. 155: Julius Kolbe; Fig. 156: Angelo Senna; Fig. 157: Richard Kleine.

CLASSIFICATION

Several authors have provided historical reviews on brentid classification (Kleine, 1938d; Haedo Rossi, 1961; Damoiseau, 1963b; Scivittaro, 1975; Zimmerman, 1994). However, in our opinion a summary is beyond the scope of this book.

Lacordaire (1866) was the first to split the brentids into groups. He divided the family into two tribes: "true" Brenthides and Ulocerides. The "true" Brenthides tribe was subsequently divided into twelve groups:

- Taphrodérides
- Ischnomérides
- Héphébocérides
- Trachélizides
- Arrhénodides
- Bélophérides
- Bélorhynchides
- Eutrachélides
- Brenthides vrais
- Céocéphalides
- Némocéphalides
- Ithysténides

Sharp, in his "Biologia Centrali Americana" (1895a), treated the Brentidae as two subfamilies: the Brenthinae and Ulocerinae. He also divided Lacordaire's Trachélizides into two groups, the Stereodermina and Trachelizina, and deleted the Héphébocérides. Senna (1896a) adopted Sharp's classification, but considered the Stereoderminae and Taphroderinae as subfamilies, the latter comprising the Cyphagogi and Taphroderi groups. Schönfeldt, in his Genera Insectorum checklist (1908) and Coleptorum Catalogus (1910a), followed Lacordaire's grouping with a few modifications, but did not take Sharp's or Senna's papers into consideration:

Group Brenthini

- Tribe Taphroderidae
- Ischnomeridae
- Epheboceridae
- Trachelizidae
- Arrhenodidae
- Belopheridae
- Tychaeidae

Eutrachelidae
 Brenthinidae
 Ceocephalidae
 Nematocephalidae
 Leptorrhynchidae
 Eremoxenidae
 Group Ulocerini

Senna and Calabresi (1919) criticised Schönfeldt's work and proposed adding the Hoplopisthi group to the Taphroderinae subfamily, next to the Cyphagogi and Taphroderi.

In his paper "Neuklassifizierung der Brentidae" (1922m) Kleine proposed a new classification for brentids, subdividing them into 15 tribes:

Calodromini
 Stereodermini
 Trachelizini
 Amorphotocephalini
 Arrhenodini
 Belopherini
 Eutrachelini
 Tychaeini
 Ithystenini
 Ulocerini
 Pseudoceocephalini
 Taphroderini
 Rhyticephalini
 Nemocephalini
 Brenthini

Kleine, in his *Coleopterorum Catalogus* (1927a) and *Genera Insectorum* (1938d) checklists followed the same pattern. Both Damoiseau (1963b) and Zimmerman (1994) pointed out that Kleine's classification was totally unsatisfactory, lacking reason or comment.

Morimoto (1962b), in his paper on Japanese Curculionoidea, merged the Apionidae and Brentidae into a single family.

Damoiseau, in his *Monograph on African Brentidae* (1967a), divided the African Brentidae into four subfamilies with various tribes: Calodrominae (Calodromini, Stereodermini, Hoplopisthiini and Atopobrentini), Ceocephalinae (Ceocephalini), Brentinae (Arrhenodini, Amorphotocephalini), and Taphroderinae (Taphroderini).

Morimoto (1976a) arranged the Japanese brentid fauna into two subfam-

ilies, Brentinae and Cyladinae. The Brentinae comprised the tribes Calodromini, Ceocephalini, Stereodermini, Arrhenodini, and Amorphocephalini. Morimoto did not follow Damoiseau's classification, and merged the Trachelizini with the Ceocephalini.

Both Damoiseau's and Morimoto's papers dealt with local fauna, so they did not consider the whole brentid family in their classifications.

At present controversy over the classification of the members of the Brentidae still reigns (Anderson & Kissinger, 2002).

Sanborne (1981) regarded the Antliarhininae as a distinct family but very close to Apionidae. He stated that if the two were united it would probably be necessary to also include the Brentidae.

Lawrence (1982) recognised the Brentidae (comprising Brentinae, Cyphagoginae and Trachelizinae) and Apionidae (comprising Apioninae, Nanophyinae and Cyladinae) as separate families.

Calder (1989), on the basis of internal characters (alimentary canal and central nervous system) wrote that *Cylas* may be allied to both the Brentidae and Apionidae, and that *Eurhynchus* and *Anthliarhinus* are primitive apionids.

Later, Calder (1990), on the grounds of the gross morphology of male soft parts and female internal reproductive organs, stated that *Cylas* is more closely related to the Apionidae, but confirmed the positions of *Eurhynchus* and *Anthliarhinus*. He also doubted Morimoto's amalgamation of the Apionidae and Brentidae into a single family.

Kuschel (1990) again merged Brentinae and Apioninae in the Brentidae.

Thompson (1992) studied the following characters for classifying the Curculionoidea: venter, abdominal tergites, sternite 8 of the male, apex of the hind tibia and deciduous mandibular processes. According to him, the family Brentidae comprises the Eurhynchinae, Apioninae, Nanophyinae, Tanainae, and Antliarhininae.

Zimmerman (1994), working on the Australian Brentidae, considers the family to be divided into two subfamilies (Brentinae and Cyladinae) and writes that "it is obvious from an examination of published works that various features of taxonomic importance have been ignored or examined superficially, and too much emphasis has been placed in isolation of some details".

According to Kuschel (1995), who submitted adult and larval characters to cladistic analysis, the Brentidae comprehend 7 taxa at tribal or subfamily level. The Brentini tribe falls under the same concept of Brentidae *sensu Auctorum*. Kuschel's classification is the following:

Brentidae

Carinae

Brentinae

Eurhynchini

- Antliarhinini
- Brentini
- Cyladinae
- Apioninae
 - Apionini
 - Nanophyini

Lawrence & Britton (1994) and Lawrence & Newton (1995) also include the Apioninae under the Brentidae.

Zherikhin & Gratshev (1995) studied the hind wing venation in the Curculionoidea and considered the family to have four subfamilies: Brentinae, Apioninae, Cyladinae and Nanophyinae.

Morrone (1998) provided an annotated consensus classification to families and subfamilies of Curculionoidea as follows:

- Brentidae
 - Brentinae
 - Brentini
 - Arrhenodini
 - Amorphocephalini
 - Cyphagogini
 - Hoplopisthini
 - Stereodermini
 - Trachelizini
 - Microtrachelizini
 - Cyladinae
 - Antliarhininae
 - Eurhynchinae
 - Apioninae
 - Apionini
 - Rhinorhynchidiini
 - Myrmacielini
 - Aplemonini
 - Notapionini
 - Nanophyinae

Oberprieler (2000), after phylogenetic analysis of several larval characters, included the Microcerini under the Brentidae as a Brentinae tribe and subdivided the Brentidae into three subfamilies: Ithycerinae, Brentinae and Apioninae. However, Marvaldi *et al.* (2002) wrote that “the combined analysis places the monotypic (and enigmatic) genus *Ithycerus* in the Brentidae, in accordance with Oberprieler (2000), but independent analyses of morphology and molecules do not support this grouping”. Marvaldi & Morrone (2000) consider the Ithycerinae as a subfamily of the Curculionidae.

Wanat (2001), after cladistic analysis of all family level taxa hitherto classified with the family Brentidae, subdivides the Brentidae *sensu* Kuschel (1995) into several families: Brentidae (with two subfamilies Brentinae and Cyladinae), Caridae, Nanophyidae, Eurhynchidae, and Apionidae (including Anthliarhininae). According to Wanat (2001) the monophyly of the Brentinae and Cyladinae is supported by a series of unique apomorphic characters.

Marvaldi *et al.* (2002) performed molecular and morphological phylogenetic research on Curculionoidea, but did not use any of the Brentidae *sensu* Auctorum species in their analysis. With respect to the paper by Kuschel (1995) Oberprieler (2000) and Marvaldi *et al.* (2002) exclude the Carinae from the Brentidae; Kuschel himself (2003) adopts this classification.

From the above, the deep divergences between the various authors are plain to see. We agree with Zimmerman (1994) when he writes "... a new classification of the Brentidae is urgently required". Hopefully, future phylogenetic studies, for example in the field of molecular biology, will finally clarify the systematic position of the different subfamilies and tribes.

In this book we follow the classification proposed by Alonso-Zarazaga & Lyal (1999), with some modifications (Alonso-Zarazaga & Lyal, 2002):

BRENTINAE Billberg, 1820

BRENTINI Billberg, 1820 (Fig. 158)

ARRHENODINI Lacordaire, 1866 (Figs 159-160)

EREMOXENINI Semenov-Tian-Shanskij, 1892 [= Amorphocephalini] (Figs 161-162)

ANTLIARHININAE Schoenherr, 1823

CYLADINAE Schoenherr, 1823

CYPHAGOGINAE Kolbe, 1892

CYPHAGOGINI Kolbe, 1892 [= Calodromini] (Figs 163-165)

ATOPOBRENTINI Damoiseau, 1965 (Fig. 166)

HOPLOPISTHIINI Senna & Calabresi, 1919 (Figs 167-168)

STEREODERMINI Sharp, 1895 (Figs 169-170)

PHOLIDOCHLAMYDINAE Damoiseau, 1962 (Pl. 1)

TAPHRODERINAE Lacordaire, 1866 (Fig. 171)

TRACHELIZINAE Lacordaire, 1866

TRACHELIZINI Lacordaire, 1866 (Figs 172-173)

ACRATINI Alonso-Zarazaga, Lyal, Bartolozzi & Sforzi, 1999
[= Nemocephalini] (Fig. 174)

ITHYSTENINI Lacordaire, 1866 (Figs 175-176)

MICROTRACHELIZINI Zimmerman, 1994 (Figs 177-178)

PSEUDOCEOCEPHALINI Kleine, 1922 (Figs 179-180)

RHYTICEPHALINI Kleine, 1922 (Fig. 141)

TYCHAEINI Schoenfeldt, 1908 [= Belorhynchini] (Fig. 67)

ULOCERINAE Schoenherr, 1823 (Fig. 181)

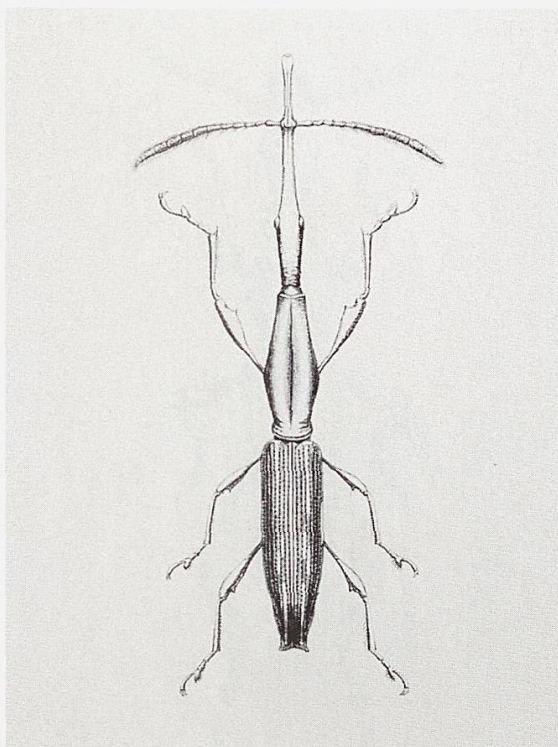


Fig. 158

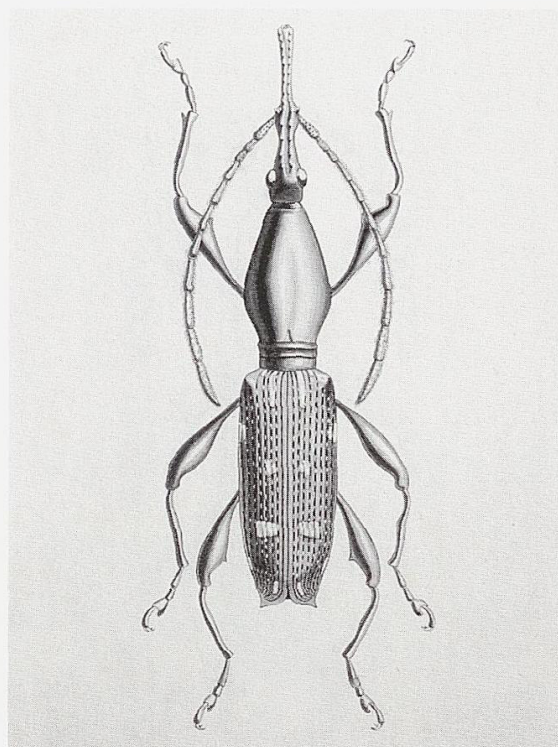


Fig. 159

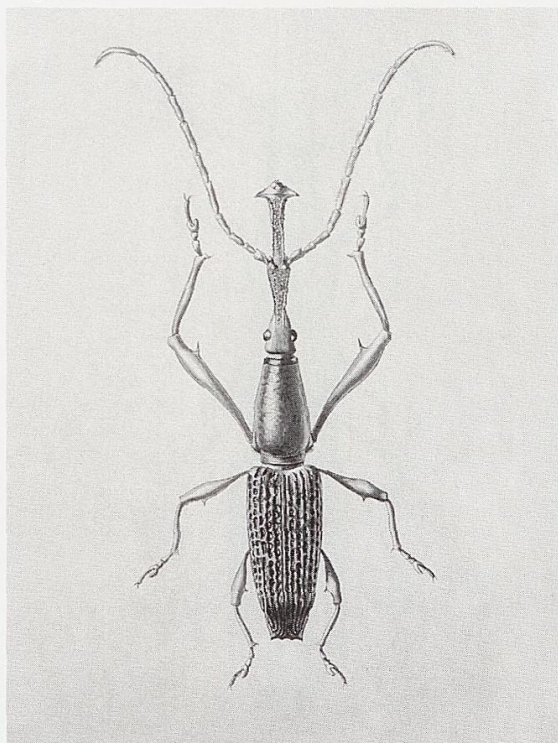


Fig. 160

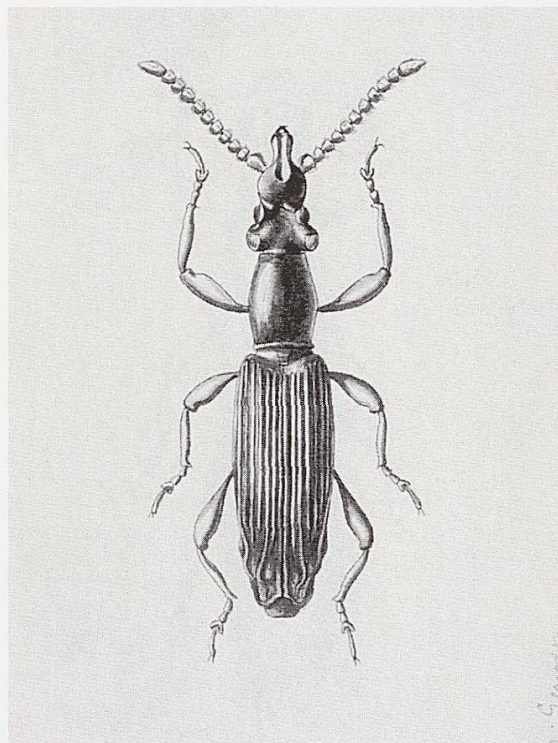


Fig. 161

Figs 158-161 - Fig. 158 - *Brentus rufiventris* (Boheman) (Brentini) (drawing by E. Calabresi); Fig. 159 - *Calabresia ceylonica* (Calabresi) (Arrhenodini) (drawing by E. Calabresi); Fig. 160 - *Pseudobelopherus orientalis* Calabresi (Arrhenodini) (drawing by E. Calabresi); Fig. 161 - *Leptamorphocephalus sumatranus* (Senna) (Eremoxenini) (drawing by E. Calabresi).

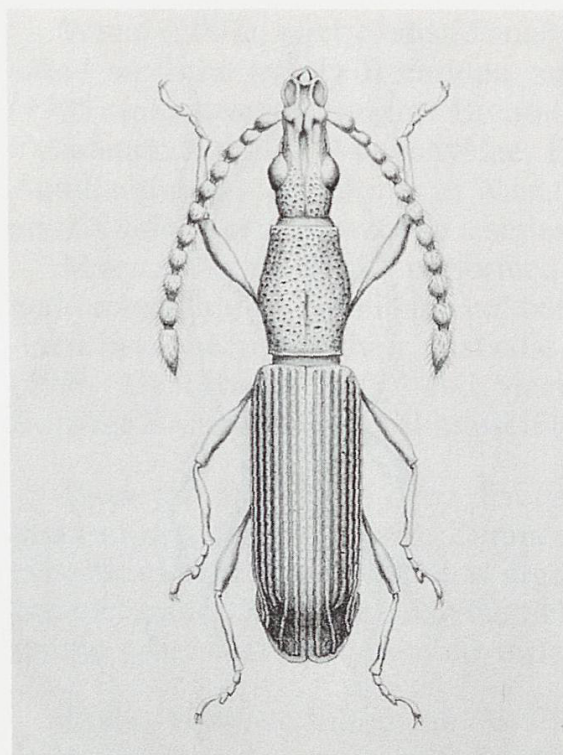


Fig. 162

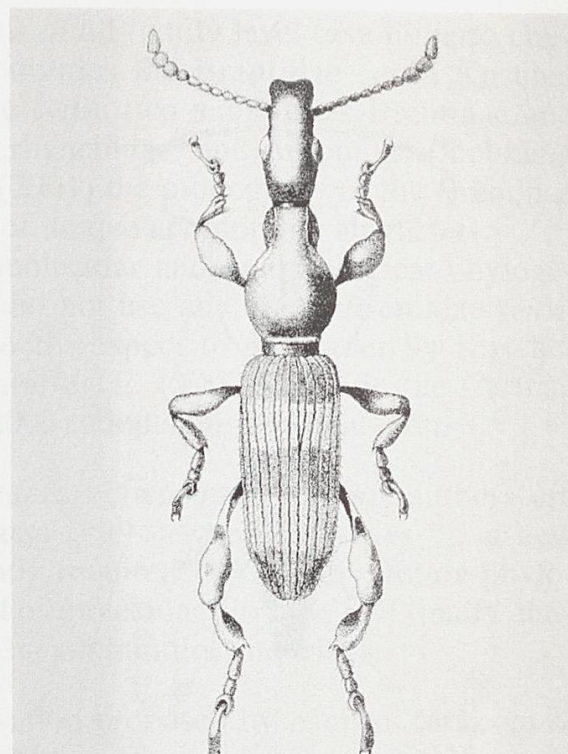


Fig. 163

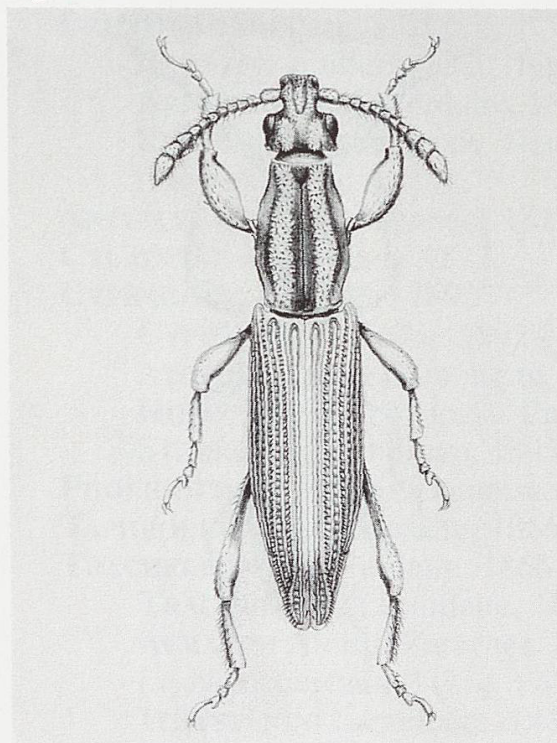


Fig. 164

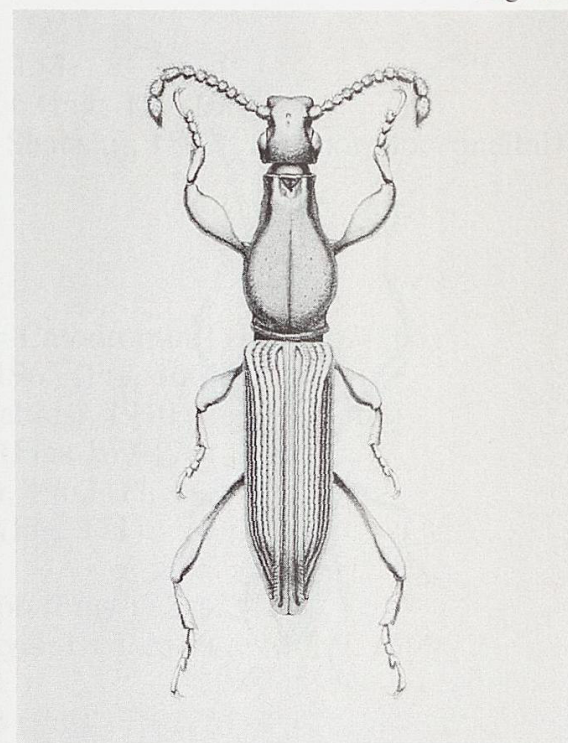


Fig. 165

Figs 162-165 - Fig. 162 - *Hemicordus guineensis* (Calabresi) (Eremoxenini) (drawing by E. Calabresi); Fig. 163 - *Cyphagogus bimaculatus* Damoiseau (Cyphagogini) (from Damoiseau, 1989a); Fig. 164 - *Megalosebus fallaciosus* Kolbe (Cyphagogini) (drawing by E. Calabresi); Fig. 165 - *Podozemius mustus* Kolbe (Cyphagogini) (drawing by E. Calabresi).

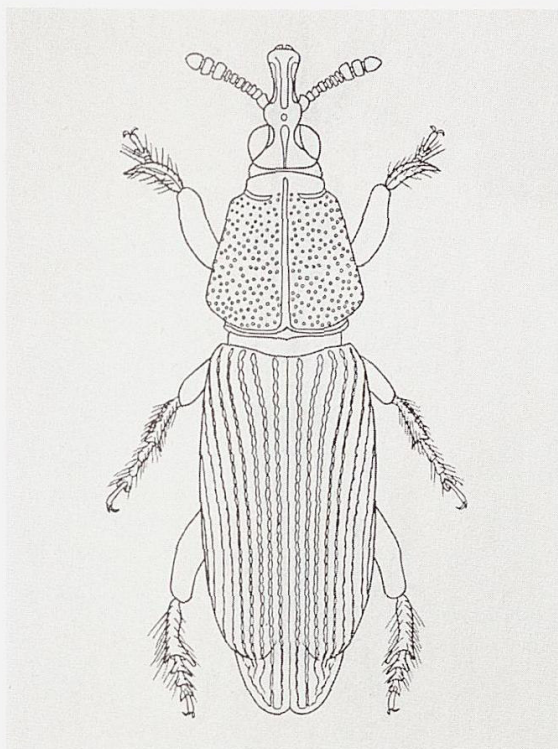


Fig. 166

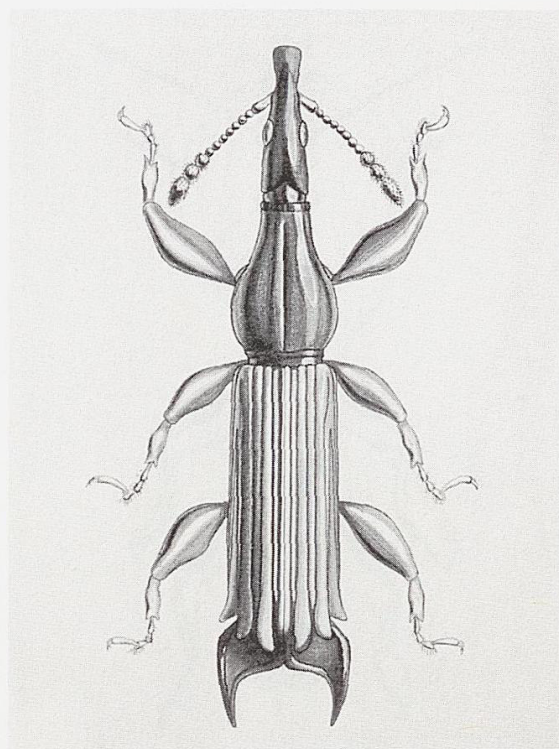


Fig. 167

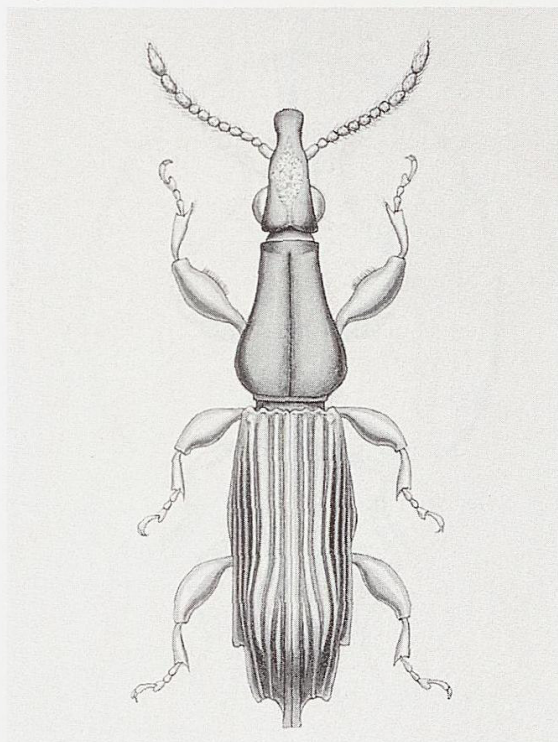


Fig. 168

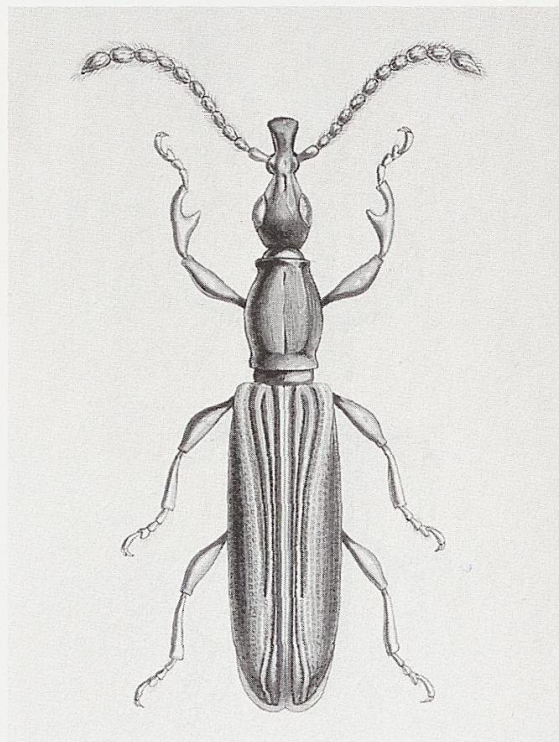


Fig. 169

Figs 166-169 - Fig. 166 - *Atopobrentus allardi* Damoiseau (Atopobrentini) (from Damoiseau, 1967a); Fig. 167 - *Carcinopisthius doriae* (Senna) (Hoplopisthiini) (drawing by E. Calabresi); Fig. 168 - *Hoplopisthius trichemerus* (Hoplopisthiini) (drawing by E. Calabresi); Fig. 169 - *Cerobates* (s. s.) *formosanus* Schönfeldt (Stereodermini) (drawing by E. Calabresi).

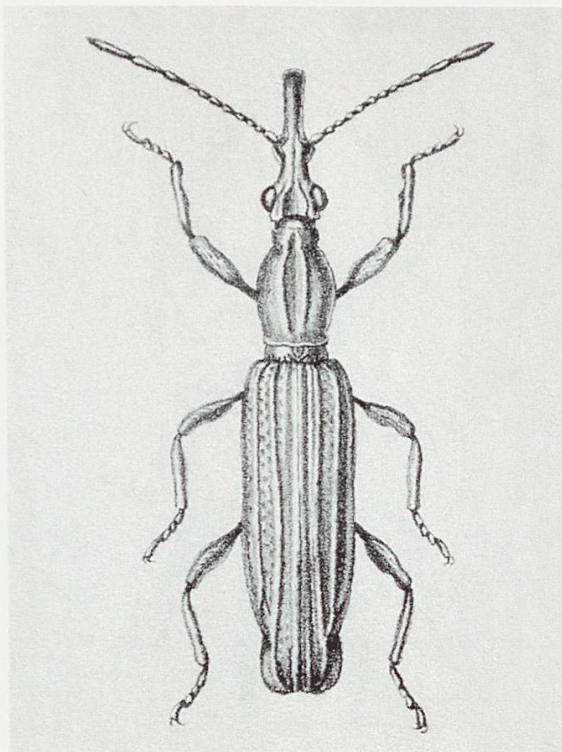


Fig. 170

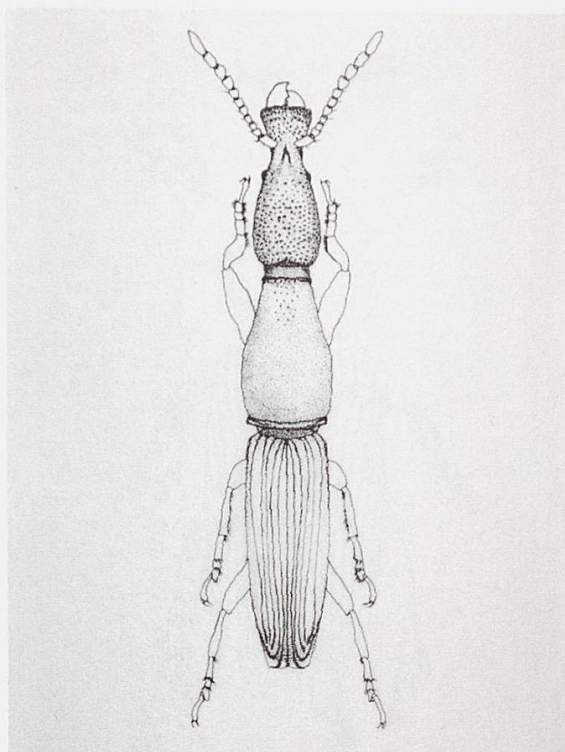


Fig. 171

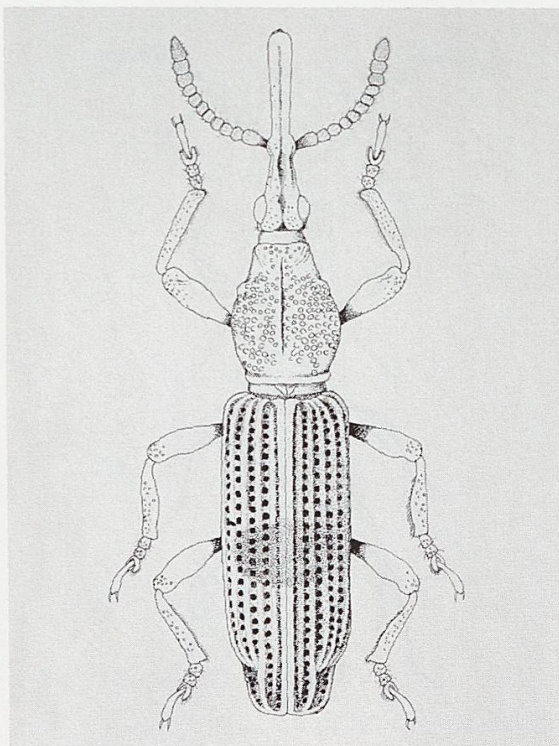


Fig. 172

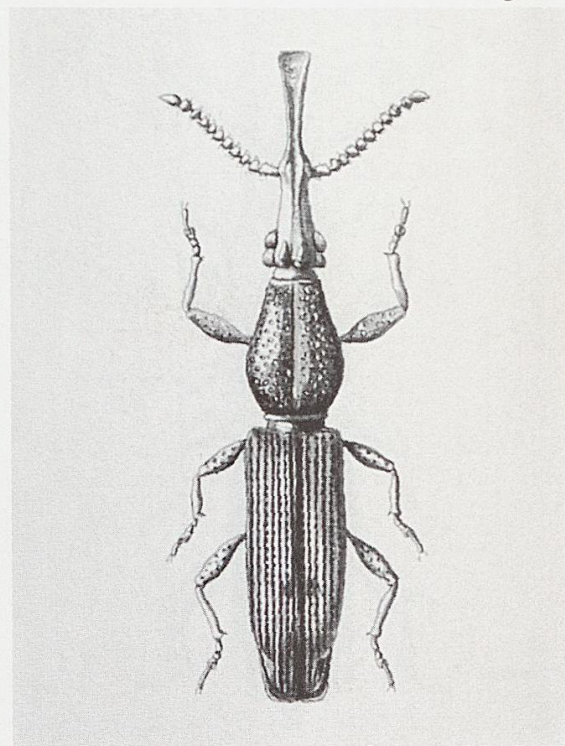


Fig. 173

Figs 170-173 - Fig. 170 - *Stereobates pedator* Sharp (Stereodermini) (from Sharp, 1895); Fig. 171 - *Plesiobolbus sagax* Kolbe (Taphroderinae) (from Sforzi, 1998b); Fig. 172 - *Hypomiolispa grandis* Damoiseau (Trachelizini) (from Damoiseau, 1989b); Fig. 173 - *Trachelizus modestus* Senna (Trachelizini) (drawing by E. Calabresi).

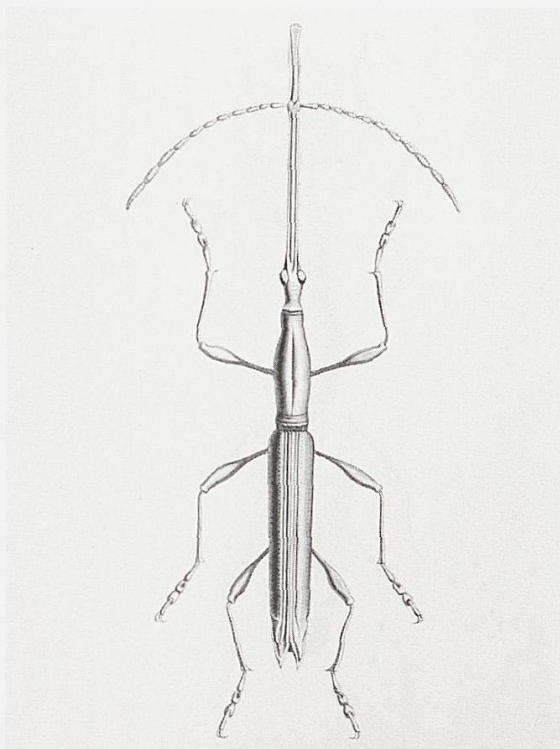


Fig. 174

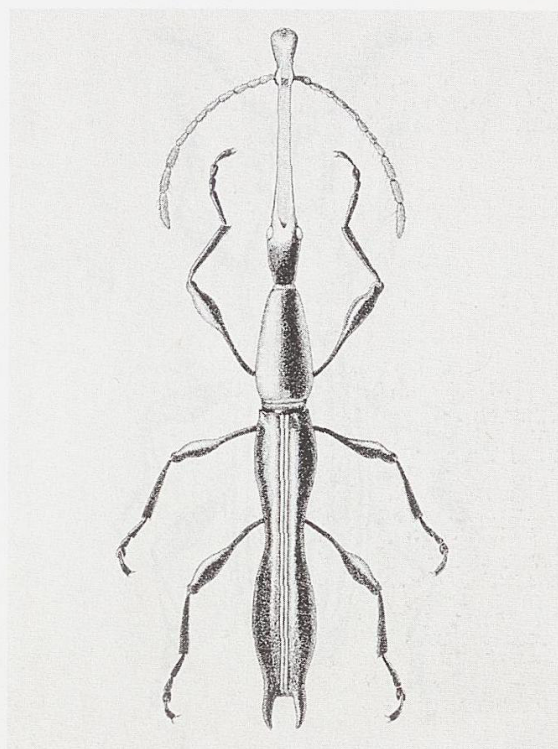


Fig. 175

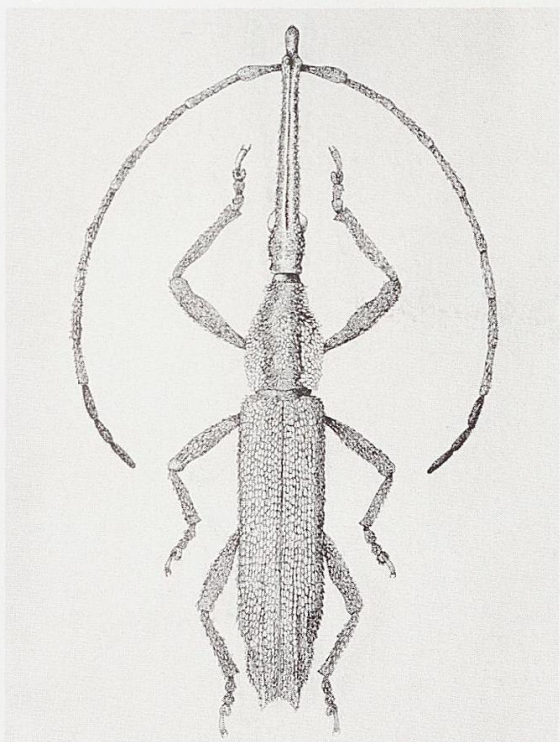


Fig. 176

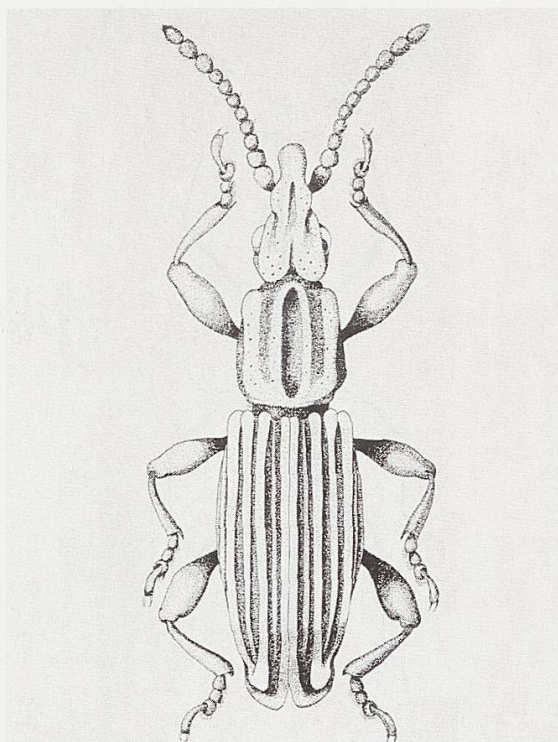


Fig. 177

Figs 174-177 - Fig. 174 - *Teramocerus* sp. (Acratini) (drawing by E. Calabresi); Fig. 175 - *Bulbogaster ctenostomoides* Lacordaire (Ithystenini) (from Damoiseau, 1989b); Fig. 176 - *Ceocephalus philippinicus* (Senna) (Ithystenini) (from Damoiseau, 1989b); Fig. 177 - *Higonius trisulcatus* Damoiseau (Microtrachelizini) (from Damoiseau, 1987).

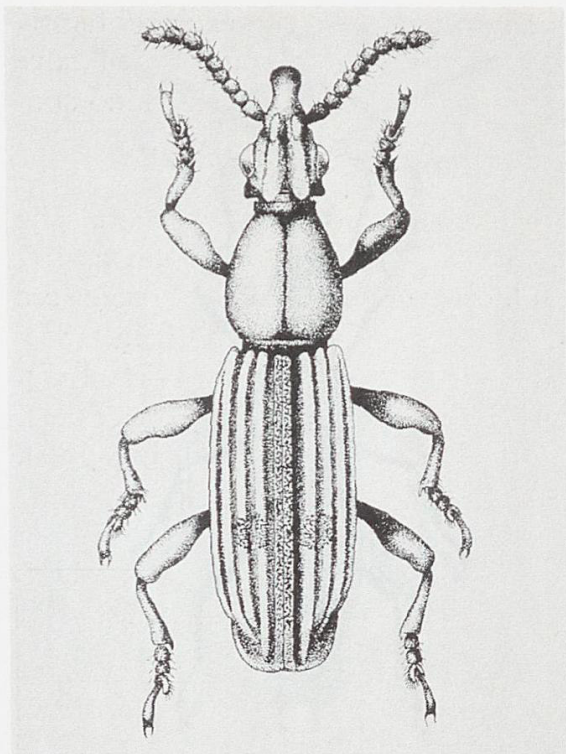


Fig. 178

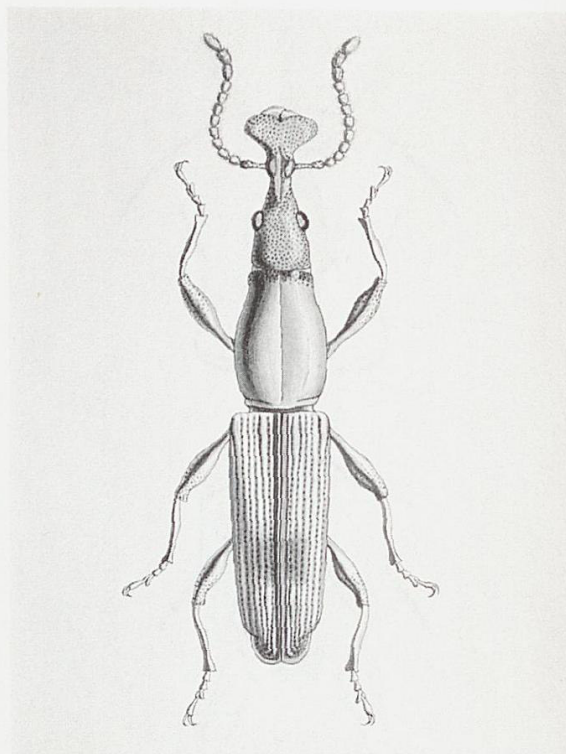


Fig. 179

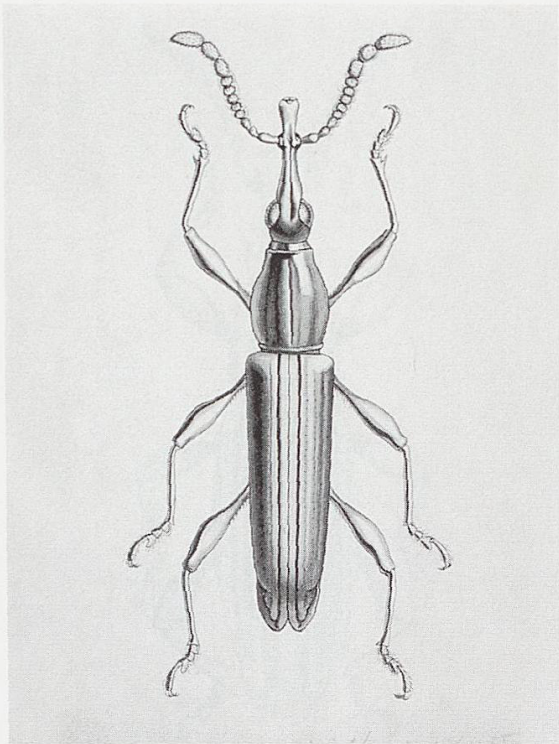


Fig. 180

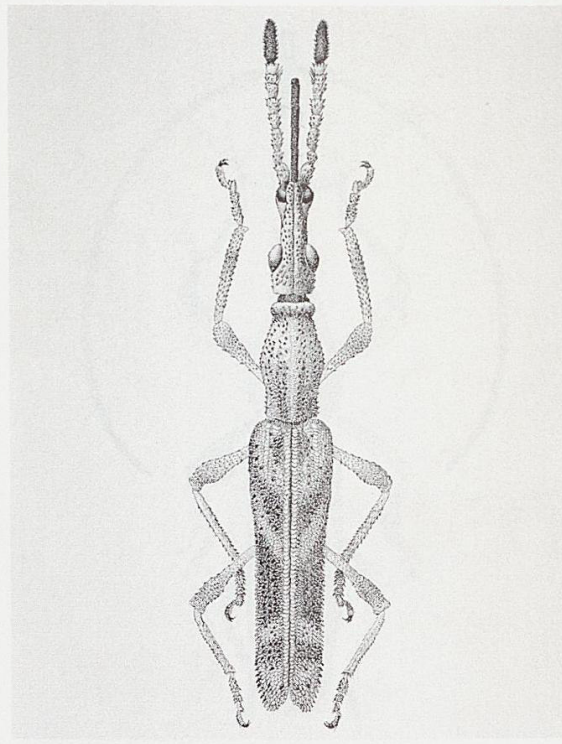


Fig. 181

Figs 178-181 - Fig. 178 - *Higonodes novenarius* (Damoiseau) (Microtrachelizini) (from Zimmerman, 1994); Fig. 179 - *Heterothesis elegans* Kleine (Pseudoceocephalini) (drawing by E. Calabresi); Fig. 180 - *Proephebocerus populeus* (Boheman) (Pseudoceocephalini) (drawing by E. Calabresi); Fig. 181 - *Ulocerus amazonicus* Damoiseau (Ulocerinae) (from Damoiseau, 1962b).

We believe that illustrating at least one representative from each subfamily and tribe will assist identification and classification. For this purpose we have employed many unpublished drawings by Enrica Calabresi, with the kind concession of her family.

The 1690 currently known species of Brentidae belong to 293 genera. The percentages of the subfamilies are given in Fig. 182. There are as many as 107 monospecific genera, i. e. 37.4% of the total. The richest genera in species are *Miolispa* (73 species) (Figs 183, 184), *Cyphagogus* (64 species) (Fig. 185), *Cerobates* (60 species) (Fig. 186), *Hypomiolispa* (46 species) (Fig. 187), *Ithystenus* (44 species) (Fig. 188), *Estenorhinus* (42 species) (Fig. 68, Pl. 1), *Brentus* (38 species) (Figs 66, 158), *Microtrachelizus* (38 species) (Fig. 189), and *Tracheloschizus* (38 species) (Fig. 190).

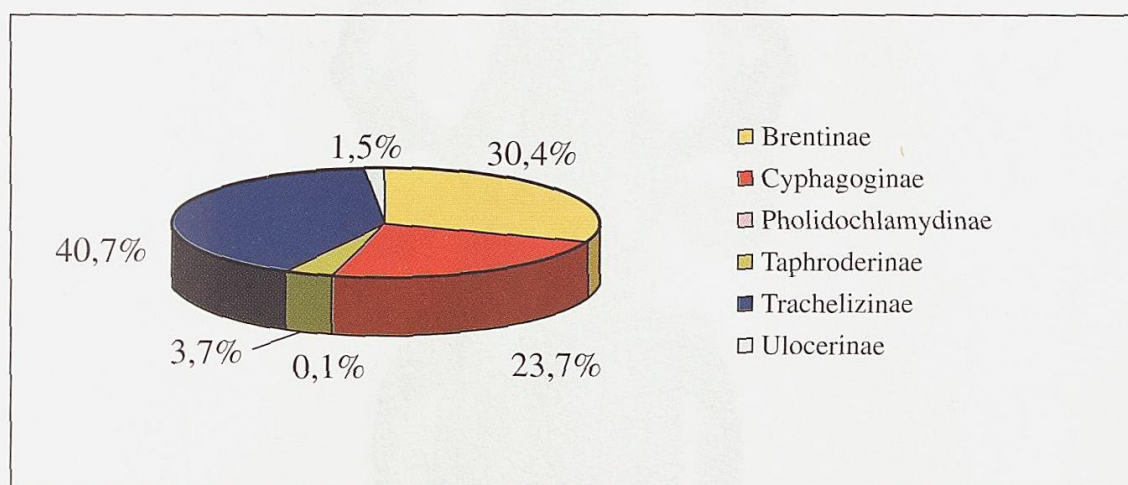


Fig. 182 - Percentages of the brentid subfamilies within the Brentidae (*sensu Auctorum*).



Fig. 183 - *Miolispa suturalis* Pascoe (photo by P. Magrini).



Fig. 184 - *Miolispa salomonensis* Senna (photo by P. Magrini).



Fig. 185 - *Cyphagogus planifrons* Kirsch (photo by P. Magrini).



Fig. 186 - *Cerobates (s. s.) sulcatus sulcatus* Boheman (photo by P. Magrini).

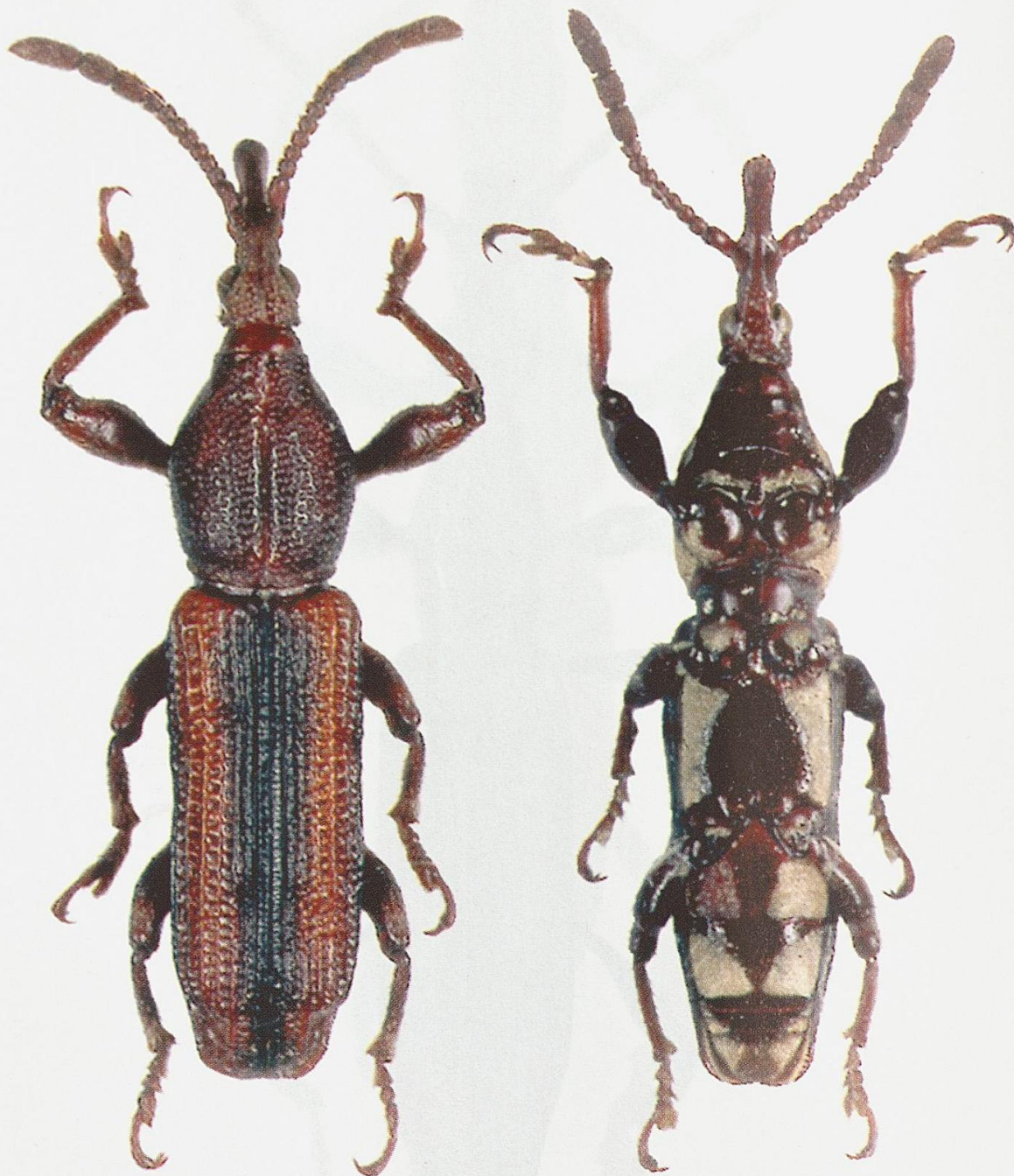


Fig. 187 - *Hypomiolispa exarata* (Desbrochers des Loges), dorsal and ventral view (photo by P. Margrini).

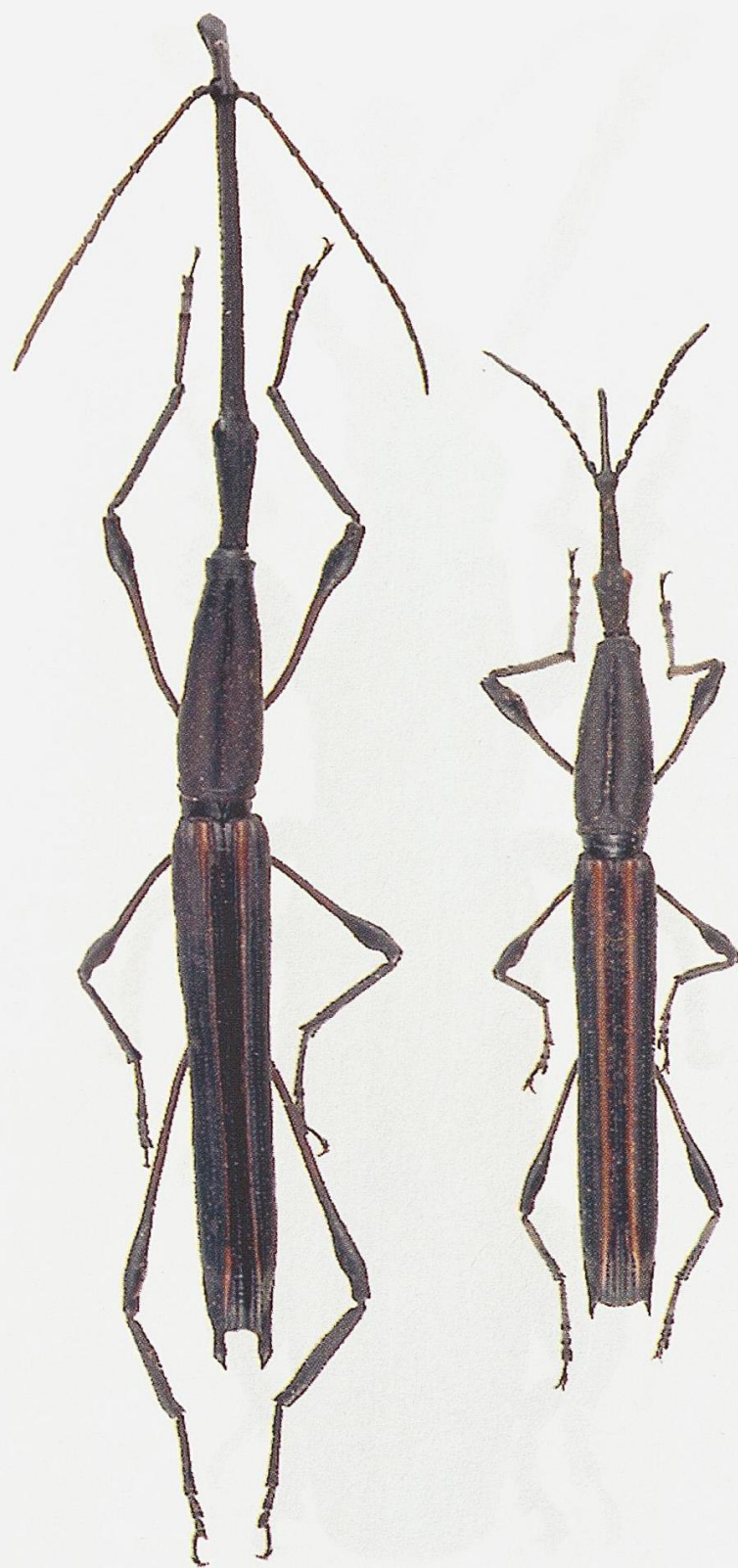


Fig. 188 - *Ithystenus* sp. (photo by S. Bambi).



Fig. 189 - *Microtrachelizus aethiopicus* Calabresi (photo by P. Magrini).



Fig. 190 - *Tracheloschizus gloriosus* Damoiseau (Paratype) (photo by P. Magrini).

COMPILATION OF THE CHECKLIST: METHODS AND CONVENTIONS EMPLOYED

The systematic order followed in this checklist is that proposed by Alonso-Zarazaga & Lyal (1999) for the sub-families, tribes and genera. The species in each genus have been listed in alphabetical order. The following information is given for each species: author, year, and page of description, synonyms, misspellings, type depositories, geographical distribution and any notes.

When a species is now included in a different genus from the one in which it was first described, the original genus is given in brackets. For synonyms, we have given the author of the synonymy, specifying the year and page. In the case of a *lapsus*, we have only given the author who used it for the first time, specifying the year and page, plus the genus in which the species was included in brackets. In compiling this checklist, we have not considered the *nomina nuda* and the names *in litteris*. Given the great intraspecific variability in brentids, we have considered varieties, forms and races as synonyms of the nominal species.

Type depositories and the kind of type are indicated by abbreviations as listed.

Some authors (Damoiseau, Haedo Rossi and Meyer) have sometimes described a specimen of the opposite sex to the Holotype of a known species as an 'Allotype'. Obviously these "Allotypes" are not valid and therefore we have not included them in the Checklist.

If the type material is followed by the symbol "!", it means that we examined it personally. We examined types deposited in the following museums: Deutsches Entomologisches Institut, Eberswalde Finow, Germany; Institut Royal des Sciences Naturelles de Belgique, Brussels, Belgium; Museum National d'Histoire Naturelle, Paris, France; Museum für Naturkunde der Humboldt-Universität, Berlin, Germany; Musée Royal de l'Afrique Centrale, Tervuren, Belgium; Museo Regionale di Scienze Naturali, Torino, Italy; Museo Civico di Storia Naturale "Giacomo Doria", Genova, Italy; Museo di Storia Naturale, Sezione di Zoologia "La Specola", Università di Firenze, Italy; The Natural History Museum, London, United Kingdom; Naturhistorisches Museum, Basel, Switzerland; Oxford University Museum of Natural History, Oxford, Great Britain; Transvaal Museum, Pretoria, South Africa; Zoologisch Museum, Universiteit van Amsterdam, Netherlands.

In other cases we were able to verify the presence of type material thanks to personal communication from the curators or by consulting printed and on-

line catalogues. Nevertheless, we preferred not to add the “!” symbol in these cases. The museums in this category are the following: California Academy of Sciences, San Francisco, California, USA; Cornell University, Ithaca, New York, USA; Instituto Nacional de Pesquisas da Amazônia, Manaus, Brazil; Museo Civico di Storia Naturale, Trieste, Italy; Museum of Comparative Zoology, University of Harvard, Cambridge, Massachusetts, USA; Martin-Luther-Universität, Wissenschaftsbereich Zoologie, Halle a. S., Germany; Zoological Museum, University of Helsinki, Finland; Museum and Institute of Zoology, Polish Academy of Science, Warsaw, Poland; Museo de la Plata, Universidad Nacional de La Plata, Argentina; Naturkundemuseum Erfurt, Germany; National Museum (Natural History), Prague, Czech Republic; South Australian Museum, Adelaide, Australia; Staatliches Museum für Tierkunde, Dresden, Germany; Zoological Institute, St. Petersburg, Russia; Zoological Museum, Uppsala University, Uppsala, Sweden; Zoological Museum, University of Copenhagen, Denmark; Zoologische Staatssammlung, Munich, Germany.

In all other cases we deduced the location of type material from the original descriptions or from Kleine's (1928a) work on brentid types.

We can now confirm that the Kleine collection, that some authors believe was destroyed during the bombardment of Stettin in the World War II, is in fact deposited in the Museum and Institute of Zoology, Polish Academy of Science, Warsaw, Poland. We also verified that types claimed to be deposited at Dehra Dun (India) and Kuala Lumpur (Malaysia) are in fact now in the Natural History Museum, London.

During our study visits to the main European Museums to compile this volume, we decided to designate lectotypes for the species for which we were able to examine the syntypes, to establish their identities. The designation of these lectotypes is very important for taxonomic purposes (article 74.7.3 of the International Code of Zoological Nomenclature, fourth Edition) considering the great individual variability in the Brentidae.

The geographical distribution is based solely on biographical data and not on unpublished museum data. The countries are divided into Zoogeographical Regions (distinguishing between the Central American and South American countries) and listed in alphabetical order. In the case of the Island of Borneo, politically divided between Malaysia and Indonesia, we preferred to call it by the generic name “Borneo” since in most cases the precise locality is not given. For the same reason we used the generic name “New Guinea” to indicate both Irian Jaya (Indonesia) and Papua New Guinea. The symbol “I” signifies that the species has been introduced; the symbol “?” signifies that its presence is doubtful.

ABBREVIATIONS

AT	Allotype
HT	Holotype
I	Imported
LT	Lectotype
NT	Neotype
PLT	Paralectotype
PT	Paratype
ST	Syntype
AMC	Antonio Martínez collection, Argentina
AMNH	American Museum of Natural History, New York, USA
AMS	Australian Museum, New South Wales, Sydney, Australia
APC	Alberto Prosen collection, Argentina
BDVC	Barry D. Valentine collection, University of Ohio, Columbus, Ohio, USA
BPBM	Bernice P. Bishop Museum, Honolulu, Hawaii, USA
CAS	California Academy of Sciences, S. Francisco, California, USA
CJSP	J. Soukup collection, Magdalena del Mar (Lima), Peru
CMNH	Carnegie Museum of Natural History, Pittsburgh, Pennsylvania, USA
CSMBA	Colegio Máximo de San Miguel, Buenos Aires, Argentina
CUIC	Cornell University, Ithaca, New York
CUMZ	University Museum, Cambridge, United Kingdom
DEI	Deutsches Entomologisches Institut, Eberswalde Finow, Germany
DSIRN	Department of Scientific and Industrial Research, Nelson, New Zealand
DSIRW	Department of Scientific and Industrial Research, Wellington, New Zealand
DZSP	Departamento de Zoologia de Secretaria de Agricultura de São Paulo, Brazil
ELKU	Entomological Laboratory, Kyushu University, Japan
EMBT	Department of Agriculture, Bangkok, Thailand
EOC	Eylon Orbach collection, Kyriat Tivon, Israel
ETHZ	Eidgenössische Technische Hochschule, Zürich, Switzerland
EUMJ	Ehime University, Matsuyama, Japan
GDFPNG	Government Department of Forests, Boroko, National Capital District, Papua New Guinea
HNHM	Hungarian Natural History Museum, Budapest, Hungary
IBSP	Instituto Biológico, São Paulo, Brazil
IERTA	Institut d'Enseignement de Recherches Tropicales, Adiopoumé, Ivory Coast
IFRI	Indian Forest Research Institute, Dehra Dun, Uttar Pradesh, India
IMLA	Fundacion e Instituto Miguel Lillo, Universidad Nacional de Tucuman, Argentina
IOC	Instituto Oswaldo Cruz, Rio de Janeiro, Brazil
INPA	Instituto Nacional de Pesquisas da Amazônia, Manaus, Brazil
IPEACS	Instituto de Pesquisas e Experimentação Agropecuárias do Centro-Sul, Rio de Janeiro, Brazil
IPEACSM	Instituto de Pesquisas e Experimentação Agropecuárias do Centro-Sul, Manicoré, Amazonas, Brazil

IRSNB	Institut Royal des Sciences Naturelles de Belgique, Brussels, Belgium
IZAS	Institute of Zoology, Academia Sinica, Beijing, China
JHRC	José A. Haedo Rossi collection, Tucuman, Argentina
MACN	Museo Argentino de Ciencias Naturales, Buenos Aires, Argentina
MAPA	Museu Anchieta de Ciencias Naturais, Porto Alegre, Brazil
MBUC	Universidad Central de Venezuela, Caracas, Venezuela
MCCV	Museo de Ciencias, Caracas, Venezuela
MCST	Museo Civico di Storia Naturale, Trieste, Italy
MCZ	Museum of Comparative Zoology, University of Harvard, Cambridge, Massachusetts, USA
MEFP	Museu Entomológico Dr. Fritz Plaumann, Seara, Santa Catarina, Brazil
MHNG	Muséum d'Histoire Naturelle, Geneva, Switzerland
MLPA	Museo de la Plata, Universidad Nacional de La Plata, Argentina
MLUH	Martin-Luther-Universität, Wissenschaftsbereich Zoologie, Halle a. S, Germany
MMA	Melbourne Museum, Victoria, Australia
MMUS	Macleay Museum, University of Sydney, Sydney, Australia
MNHC	Museo Nacional de Historia Natural, Habana, Cuba
MNHLB	University of the Philippines at Los Baños, Museum of Natural History, Los Baños, Philippines
MNHN	Museum National d'Histoire Naturelle, Paris, France
MNHUB	Museum für Naturkunde der Humboldt-Universität, Berlin, Germany
MNRJ	Museu Nacional, Universidade Federal do Rio de Janeiro, Brazil
MNUL	Museum für Natur und Umwelt, Lübeck, Germany
MONZ	Museum of New Zealand, Te Papa Tongarewa, Wellington, New Zealand
MPEG	Museu Paraense Emilio Goeldi, Belém, Pará, Brazil
MPMP	National Museum of the Philippines, Manila, Philippines
MRAC	Musée Royal de l'Afrique Centrale, Tervuren, Belgium
MRSN	Museo Regionale di Scienze Naturali, Torino, Italy
MSNG	Museo Civico di Storia Naturale "Giacomo Doria", Genova, Italy
MVMA	Museum of Victoria, Abbotsford, Victoria, Australia
MZB	Museum Zoologicum Bogoriense, Bogor, Indonesia
MZHF	Zoological Museum, University of Helsinki, Finland
MZLU	Lund University, Lund, Sweden
MZPW	Museum and Institute of Zoology, Polish Academy of Science, Warsaw, Poland [= Museum Zoologicum Polonicum Warszawa]
MZSF	Museum Zoologique, Université de Strasbourg, France
MZSP	Museu de Zoologia da Universidade de São Paulo, São Paulo, Brazil
MZUF	Museo di Storia Naturale, Sezione di Zoologia "La Specola", Università di Firenze, Italy
MWIC	Museum Wasmannianum Ignatius College, Valkenburg, Germany
NHM	The Natural History Museum, London, United Kingdom
NHMB	Naturhistorisches Museum, Basel, Switzerland
NHRS	Naturhistoriska Riksmuseet, Stockholm, Sweden
NME	Naturkundemuseum Erfurt, Germany
NMNH	National Museum of Natural History, Washington D. C., USA

NMPC	National Museum (Natural History), Prague, Czech Republic
NMW	Naturhistorisches Museum Wien, Austria
NZSI	National Zoological Collection, Zoological Survey of India, Calcutta, India
OXUM	Oxford University Museum of Natural History, Oxford, Great Britain
PBZT	Parc Botanique et Zoologique de Tsimbazaza, Antananarivo, Madagascar
QMA	Queensland Museum, Brisbane, Queensland, Australia
RMNH	Nationaal Natuurhistorische Museum [formerly Rijksmuseum van Natuurlijke Historie], Leiden, Netherlands
SAMA	South Australian Museum, Adelaide, Australia
SEHU	Systematic Entomology, The Hokkaido University Museum, Hokkaido University, Sapporo, Japan
SMFD	Forschungsinstitut und Naturmuseum Senckenberg, Frankfurt-am-Main, Germany
SMKM	Selangor Museum, Kuala Lumpur, Malaysia
SMTD	Staatliches Museum für Tierkunde, Dresden, Germany
TARI	Taiwan Agricultural Research Institute, Taichung Hsien, Taiwan, Republic of China
TMSA	Transvaal Museum, Pretoria, South Africa
UQDZ	The University of Queensland, Department of Zoology and Entomology, Brisbane, Queensland, Australia
URMU	Museo Nacional de Historia Natural, Montevideo, Uruguay
UZI	Zoological Museum, Uppsala University, Uppsala, Sweden
WSC	Wolfgang Schedl collection, Innsbruck, Austria
ZFMK	Zoologische Forschungsinstitut und Museum "Alexander Koenig", Bonn, Germany
ZIN	Zoological Institute, St. Petersburg, Russia
ZMAN	Zoologisch Museum, Universiteit van Amsterdam, Netherlands
ZMUC	Zoological Museum, University of Copenhagen, Denmark
ZMUH	Zoologisches Institut und Zoologisches Museum, Universität von Hamburg, Germany
ZMUK	Zoologisches Museum, Universität Kiel, Germany
ZMUM	Zoological Museum of the Moscow State University, Moscow, Russia
ZRC	Zoological Reference Collection, Raffles Museum of Biodiversity Research, National University of Singapore, Singapore
ZSM	Zoologische Staatssammlung, Munich, Germany

CHECKLIST

BRENTIDAE Billberg, 1820

- Brenthides Billberg, 1820: 40.
 Brentusidae: Fleming, 1821: 51 [lapsus].
 Brenthoidae: Agassiz, 1846b: 53 [lapsus].
 Brenthidae: Imhoff, 1856: XVIII [lapsus].
 Brenthitae: Thomson, 1858: 116 [lapsus].
 Brenthi: Kolbe, 1883c: 75 [lapsus].
 Brenthii: Acloque, 1896: 351 [lapsus].
 Brenthinidae: Schönfeldt, 1908: 50 [lapsus].
 Brentoidea Pierce, 1916: 462.
 Brenthoidea: Ienistea, 1986: 32 [lapsus].
 Brenusidae: Alonso-Zarazaga & Lyal, 1999: 47 [lapsus].

BRENTINAE Billberg, 1820

- Brenthinae: Leconte & Horn, 1883: 501 [lapsus].
 Brenthina: Sharp, 1895a: 63 [lapsus].

BRENTINI Billberg, 1820

- Brenthini: Imhoff, 1856: XVIII [lapsus].

Brentus Fabricius, 1787

- Brentus* Fabricius, 1787: 95.
Brenthus Illiger, 1801: 130; syn. Schoenherr, 1833: 342.
Nabrenthus Kleine, 1929a: 51 [not available, no description].
Brentis: Zherikhin & Gratshev, 1995: 637 [lapsus].

Type species: *Curculio anchorago* Linné, 1758.

Distribution. USA (California, Florida) south to Argentina; West Indies; Polynesia (Marquesas, Tahiti) (I?).

anchorago (Linné, 1758)

anchorago Linné, 1758: 383 (*Curculio*).

longicollis De Geer, 1775: 274 (*Curculio*); syn. Schoenherr, 1833: 344.

anchoraco: De Geer, 1775: 274 (*Curculio*) [lapsus].

hastile Pallas, 1781: 25 (*Curculio*); syn. Schoenherr, 1833: 344.

canaliculatus Olivier, 1790: 191 (*Brentus*); syn. Schoenherr, 1833: 344.

peregrinus Herbst, 1797: 190 (*Brentus*); syn. Schoenherr, 1833: 344.

anchora: Schoenherr, 1826: 69 (*Brenthus*) [lapsus].

lucanus Horn, 1872: 129 (*Brenthus*); syn. Kleine, 1922l: 91.

severa Senna, 1890: 108 (*Brenthus anchorago* var.); syn. Schönfeldt, 1908: 52.

lutans: Beyer, 1904: 168 (*Brenthus*) [lapsus].

anchorage: Tanner, 1927: 50 (*Brentus*) [lapsus].

hastilis: Kleine, 1938d: 173 (*Brenthus*) [lapsus].

Type depositories. *C. anchorago*: UZIU (?); *C. longicollis*: unknown; *C. hastile*: unknown; *B. canaliculatus*: unknown; *B. peregrinus*: MNHUB (Herbst coll.) (?) (HT); *B. lucanus*: MCZ (ST); *B. anchorago* var. *severa*: unknown.

Distribution. USA (California, Florida) to Paraguay.

Notes. We did not find the type for *B. peregrinus* in MNHUB where the Herbst collection is held.

In the collection of Senna (MZUF) there are no specimens corresponding to the description of the *severa* variety.

approximatus Erichson, 1847

approximatus Erichson, 1847: 126 (*Brenthus*).

unidentatus Perroud, 1853: 430 (*Brenthus*); syn. Kleine, 1922l: 90.

unidentatus Kirsch, 1868: 221 (non Perroud, 1853) (*Brenthus*); syn. Kleine, 1922l: 90.

multilineatus Sharp, 1895a: 69 (*Brenthus*); syn. Kleine, 1922l: 90.

Type depositories. *B. approximatus*: MNHUB (HT!); *B. unidentatus* Perroud: MNHN (LT!, PLT!), *B. unidentatus* Kirsch: SMTD (ST); *B. multilineatus*: NHM (LT!, PLT!).

Distribution. Costa Rica, El Salvador, Guatemala, Mexico, Panama; Argentina, Bolivia, Colombia, Ecuador, Guyana, Peru, Venezuela.

Notes. Lectotype of *B. unidentatus* Perroud (MNHN) designated on a male specimen with the following labels: Venezuela, Caracas. Paralectotype: 1 female specimen (MNHN) with no label.

Lectotype of *B. multilineatus* (NHM) designated on a male specimen with the following labels: Cubilguitz, Verapaz, Champion/ B. C. A., coll. IV, 6. *Brenthus multilineatus* Sharp/ *Brenthus multilineatus* type D. S. Paralectotypes: 1 male (NHM), B. C. A., coll. IV, 6. *Brenthus multilineatus* Sharp/ Sharp coll. 1905-313/ *Brenthus multilineatus* mihi, Bogotá; 1 male (NHM), Chiriqui, Ribbe, *Brenthus multilineatus* D. S./ B. C. A. coll. IV, 6. *Brenthus multilineatus* Sharp/ Sharp coll. 1905-313.

armiger Herbst, 1797

armiger Herbst, 1797: 196.

interruptus Herbst, 1797: 197 (*Brenthus*); syn. Schoenherr, 1833: 346.

canaliculatus Lund, 1800: 78 (non Olivier, 1790) (*Brenthus*); syn. Schoenherr, 1833: 346.

insubidus Kirsch, 1868: 221 (*Brenthus canaliculatus* var.); syn. Gemminger & Harold, 1872: 2713.

championi Sharp, 1895a: 65 (*Brenthus*); syn. Kleine, 1927a: 75.

Type depositories. *B. armiger*: MNHUB (Herbst coll.) (?) (HT); *B. interruptus*: MNHUB (Herbst coll.) (?) (HT); *B. canaliculatus*: ZMUC (ST); *B. canaliculatus* var. *insubidus*: SMTD (ST); *B. championi*: NHM (HT!).

Distribution. "Antilles", Costa Rica, Belize, El Salvador, Guatemala, Honduras, Mexico, Nicaragua, Panama, Trinidad; Argentina, Bolivia, Brazil, Colombia, Ecuador, Guiana, Paraguay, Peru, Suriname, Venezuela.

Notes. We did not find any types of *B. armiger* and *B. interruptus* in MNHUB where the Herbst collection is held.

armillatus Kirsch, 1868

armillatus Kirsch, 1868: 219 (*Brenthus*).

erythromerus Kirsch, 1868: 220 (*Brenthus armillatus* var.); syn. n.

Type depositories. *B. armillatus*: SMTD (ST); *B. armillatus* var. *erythromerus*: unknown.

Distribution. Colombia, Peru, Venezuela.

calcar Lund, 1800

calcar Lund, 1800: 84.

Type depository. ZMUC (ST).

Distribution. Brazil, French Guyana, Paraguay, Peru.

caudatus Herbst, 1797

caudatus Herbst, 1797: 201.

caudatus Olivier, 1807: 440 (non Herbst, 1797) (*Curculio*); syn. Schoenherr, 1833: 343.

deplanatus Gyllenhal (in Schoenherr, 1840: 528) (*Brenthus*); syn. Gemminger & Harold, 1872: 2713.

Type depositories. *B. caudatus*: MNHUB (Herbst coll.) (?) (HT); *B. deplanatus*: NHRS (ST).

Distribution. Costa Rica, Guatemala, Mexico, Nicaragua, Panama; Argentina, Bolivia, Brazil, Colombia, Ecuador, Paraguay, Peru, Suriname.

Notes. We did not find the type of *B. caudatus* in MNHUB, where the Herbst collection is held.

clavipes Sharp, 1895

clavipes Sharp, 1895a: 69 (*Brenthus*).

Type depository. NHM (LT!, PLT!).

Distribution. Guatemala, Costa Rica, Panama.

Notes. Lectotype of *B. clavipes* (NHM) designated on a male specimen with the following labels: Bugaba, 800-1,500 ft. Champion/ *Brenthus clavipes* Types D. S./ Syntype/ B. C. A., coll. IV, 6. *Brenthus clavipes* Sharp. Paralectotypes: 1 female specimen (NHM) [glued to the same label as the lectotype], with the same label data; 1 male specimen (NHM), Tolé, Panama, Champion/ syntype/ B. C. A., coll. IV, 6. *Brenthus clavipes* Sharp; 2 male and 3 female specimens (NHM), Bugaba, Panama, Champion/ B. C. A., coll. IV, 6. *Brenthus clavipes* Sharp; 1 female specimen (NHM), V. de Chiriqui, 24-4000 ft., Champion/ B. C. A., coll. IV, 6. *Brenthus clavipes* Sharp; 2 female specimens (NHM), V. de Chiriqui, 3-4000 ft., Champion/ B. C. A., coll. IV, 6. *Brenthus clavipes* Sharp.

consentaneus Perroud, 1853

consentaneus Perroud, 1853: 434 (*Brenthus*).

Type depository. MNHN (LT!, PLT!).

Distribution. Costa Rica; Brazil, Colombia, French Guyana, Peru.

Notes. Lectotype of *B. consentaneus* (MNHN) designated on a male specimen with the following label: Cayenne. Paralectotype: 1 female specimen (MNHN), with no label.

cylindrus Lund, 1800

cylindrus Lund, 1800: 77.

bidentatus Lund, 1800: 80 (*Brentus*); syn. Schönfeldt, 1908: 53.

bicalcaratus Boheman (in Schoenherr, 1840: 534) (*Brenthus*); syn. Kleine, 1922l: 90.

capulus Boheman (in Schoenherr, 1840: 536) (*Brenthus*); syn. Kleine, 1927a: 76.

bicarinatus: Schönfeldt, 1908: 52 (*Brenthus*) [lapsus].

efferatus Kleine, 1927b: 473 (*Brenthus*); syn. Zimmerman, 1994: 18.

cylindricus: Kleine, 1927b: 475 (*Brenthus*) [lapsus].

effrenatus: Kleine, 1923g: 622 (*Brenthus*) [lapsus].

Type depositories. *B. cylindrus*: ZMUC (ST), ZMUK (ST); *B. bidentatus*: ZMUC (ST); *B. bicalcaratus*: MNHN (HT!); *B. capulus*: NHRS (HT); *B. efferatus*: NHM (LT!, PLT!).

Distribution. Costa Rica, Cuba, Guatemala, Trinidad and Tobago (Trinidad); Argentina (?), Brazil, Chile, Colombia, Ecuador, Paraguay, Peru, Venezuela; Polynesia (Marquesas, Tahiti) (I?).

Notes. Lectotype of *B. efferatus* (NHM) designated on a male specimen with the following labels: syntype/ Nuka Hiva Isl., Marquesas Isls., Walker/ Sharp coll., 1905-313. Paralectotypes: 7 male and 7 female specimens (NHM), syntype/ Nuka Hiva Isl., Marquesas Isls., Walker/ Sharp coll., 1905-313.

The synonymy stated by Zimmerman (1994: 18) between *B. cylindrus* and *B. efferatus* is not clear and requires further study.

Following Damoiseau (1966f: 308) the species does not occur in Argentina.

difficilis Boheman, 1840

difficilis Boheman (in Schoenherr, 1840: 530) (*Brenthus*).

Type depository. NHRS (ST).

Distribution. Costa Rica, Mexico; Brazil, Colombia.

dignus Kleine, 1943

dignus Kleine, 1943b: 143 (*Brenthus*).

Type depository. HNHM (HT).

Distribution. Peru.

doellojuradoi Haedo Rossi, 1948

doellojuradoi Haedo Rossi, 1948: 158 (*Brenthus*).

Type depositories. MLPA (HT), IMLA (PT).

Distribution. Argentina, Bolivia, Brazil, Ecuador, Paraguay, Uruguay.

exoptatus Kleine, 1927

exoptatus Kleine, 1927b: 477 (*Brenthus*).

Type depository. NHM (HT!).

Distribution. Colombia.

extrinsecus Kleine, 1927

extrinsecus Kleine, 1927b: 476 (*Brenthus*).

Type depository. NHM (LT!, PLT!).

Distribution. Bahamas Is., Cuba.

Notes. Lectotype of *B. extrinsecus* (NHM) designated on a male specimen with the following labels: syntype/ 171/ Cuba, Dupont/ Bowring 63 47/ *Brenthus extrinsecus* Kln ♂ typus, 1923, R. Kleine det. Paralectotype: 1 female specimen (NHM), syntype/ Bahamas, J. L. Bonhote, 1902-299/ *Brenthus extrinsecus* Kln ♀ typus, 1923, R. Kleine det.

festae Griffini, 1899

festae Griffini, 1899: 2 (*Brenthus*).

Type depository. MRSN (HT).

Distribution. Ecuador.

festivus Senna, 1897

festivus Senna, 1897a: 232 (*Brenthus*).

Type depository. MZUF (HT!, PT!).

Distribution. Panama; Brazil, Colombia, French Guyana, Peru, Venezuela.

finitimus Kleine, 1927

finitimus Kleine, 1927b: 475 (*Brenthus*).

Type depository. NHM (LT!, PLT!).

Distribution. Colombia, Guiana, Paraguay.

Notes. Lectotype of *B. finitimus* (NHM) designated on a male specimen with the following labels: Demerara Summer, 1878/ syntype/ Sharp coll. 1905-313/ *B. finitimus* typus! Kln., 1923 R. Kleine det. Paralectotype: 1 male specimen (NHM), Demerara Summer, 1878/ syntype/ Sharp coll. 1905-313.

firmus Kleine, 1927

firmus Kleine, 1927b: 478 (*Brenthus*).

Type depository. SMTD (HT).

Distribution. Costa Rica.

forcipitigerus (Gyllenhal, 1840)

forcipitigerus Gyllenhal (in Schoenherr, 1840: 478) (*Arrhenodes*).

forcipitiger: Denier, 1922: 23 (*Estenorrhinus*) [lapsus].

Type depository. IRSNB (NT!).

Distribution. Cuba, Dominican Republic.

Notes. Kleine did not include this species in his *Genera Insectorum* (1938d). Damoiseau (1989b: 62) designated the neotype.

Blackwelder (1947: 773; 1982: 773) included this species in the genus *Estenorrhinus*.

guatemalenus Sharp, 1895

guatemalenus Sharp, 1895a: 68 (*Brenthus*).

chiriquensis Sharp, 1895a: 68 (*Brenthus*); syn. Kleine, 1922l: 91.

Type depositories. *B. guatemalenus*: NHM (LT!, PLT!), IRSNB (PLT!), MZUF (PLT!), NHRS (PLT); *B. chiriquensis*: NHM (LT!, PLT!).

Distribution. Costa Rica, Guatemala, Nicaragua, Panama; Colombia.

Notes. Lectotype of *B. guatemalenus* (NHM) designated on a male specimen with the following labels: Cerro Zunil/ 4-5000 ft., Champion/ syntype/ B. C. A., coll. IV, 6. *Brenthus guatemalenus* Sharp/ Sharp coll. 1905-313. Paralectotypes: 13 male and 11 female specimens (NHM), Cerro Zunil/ 4-5000 ft., Champion/ syntype/ B. C. A., coll. IV, 6. *Brenthus guatemalenus* Sharp; 2 male and 2 female specimens (NHM), Zapote, Guatemala, G. C. Champion/ syntype/ B. C. A., coll. IV, 6. *Brenthus guatemalenus* Sharp; 3 male specimens (NHM), Pantaleon, 1700 ft., G. C. Champion/ syntype/ B. C. A., coll. IV, 6. *Brenthus guatemalenus* Sharp; 1 male specimen (NHM), Capetillo, Guatemala, G. C. Champion/ syntype/ B. C. A., coll. IV, 6. *Brenthus guatemalenus* Sharp; 1 male specimen (MZUF), Zapote, Guatemala, G. C. Champion, dono D. Sharp/ *Brenthus guatemalenus* A. S./ *Brenthus guatemalenus* Sharp, cotypus! ex-coll. Senna/ "La Specola" Firenze, 9572; 1 female specimen (MZUF), Cerro Zunil/ 4000 ft., Champion/ *Brenthus guatemalenus* Sharp, cotypus! ♀/ ex-coll. Senna/ "La Specola" Firenze, 9573; 1 male and 1 female specimen (IRSNB), Cerro Zunil/ 4000 ft., Champion/ Bio. C. Amer. Don Godman et Salvin/ *Brenthus guatemalenus*/ Paratype.

Lectotype of *B. chiriquensis* (NHM) designated on a male specimen with the following labels: V. de Chiriqui 25-4000 ft., Champion/ syntype/ B. C. A., coll. IV, 6. *Brenthus chiriquensis* Sharp. Paralectotypes: 9 male and 7 female specimens (NHM), V. de Chiriqui, 25/4000 ft., Champion/ syntype/ B. C. A., coll. IV, 6; 2 male and 2 female specimens (NHM), Bugaba, 800-1500 ft., Champion/ syntype/ B. C. A., coll. IV, 6. *Brenthus chiriquensis* Sharp; 1 male

and 3 female specimens (NHM), Chontales, Nicaragua, T. Belt/ syntype/ B. C. A., coll. IV, 6. *Brenthus chiriquensis*, Sharp; 1 female specimen (NHM), Costa Rica, Caché/ 800-1500 ft., Champion/ syntype/ B. C. A., coll. IV, 6. *Brenthus chiriquensis* Sharp; 1 female specimen (NHM), Costa Rica, Sallé coll.; 1 male specimen (NHM), Irazu, 6-7000 ft., H. Rogers/ syntype/ B. C. A., coll. IV, 6. *Brenthus chiriquensis*, Sharp.

insperatus Soares & Meyer, 1959

insperatus Soares & Meyer, 1959: 170.

Type depository. MNRJ (Campos Seabra coll.) (HT).

Distribution. Peru.

jaegeri Soares, 1959

jaegeri Soares (in Soares & Meyer, 1959: 174).

Type depository. Scivittaro coll. (Instituto Biológico, Sorocaba, Sao Paulo, Brazil) (HT).

Distribution. Brazil.

Notes. *B. jaegeri* Soares was included in a key supplied by Soares & Meyer (1959: 174). Soares never described this taxon, but the characters given in the key are sufficient to consider the species as valid.

laticornis Sharp, 1895

laticornis Sharp, 1895a: 71 (*Brenthus*).

Type depository. NHM (LT!, PLT!).

Distribution. Costa Rica, Nicaragua, Panama.

Notes. Lectotype of *B. laticornis* (NHM) designated on a female specimen with the following labels: Chontales, Janson, *Brenthus laticornis* type D. S./ syntype/ B. C. A., coll. IV, 6. *Brenthus laticornis* Sharp. Paralectotype: 1 female specimen (NHM), Bugaba, 800-1500 ft., Champion *Brenthus laticornis* D. S./ syntype/ B. C. A., coll. IV, 6. *Brenthus laticornis* Sharp.

***linearis* Olivier, 1790 stat. res.**

linearis Olivier, 1790: 192 .

anchorago Pallas, 1781: 25 (non Linné, 1758) (*Curculio*); syn. Schoenherr, 1833: 345.

volvulus Fabricius, 1792: 493 (*Brentus*); syn. Schoenherr, 1833: 345.

Type depositories. *B. linearis*: unknown; *B. volvulus*: ZMUC (ST).

Distribution. Cuba, Dominican Republic, Guadeloupe Is., Panama, Puerto Rico; Argentina.

Notes. *B. linearis* was described by Olivier in 1790, but strangely, in mentioning this species of Olivier, Fabricius (1792) changed its name to *volvulus*. All following authors (e. g. Schoenherr, 1833, Schönfeldt, 1908, Kleine, 1938d, Haedo Rossi, 1961) considered *B. linearis* as a synonym of *B. volvulus*, but - out of priority - the name *linearis* has precedence over *volvulus* and should be considered valid.

***lineicollis* Boheman, 1833**

lineicollis Boheman (in Schoenherr, 1833: 347) (*Brenthus*).

pyctes Gyllenhal (in Schoenherr, 1833: 348) (*Brenthus*); syn. Kleine, 1922l: 91.

Type depositories. *B. lineicollis*: NHRS (ST); *B. pyctes*: NHRS (HT).

Distribution. Mexico; Argentina, Brazil, Colombia, French Guyana, Paraguay, Peru, Uruguay.

***maculipennis* Senna, 1897**

maculipennis Senna, 1897a: 234 (*Brenthus*).

Type depositories. MZUF (HT!), IRSNB (?) (PT), NHM (?) (PT).

Distribution. Colombia, Ecuador, Peru.

Notes. In his original description Senna wrote that paratypes were in IRSNB and in the Tring Museum (now Tring types are in NHM). We did not find any paratype in IRSNB neither in NHM.

***mexicanus* Boheman, 1840**

mexicanus Boheman (in Schoenherr, 1840: 532) (*Brenthus*).

peninsularis Horn, 1872: 128 (*Brenthus*); syn. Kleine, 1922l: 91.

penninsularis: Horn, 1894: 359 (*Brenthus*) [lapsus].

Type depositories. *B. mexicanus*: NHRS (ST); *B. peninsularis*: MCZ (ST).

Distribution. USA (California); Belize, Costa Rica, El Salvador, Guatemala, Mexico, Nicaragua, Panama, Trinidad and Tobago (Tobago).

***peruvianus* Kleine, 1928**

peruvianus Kleine, 1928k: 226 (*Brenthus*).

Type depositories. SMFD (HT, PT), SMTD (van Emden coll.) (PT).

Distribution. Peru.

***quadrilineatus* Sharp, 1895**

quadrilineatus Sharp, 1895a: 70 (*Brenthus*).

Type depository. NHM (LT!, PLT!).

Distribution. Mexico, Nicaragua.

Notes. Lectotype of *B. quadrilineatus* (NHM) designated on a male specimen with the following labels: Toxpam, Mexico, Salle coll., *Brenthus quadrilineatus* type D. S./ B. C. A., coll. IV, 6. *Brenthus quadrilineatus* Sharp. Paralectotypes: 1 female specimen [glued to the same label of the lectotype] (NHM), with the same data as the lectotype; 1 male specimen (NHM), Bobo, Mexico, Hoege, *Brenthus quadrilineatus* type D. S./ syntype/ B. C. A., coll. IV, 6. *Brenthus quadrilineatus* Sharp; 1 female specimen (NHM), Chontales, Janson, *Brenthus quadrilineatus* type D. S./ syntype/ B. C. A., coll. IV, 6. *Brenthus quadrilineatus* Sharp.

***rufescens* Sharp, 1895**

rufescens Sharp, 1895a: 67 (*Brenthus*).

Type depository. NHM (LT!, PLT!).

Distribution. Costa Rica.

Notes. Lectotype of *B. rufescens* (NHM) designated on a male specimen with the following labels: Irazu, 6-7000 ft., H. Rogers, *Brenthus rufescens* type D. S./ syntype/ B. C. A., coll. IV, 6. *Brenthus rufescens* Sharp. Paralectotype: 1 female specimen (NHM), R. Susio, Costa Rica, H. Rogers *Brenthus rufescens* D. S./ syntype/ B. C. A., coll. IV, 6. *Brenthus rufescens* Sharp/ Sharp coll. 1905-313.

rufiventris Boheman, 1840

rufiventris Boheman (in Schoenherr, 1840: 538) (*Brenthus*).

Type depository. NHRS (ST).

Distribution. Costa Rica; Argentina, Brazil, Uruguay.

saltensis Haedo Rossi, 1948

saltensis Haedo Rossi, 1948: 153 (*Brenthus*).

Type depository. MLPA (HT, AT).

Distribution. Argentina.

sculptipennis Sharp, 1895

sculptipennis Sharp, 1895a: 70 (*Brenthus*).

Type depository. NHM (LT!, PLT!).

Distribution. Nicaragua, Panama.

Notes. Lectotype of *B. sculptipennis* (NHM) designated on a male specimen with the following labels: Chontales, Nicaragua, T. Belt, *Brenthus sculptipennis* type D. S./ syntype/ B. C. A., coll. IV, 6. *Brenthus sculptipennis* Sharp. Paralectotypes: 1 male and 1 female specimen (NHM), V. de Chiriqui, 25-4000 ft., Champion *Brenthus sculptipennis* D. S. type/ syntype/ B. C. A., coll. IV, 6. *Brenthus sculptipennis* Sharp/ Sharp coll. 1905-313; 2 male specimens (NHM), V. de Chiriqui, 25-4000 ft., Champion, *Brenthus sculptipennis* D. S./ syntype/ B. C. A., coll. IV, 6. *Brenthus sculptipennis* Sharp/ Sharp coll.

1905-313; 1 female specimen (NHM), syntype/ V. de Chiriqui, 25-4000 ft., Champion/ B. C. A., coll. IV, 6. *Brenthus sculptipennis* Sharp; 1 female specimen (NHM), V. de Chiriqui, 3-4000 ft., Champion *Brenthus sculptipennis* D. S./ syntype/ B. C. A., coll. IV, 6. *Brenthus sculptipennis* Sharp; 1 female specimen (NHM), Chontales, Nicaragua, T. Belt, B. C. A., coll. IV, 6. *Brenthus sculptipennis* Sharp.

signatus Dalman, 1823

signatus Dalman, 1823: 86.

calcar Boheman (in Schoenherr, 1840: 540) (non Lund, 1800) (*Brenthus*); syn. Schönfeldt, 1908: 53.
argentinicus Kleine, 1944: 158 (*Brenthus*); syn. Damoiseau, 1989b: 63.

Type depositories. *B. signatus*: NHRS (?) (HT); *B. argentinicus*: MNHN (HT!).

Distribution. Puerto Rico; Argentina, Bolivia, Paraguay.

Notes. Following Damoiseau (1989b: 63) *B. signatus* is a valid species and not a synonym of *B. calcar* as stated by Kleine (1938d: 173) and Haedo Rossi (1961: 303).

Kleine (1928a: 95) reported that the type for *B. signatus* was in NHRS but it is not listed in the NHRS web site.

soukupi Haedo Rossi, 1959

soukupi Haedo Rossi, 1959a: 19 (*Brenthus*).

Type depositories. JHRC (HT, AT, PT), AMC (PT), CJSP (PT), CSMBA (PT).

Distribution. Bolivia, Peru.

turbatus Boheman, 1840 stat. res.

turbatus Boheman (in Schoenherr, 1840: 533) (*Brenthus*).

volvulus Olivier, 1807: 441 (non Fabricius, 1792) (*Brentus*); syn. Schönfeldt, 1908: 53.

Type depositories. *B. turbatus*: MNHN (ST!), NHRS (ST!).

Distribution. Cuba, Dominican Republic, Puerto Rico.

Notes. This species was included among the synonyms of *B. vulneratus* (Gyllenhal), but after studying of the type material we believe that *turbatus* is a valid species. The metafemora are unarmed in small males (as quoted in the original description), but become toothed in larger ones. The elytral apex is spatulate in *B. turbatus*, truncate in *B. vulneratus*.

variegatus Sharp, 1895

variegatus Sharp, 1895a: 68 (*Brenthus*).

Type depository. NHM (LT!, PLT!).

Distribution. El Salvador, Guatemala, Mexico.

Notes. Lectotype of *B. variegatus* (NHM) designated on a male specimen with the following labels: San Lucas, Guatemala, Champion/ syntype/ B. C. A., coll. IV, 6. *Brenthus variegatus* Sharp/ Sharp coll. 1905-313. Paralectotypes: 1 male and 2 female specimens (NHM), San Juan, Vera Paz, Champion/ syntype/ B. C. A., coll. IV, 6. *Brenthus variegatus* Sharp; 3 male and 4 female specimens (NHM), Purula, Vera Paz, Champion/ syntype/ B. C. A., coll. IV, 6. *Brenthus variegatus* Sharp; 2 male and 3 female specimens (NHM), Capetillo, Guatemala, G. C. Champion/ syntype/ B. C. A., coll. IV, 6. *Brenthus variegatus* Sharp; 1 male specimen (NHM), Zapote, Guatemala, G. C. Champion/ syntype/ B. C. A., coll. IV, 6. *Brenthus variegatus* Sharp.

vulneratus Gyllenhal, 1833

vulneratus Gyllenhal (in Schoenherr, 1833: 345) (*Brenthus*).

volubilis Boheman (in Schoenherr, 1840: 535) (*Brenthus*); syn. Kleine, 1922l: 91.

Type depositories. *B. vulneratus*: NHRS (HT!); *B. volubilis*: MNHN (ST!), NHRS (ST!).

Distribution. Mexico; Argentina, Brazil, Ecuador, Paraguay.

Cephalobarus Schoenherr, 1840

Cephalobarus Schoenherr, 1840: 517.

Brachycephalobarus Kleine, 1914a: 193; syn. Kleine, 1927a: 75.

Type species: *Cephalobarus macrocephalus* Boheman, 1840.

Distribution. Bolivia, Brazil, Colombia, Ecuador, Venezuela (Madagascar in error).

macrocephalus Boheman, 1840

macrocephalus Boheman (in Schoenherr, 1840: 519).

flavoornatus Senna, 1889: 35 (*Cephalobarus macrocephalus* var.); syn. Schönfeldt, 1908: 52.

maculatus Senna, 1889: 35 (*Cephalobarus macrocephalus* var.); syn. Schönfeldt, 1908: 52.

enderleini Kleine, 1914a: 193 (*Brachycephalobarus*); syn. Kleine, 1927a: 75.

Type depositories. *C. macrocephalus*: NHRS (ST); *C. macrocephalus* var. *flavoornatus*: MZUF (LT!, PLT!); *C. macrocephalus* var. *maculatus*: unknown; *B. enderleini*: unknown.

Distribution. Bolivia, Brazil, Colombia, Ecuador, Venezuela (Madagascar in error).

Notes. Lectotype of *C. macrocephalus* var. *flavoornatus* (MZUF) designated on a male specimen with the following labels: Cauca, Colombia, d. Staudinger/ *Cephalobarus macrocephalus* Boh. var. *flavoornatus* cotypi! Senna/ Bull. Soc. Ent. It., XXI, 1889, cotypi ♂ ♀ / ex-coll. Senna/ "La Specola" Firenze, 7073. Paralectotype: 1 female specimen (MZUF), Cauca, Colombia, d. Staudinger/ var. *flavoornatus* cotypi! Senna/ ex-coll. Senna/ "La Specola" Firenze, 7073.

ARRHENODINI Lacordaire, 1866

Arrhénodides Lacordaire, 1866: 425.

Bélophérides Lacordaire, 1866: 433.

Bélorhynchides Lacordaire, 1866: 437.

Eutrachélides Lacordaire, 1866: 438.

Arrhenodinae Pascoe, 1872: 317.

Belopherinae Pascoe, 1872: 317.

Arrhenodini Leconte, 1876: 325.

Eutrachelinae Senna, 1892a: 179.

Belophorini: Senna, 1892b: 27 [lapsus].

Belophorinae: Senna, 1892c: 49 [lapsus].

Arrhenodina Sharp, 1895a: 35.

Belopherina Sharp, 1895a: 53.

Bélorhynchina Sharp, 1895a: 63.

Orychodi Senna, 1895b: 213.

- Eutrachelini Heyne & Taschenberg, 1907a: 234.
 Arrhenodidae Schönfeldt, 1908: 31.
 Belopheridae Schönfeldt, 1908: 42.
 Eutrachelidae Schönfeldt, 1908: 49.
 Belopherini Schönfeldt, 1910a: 27.
 Entrachilini: Kleine, 1921a: 78 [lapsus].
 Eupsalini De Muizon, 1960: 170.
 Eutrachelidae: Ienistea, 1986: 32 [lapsus].
 Belorhynchidae Ienistea, 1986: 32.
 Belorhynchini Zimmerman, 1994: 34.
 Beopherina: Alonso-Zarazaga & Lyal, 1999: 47 [lapsus].
 Beloferinae: Poggesi & Sforzi, 2001: 228 [lapsus].

***Abrentodes* Sharp, 1895**

Abrentodes Sharp, 1895a: 48.

Type species: *Abrentodes auratus* Sharp, 1895.

Distribution. Costa Rica, Guatemala, Mexico.

***auratus* Sharp, 1895**

auratus Sharp, 1895a: 48.

Type depository. NHM (LT!, PLT!).

Distribution. Guatemala, Mexico.

Notes. Lectotype of *A. auratus* (NHM) designated on a male specimen with the following labels: Coban, Vera Paz., Champion, *Abrentodes auratus* ♂ type D. S./ syntype/ B. C. A., coll. IV, 6. *Abrentodes auratus* Sharp/ Sharp coll. 1905-313. Paralectotypes: 1 male specimen (NHM), Coban, Vera Paz., Champion, *Abrentodes auratus* D. S. ♂ syntype/ B. C. A., coll. IV, 6. *Abrentodes auratus*, Sharp; 1 female specimen (NHM), S. Juan, Vera Paz., Champion, *Abrentodes auratus* ♀ type D. S./ syntype/ B. C. A., coll. IV, 6. *Abrentodes auratus* Sharp/ Sharp coll. 1905-313; 3 female specimens (NHM), syntype/ Senahu, Vera Paz., Champion/ Sharp coll. 1905-313/ B. C. A., coll. IV, 6. *Abrentodes auratus* Sharp; 2 female specimens (NHM), syntype/ Juquila, Mexico/ Sharp coll. 1905-313/ B. C. A., coll. IV, 6. *Abrentodes auratus* Sharp; 1 male specimen (NHM), Sinaja, Vera Paz., Champion, *Abrentodes auratus* D. S./ syntype/ B. C. A., coll. IV, 6. *Abrentodes auratus* Sharp.

breviceps Sharp, 1895

breviceps Sharp, 1895a: 49.

Type depository. NHM (HT!).

Distribution. Guatemala.

pusillus Sharp, 1895

pusillus Sharp, 1895a: 49.

Type depository. NHM (LT!, PLT!).

Distribution. Guatemala.

Notes. Lectotype of *A. pusillus* (NHM) designated on a male specimen with the following labels: *Abrentodes pusillus* D. S. Type D. S. ♂ / syntype/ Cerro Zunil, 4-5000 ft., Champion/ B. C. A., coll. IV, 6. *Abrentodes pusillus*, Sharp/ Sharp coll. 1905-313. Paralectotypes: 1 male specimen (NHM), *Abrentodes pusillus* D. S., Sinanja, Vera Paz., Champion/ syntype/ B. C. A., coll. IV, 6. *Abrentodes pusillus* Sharp/ Sharp coll. 1905-313; 1 female specimen (NHM), *Abrentodes pusillus* D. S., Perula, Vera Paz., Champion/ syntype/ B. C. A., coll. IV, 6. *Abrentodes pusillus* Sharp/ Sharp coll. 1905-313.

robustus Sharp, 1895

robustus Sharp, 1895a: 51.

Type depository. NHM (LT!, PLT!).

Distribution. Costa Rica, Guatemala.

Notes. Lectotype of *A. robustus* (NHM) designated on a male specimen with the following labels: *Abrentodes robustus* Type D. S. ♂ / Perula, Vera Paz., Champion/ Syntype/ B. C. A., coll. IV, 6. *Abrentodes robustus*, Sharp/ Sharp coll. 1905-313. Paralectotype: 1 female specimen (NHM), with the same label data.

sulcipennis Sharp, 1895

sulcipennis Sharp, 1895a: 50.

Type depository. NHM (LT!, PLT!).

Distribution. Guatemala.

Notes. Lectotype of *A. sulcipennis* (NHM) designated on a male specimen with the following labels: *Abrentodes sulcipennis* D. S. ♂ / Cerro Zunil, 4000 ft., Champion/ syntype/ B. C. A., coll. IV, 6. *Abrentodes sulcipennis* Sharp/ Sharp coll. 1905-313. Paralectotypes: 1 female specimen (NHM), *Abrentodes sulcipennis* Type D. S. ♂ / Cerro Zunil, 4000 ft., Champion/ syntype/ B. C. A., coll. IV, 6. *Abrentodes sulcipennis* Sharp/ Sharp coll. 1905-313; 1 female specimen (NHM), *Abrentodes sulcipennis* Type D. S. ♀ / Cerro Zunil, 4-5000 ft., Champion/ syntype/ B. C. A., coll. IV, 6. *Abrentodes sulcipennis* Sharp/ Sharp coll. 1905-313.

Agriorrhynchus Power, 1878

Agriorrhynchus Power, 1878b: 241.

Prosorychodes Kleine, 1923j: 409; syn. Damoiseau, 1989b: 45.

Type species: *Agriorrhynchus borrei* Power, 1878.

Distribution. China; Borneo, India (including Assam, Darjeeling), Indonesia (Java, Sumatra), Laos, Malaysia, Myanmar [= Burma], Philippines (Luzon), Thailand, Vietnam.

Notes. Kleine (1922m: 162) mentioned *Prosorychodes* in a list of genera, but did not describe it until one year later (1923j: 409).

borrei Power, 1878

borrei Power, 1878b: 241.

consonus Kleine, 1923j: 410 (*Prosorychodes*); syn. Damoiseau, 1989b: 45.

Type depositories. *A. borrei*: IRSNB (LT!, PLT!), MCZ (PLT); *P. consonus*: Buitenzorg Museum (?) (HT).

Distribution. Borneo, Burma, India (Assam, Darjeeling), Indonesia (Java, Sumatra), Malaysia, Thailand, Vietnam.

Notes. Damoiseau (1989b: 48) designated the lectotypes of *A. borrei* from material in IRSNB.

***cynicus* Damoiseau, 1966**

cynicus Damoiseau, 1966b: 452.

Type depositories. BPBM (HT), IRSNB (PT!).

Distribution. S China (Guangdong).

***ignarus* Kleine, 1925**

ignarus Kleine, 1925c: 598.

ignarius: Kleine, 1931h: 418 (*Agriorrhynchus*) [lapsus].

Type depository. MNHUB (NT!).

Distribution. Philippines (Luzon).

***klapperichi* Kleine, 1950**

klapperichi Kleine, 1950: 24.

Type depositories. ZFMK (HT), DEI (PT!).

Distribution. China (Fujian).

***poweri* Damoiseau, 1989**

poweri Damoiseau, 1989b: 48.

Type depositories. MNHN (HT!, PT!), IRSNB (PT!).

Distribution. Borneo, Indonesia (Sumatra), Malaysia.

quadrituberculatus Senna, 1892

quadrituberculatus Senna, 1892e: 470.

quadrituberculatus: Kleine, 1918a: 120 (*Agriorrhynchus*) [lapsus].

Type depositories. MZUF (LT!), MSNG (PLT!).

Distribution. India (including Darjeeling), Laos, Myanmar [= Burma], Thailand, Vietnam.

Notes. Damoiseau (1989b: 50) reported that the specimen held in MSNG is the holotype and the one in MNHUB is the allotype, but in his original description Senna did not designate any of the four syntypes (three in MSNG and one in MZUF) as holotype.

Lectotype designated by Bartolozzi *et al.* (1985: 100) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

scalpratus Damoiseau, 1989

scalpratus Damoiseau, 1989b: 50.

Type depositories. MNHUB (HT!, AT!), IRSNB (PT!), MNHN (PT!).

Distribution. Indonesia (Sumatra).

undulatus Power, 1878

undulatus Power, 1878b: 241.

Type depository. MNHN (HT!).

Distribution. Borneo, India (Assam), Indonesia (Java, Sumatra), Malaysia, Myanmar [= Burma].

venustus Kleine, 1938

venustus Kleine, 1938d: 89.

Type depositories. EMBT (HT, PT), DEI (PT!).

Distribution. Thailand, Vietnam (?).

Notes. Damoiseau (1989b) did not mention this species in his revision of the genus.

Allacompsus Kleine, 1921

Allacompsus Kleine, 1921k: 286.

Type species: *Allacompsus separator* Kleine, 1921.

Distribution. Cocos Is.

separator Kleine, 1921

separator Kleine, 1921k: 287.

Type depository. MNUL (?) (HT) (probably lost).

Distribution. Cocos Is.

Notes. Kleine wrote that the type was held in the Museum of Lübeck, but according to Susanne Fütting (Museum für Natur und Umwelt, Lübeck) (2002, personal communication) the former Naturhistorisches Museum der Hansestadt Lübeck was destroyed during war in 1942 and the collections lost.

Amphicordus Heller, 1913

Amphicordus Heller, 1913: 151.

Oncodetichus Damoiseau, 1967c: 17; syn. Damoiseau, 1989b: 50.

Type species: *Amphicordus improportionatus* Heller, 1913.

Distribution. Laos, Philippines (Luzon, Mindanao).

dentirostris Damoiseau, 1989

dentirostris Damoiseau, 1989b: 51.

Type depositories. IRSNB (HT!, AT!, PT!), BPBM (PT).

Distribution. Laos.

inproportionatus Heller, 1913

inproportionatus Heller, 1913: 152.

improportionalis: Kleine, 1925c: 600 (*Amphicordus*) [lapsus].

lacertosus Damoiseau, 1967c: 18 (*Oncodetichus*); syn. Damoiseau, 1989b: 51.

improportionatus: Damoiseau, 1989b: 51 (*Amphicordus*) [lapsus].

Type depositories. *A. inproportionatus*: BPBM (?) (NT); *O. lacertosus*: IRSNB (HT!, PT!).

Distribution. Laos, Philippines (Luzon, Mindanao).

Notes. According to A. Samuelson (BPBM) (2001, personal communication) the neotype (Damoiseau, 1989b: 51) of *A. inproportionatus* is not in BPBM now because it was never returned after the untimely death of Damoiseau.

Ananesiotes Kleine, 1922

Ananesiotes Kleine, 1922p: 132.

Nesiotes Kleine, 1921a: 43 (non Martens, 1860, nec Wollaston, 1861).

Type species: *Ananesiotes insularis* Kleine, 1922.

Distribution. Taiwan.

Notes. Kleine mentioned the name *Nesiotes* in 1921 in a paper on brentid elytral patterns, but he described it under the name *Ananesiotes* only in 1922.

insularis Kleine, 1922

insularis Kleine, 1922p: 133.

Type depository. DEI (LT!, PLT!).

Distribution. Taiwan.

Notes. Lectotype of *A. insularis* (DEI) designated on a male specimen with the following labels: Formosa, Fuhosho, VI.1909, H. Sauter/ Syntypus/ Kleine det./ *Ananesiotes insularis* Kleine., det. Autor. Paralectotypes: 6 male and 2 female specimens (DEI), Formosa, Fuhosho, VI.1909, H. Sauter/ Synty-

pus/ Kleine det.; 1 male specimen (DEI), Formosa, Taihorin, 1911, H. Sauter/
7.XI/ Syntypus/ Kleine det.; 1 female specimen (DEI), Formosa, Kosempo,
1909, H. Sauter/ 7.VII/ Syntypus/ Kleine det.

Anepsiotes Kleine, 1917

Anepsiotes Kleine, 1917f: 318.

Type species: *Anepsiotes schenklingi* Kleine, 1917.

Distribution. S China; Borneo, India, Malaysia, Philippines (Luzon), Sri Lanka, Vietnam.

bellus Kleine, 1925

bellus Kleine, 1925b: 159.

Type depository. NHM (HT!).

Distribution. India.

commendabilis Kleine, 1925

commendabilis Kleine, 1925b: 159.

Type depository. NHM (HT!).

Distribution. India.

elegans Calabresi, 1921

elegans Calabresi, 1921c: 68.

Type depository. SMTD (ST).

Distribution. Borneo.

kleinei Calabresi, 1920*kleinei* Calabresi, 1920b: 68.**Type depositories.** MSNG (LT!), MZUF (PLT!).**Distribution.** Borneo, Malaysia.

Notes. Lectotype of *A. kleinei* (MSNG) designated on a male specimen with the following labels: Sarawak, G. Doria, 1866/ type/ *Belopherus sarawacus* Pow./ *Anepsiotes kleinei* typus! Calabresi/ lectotype/ Prép. génit n° 76 J 31, R. Damoiseau/ *Heteroblysmia kleinei* (Calabr.) lectotype R. Damoiseau det. [*in litteris*]. Paralectotypes: 1 male specimen (MZUF), Is. Penang, A. Boucard/ *Anepsiotes kleinei* n. sp./ Bull. Soc. Ent. It., LI, 1919-20, cotypi! Calabresi/ Prép. génit n° 76 J 12 R. Damoiseau/ ex-coll. Senna/ "La Specola" Firenze, 7036; 1 male specimen (MZUF), Is. Penang, A. Boucard/ ex-coll. Senna/ "La Specola" Firenze, 7036/ *Heteroblysmia kleinei* (Calabresi), PT ♂/ Prép. Micr. n° 75 C 01.

luzonicus Calabresi, 1920*luzonicus* Calabresi, 1920b: 66.**Type depository.** MZUF (HT!).**Distribution.** Philippines (Luzon).*nitidicollis* Calabresi, 1920*nitidicollis* Calabresi, 1920b: 69.**Type depository.** MZUF (HT!).**Distribution.** Philippines (Luzon).*pahanganus* Kleine, 1938*pahanganus* Kleine, 1938d: 122.**Type depository.** NHM (HT!).**Distribution.** Malaysia.

schenklingi Kleine, 1917*schenklingi* Kleine, 1917f: 321.**Type depository.** DEI (LT!, PLT!).**Distribution.** S China, Sri Lanka, Vietnam.**Notes.** Lectotype of *A. schenklingi* (DEI) designated on a male specimen with the following labels: Nalanda, Ceylon, W. Horn, 1899/ Syntypus/ Kleine det. Paralectotype: 1 male specimen (DEI), with the same label data as the lectotype.*Anomobrenthus* Fairmaire, 1881*Anomobrenthus* Fairmaire, 1881a: 349.**Type species:** *Anomobrenthus hamatirostris* Fairmaire, 1881.**Distribution.** Fiji Is., Vanuatu [= New Hebrides].*hamatirostris* Fairmaire, 1881*hamatirostris* Fairmaire, 1881a: 349.**Type depository.** NHM (NT!).**Distribution.** Fiji Is., Vanuatu [= New Hebrides].*kuscheli* Damoiseau, 1989*kuscheli* Damoiseau, 1989b: 55.**Type depositories.** DSIRW (HT, AT, PT), IRSNB (PT!).**Distribution.** Fiji Is.

Apocemus Calabresi, 1921

Apocemus Calabresi, 1921c: 58.

Apocemis: Kleine, 1925c: 602 [lapsus].

Type species: *Megacerus conciliator* Kirsch, 1875.

Distribution. Malaysia, Philippines (Luzon).

conciliator (Kirsch, 1875)

conciliator Kirsch, 1875b: 46 (*Megacerus*).

Type depository. SMTD (ST).

Distribution. Malaysia.

ignobilis Kleine, 1925

ignobilis Kleine, 1925c: 602.

Type depository. MPMP (HT).

Distribution. Philippines (Luzon).

Arrenodes Schoenherr, 1823

Arrenodes Schoenherr, 1823: 1137.

Brentus Panzer, 1788: 159 (non Fabricius, 1787); syn. Alonso-Zarazaga & Lyal, 1999: 47.

Arrhenodes Schoenherr, 1826: 70; syn. Alonso-Zarazaga & Lyal, 1999: 47.

Eupsalis Lacordaire, 1866: 430; syn. Kuntzen, 1937: 193.

Platysystrophus Kleine, 1917g: 139; syn. Kuntzen, 1937: 193.

Arrenhodes: Calder, 1990: 457 [lapsus].

Type species: *Brentus maxillosus* Olivier, 1807 (non Herbst, 1797) = *Curculio minutus* Drury, 1773.

Distribution. Canada to Mexico.

minutus (Drury, 1773)

- minutus* Drury, 1773 [in Index, non-numbered pages] (*Curculio*).
dispar Fabricius, 1781: 182 (non Linné, 1758) (*Curculio*); syn. Kuntzen, 1937: 193.
druryanus Pallas, 1781: 26 (*Curculio*); **syn. n.**
septemtrionis Herbst, 1797: 183 (*Brentus*); syn. Schoenherr, 1833: 326.
distans Panzer, 1798 [in Forward, non-numbered pages, fig. 5 on title plate] (*Brentus*); syn. Gemminger & Harold, 1872: 2709.
brunneus Panzer, 1798: 44 (non Olivier, 1790) (*Curculio*); syn. Gemminger & Harold, 1872: 2709.
maxillosus Olivier, 1807: 443 (*Brentus*); syn. Gemminger & Harold, 1872: 2709.
septentrionis: Schoenherr, 1833: 326 (*Arrhenodes*) [lapsus].
lecontei Power, 1878a: 494 (*Eupsalis*); syn. Damoiseau, 1963f: 5.
sallei Power, 1878a: 494 (*Eupsalis*); syn. Damoiseau, 1963f: 5.
paupercula Blatchley & Leng, 1916: 21 (*Eupsalis minuta* var.); syn. Kuntzen, 1937: 194.

Type depositories. *C. minutus*: unknown; *B. druryanus*: unknown; *B. septemtrionis*: MNHUB (Herbst coll.) (?) (HT); *B. maxillosus*: unknown; *E. lecontei*: MNHN (LT!); *E. sallei*: MNHN (LT!, PLT!); *E. minuta* var. *paupercula*: unknown.

Distribution. Canada (Ontario), USA (Arkansas, District of Columbia, Florida, Georgia, Illinois, Indiana, Kansas, Louisiana, Massachusetts, Minnesota, Mississippi, Missouri, Nebraska, New England, New Jersey, New York, North Carolina, Ohio, Oklahoma, Tennessee, Texas, Virginia, West Virginia, Wisconsin); Mexico.

Notes. Lectotype of *E. lecontei* (MNHN) designated on a female specimen with the following label: ex Musaeo Power (without locality).

Lectotype of *E. sallei* (MNHN) designated on a specimen with the following labels: Texas/ Chevrolat 124/ type/ ex Musaeo Power/ Museum Paris 1952, coll. R. Oberthür. Paralectotype: 1 specimen (MNHN), Madagascar?/ Chevr. 128/ type/ Museum Paris 1952, coll. R. Oberthür.

The species *B. minutus* was described and drawn by Drury in 1770 with no name, it appears in the Index of volume 2 published in 1773. Pallas (1781) redescribed the species figured by Drury under the name of *Curculio druryanus*.

We did not find the type for *B. septemtrionis* in MNHUB, where the Herbst collection is held.

According to R. Brett (CAS) (2002, personal communication) the type for *E. minuta* var. *paupercula* is not in CAS, where the Blatchley collection is held.

***Baryrhynchus* Lacordaire, 1866**

Baryrhynchus Lacordaire, 1866: 428.

Type species: *Arrhenodes latirostris* Gyllenhal, 1833.

Distribution. China, Japan, Russia (Kuriles), Taiwan; Bangladesh, Borneo, Kampuchea [= Cambodia], India (including Andaman Is., Assam, Darjeeling, Nicobar Is., Sikkim), Indonesia (Engano; Java; Mentawai; Misool; Moluccas: Ambon, Aru, Bacan [= Batjan], Buru, Halmahera [= Gilolo], Seram [= Ceram]; Nias; Salawati; Sulawesi; Sumbawa; Sumatra), Laos, Malaysia, Myanmar [= Burma], Nepal, Philippines (Luzon, Mindanao, Negros, Palawan, Samar, Sibuyan, Tawi Tawi), Sri Lanka, Thailand, Vietnam; Australia, Melanesia, New Guinea (including Bismarck Archipelago: Duke of York, New Britain), Solomon Is.

subgenus ***Baryrhynchus* s. s.**

Baryrhynchus: Gemminger & Harold, 1872: 2707 [lapsus].

Pachyrhynchus: Desbrochers des Loges, 1891: 4 [lapsus].

Baryrhynchus: Kleine, 1922m: 161 [lapsus].

Baryrhynchus: Miwa, 1933: 13 [lapsus].

Baryrhynchus: Azuma, 2001: 347 [lapsus].

***andamanicus* Power, 1879**

andamanicus Power, 1879b: 298 (*Baryrhynchus*).

Type depository. MNHN (ST!).

Distribution. India (Andaman Is., Assam, Nicobar Is.), Laos, Vietnam.

Notes. Damoiseau labelled the two MNHN syntypes as lectotype and paralectotype (Andaman/ type/ Mniszech 5/ ex Musaeo Power/ Museum Paris 1952, coll. R. Oberthür) but actually they are *in litteris*.

***angulatus* Zhang, 1993**

angulatus Zhang, 1993: 161 (*Baryrhynchus*).

Type depository. IZAS (HT, AT, PT).

Distribution. China (Sichuan, Yunnan).

***anthracinus* Kleine, 1916**

anthracinus Kleine, 1916f: 164 (*Baryrrhynchus*).

Type depository. MNHUB (LT!, PLT!).

Distribution. Borneo, Indonesia (Java, Sumatra).

Notes. Lectotype of *B. anthracinus* (MNHUB) designated on a male specimen with the following labels: Mittel Java, Axel Preyer S. G./ *Baryrhynchus anthracinus* Kl. type/ *Baryrhynchus anthracinus* R. Damoiseau det. Paralectotype: 1 male specimen (MNHUB), Mittel Java, Axel Preyer S. G./ Typus.

***bengalensis* Kleine, 1937**

bengalensis Kleine, 1937i: 203 (*Baryrrhynchus*).

Type depository. IFRI (HT).

Distribution. India (Darjeeling).

***concretus* Zhang, 1993**

concretus Zhang, 1993: 164 (*Baryrrhynchus*).

Type depository. IZAS (HT).

Distribution. China (Yunnan).

***convexus* Zhang, 1993**

convexus Zhang, 1993: 168 (*Baryrrhynchus*).

Type depository. IZAS (HT).

Distribution. China (Hainan).

cratus Zhang, 1993

cratus Zhang, 1993: 165 (*Baryrrhynchus*).

Type depository. IZAS (HT, AT, PT).

Distribution. China (Yunnan).

dehiscens (Gyllenhal, 1833)

dehiscens Gyllenhal (in Schoenherr, 1833: 324) (*Arrhenodes*).

Type depository. NHRS (ST).

Distribution. S China; Borneo, India (Assam), Indonesia (Java, Mentawai, Nias, Sulawesi, Sumatra), Malaysia, Myanmar [= Burma], Sri Lanka, Vietnam.

discolor Kleine, 1916

discolor Kleine, 1916f: 157 (*Baryrrhynchus*).

Type depository. MZPW (HT).

Distribution. Indonesia (Moluccas: Aru; Sumbawa).

latirostris (Gyllenhal, 1833)

latirostris Gyllenhal (in Schoenherr, 1833: 323) (*Arrhenodes*).

Type depositories. IRSNB (LT!, PLT!), NHRS (PLT).

Distribution. Borneo, India (Andaman Is., Assam, Sikkim), Indonesia (Java, Nias, Sumatra), Malaysia; New Guinea.

Notes. Lectotype of *A. latirostris* (IRSNB) designated on a male specimen [incorrectly labelled as holotype] with the following labels: Java/ Coll. Dejean, Coll. Roelofs/ *latirostris* mihi, h. in int. Java/ *Baryrrhynchus latirostris* Gyll., Type ♂, R. Damoiseau rev. 1961/ Holotype. Paralectotypes: 1 male specimen (IRSNB), Java/ Coll. Dejean, Coll. Roelofs/ *latirostris* Dej./ Paratype; 1 male specimen (IRSNB), Java/ Coll. Roelofs/ M. R. Belg./ Paratype; 1 female specimen (IRSNB), Coll. Dejean, Coll. Roelofs/ M. R. Belg./ Paratype; 1 female specimen (IRSNB), Java, Coll. Dejean, Coll. Roelofs/ *crenulatus* Dej./ Paratype; 1 female specimen (IRSNB), Coll. Dejean, Coll. Roelofs/ *Baryrrhynchus latirostris* Gyll., Allotype ♀, R. Damoiseau rev. 1961/ Allotype.

merocephalus Kleine, 1916

merocephalus Kleine, 1916f: 178 (*Baryrrhynchus*).

Type depositories. DEI (LT!, PLT), MNHUB (PLT!), SMTD (PLT), ZSM (PLT).

Distribution. China (Fujian); India (including Assam, Darjeeling), Kampuchea [= Cambodia], Vietnam.

Notes. Lectotype of *B. merocephalus* (DEI) designated on a male specimen with the following labels: Assam, India/ Kleine vidit, 1916/ Kleine det./ Syntypus/ *Baryrrhynchus merocephalus* Kl. cotype/ Syntypus. Paralectotypes: 1 female specimen (DEI), Assam, India/ Kleine vidit, 1916/ Kleine det./ Syntypus; 1 female specimen (MNHUB), Darjeeling/ 82234/ 6/ *Baryrrhynchus merocephalus* Kleine, cotypus; 1 female specimen (MNHUB), Darjeeling/ 82225/ 4.

miles (Boheman, 1845)

miles Boheman (in Schoenherr, 1845: 372) (*Arrhenodes*).

approximans F. Walker, 1859: 262 (*Arrhenodes*); syn. Kleine, 1923f: 90.

approximens: Motschulsky, 1863: 524 (*Arrhenodes*) [lapsus].

approximatus: Schönfeldt, 1908: 35 (*Arrhenodes*) [lapsus].

Type depositories. *A. miles*: NHRS (ST); *A. approximans*: NHM (LT!, PLT!).

Distribution. S China (Hainan, Shanghai), Taiwan; Bangladesh, Borneo, Laos, India (Assam, Darjeeling, Sikkim), Indonesia (Java), Malaysia, Myanmar [= Burma], Nepal, Sri Lanka, Thailand, Vietnam.

Notes. Lectotype of *A. approximans* (NHM) designated on a male specimen with the following labels: syntype/ Ceylon/ *approximans*/ *Arrhenodes approximans* a. n. H. (Type). Paralectotype: 1 female specimen (NHM), syntype/ 59.106/ *Arrhenodes approximans* a. n. H. (Type)/ *Baryrhynchus miles* Boh. ♀, R. Kleine det., 1922.

minisulus Zhang, 1993

minisulus Zhang, 1993: 168 (*Baryrrhynchus*).

minisculus: Zhang, 1993: 161 (*Baryrhynchus*) [lapsus].

Type depository. IZAS (HT).

Distribution. China (Yunnan).

***nitidus* Zhang, 1993**

nitidus Zhang, 1993: 169 (*Baryrrhynchus*).

Type depository. IZAS (HT, AT).

Distribution. China (Hainan).

***odontus* Zhang, 1993**

odontus Zhang, 1993: 163 (*Baryrrhynchus*).

Type depository. IZAS (HT, AT, PT).

Distribution. China (Sichuan).

***phaeus* Zhang, 1993**

phaeus Zhang, 1993: 162 (*Baryrrhynchus*).

Type depository. IZAS (HT, AT, PT).

Distribution. China (Fujian, Zhejiang).

***planus* Zhang, 1993**

planus Zhang, 1993: 163 (*Baryrrhynchus*).

Type depository. IZAS (HT, AT, PT).

Distribution. China (Yunnan).

***rudis* Senna, 1893**

rudis Senna, 1893b: 280 (*Baryrrhynchus*).

Type depositories. MSNG (LT!, PLT!), DEI (PLT!), IRSNB (PLT!), MNHN (PLT!), MZUF (PLT!), NHM (PLT!), MZPW (ST), SMTD (ST).

Distribution. Bangladesh, India, Indonesia (Engano, Java, Sumatra), Philippines (Palawan), Vietnam.

Notes. Lectotype of *B. rudis* (MSNG) designated on a male specimen with the following labels: Engano, Bua Bua, V.VI.1891, Modigliani/ *Baryrrhynchus rudis* Senna typi! Paralectotypes: 5 male and 7 female specimens (MSNG), 1 specimen (MNHN), 2 male and 1 female specimens (NHM), 2 male and 1 female specimens (DEI), 2 male specimens (MNHUB), 2 male and 2 female specimens (IRSNB), Engano, Bua Bua, V.VI.1891, Modigliani; 5 male and 5 female specimens (MSNG), 1 male specimen (NHM), 1 male specimen (IRSNB), Engano, Kifa-juc, V.1891, Modigliani; 1 male specimen (MZUF), Engano, Bua Bua, V-VI.1891, Modigliani/ *Baryrrhynchus rudis* Senna, typus!/ *Baryrrhynchus rudis* Senna, cotypi!/ Ann. Mus. Genov., ser. 2, VIII, 1893, cotypi ♂♂ ♀♀/ ex-coll. Senna/ "La Specola" Firenze, 6968; 3 male and 5 female specimens (MZUF), 1 male and 1 female specimens (DEI), Engano, Bua Bua, V-VI.1891, Modigliani/ ex-coll. Senna/ 6968; 1 male and 1 female specimen (MNHN), Engano, Bua Bua, V-VI.1891, Modigliani/ *Baryrrhynchus rudis* Senna, R. Damoiseau det./ ex-coll. Senna/ 6968.

setulosus Zhang, 1993

setulosus Zhang, 1993: 166 (*Baryrrhynchus*).

Type depository. IZAS (HT).

Distribution. China (Yunnan).

speciosissimus Kleine, 1916

speciosissimus Kleine, 1916f: 180 (*Baryrrhynchus*).

speciosissimus: Kleine, 1916f: 180 (*Baryrrhynchus*) [lapsus].

speciosissimus: Kleine, 1938b: 68 (*Baryrrhynchus*) [lapsus].

Type depositories. DEI (LT!, PLT!), MNHUB (PLT!), SMTD (PLT).

Distribution. China (Fujian); India, Kampuchea (= Cambodia), Laos, Vietnam.

Notes. Lectotype of *B. speciosissimus* (DEI) designated on a male specimen with the following labels: Tonkin, Than-Moi, Juni-Juli, H. Fruhstorfer/ *Baryrrhynchus speciosissimus* Kl., Typus. Paralectotypes: 1 female specimen (DEI), Laos/ Kleine vidit 1916/ Kleine det./ Syntypus; 1 female specimen (MNHUB), Madura, Ind. or./ *Baryrrhynchus speciosissimus* Kleine, cotype/ Lectotype [*in litteris*].

***umbraticus* Kleine, 1916**

umbraticus Kleine, 1916f: 168 (*Baryrrhynchus*).

Type depositories. DEI (LT!, PLT!), MNHUB (PLT!), SMTD (PLT).

Distribution. Borneo, Indonesia, Laos.

Notes. Lectotype of *B. umbraticus* (DEI) designated on a male specimen with the following labels: Wahnes, Borneo/ col. Kraatz/ Kleine det./ *Baryrrhynchus umbraticus* Kl. Cotype. Paralectotypes: 1 female specimen (DEI), with the same label data as the lectotype; 1 male specimen (MNHUB), Lectotype [*in litteris*]/ Sunda Ins., ex coll. v. Studt G./ *Baryrrhynchus umbraticus* Kl., cotype/ *Baryrrhynchus umbraticus* Kleine LT, R. Damoiseau det.

***yaeyamensis* Morimoto, 1979**

yaeyamensis Morimoto, 1979: 27 (*Baryrhynchus*).

miles Morimoto, 1976a: 278 (non Boheman, 1845) (*Baryrrhynchus*); syn. Morimoto, 1979: 27.

Type depository. ELKU (HT, PT).

Distribution. Japan.

subgenus *Eupsalomimus* Kleine, 1916

Eupsalomimus Kleine, 1916f: 151.

Eupsalominus: Kleine, 1916f: 187 [lapsus].

***lineicollis* Power, 1879**

lineicollis Power, 1879b: 297 (*Baryrrhynchus*).

indocilis Fairmaire, 1883: 41 (*Baryrhynchus*); syn. Damoiseau, 1972b: 274.

schroederi Kleine, 1914b: 172 (*Baryrrhynchus*); syn. Damoiseau, 1972b: 274.

lineicollis Kleine, 1916f: 186 (non Power, 1879) (*Baryrrhynchus*); syn. Damoiseau, 1972b: 275.

pictipennis Lea, 1916: 367 (*Eupsalis*); syn. K. E. Schedl, 1959: 61.

solidus Kleine, 1920n: 44 (*Baryrhynchus*); syn. Damoiseau, 1972b: 274.

compositus Kleine, 1923j: 407 (*Baryrrhynchus*); syn. Damoiseau, 1972b: 274.

setosellus Kleine, 1928b: 49 (*Baryrrhynchus*); syn. Damoiseau, 1972b: 274.

kleinei Damoiseau, 1972b: 274 (*Baryrrhynchus lineicollis* f.); **syn. n.**

Type depositories. *B. lineicollis*: IRSNB (HT!); *B. indocilis*: MNHN (?) (HT); *B. schroederi*: MZPW (ST); *E. pictipennis*: MVMA (ST); *B. solidus*: ZMAN (HT!); *B. compositus*: Buitenzorg Museum (?) (HT); *B. setosellus*: BPBM (HT, PT), SMTD (PT); *B. lineicollis* f. *kleinei*: unknown.

Distribution. Indonesia (Misool; Moluccas: Ambon, Aru, Bacan [= Batjan], Buru, Halmahera [= Gilolo], Seram [= Ceram]; Salawati; Sulawesi), Philippines (Luzon, Mindanao, Negros, Samar, Sibuyan, Tawi Tawi), Thailand; Australia, New Guinea (including Bismarck Archipelago: Duke of York, New Britain), Solomon Is.

Notes. We did not find the holotype of *B. indocilis* in MNHN, where many types of Fairmaire are held.

poweri Roelofs, 1879

poweri Roelofs, 1879: LIV (*Baryrrhynchus*).

poweii: Kojima, 1955a: 36 (*Baryrrhynchus*) [lapsus].

Type depository. IRSNB (LT!, PLT!).

Distribution. China (Hainan, Hubei, Hunan, Jiangxi, Guangdong, Guangxi, Sichuan, Yunnan), Japan, Russia (Kuriles), Taiwan; Borneo, India, Indonesia (Moluccas), Vietnam; Australia.

Notes. Lectotype of *B. poweri* (IRSNB) designated on a male specimen [incorrectly labelled as holotype] with the following labels: Hiller, Tokyo (Jap)/ Coll. Roelofs/ TYPE/ M. R. Belg./ *Baryrrhynchus poweri* R./ *Baryrrhynchus poweri* Roelofs type ♂ R. Damoiseau rev. 1961/ Holotype. Paralectotype: 1 female specimen (IRSNB), Hiller, Tokyo (Jap)/ Coll. Roelofs/ TYPE/ M. R. Belg./ *Baryrrhynchus poweri* Roelofs paratype ♀ R. Damoiseau rev. 1961/ Allotype.

tokarensis Ohbayashi & Satô, 1966

tokarensis Ohbayashi & Satô, 1966: 1 (*Baryrrhynchus*).

Type depositories. EUMJ (HT), Satô coll., Nagoya (?) (PT).

Distribution. Japan.

***Belopherus* Schoenherr, 1833 stat. res.**

Belopherus Schoenherr, 1833: 334.

Belophorus: Agassiz, 1846a: 20 [lapsus].

Belophorus Suffrian, 1848: 52 [unjustified emendation].

Belophorus Gemminger & Harold, 1872: 2711 (non Suffrian, 1848) [unjustified emendation].

Type species: *Brentus militaris* Olivier, 1807.

Distribution. "Antilles", Cuba, Dominican Republic, Haiti, Jamaica, Puerto Rico.

Notes. Alonso-Zarazaga & Lyal (2002: 10) placed *Belopherus* Schoenherr in synonymy with *Belorhynchus* Berthold, 1827, but for us the two genera are distinct.

***eximus* Kleine, 1927**

eximus Kleine, 1927b: 442.

Type depository. NHM (LT!, PLT!).

Distribution. Jamaica.

Notes. Lectotype of *B. eximus* (NHM) designated on a male specimen with the following labels: syntype/ Jamaica/ *Belopherus eximus* ♂ typus! Kln 1923, R. Kleine det. Paralectotype: 1 female specimen (NHM), syntype/ Jamaica/ *Belopherus eximus* ♀ type 1923, R. Kleine det.

***maculatus* (Olivier, 1790)**

maculatus Olivier, 1790: 192 (*Brentus*).

Type depository. Unknown.

Distribution. Cuba, Dominican Republic, Puerto Rico.

***militaris* (Olivier, 1807)**

militaris Olivier, 1807: 439 (*Brentus*).

spinosus Gyllenhal (in Schoenherr, 1833: 336) (*Belopherus*); syn. Damoiseau, 1962b: 31.

Type depositories. *B. militaris*: MNHN (ST!); *B. spinosus*: NHRS (HT).

Distribution. "Antilles", Cuba, Dominican Republic, Puerto Rico.

rostratus Damoiseau, 1989

rostratus Damoiseau, 1989b: 53.

Type depository. NHMB (HT!).

Distribution. Dominican Republic.

schoenherrii Mannerheim, 1840

schoenherrii Mannerheim (in Schoenherr, 1840: 509).

schoenherrii: Schönfeldt, 1908: 45 (*Belopherus*) [lapsus].

Type depositories. NHRS (?) (ST), MZHF (ST).

Distribution. Cuba, Dominican Republic, Haiti.

Notes. Kleine (1928a: 87) reported that the type for *B. schoenherrii* is in NHRS but the species is not listed in the NHRS web site.

simplicicollis Suffrian, 1870

simplicicollis Suffrian, 1870: 212.

Type depository. MNHC (HT).

Distribution. Cuba, Jamaica.

Blysmia Pascoe, 1872

Blysmia Pascoe, 1872: 323.

Ypselagonia Kleine, 1922r: 157; syn. Damoiseau, 1989b: 54.

Type species: *Blysmia ruficollis* Pascoe, 1872.

Distribution. Taiwan; Borneo, Indonesia (Java; Moluccas: Bacan [= Batjan]), Malaysia, Philippines (Mindanao).

decens (Kleine, 1932)

decens Kleine, 1932b: 116 (*Ypselagonia*).

Type depository. NHM (HT!).

Distribution. Malaysia.

drescheri (Kleine, 1938)

drescheri Kleine, 1938f: 289 (*Ypselagonia*).

Type depositories. ZMAN (Drescher coll.) (?) (HT, PT), DEI (PT!).

Distribution. Indonesia (Java).

Notes. We did not find the holotype and paratype in ZMAN, where many types from Drescher's collection are held. It is possible that they are in MZB.

guttata (Kleine, 1928)

guttata Kleine, 1928e: 102 (*Ypselagonia*).

Type depository. SMFD (HT).

Distribution. Taiwan; Borneo.

peregrina (Kleine, 1922)

peregrina Kleine, 1922r: 158 (*Ypselagonia*).

Type depository. SMTD (HT).

Distribution. Taiwan; Borneo, Philippines (Mindanao).

ruficollis Pascoe, 1872

ruficollis Pascoe, 1872: 324.

Type depository. NHM (HT!).

Distribution. Indonesia (Moluccas: Bacan [= Batjan]).

Cacopsalis Sharp, 1895

Cacopsalis Sharp, 1895a: 33.

Type species: *Cacopsalis rudis* Sharp, 1895.

Distribution. Belize, Costa Rica, Mexico.

rudis Sharp, 1895

rudis Sharp, 1895a: 34.

Type depository. NHM (LT!, PLT!).

Distribution. Belize, Costa Rica, Mexico.

Notes. Lectotype of *C. rudis* (NHM) designated on a male specimen with the following labels: *Cacopsalis rudis* Type D. S. ♂ Toxpam, Mexico, Salle Coll./ syntype/ 2520/ B. C. A., Col. IV, 6. *Cacopsalis rudis* Sharp. Paralectotypes: 1 female specimen (NHM), *Cacopsalis rudis* Type D. S. ♀, Toxpam, Mexico, syntype/ Sharp Coll. 1905-313; 1 female specimen (NHM), *Cacopsalis rudis* Type D. S. Toxpam, Mexico, Salle Coll./ syntype/ B. C. A., Col. IV, 6. *Cacopsalis rudis* Sharp.

Calabresia Alonso-Zarazaga, Lyal, Bartolozzi & Sforzi, 1999

Calabresia Alonso-Zarazaga, Lyal, Bartolozzi & Sforzi (in: Alonso-Zarazaga & Lyal, 1999: 47)
[replacement name for *Euphenges*].

Euphenges Calabresi, 1920b: 62 (non Clark, 1860).

Type species: *Euphenges ceylonicus* Calabresi, 1920.

Distribution. India, Indonesia (Java), Malaysia, Sri Lanka, Vietnam.

aciculata (F. Walker, 1859)

aciculatus F. Walker, 1859: 262 (*Cerobates*).

Type depository. NHM (HT!).

Distribution. Sri Lanka.

ceylonica (Calabresi, 1920)

ceylonicus Calabresi, 1920b: 63 (*Euphenges*).

Type depository. MZUF (HT!).

Distribution. Sri Lanka.

deliberata (Kleine, 1925)

deliberata Kleine, 1925b: 154 (*Euphenges*).

delibratus: Kleine, 1916 (*Euphenges*) [lapsus].

Type depository. NHM (HT!).

Distribution. India.

distributa (Kleine, 1925)

distributa Kleine, 1925b: 154 (*Euphenges*).

Type depository. NHM (HT!).

Distribution. Malaysia.

iridescens (Calabresi, 1920)

iridescens Calabresi, 1920b: 65 (*Euphenges*).

Type depository. MZUF (LT!, PLT!).

Distribution. Malaysia.

Notes. Lectotype of *E. iridescens* (MZUF) designated on a male specimen with the following labels: Is. Penang, A. Boucard/ *Euphenges iridescens* ♂ ♀ typi!/ n. sp./ Bull. Soc. Ent. It., LI, 1920, typi! ♂ ♀ / Prép. génit N° 76 J 10 R. Damoiseau/ ex-coll. Senna/ "La Specola" Firenze, 7016. Paralectotype: 1 female specimen (MZUF), Is. Penang, A. Boucard/ *Euphenges iridescens*/ ex-coll. Senna.

sapaensis Kabakov, 2001

sapaensis Kabakov, 2001: 219.

Type depository. ZIN (HT).

Distribution. Vietnam.

testacea (Kleine, 1942)

testacea Kleine, 1942: 130 (*Euphenges*).

Type depository. MZSF (HT).

Distribution. Indonesia (Java).

Calorychodes Kleine, 1925

Calorychodes Kleine, 1925b: 152.

Type species: *Calorychodes decens* Kleine, 1925 = *Eupsalis tuberculata* Senna, 1894.

Distribution. Myanmar [= Burma].

tuberculatus (Senna, 1894)

tuberculata Senna, 1894b: 373 (*Eupsalis*).

decens Kleine, 1925b: 153 (*Calorychodes*); **syn. n.**

Type depositories. *E. tuberculata*: MNHN (LT!); *C. decens*: NHM (LT!, PLT!).

Distribution. Myanmar [= Burma].

Notes. Lectotype of *E. tuberculata* (MNHN) designated on a male specimen with the following labels: H.te Birmanie, Mines des rubis, 1200-2300 m, Doherty 1890/ type/ Dr. Senna vidit 1893/ *Eupsalis tuberculata* typus! ♂.

Lectotype of *C. decens* (NHM) designated on a female specimen with the following labels: syntype/ Doherty/ 64922/ Birmah Ruby Mines/ Fry Coll. 1905.100/ *Calorychodes decens* Typus! R. Kleine det., 1922/ R. Damoiseau

det., *Calorychodes tuberculatus* (Senna). Paralectotype: 1 male specimen (NHM), syntype/ Doherty/ Birmah, Ruby Mines/ Fry Coll. 1905.100.

After studying the type material, we consider *C. decens* to be a synonym for *C. tuberculatus*.

Catablysmia Kleine, 1926

Catablysmia Kleine, 1926a: 49.

Type species: *Catablysmia funkei* Kleine, 1926.

Distribution. Indonesia (Moluccas: Buru, Halmahera [= Gilolo]), Singapore.

funkei Kleine, 1926

funkei Kleine, 1926a: 49.

Type depository. NHM (HT!).

Distribution. (Moluccas: Halmahera [= Gilolo]).

navigator Damoiseau, 1989

navigator Damoiseau, 1989b: 56.

Type depository. NHM (?) (HT).

Distribution. Singapore.

Notes. Damoiseau (1989b: 56) reported that the holotype is in NHM, but we did not find it there.

stoermeri Kleine, 1925

stoermeri Kleine, 1925f: 25.

Type depository. ZMAN (LT!, PLT!).

Distribution. Indonesia (Moluccas: Buru).

Notes. This species was described in the genus *Catablysmia* before the latter was published.

Lectotype of *C. stoermeri* (ZMAN) designated on a male specimen with the following labels: Toxopeus, Buru Station 9, 1-19.VII.1921. Paralectotype: 1 female specimen (ZMAN), Toxopeus, Buru between st. 1 & 16, 29.VI-VII.1921.

Claeoderes Schoenherr, 1833

Claeoderes Schoenherr, 1833: 362.

Claeoderus: Castelnau, 1840: 294 [lapsus].

Type species: *Claeoderes radulirostris* Boheman, 1833.

Distribution. Belize, Costa Rica, Guatemala, Honduras, Mexico, Nicaragua, Panama; Argentina, Bolivia, Brazil, Colombia, Ecuador, Paraguay, Peru.

radulirostris radulirostris Boheman, 1833

radulirostris radulirostris Boheman (in Schoenherr, 1833: 363).

planicollis Boheman (in Schoenherr, 1833: 365) (*Claeoderes*); syn. Schönfeldt, 1908: 51.

rimabundus Gyllenhal (in Schoenherr, 1833: 364) (*Claeoderes*); syn. Schönfeldt, 1908: 51.

radulirostris: Kleine, 1921a: 49 (*Claeoderes*) [lapsus].

radulirostris: Kleine, 1921a: 49 (*Claeoderes*) [lapsus].

Type depositories. *C. radulirostris radulirostris*: NHRS (HT); *C. planicollis*: NHRS (?) (HT); *C. rimabundus*: NHRS (ST).

Distribution. Brazil.

Notes. Kleine (1928a: 95) reported that the type for *C. planicollis* is in NHRS but the species is not listed in the NHRS web site.

radulirostris biserrirostris Boheman, 1840

radulirostris biserrirostris Boheman (in Schoenherr, 1840: 526).

Type depository. NHRS (HT).

Distribution. Mexico.

***radulirostris mexicanus* Boheman, 1840**

radulirostris mexicanus Boheman (in Schoenherr, 1840: 524).

laevicollis Boheman (in Schoenherr, 1840: 525) (*Claeoderes mexicanus* var.); syn. Gemminger & Harold, 1872: 2712.

bivittatus Boheman (in Schoenherr, 1840: 525) (*Claeoderes mexicanus* var.); syn. Gemminger & Harold, 1872: 2712.

tristis Senna, 1890: 107 (*Claeoderes mexicanus* var.); syn. Schönfeldt, 1908: 51.

guatemalensis Sharp, 1895a: 52 (*Claeoderes*); syn. Damoiseau, 1989b: 61.

guatemalus: Kleine, 1921a: 49 (*Claeoderes*) [lapsus].

kirschi Soares & Scivittaro, 1977: 94 [unjustified replacement name] (*Claeoderes*); **syn. n.**

Type depositories. *C. radulirostris mexicanus*: NHRS (ST); *C. mexicanus* var. *laevicollis*: unknown; *C. mexicanus* var. *bivittatus*: NHRS (ST); *C. mexicanus* var. *tristis*: MZUF (LT!, PLT!); *C. guatemalensis*: NHM (LT!, PLT!), IRSNB (PLT!), MZUF (PLT!).

Distribution. Belize, Costa Rica, Guatemala, Mexico, Nicaragua, Panama; Bolivia, Brazil, Colombia, Ecuador, Paraguay, Peru.

Notes. Lectotype of *C. mexicanus* var. *tristis* (MZUF) designated on a male specimen with the following labels: Amazonas, d. Staudinger/ *Claeoderes mexicanus* var. *tristis*, cotypi, Senna/ Bull. Soc. Ent. It., XXI, 1889, cotypi ♂ ♀/ ex-coll. Senna/ "La Specola" Firenze, 7077. Paralectotype: 1 female specimen (MZUF), Amazonas, d. Staudinger/ *Claeoderes mexicanus* var. *tristis* Senna, Staud. 703, Amazonas/ ex-coll. Senna/ "La Specola" Firenze, 7077.

Lectotype of *C. guatemalensis* (NHM) designated on a male specimen with the following labels: syntype/ Cerro Zunil, 4-5000 ft., Champion/ B. C. A., Col., IV, 6. *Claeoderes guatemalensis*, Sharp/ *Claeoderes guatemalensis* ♂ D. S. Paralectotypes: 25 male and 19 female specimens (NHM), coming from Balheu, San Gerónimo, Purula, Coban, Senahu, San Juan in Vera Paz, El Tumbador, Cerro Zunil, Zapote, Capetillo; 1 male specimen (MZUF), Capetillo, Guatemala, G. C. Champion/ *Claeoderes guatemalensis*/ *Claeoderes guatemalensis* cotypi ♂ ♀ da Sharp/ ex-coll. Senna/ "La Specola" Firenze, 7074; 1 female specimen (MZUF), Cerro Zunil, 4-5000 ft., Champion/ ex-coll. Senna/ "La Specola" Firenze, 7124; 2 male and 1 female specimens (IRSNB), Cerro Zunil, 4-5000 ft., Champion/ Biol. C. Amer., Don Godman et Salvin/ Paratype.

Soares & Scivittaro (1977: 94) gave the new name *kirschi* to *Claeoderes bivittatus* Kirsch, 1867 because the name *bivittatus* was preoccupied by *bivittatus* Boheman, 1840. Kirsch (1867: 219) was not describing a new species but simply quoting the species of Boheman (incorrectly ascribed to Schoenherr). Thus *kirschi* Soares & Scivittaro is an unjustified replacement name.

Corporaalia Kleine, 1921

Corporaalia Kleine, 1921o: 31.

Type species: *Corporaalia baryrrhynchoides* Kleine, 1921.

Distribution. Indonesia (Sumatra).

baryrrhynchoides Kleine, 1921

baryrrhynchoides Kleine, 1921o: 33.

Type depository. MZPW (ST).

Distribution. Indonesia (Sumatra).

Notes. According to the original description (Kleine, 1921o: 33) the types were held in Kleine's collection. Later Kleine (1928a: 84) reported that the types were in ZMAN. We found these specimens in ZMAN: they are indeed labelled as types but the dates and localities do not correspond to the description. According to T. Huflejt (MZPW) (2002, personal communication) a male syntype is held in MZPW.

Cyriodontus Kirsch, 1868

Cyriodontus Kirsch, 1868: 216.

Type species: *Arrhenodes lineatus* Gyllenhal, 1840.

Distribution. Bolivia, Brazil, Colombia, Ecuador, Paraguay, Peru, Venezuela.

certus Kleine, 1927

certus Kleine, 1927b: 429.

sertus: Kleine, 1938d: 107 (*Cyriodontus*) [lapsus].

Type depository. ZMUH (HT).

Distribution. Brazil.

circumscriptus Kleine, 1927*circumscriptus* Kleine, 1927b: 430.**Type depository.** ZMUH (HT).**Distribution.** Brazil.*erraticus* Kleine, 1927*erraticus* Kleine, 1927b: 431.**Type depository.** NHM (HT!).**Distribution.** "S America".*evanidus* Kleine, 1927*evanidus* Kleine, 1927b: 433.**Type depository.** NHM (LT!, PLT!).**Distribution.** Ecuador, Peru.

Notes. Lectotype of *C. evanidus* (NHM) designated on a male specimen with the following labels: syntype/ Sarayacu, Peru/ Pascoe coll., 93-60/ *Cyriodontus evanidus* ♂ type Kln., 1923, R. Kleine det./ *Cyriodontus inermicollis* ♂ Pow. Paralectotype: 1 female specimen (NHM), syntype/ cotype/ 52275/ Buckley/ Ecuador/ Fry Coll. 1905.100/ *Cyriodontus evanidus* ♀ typus! Kln., 1923, R. Kleine det.

exactus Kleine, 1927*exactus* Kleine, 1927b: 432.**Type depository.** NHM (LT!, PLT!).**Distribution.** Ecuador.

Notes. Lectotype of *C. exactus* (NHM) designated on a male specimen with the following labels: syntype/ Buckley/ Ecuador/ *Cyriodontus exactus*

typus ♂ Kln., 1923, R. Kleine det./ *Cyriodontus aureonotatus* ♂, T. Pow.
 Paralectotype: 1 female specimen (NHM), syntype/ Buckley/ Ecuador/ Fry
 Coll.1905.100/ *Cyriodontus exactus* ♀ Kln., 1923, R. Kleine det.

***guttatus* Kleine, 1917**

guttatus Kleine, 1917a: 229.

Type depositories. MNHUB (ST!), ZMUH (ST).

Distribution. Ecuador.

Notes. Heads are missing from the two syntypes of MNHUB.

***lineatus* (Gyllenhal, 1840)**

lineatus Gyllenhal (in Schoenherr, 1840: 476) (*Arrhenodes*).

Type depository. NHRS (HT).

Distribution. Bolivia, Colombia, Ecuador, Paraguay, Peru, Venezuela.

***weyrauchi* Haedo Rossi, 1965**

weyrauchi Haedo Rossi, 1965: 324.

Type depository. IMLA (HT, AT).

Distribution. Peru.

***Damoiysiella* Alonso-Zarazaga, Lyal,
 Bartolozzi & Sforzi, 1999**

Damoiysiella Alonso-Zarazaga, Lyal, Bartolozzi & Sforzi (in Alonso-Zarazaga & Lyal, 1999: 48).
Orychodes Kleine, 1921b: 62 [misidentification; Alonso Zarazaga & Lyal, 1999: 48].

Type species: *Orychodes pictus* Pascoe, 1862.

Distribution. Borneo, Indonesia (Moluccas: Bacan [= Batjan]; Sulawesi),
 Malaysia, Myanmar [= Burma], Vietnam.

breviceps (Senna, 1894)

breviceps Senna, 1894b: 380 (*Orychodes*).

Type depository. MNHN (HT!).

Distribution. Malaysia, Myanmar [= Burma], Vietnam.

continua (Kleine, 1926)

continuus Kleine, 1926a: 46 (*Orychodes*).

continuis: Kleine, 1938d: 105 (*Orychodes*) [lapsus].

Type depository. NHM (HT!).

Distribution. Borneo.

picta (Pascoe, 1862)

pictus Pascoe, 1862: 389 (*Orychodes*).

Type depository. NHM (LT!, PLT!).

Distribution. Indonesia (Moluccas: Bacan [= Batjan]).

Notes. Lectotype of *O. pictus* (NHM) designated on a male specimen with the following labels: syntype/ Batchian/ Pascoe Coll. 93-60/ *Orychodes pictus* Pascoe. Paralectotypes: 1 male and 2 female specimens (NHM), syntype/ Batchian/ Pascoe Coll. 93-60.

Debora Power, 1879

Debora Power, 1879a: 490.

Type species: *Debora bocandei* Power, 1879.

Distribution. Angola, Cameroon, Central African Republic, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea, Gabon, Ghana, Ivory Coast, Nigeria, Republic of the Congo, Republic of Guinea, Sierra Leone, Togo.

bocandei Power, 1879

bocandei Power, 1879a: 490.

thomsoni Power, 1879a: 491 (*Debora*); syn. Kleine, 1927a: 38.

Type depositories. *D. bocandei*: IRSNB (LT!, PLT!); *D. thomsoni*: MNHN (?) (ST).

Distribution. Angola, Cameroon, Central African Republic, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea, Gabon, Ghana, Ivory Coast, Nigeria, Republic of the Congo, Republic of Guinea, Sierra Leone, Togo.

Notes. Lectotype of *D. bocandei* designated by Damoiseau (1967a: 342).

We did not find any syntype of *D. thomsoni* in MNHN neither in IRSNB, where many types of the Oberthür collection are held.

Damoiseau (1967a: 342) did not mention Nigeria (Senna, 1894a: 410) when he gave the species distribution.

forficata (J. Thomson, 1858)

forficatus J. Thomson, 1858: 118 (*Arrhenodes*).

Type depository. MNHN (LT!, PLT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea, Gabon, Ghana, Ivory Coast, Nigeria.

Notes. Damoiseau (1967a: 343) reported that the specimen held in MNHN is the holotype, but in his original description J. Thomson did not designate any of the syntypes as holotype, however, as there is more than one syntype in MNHN, and as Damoiseau did not specifically select one as holotype, we designate the lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Lectotype of *A. forficatus* (MNHN) designated on a male specimen with the following labels: Gabon/ Mniszech/ type/ ex Musaeo Power. Paralectotypes: 1 female specimen (MNHN), Gabon/ Thomson ♀ 17/ type/ ex Musaeo Power; 1 male specimen, Gabon/ Mniszech 33/ ex Musaeo Power.

Damoiseau (1967a: 343) did not mention Nigeria (Senna, 1894a: 412) when he gave the species distribution. The quotation for the People's Republic of Congo (Sforzi, 1998b: 268) is incorrect and should be referred to as the Democratic Republic of the Congo (ex Zaire).

Desgodinsia Senna, 1894

Desgodinsia Senna, 1894b: 381.

Type species: *Desgodinsia spinirostris* Senna, 1894.

Distribution. Borneo, India (including Assam), Malaysia, Myanmar [= Burma], Vietnam (?).

Notes. Kabakov (2001: 213) quotes the genus *Desgodinsia* as a query for Vietnam.

ambigua Kleine, 1923

ambigua Kleine, 1923c: 130.

Type depository. NHM (HT!).

Distribution. Borneo, Malaysia.

Notes. Kleine (1928a: 86) incorrectly reported that the holotype was in CUMZ.

spinirostris Senna, 1894

spinirostris Senna, 1894b: 382.

Type depository. MNHN (HT!).

Distribution. India (including Assam), Malaysia, Myanmar [= Burma].

Ectocemus Pascoe, 1862

Ectocemus Pascoe, 1862: 388.

Megacerus Montrouzier, 1855: 36 (non Burmeister, 1847); syn. Lacordaire, 1966: 434.

Megaterus: Kleine, 1927j: 158 [lapsus].

Ectocnemus: Zimmerman, 1994: 1 [lapsus].

Ectochemus: Zborowki & Storey, 2003: 128 [lapsus]

Type species: *Ectocemus wallacei* Pascoe, 1862 = *Megacerus decemmaculatus* Montrouzier, 1855.

Distribution. Borneo, India (including Andaman Is.), Indonesia (Batu; Java; Mentawai; Moluccas: Aru, Bacan [= Batjan], Halmahera [= Gilolo], Morotai; Nias; Sumatra; Sulawesi), Kampuchea [= Cambodia], Malaysia, Philippines (Luzon, Mindanao); Sri Lanka, Thailand, Vietnam; Australia, New Caledonia, New Guinea (including Bismarck Archipelago: Duke of York, New Britain, New Ireland; Saint Matthias Group, Woodlark), Solomon Is.

cinnamomi (Herbst, 1783)

cinnamomi Herbst, 1783: 76 (*Curculio*).

cinnamomeus: Herbst, 1797: 174 (*Brentus*) [lapsus].

spinirostris F. Walker, 1859: 262 (*Nemocephalus*); syn. Kleine, 1923f: 90.

spatulirostris Chevrolat, 1866: pl. 77, fig. 3 (*Ectocemus*); syn. Kleine, 1926a: 51.

pubescens Kirsch, 1875b: 47 (*Megacerus*); syn. Calabresi, 1921a: 10.

quatuordentulus Desbrochers des Loges, 1890: 219 (*Megacerus*); syn. Calabresi, 1921a: 10.

cynnamomi: Senna, 1892a: 177 (*Orychodes*) [lapsus].

tuberculatus Senna, 1903: 175 (*Orychodes cinnamomi* var.); syn. Schönfeldt, 1908: 41.

quattuordentulus: Schönfeldt, 1908: 43 (*Ectocemus*) [lapsus].

quatuordentatus: Calabresi, 1921a: 10 (*Ectocemus*) [lapsus].

condensa Kleine, 1926a: 48 (*Heteroblysmia*); **syn. n.**

quatuornotatus: Kleine, 1927a: 51 (*Ectocemus*) [lapsus].

pubens: Kleine, 1938d: 117 (*Ectocemus*) [lapsus].

Type depositories. *C. cinnamomi*: MNHUB (HT!); *N. spinirostris*: NHM (LT!, PLT!); *E. spatulirostris*: MNHN (?) (HT); *M. pubescens*: SMTD (ST); *M. quatuordentulus*: MNHN (?) (ST), NZSI (?) (ST); *O. cinnamomi* var. *tuberculatus*: MSNG (LT!, PLT!); *H. condensa*: NHM (HT!).

Distribution. Borneo, India (Andaman Is.), Indonesia (Batu, Java, Mentawai, Nias, Sumatra, Sulawesi), Malaysia, Sri Lanka, Thailand, Vietnam.

Notes. Lectotype of *N. spinirostris* (NHM) designated on a male specimen with the following labels: syntype/ Ceylon/ *Nemocephalus spinirostris* Walker a. n. H. (Type)/ *Ectocemus cinnamomi* Herbst, R. Kleine det. 1922. Paralectotype: 1 male specimen (NHM), syntype/ Ceylon/ *spinirostris*/ *Nemocephalus spinirostris* Walker a. n. H. (Type).

Lectotype of *O. cinnamomi* var. *tuberculatus* (MSNG) designated on a male specimen with the following labels: Sumatra, Siboga X.90 e III.91, E. Modigliani/ *Orychodes cinnamomi* (Herbst) *tuberculatus* Senna. Paralectotypes: 3 male and 2 female specimens (MSNG), Sumatra, Siboga, X.90 and III.91, E. Modigliani.

We did not find any types of *E. spathulirostris* and *M. quatuordentulus* in MNHN neither in IRSNB, where many types from Oberthür collection are held.

After studying the type material, we consider *H. condensa* to be a synonym of *E. cinnamomi*.

contractus Kleine, 1925

contractus Kleine, 1925b: 158.

Type depository. NHM (HT!).

Distribution. Kampuchea [= Cambodia], Thailand.

decemmaculatus (Montrouzier, 1855)

decemmaculatus Montrouzier, 1855: 37 (*Megacerus*).

wallacei Pascoe, 1862: 388 (*Ectocemus*); syn. Senna, 1892c: 54.

badenii Kirsch, 1875b: 48 (*Megacerus*); syn. Damoiseau, 1966a: 15.

pulchellus Kirsch, 1875b: 49 (*Megacerus*); syn. Schönfeldt, 1908: 44.

granulirostris Gestro, 1876: 519 (*Ectocemus*); syn. Damoiseau, 1966a: 15.

pterygorhinus Gestro, 1876: 519 (*Ectocemus*); syn. Schönfeldt, 1908: 44.

ruficauda Bates, 1877: 156 (*Ectocemus*); syn. Fairmaire, 1883: 42.

decemmaculatus Fairmaire, 1883: 42 (non Montrouzier, 1855) (*Ectocemus*); syn. Senna, 1892c: 54.

pterygorrhinus: Schönfeldt, 1908: 44 (*Ectocemus*) [lapsus].

badeni: Schönfeldt, 1908: 43 (*Ectocemus*) [lapsus].

decimmaculatus: Zimmerman, 1994: 17 (*Ectocnemus*) (sic!) [lapsus].

Type depositories. *M. decemmaculatus*: IRSNB (HT!); *E. wallacei*: NHM (LT!, PLT!); *M. badenii*: SMTD (HT); *M. pulchellus*: SMTD (HT); *E. granulirostris*: MSNG (LT!, PLT!); *E. pterygorrhinus*: MSNG (LT!, PLT!); *E. ruficauda*: unknown.

Distribution. Borneo, Indonesia (Moluccas: Aru, Bacan [= Batjan], Halmahera [= Gilolo], Morotai; Sulawesi), Philippines (Luzon, Mindanao); Australia, New Caledonia, New Guinea (including Bismarck Archipelago: Duke of York, New Britain, New Ireland; Saint Matthias Group, Woodlark), Solomon Is.

Notes. Lectotype of *E. wallacei* (NHM) designated on a male specimen with the following labels: syntype/ Batchian/ Pascoe Coll. 93-60/ *Ectocemus*

Wallacei Pasc. Type/ *Ectocemus Wallacei* Pascoe, Moluccas. Paralectotypes: 1 male specimen (NHM), syntype/ Batchian/ Pascoe Coll. 93-60; 1 female specimen (NHM), syntype/ Pascoe Coll. 93-60.

Lectotype of *E. granulirostris* (MSNG) designated on a male specimen with the following labels: Molucche, Halmahera, coll. Bruijn 1875/ *Ectocemus granulirostris* Gestro. Paralectotypes: 1 male and 1 female specimen (MSNG), Morotai, L. Deys, 1867.

Lectotype of *E. pterygorhinus* (MSNG) designated on a male specimen with the following labels: Somerset, I.75, L. M. D'Albertis/ *Ectocemus pterygorhinus* Gestro tipo! Paralectotypes: 7 male and 8 female specimens (MSNG), Australia, Somerset, I.75, L. M. D'Albertis.

madrasanus Kleine, 1931

madrasanus Kleine, 1931a: 96.

Type depository. NHM (LT!, PLT!).

Distribution. S India.

Notes. Lectotype of *E. madrasanus* (NHM) designated on a male specimen with the following labels: syntype/ S. Mangalore, 1200, Madras, J. C. M. Gardner, 24.V.1930/ India, B. M. 1931-84/ *Ectocemus madrasanus* Kln Type, R. Kleine det.; Paralectotype: 1 female specimen (NHM), syntype/ S. Mangalore, 1200, Madras, J. C. M. Gardner, 24.V.1930/ India, B.M. 1931-84/ *Ectocemus madrasanus* Kln Type, R. Kleine det.

malcheri Kleine, 1935

malcheri Kleine, 1935b: 175.

Type depository. SMTD (HT).

Distribution. Solomon Is.

Elytracantha Kleine, 1915

Elytracantha Kleine, 1915b: 239.

Type species: *Belopherus pogonocerus* Montrouzier, 1855.

Distribution. Indonesia (Moluccas: Bacan [= Batjan], Buru, Halmahera, Morotai, Seram [= Ceram]); New Guinea (including Bismarck Archipelago: Duke of York, New Britain; Woodlark; Yule Is.), Solomon Is.

cerbera Kleine, 1919

cerbera Kleine, 1919a: 37.

Type depository. MNHUB (LT!, PLT!).

Distribution. Indonesia (Moluccas: Buru); New Guinea (including Bismarck Archipelago: New Britain).

Notes. Lectotype of *E. cerbera* (MNHUB) designated on a male specimen with the following labels: Sattelberg/ Typus/ 111491/ *Elytracantha cerberus* Kl. Paralectotype: 1 female specimen (MNHUB), Neu Pommern/ Typus/ 111491.

contraria Kleine, 1926

contraria Kleine, 1926a: 51.

Type depository. NHM (LT!, PLT!).

Distribution. Indonesia (Moluccas: Bacan [= Batjan], Seram [= Ceram]).

Notes. Lectotype of *E. contraria* (NHM) designated on a male specimen with the following labels: syntype/ Batchian/ Pascoe Coll. 93-60/ *Elytracantha contraria* Kln. ♂ Type 1922, R. Kleine det. Paralectotype: 1 female specimen (NHM), with the same label data as the lectotype.

pogonocera (Montrouzier, 1855)

pogonocerus Montrouzier, 1855: 37 (*Belopherus*).

spinipennis Fairmaire, 1881a: 349 (*Ectocemus*); syn. Fairmaire, 1883: 43.

Type depositories. *B. pogonocera*: IRSNB (HT!); *E. spinipennis*: unknown.

Distribution. Indonesia (Moluccas: Halmahera, Morotai); New Guinea (including Bismarck Archipelago: Duke of York, New Britain; Woodlark; Yule Is.), Solomon Is.

Epicoinoneus Senna, 1892

Epicoinoneus Senna, 1892e: 476.

Epicoenoneus Senna, 1898d: 62 [unjustified emendation].

Epicononeus: Kabakov, 2001: 207 [lapsus].

Type species: *Epicoinoneus femoralis* Senna, 1892.

Distribution. Borneo, Indonesia (Java, Sumatra), Myanmar [= Burma], Vietnam.

femoralis Senna, 1892

femoralis Senna, 1892e: 478.

Type depository. MSNG (HT!).

Distribution. Borneo, Indonesia (Java, Sumatra), Myanmar [= Burma], Vietnam.

Episphales Kirsch, 1871

Episphales Kirsch, 1871: 378 [replacement name for *Automolus*].

Automolus Kirsch, 1868: 218 (non Reichenbach, 1853, nec Burmeister, 1855).

Eusphales: Sharp, 1895a: 36 [lapsus].

Episphalis: Kleine, 1920a: 43 [lapsus].

Type species: *Automolus pictus* Kirsch, 1868.

Distribution. Colombia, Venezuela.

crucifer Damoiseau, 1963

crucifer Damoiseau, 1963f: 17.

Type depository. IRSNB (HT!, AT!, PT!).

Distribution. Venezuela.

pictus (Kirsch, 1868)

pictus Kirsch, 1868: 218 (*Automolus*).

Type depository. SMTD (ST).

Distribution. Colombia, Venezuela.

Estenorhinus Lacordaire, 1866

Estenorhinus Lacordaire, 1866: 431.

Arrhenodes Schoenherr, 1833: 36 (non Schoenherr, 1826); syn. Damoiseau, 1963f: 9.

Arrhenodes Lacordaire, 1866: 429 [not available, misidentification].

Estenorrhinus: Gemminger & Harold, 1872: 2709 [lapsus].

Hosendarre Kuntzen, 1937: 197 [not available, see Alonso-Zarazaga & Lyal, 1999: 48].

Errhanodes Kuntzen, 1937: 197; syn. Damoiseau, 1963f: 9.

Pararrhenodes Haedo Rossi, 1954a: 2; syn. Soares, 1966a: 271.

Vianodes Haedo Rossi, 1955: 62; syn. Damoiseau, 1963f: 9.

Type species: *Arrhenodes forficatus* Gyllenhal, 1833.

Distribution. Belize, Costa Rica, Dominican Republic, Guatemala, Mexico, Nicaragua, Panama, Trinidad; Argentina, Bolivia, Brazil, Colombia, Ecuador, French Guyana, Paraguay, Peru, Venezuela.

Notes. Alonso-Zarazaga & Lyal (1999: 48) listed *Pararrhenodes* Haedo Rossi among the valid genera, but we consider it as a synonym of *Estenorhinus* Lacordaire, as already indicated by Soares (1966a: 271).

angulicollis (Gyllenhal, 1833)

angulicollis Gyllenhal (in Schoenherr, 1833: 318) (*Arrhenodes*).

Type depository. NHRS (HT).

Distribution. Mexico, Trinidad; Argentina, Brazil, Ecuador, French Guyana, Paraguay, Peru.

angustirostris (Haedo Rossi, 1954)

angustirostris Haedo Rossi, 1954a: 2 (*Arrhenodes*).

Type depository. MLPA (HT).

Distribution. Argentina, Brazil.

appositus Kleine, 1923

appositus Kleine, 1923h: 51.

misionensis Haedo Rossi, 1954a: 2 (*Pararrhenodes*); syn. Soares, 1966a: 271.

misionera: Haedo Rossi, 1961: 160 (*Pararrhenodes*) [lapsus].

Type depositories. *E. appositus*: NMPC (HT); *P. misionensis*: IMLA (HT, PT).

Distribution. Argentina, Brazil, Paraguay.

Notes. In all papers published after the original description, Haedo Rossi quoted *Pararrhenodes misionensis* under the name of *P. misionera*.

bomansi Damoiseau, 1989

bomansi Damoiseau, 1989b: 58.

Type depository. IRSNB (HT!, AT!, PT!).

Distribution. Peru.

columbianus Damoiseau, 1989

columbianus Damoiseau, 1989b: 57.

Type depositories. MNHUB (HT!), IRSNB (AT!).

Distribution. Colombia.

concolor (Sharp, 1895)

concolor Sharp, 1895a: 45 (*Arrhenodes*).

Type depository. NHM (LT!, PLT!).

Distribution. Costa Rica, Mexico, Nicaragua, Panama.

Notes. Lectotype of *A. concolor* (NHM) designated on a male specimen with the following labels: *Arrhenodes concolor* ♂ Type D. S./ Chontales, Janson/ syntype/ type/ B. C. A., Col., IV, 6. *Arrhenodes concolor* Sharp/ Sharp Coll. 1905-313. Paralectotypes: 14 male and 10 female specimens (NHM), coming from Mexico, Toxpan and Bobo; Nicaragua, Chontales; Panama, Tolé.

confrater (Kleine, 1944)

confrater Kleine, 1944: 154 (*Arrhenodes*).

Type depositories. MNHN (LT!, PLT!), DEI (PLT!).

Distribution. Brazil, French Guyana.

Notes. Döbler & Gaedike (1995: 442) reported that the specimens held in MNHN and in Kleine's collection are holotype and paratypes, but in the original paper the species was described from 2 males and 3 females in MNHN, therefore they must be considered as syntypes; however, as there is more than one syntype in MNHN, and as Döbler & Gaedike did not specifically select one as holotype, we designate the lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Lectotype of *A. confrater* (MNHN) designated on a male specimen with the following labels: Museum Paris, Guyane Française, Rey 1909/ type/ *Arrhenodes confrater* Kln. R. Kleine det. 1941. Paralectotypes: 1 female specimen (MNHN), Brésil, Ét. de Sao Paulo, Val. du Rio Pardo, E. Gounelle, 12-98/ cotype/ Museum Paris coll. E. Gounelle 1915/ *Arrhenodes confrater* Kln. R. Kleine det. 1941; 1 male specimen (DEI), Brésil, Ét. de Sao Paulo, Val. du Rio Pardo, E. Gounelle, 12-98/ paratypus/ *Arrhenodes confrater* Kl.; 1 female specimen (DEI), Brésil, Ét. de Sao Paulo, Ribeirao Pires, E. Gounelle, 11-98/ paratype.

cyriodontoides (Soares, 1966)

cyriodontoides Soares, 1966a: 272 (*Vianodes*).

Type depository. MNRJ (HT).

Distribution. Brazil.

***denticollis* (Gyllenhal, 1833)**

denticollis Gyllenhal (in Schoenherr, 1833: 320) (*Arrhenodes*).

Type depository. NHRS (HT).

Distribution. Costa Rica; Argentina, Brazil, French Guyana.

***depressus* (Meyer, 1960)**

depressus Meyer, 1960: 143 (*Vianodes*).

crassus Haedo Rossi, 1965: 335 (*Vianodes*); syn. Soares & Scivittaro, 1979a: 165.

Type depositories. *V. depressus*: MNRJ (Campos Seabra coll.) (HT, PT); *V. crassus*: CSMBA (HT).

Distribution. Bolivia, Brazil.

***designatus* (Boheman, 1840)**

designatus Boheman (in Schoenherr, 1840: 466) (*Arrhenodes*).

Type depositories. IRSNB (LT!), NHRS (PLT).

Distribution. Colombia, Ecuador, Peru, Venezuela.

Notes. Lectotype of *A. designatus* (IRSNB) designated on a male specimen with the following labels: n. Gr./ Coll. Dej. an Coll. Roelofs/ ♂/ *designatus* Schoenherr/ M. R. Belg./ *Arrhenodes designatus* Schoenherr, h. in Colombia, D. Schoenherr/ *Estenorrhinus designatus* Bohem typique suivant la description/ *Estenorrhinus designatus* Boh. R. Damoiseau det., 1962/ Type.

***dispar* (Linné, 1758)**

dispar Linné, 1758: 382 (*Curculio*).

anomaloceps Pallas, 1781: 24 (*Curculio*); syn. Schoenherr, 1840: 470.

bifrons Fabricius, 1787: 96 (*Brentus*); syn. Schoenherr, 1833: 315.

maxillosus Herbst, 1797: 177 (*Brentus*); syn. Schoenherr, 1833: 315.

volvulus Panzer, 1798: 44 (non Fabricius, 1792) (*Brentus*); syn. Schoenherr, 1833: 315.

decoratus Perty, 1832: 68 (*Arrhenodes*); syn. Schoenherr, 1840: 470.

chevrolati Boheman (in Schoenherr, 1840: 469) (*Arrhenodes*); syn. Kleine, 1927a: 40.

anomalocephalus: Schönfeldt, 1908: 35 (*Arrhenodes*) [lapsus].

mourei Haedo Rossi, 1965: 344 (*Vianodes*); syn. Soares & Scivittaro, 1979a: 167.

signaticollis Haedo Rossi, 1965: 348 (*Vianodes*); syn. Soares & Scivittaro, 1979a: 167.

Type depositories. *C. dispar*: UZIU (?); *C. anomalocephalus*: unknown; *B. bifrons*: unknown; *B. maxillosus*: MNHUB (Herbst coll.) (?) (HT); *A. decoratus*: ZSM (?) (HT); *A. chevrolati*: NHRS (ST); *V. mourei*: JHRC (HT); *V. signaticollis*: JHRC (HT).

Distribution. "Antilles", Costa Rica, Mexico; Argentina, Bolivia, Brazil, Ecuador, French Guyana, Paraguay, Peru, Venezuela.

Notes. We did not find any types of *B. maxillosus* in MNHUB, where Herbst collection is held. Concerning *B. bifrons*, Zimsen (1964: 223) does not indicate where the type is held.

Kleine (1928a: 82) reported that the type for *A. decoratus* was in ZSM, but according to M. Baehr (ZSM) (2001, personal communication) these types are missing; they were probably lost during World War II like many other Peruvian types.

ecuadorensis Kleine, 1931

ecuadorensis Kleine, 1931a: 95 (*Estenorrhinus*).

Type depository. SMTD (HT, PT).

Distribution. Ecuador.

elegans (Erichson, 1847)

elegans Erichson, 1847: 126 (*Arrhenodes*).

sexvittatus Senna, 1890: 104 (*Arrhenodes elegans* var.); syn. Schönfeldt, 1908: 35.

Type depositories. *A. elegans*: MNHUB (?) (HT); *A. elegans* var. *sexvittatus*: unknown.

Distribution. Panama; Bolivia, Brazil, Colombia, Peru.

Notes. Kleine (1928a: 83) reported that the type for *A. elegans* was in MNHUB, but we did not find it there.

evidens Kleine, 1927

evidens Kleine, 1927b: 427.

Type depositories. NHM (LT!, PLT!), SMTD (PLT).

Distribution. Brazil.

Notes. Lectotype of *E. evidens* (NHM) designated on a male specimen with the following labels: syntype/ Ilha Santo Amaro nr. Santos, Brazil, G. E. Bryant, 25.III.1912/ *Estenorhinus evidens* Kln ♂ Typus!, R. Kleine det., 1923. Paralectotypes: 1 female specimen (NHM), syntype/ Ilha Santo Amaro nr. Santos, Brazil, G. E. Bryant, 3.IV.1912/ *Estenorhinus evidens* Kln, Typus!, R. Kleine det., 1923; 1 female specimen (NHM), syntype/ Corcovado, Rio de Janeiro, G. E. Bryant, 8.V.1912.

expilator Damoiseau, 1989

expilator Damoiseau, 1989b: 58.

Type depositories. IRSNB (HT!, AT!, PT!), HNHM (PT), MZLU (PT).

Distribution. Bolivia, Peru.

exsertus (Gyllenhal, 1833)

exsertus Gyllenhal (in Schoenherr, 1833: 319) (*Arrhenodes*).

rugosus Gyllenhal (in Schoenherr, 1840: 472) (*Arrhenodes*); syn. Kleine, 1927a: 40.

Type depositories. *A. exsertus*: IRSNB (ST!), NHRS (ST); *A. rugosus*: NHRS (HT).

Distribution. Trinidad; Brazil, French Guyana, Peru.

facetus (Kleine, 1927)

facetus Kleine, 1927b: 426 (*Arrhenodes*).

Type depository. NHM (HT!).

Distribution. Brazil.

faldermanni (Gyllenhal, 1840)

faldermanni Gyllenhal (in Schoenherr, 1840: 483) (*Arrhenodes*).

corniger Gyllenhal (in Schoenherr, 1840: 484) (*Arrhenodes*); syn. Gemminger & Harold, 1872: 2709.

Type depositories. *A. faldermanni*: NHRS (?) (HT); *A. corniger*: NHRS (HT).

Distribution. Mexico; Brazil, Paraguay.

Nota: Blackwelder (1947: 773, 1982: 773) listed *singularis* Lacordaire, 1866 as a synonym, but this name is a *nomen nudum*, in that Lacordaire simply quoted *singularis* Dejean (in turn *nomen nudum*) as a synonym, without any description.

Kleine (1928a: 83) reported that the type for *A. faldermanni* was in NHRS but it is not listed in the NHRS web site.

flaccus (Meyer, 1959)

flaccus Meyer, 1959a: 125 (*Vianodes*).

Type depository. MNRJ (Campos Seabra coll.) (HT).

Distribution. Brazil.

flavolineatus (Gyllenhal, 1840)

flavolineatus Gyllenhal (in Schoenherr, 1840: 473) (*Arrhenodes*).

Type depositories. NHRS (ST).

Distribution. Belize, Costa Rica, Guatemala, Mexico, Nicaragua, Panama; Ecuador.

forficatus (Gyllenhal, 1833)

forficatus Gyllenhal (in Schoenherr, 1833: 314) (*Arrhenodes*).

monilifer Boheman (in Schoenherr, 1840: 467) (*Arrhenodes*); syn. Kleine, 1921d: 281.

Type depositories. *A. forficatus*: NHRS (HT); *A. monilifer*: NHRS (ST).

Distribution. Costa Rica; Bolivia, Brazil, Colombia, Ecuador, French Guyana, Peru.

formosus Senna, 1897

formosus Senna, 1897a: 231 (*Estenorrhinus*).

Type depository. IRSNB (HT!).

Distribution. Colombia, Ecuador.

funebis (Sharp, 1895)

funebis Sharp, 1895a: 44 (*Arrhenodes*).

Type depositories. NHM (LT!, PLT!), IRSNB (PLT!), MZUF (PLT!).

Distribution. Panama, Guatemala; Ecuador.

Notes. Lectotype of *A. funebis* (NHM) designated on a male specimen with the following labels: *Arrhenodes funebis* Type D. S./ V. de Chiriqui, 25-4000 ft., Champion/ syntype/ type/ Sharp Coll. 1905-313/ B. C. A., Col., IV, 6. *Arrhenodes funebis* Sharp. Paralectotypes: 35 male and 23 female specimens (NHM), coming from Panama, Bugaba and V. de Chiriqui; 1 male specimen (MZUF), V. de Chiriqui, 25-4000 ft., Champion/ *Claeoderes funebis*/ Biol. Centr. Am., IV 1895, cotypi, D. Sharp ♂ ♀/ ex-coll. Senna/ "La Specola" Firenze, 7002; 1 female specimen (MZUF), V. de Chiriqui, 25-4000 ft., Champion/ ex-coll. Senna/ "La Specola" Firenze, 7002; 1 male and 1 female specimen (MZUF), Chiriqui, Amer. centr., ex Staudinger/ ex-coll. Senna/ "La Specola" Firenze, 7002; 1 male specimen (IRSNB), V. de Chiriqui, 25-4000 ft., Champion/ *Arrhenodes funebis*/ Biol. C. Amer., Don Godman et Salvin/ Prépar. génit. No 787 R. Damoiseau/ Paratype; 1 male and 1 female specimen (IRSNB), V. de Chiriqui, 25-4000 ft., Champion/ Biol. C. Amer., Don Godman et Salvin/ Paratype.

gnatho (Lichtenstein, 1796)

gnatho Lichtenstein, 1796: 53 (*Brentus*).

ruficollis Lichtenstein, 1796: 53 (*Brentus*); syn. Gemminger & Harold, 1872: 2708.

Type depositories. *B. gnatho*: MNHUB (?) (ST); *B. ruficollis*: MNHUB (?) (ST).

Distribution. Argentina, Bolivia, Brazil, Colombia, Ecuador, Peru.

Notes. We did not find any types of *B. gnatho* and *B. ruficollis* in MNHUB, where Lichtenstein collection is held.

goudoti (Kirsch, 1867)

goudoti Kirsch, 1867: 215 (*Arrhenodes*).

willineri Haedo Rossi, 1965: 339 (*Vianodes*); syn. Soares & Scivittaro, 1979a: 165.

willneri: Soares & Scivittaro, 1979a: 165 (*Vianodes*) [lapsus].

Type depositories. *A. goudoti*: SMTD (ST); *V. willineri*: JHRC (HT, PT), CSMBA (PT).

Distribution. Costa Rica, Guatemala, Mexico, Panama; Bolivia, Brazil, Colombia, Ecuador, Peru.

guttatus Sharp, 1895

guttatus Sharp, 1895a: 43.

Type depository. NHM (LT!, PLT!).

Distribution. Costa Rica, Nicaragua, Panama.

Notes. Lectotype of *E. guttatus* (NHM) designated on a male specimen with the following labels: *Estenorhinus guttatus* ♂ Type D. S./ V. de Chiriqui, 2-3000 ft., Champion/ syntype/ type/ B. C. A., Col., IV, 6. *Estenorhinus guttatus* Sharp/ Sharp Coll. 1905-313. Paralectotypes: 7 male and 8 female specimens (NHM), coming from Nicaragua, Chontales and Panama, V. de Chiriqui.

longirostris (Meyer, 1961)

longirostris Meyer, 1961: 61 (*Vianodes*).

Type depository. MNRJ (Campos Seabra coll.) (HT).

Distribution. Brazil.

melancholicus (Gyllenhal, 1833)

melancholicus Gyllenhal (in Schoenherr, 1833: 321) (*Arrhenodes*).

trilineatus Kirsch, 1868: 215 (*Arrhenodes*); syn. Kleine, 1927a: 41.

Type depositories. *A. melancholicus*: NHRS (ST); *A. trilineatus*: SMTD (HT).

Distribution. Brazil, Colombia, Ecuador.

Notes. Kleine (1938d) did not mention *A. trilineatus* among the species listed in the Genera Insectorum.

nitidissimus (Meyer, 1959)

nitidissimus Meyer, 1959b: 299 (*Pararrhenodes*).

Type depository. MNRJ (Campos Seabra coll.) (HT).

Distribution. Brazil.

nordestinus (Meyer, 1960)

nordestinus Meyer, 1960: 145 (*Vianodes*).

Type depository. MNRJ (Campos Seabra coll.) (HT, AT).

Distribution. Brazil.

ornatus (Gyllenhal, 1840)

ornatus Gyllenhal (in Schoenherr, 1840: 475) (*Arrhenodes*).

Type depository. NHRS (HT).

Distribution. Colombia.

***perforatus* (Blanchard, 1843)**

perforatus Blanchard, 1843: 201 (*Arrhenodes*).

Type depository. MNHN (?) (HT).

Distribution. Bolivia.

Notes. Kleine (1938d) did not mention *A. perforatus* among the species listed in the Genera Insectorum. We did not find the holotype of this species in MNHN, but we are not sure that this Museum is the true type depository.

***peruvianus* Damoiseau, 1989**

peruvianus Damoiseau, 1989b: 58.

Type depository. HNHM (HT).

Distribution. Peru.

***quadridentaticollis* (Soares, 1966)**

quadridentaticollis Soares, 1966a: 273 (*Vianodes*).

Type depositories. IPEACSM (HT), MNRJ (Campos Seabra coll.) (AT, PT).

Distribution. Brazil.

***quadrifasciatus* Senna, 1897**

quadrifasciatus Senna, 1897a: 232 (*Estenorrhinus*).

nuadrisignatus: Kleine, 1931n: 238 (*Estenorrhinus*) [lapsus].

quadrifasciatus: Kleine, 1938b: 72 (*Estenorrhinus*) [lapsus].

Type depository. MZUF (HT!).

Distribution. Colombia.

***similis* (Haedo Rossi, 1954)**

similis Haedo Rossi, 1954a: 2 (*Arrhenodes*).

angulicollis Haedo Rossi, 1961: 141 (non Gyllenhal, 1833) (*Vianodes*); syn. Soares & Scivittaro, 1979a: 166.

liae Meyer, 1961: 62 (*Vianodes*); syn. Soares & Scivittaro, 1979a: 166.

paraguayana Haedo Rossi, 1967: 161 (*Vianodes*); syn. Soares & Scivittaro, 1979a: 166.

Type depositories. *A. similis*: IMLA (HT), JHRC (PT); *V. liae*: MNRJ (Campos Seabra coll.) (HT); *V. paraguayana*: JHRC (HT), MACN (PT), IMLA (PT).

Distribution. Argentina, Brazil, Paraguay.

***spinipennis* (Soares, 1966)**

spinipennis Soares, 1966a: 274 (*Vianodes*).

Type depositories. MNRJ (Campos Seabra coll.) (HT), IPEACSM (AT, PT).

Distribution. Brazil.

***transversesignatus* (Gyllenhal, 1840)**

transversesignatus Gyllenhal (in Schoenherr, 1840: 474) (*Arrhenodes*).

Type depository. NHRS (ST).

Distribution. Colombia.

***turbatus* (Gyllenhal, 1833)**

turbatus Gyllenhal (in Schoenherr, 1833: 317) (*Arrhenodes*).

dispar Germar, 1824: 190 (non Linné, 1758) (*Brentus*); syn. Schoenherr, 1833: 317.

Type depository of *A. turbatus*: NHRS (HT).

Distribution. Dominican Republic; Argentina.

Notes Kleine (1938d) did not mention *A. dispar* Germar among the species listed in his *Genera Insectorum*.

***vitticollis* (Gyllenhal, 1840)**

vitticollis Gyllenhal (in Schoenherr, 1840: 471) (*Arrhenodes*).

Type depositories. MNHN (ST!), NHRS (ST).

Distribution. Brazil.

xanthozonatus (Jekel, 1858)

xanthozonatus Jekel, 1858: 356 (*Arrhenodes*).

Type depository. Unknown.

Distribution. "C America".

Notes. According to Sharp (1895a: 47) this species could be included in the genus *Cyriodontus*.

Kleine (1938d) did not mention *A. xanthozonatus* among the species listed in his *Genera Insectorum*.

Eupeithes Senna, 1898

Eupeithes Senna, 1898b: 381.

Type species: *Eupeithes dux* Senna, 1898.

Distribution. Borneo, Indonesia (Batu, Nias, Sumatra), Malaysia, Philippines (Mindanao, Samar, Tawi Tawi).

barbarus Kleine, 1921

barbarus Kleine, 1921g: 125.

Type depository. SMTD (HT).

Distribution. Borneo, Indonesia (Sumatra).

bardus Kleine, 1921

bardus Kleine, 1921g: 124.

Type depositories. MZPW (HT), SMTD (AT, PT).

Distribution. Borneo, Indonesia (Sumatra), Malaysia.

***dominator* Kleine, 1921**

dominator Kleine, 1921g: 125.

Type depository. SMTD (HT).

Distribution. Philippines (Mindanao, Samar, Tawi Tawi).

***dux* Senna, 1898**

dux Senna, 1898b: 382.

Type depository. MSNG (HT!).

Distribution. Borneo, Indonesia (Batu, Nias, Sumatra), Malaysia.

***Eupsalomorphus* Kleine, 1926**

Eupsalomorphus Kleine, 1926a: 41.

Type species: *Eupsalomorphus decorus* Kleine, 1926.

Distribution. Indonesia (Sumatra), Laos, Malaysia, Vietnam.

***decorus* Kleine, 1926**

decorus Kleine, 1926a: 41.

Type depository. NHM (HT!).

Distribution. Indonesia (Sumatra), Malaysia, Vietnam.

***laoticus* Damoiseau, 1989**

laoticus Damoiseau, 1989b: 52.

Type depository. BPBM (HT).

Distribution. Laos.

Eutrachelus Berthold, 1827

Eutrachelus Berthold, 1827: 383.

Entrachelus: Berthold, 1827: 383 [lapsus].

Eutracheles Latreille, 1828: 593; syn. Alonso-Zarazaga & Lyal, 1999: 48.

Eutrachelus Schoenherr, 1833: 337 (non Berthold, 1827) [unjustified replacement name].

Eutrachela: Chevrolat, 1839: 172 [lapsus].

Type species: *Brentus temminckii* Latreille, 1825.

Distribution. Borneo, Indonesia (Java, Sumatra), Malaysia, Vietnam.

Notes. Kabakov (2001: 213) quotes *Eutrachelus* sp. for Vietnam.

morio Kleine, 1936

morio Kleine, 1936b: 124.

Type depository. NHM (LT!, PLT!).

Distribution. Malaysia.

Notes. Lectotype of *E. morio* (NHM) designated on a male specimen with the following labels: syntype/ Typus ♂ / Pahang, F. M. S. Cameron's Highlands, 4500-5000 ft., 18.VI.1935, H. M. Pendlebury/ *Eutrachelus morio* Kln., R. Kleine det., 1935. Paralectotype: 1 female specimen (NHM), syntype/ Typus ♀ / Pahang, F. M. S. Cameron's Highlands, 4800 ft., 26.VI.1935, H. M. Pendlebury.

temminckii (Latreille, 1825)

temminckii Latreille, 1825: 389 (*Brentus*).

temmincki: Gyllenhal (in Schoenherr, 1833: 338) (*Eutrachelus*) [lapsus].

sumatrensis Waterhouse, 1881: 489 (*Eutrachelus*); syn. Schönfeldt, 1908: 50.

achilles Rothschild & Jordan, 1893: 454 (*Eutrachelus*); syn. Kleine, 1926h: 38.

borneensis Rothschild & Jordan, 1893: 454 (*Eutrachelus*); syn. Kleine, 1926c: 172.

Type depositories. *B. temminckii*: unknown; *E. sumatrensis*: NHM (HT!); *E. achilles*: unknown; *E. borneensis*: unknown.

Distribution. Borneo, Indonesia (Java, Sumatra), Malaysia.

Notes. According to Kleine (1928a: 87) the types of *E. achilles* and *E. borneensis* were in the Oberthür collection, but we did not find them in MNHN, NHM nor IRSNB.

Gyalostoma Kleine, 1914

Gyalostoma Kleine, 1914b: 174.

Type species: *Gyalostoma jucundum* Kleine, 1914 = *Baryrrhynchus deyrollei* Power, 1879.

Distribution. Indonesia (Moluccas: Aru; Sulawesi); New Guinea.

deyrollei (Power, 1879)

deyrollei Power, 1879b: 298 (*Baryrrhynchus*).

jucundum Kleine, 1914b: 176 (*Gyalostoma*); syn. Kleine, 1916e: 127.

Type depositories. *B. deyrollei*: MNHN (LT!, PLT!), IRSNB (PLT!); *G. jucundum*: MZPW (ST).

Distribution. Indonesia (Moluccas: Aru; Sulawesi).

Notes. Lectotype of *B. deyrollei* (MNHN) designated on a male specimen with the following labels: Menado/ type/ Mniszech 10/ ex Musaeo G. Power/ Museum Paris 1952, coll. R. Oberthür. Paralectotypes: 1 female specimen (MNHN), Menado/ type/ Mniszech 10/ ex Musaeo G. Power/ Museum Paris 1952, coll. R. Oberthür; 1 male specimen [incorrectly labelled as holotype] (IRSNB), ♂/ Menado, Wallace/ Coll. Castelnau, Coll. Roelofs/ *Baryrrhynchus Deyollei* Pow. ♂/ *Gyalostoma deyrollei* (Power) R. Damoiseau det. 1967/ Holotype; 1 female specimen (IRSNB), ♀/ Coll. Roelofs/ Type/ *Baryrrhynchus Deyollei* Pow. ♀/ *Gyalostoma deyrollei* (Power) R. Damoiseau det. 1967/ Allotype.

elegans Kleine, 1916

elegans Kleine, 1916a: 184.

ochraceus Kleine, 1916f: 184 (*Baryrrhynchus*); syn. Damoiseau, 1989b: 52.

Type depositories. *G. elegans*: MNHUB (LT!, PLT!), SMTD (PLT); *B. ochraceus*: DEI (LT!, PLT!), IRSNB (PLT!).

Distribution. Indonesia (Moluccas, Sulawesi); New Guinea.

Notes. Lectotype of *G. elegans* (MNHUB) designated on a male specimen with the following labels: S. Celebes, Borithain, C. Ribbe, 1882/ Typus/ 117126/ *Gyalostoma elegans* Kl., Type. Paralectotype: 1 female specimen (MNHUB), with the same locality as the lectotype.

Lectotype of *B. ochraceus* (DEI) designated on a male specimen with the following labels: N Guinea/ coll. Kraatz/ *Baryrrhynchus ochraceus* Kl./ Kleine det./ Syntypus. Paralectotypes: 1 female specimen (DEI), Neuguinea/ coll. Kraatz/ *Baryrrhynchus ochraceus* Kl., Typus/ Syntypus; 1 male specimen (IRSNB), S Celebes, Bonthain, C. Ribbe 1882/ *Baryrrhynchus ochraceus* Kl. cotype/ R. I. Sc. N. B., I. G. 23.643/ *Gyalostoma elegans* Kleine R. Damoiseau det., 1967/ Paratype.

Haywardiales Haedo Rossi, 1965

Haywardiales Haedo Rossi, 1965: 308.

Type species: *Haywardiales brasileria* Haedo Rossi, 1965.

Distribution. Argentina, Brazil.

brasileria Haedo Rossi, 1965

brasileria Haedo Rossi, 1965: 309.

Type depositories. MLPA (HT), JHRC (AT, PT), MACN (PT).

Distribution. Argentina, Brazil.

Hemiorychodes Kleine, 1921

Hemiorychodes Kleine, 1921b: 83.

Type species: *Hemiorychodes cambodjensis* Kleine, 1921.

Distribution. S China; Borneo, India, Indonesia (Java, Sumatra), Kampuchea [= Cambodia], Malaysia, Myanmar [= Burma], Vietnam.

***cambodjensis* Kleine, 1921**

cambodjensis Kleine, 1921b: 84.

Type depository. SMTD (HT).

Distribution. S China; Kampuchea [= Cambodia], Vietnam.

***compactus* Kleine, 1931**

compactus Kleine, 1931a: 95.

conspectus: Kleine, 1933c: 290 (*Hemiorychodes*) [lapsus].

Type depositories. NHM (LT!, PLT!), SMTD (PLT).

Distribution. India.

Notes. Lectotype of *H. compactus* (NHM) designated on a male specimen with the following labels: syntype/ Type/ ex *Hopea parviflora*/ typus ♂ / 195/ Kasaragod R., S. Bangalore, Madras, R. O. Coll., 26.VII.1930/ Prépar. génit. N° BM 120 R. Damoiseau. Paralectotype: 1 female specimen (NHM), Type/ ex *Hopea parviflora*/ typus ♂ / 193/ S. Bangalore 400, Madras, J. C. M. Gardner, 25.V.1930/ *Hemiorychodes compactus* Kleine ♀ type, R. Kleine det.

***contextus* Kleine, 1925**

contextus Kleine, 1925b: 148.

Type depository. NHM (HT!).

Distribution. Kampuchea [= Cambodia].

***continens* Kleine, 1925**

continens Kleine, 1925b: 149.

Type depository. NHM (HT!).

Distribution. Malaysia.

curvus Kleine, 1925

curvus Kleine, 1925b: 147.

Type depository. NHM (LT!, PLT!).

Distribution. India, Malaysia, Myanmar [= Burma].

Notes. Lectotype of *H. curvus* (NHM) designated on a male specimen with the following labels: syntype/ Typus!/ Doherty/ India Or. Manipur/ Fry coll. 1905.100/ *Hemiorchodes curvus* Kln. typus ♂, R. Kleine det., 1922/ synonymous of *Pseud. tenuirostris* Senna comp. type, R. Damoiseau det., 1978. Paralectotypes: 1 female specimen (NHM), syntype/ Typus!/ Doherty/ India Or. Manipur/ Fry coll. 1905.100/ *Hemiorchodes curvus* Kln. typus ♀, R. Kleine det., 1922; 1 female specimen (NHM), syntype/ Doherty/ India Or., Manipur/ Fry coll. 1905.100; 1 male specimen (NHM), syntype/ Doherty/ Perak/ Fry coll. 1905.100; 3 male specimens (NHM), syntype/ Doherty/ Birmah, Ruby Mines/ Fry coll. 1905.100.

dentipennis (Senna, 1898)

dentipennis Senna, 1898d: 60 (*Pseudorychodes*).

Type depository. MZUF (HT!).

Distribution. Borneo, Indonesia (Java, Sumatra), Malaysia.

dissonus Kleine, 1925

dissonus Kleine, 1925b: 146.

dissonous: Kleine, 1936b: 120 (*Hemiorchodes*) [lapsus].

Type depository. NHM (LT!, PLT!).

Distribution. Borneo, Malaysia, Vietnam.

Notes. Lectotype of *H. dissonus* (NHM) designated on a male specimen with the following labels: syntype/ Doherty/ Perak/ Fry Coll. 1905.100/ *Hemiorchodes dissonus* Kln. typus! R. Kleine det. Paralectotypes: 1 female specimen (NHM), syntype/ Type ♀/ Doherty/ Perak/ Fry Coll. 1905.100; 3 female specimens (NHM), syntype/ Doherty/ Perak/ Fry Coll. 1905.100.

drescheri Kleine, 1926

drescheri Kleine, 1926a: 45.

Type depository. ZMAN (HT!).

Distribution. "Indonesia".

modestus Kleine, 1921

modestus Kleine, 1921b: 85.

Type depositories. SMTD (?) (HT), MNHUB (?) (PT), RMNH (PT), ZSM (PT).

Distribution. S China; Indonesia (Sumatra), "Indochina", Kampuchea [= Cambodia].

Notes. In the original description Kleine (1921b: 85) reported that the holotype was in SMTD, but according to O. Jaeger (SMTD) (2002, personal communication) it is now missing from the collection. Kleine (1921b: 85) also reported two paratypes in MNHUB, but we did not find them there.

Hemipsalis Sharp, 1895

Hemipsalis Sharp, 1895a: 34.

Type species: *Hemipsalis crassa* Sharp, 1895.

Distribution. Nicaragua; Brazil, Colombia, French Guyana, Paraguay.

benedictoi Naliato, 1986

benedictoi Naliato, 1986: 390.

Type depository. MNRJ (HT, PT).

Distribution. Brazil.

brasiliensis Kleine, 1922*brasiliensis* Kleine, 1922p: 146.**Type depository.** ZMUH (ST).**Distribution.** Brazil, Colombia, Paraguay.*crassa* Sharp, 1895*crassus* Sharp, 1895a: 35.**Type depository.** NHM (LT!, PLT!).**Distribution.** Nicaragua.

Notes. Lectotype of *H. crassa* (NHM) designated on a male specimen with the following labels: *Hemipsalis crassus* Type D. S. ♂ Chontales, Nicaragua, T. Belt/ syntype/ B. C. A., Col. IV, 6. *Hemipsalis crassus*, Sharp. Paralectotype: 1 female specimen (NHM), with the same label data as the lectotype.

fausta Kleine, 1927*fausta* Kleine, 1927b: 436.**Type depository.** NHM (HT!).**Distribution.** French Guyana.*Henarrhodes* Heller, 1913*Henarrhodes* Heller, 1913: 152.*Henarrhenodes*: Kleine, 1917f: 318 [lapsus].**Type species:** *Henarrhodes macgregori* Heller, 1913.

Distribution. Borneo, Indonesia (Sumatra), Philippines (Luzon, Mindanao, Negros, Polillo, Siargao).

***contortus* Kleine, 1926**

contortus Kleine, 1926a: 50 (*Henarrhenodes*).

Type depository. NHM (HT!).

Distribution. Borneo, Indonesia (Sumatra).

***macgregori* Heller, 1913**

macgregori Heller, 1913: 153.

Type depository. SMTD (ST).

Distribution. Philippines (Luzon, Mindanao, Negros, Polillo, Siargao).

***roseni* Kleine, 1920**

roseni Kleine, 1920m: 9 (*Henarrhenodes*).

Type depository. ZSM (HT).

Distribution. Borneo, Indonesia (Sumatra).

***Henorychodes* Kleine, 1921**

Henorychodes Kleine, 1921b: 85.

Type species: *Henorychodes pretiosus* Kleine, 1921.

Distribution. New Guinea.

***pretiosus* Kleine, 1921**

pretiosus Kleine, 1921b: 87.

Type depository. MNHUB (HT).

Distribution. New Guinea.

Notes. The type specimen is missing from the card.

Heteroblysmia Kleine, 1917

Heteroblysmia Kleine, 1917c: 285.

Type species: *Heteroblysmia borneensis* Kleine, 1917.

Distribution. Borneo, Laos, Malaysia, Philippines (Mindanao, Negros), Vietnam.

Notes. Kabakov (2001: 213) quotes *Heteroblysmia* sp. for Vietnam.

accurata Kleine, 1922

accurata Kleine, 1922f: 215.

accurate: Kleine, 1926a: 48 (*Heteroblysmia*) [lapsus].

Type depository. SMTD (HT).

Distribution. Borneo, Philippines (Mindanao).

borneensis Kleine, 1917

borneensis Kleine, 1917c: 289.

Type depository. SMTD (?) (HT).

Distribution. Borneo.

Notes. In the original description Kleine (1917c: 289) reported that the holotype was in SMTD, but according to O. Jaeger (SMTD) (2002, personal communication) it is now missing from the collection.

cava Kleine, 1925

cava Kleine, 1925b: 155.

Type depository. NHM (HT!).

Distribution. Malaysia.

electa Kleine, 1923*electa* Kleine, 1923e: 164.**Type depository.** NHM (HT!).**Distribution.** Philippines.*formidolosa* Kleine, 1923*formidolosa* Kleine, 1923e: 165.**Type depository.** SMTD (HT).**Distribution.** Philippines (Negros).*fusca* Calabresi, 1921*fusca* Calabresi, 1921c: 66.**Type depository.** MZUF (LT!).**Distribution.** Borneo.**Notes.** Lectotype designated by Bartolozzi *et al.* (1985: 97) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).*laos* Damoiseau, 1965*laos* Damoiseau, 1965b: 5.**Type depositories.** IRSNB (HT!), BPBM (?) (AT, PT).**Distribution.** Laos.**Notes.** In his original description Damoiseau also designated the allotype and a paratype, which are not held in IRSNB. According to P. Grootaert (IRSNB) (2001, personal communication) they are probably in BPBM.

rondoni Damoiseau, 1989

rondoni Damoiseau, 1989b: 59.

Type depository. IRSNB (HT!, AT!, PT!).

Distribution. Laos.

vittata Calabresi, 1921

vittata Calabresi, 1921c: 65.

Type depository. MZUF (HT!).

Distribution. Borneo.

wapi Damoiseau, 1989

wapi Damoiseau, 1989b: 60.

Type depository. IRSNB (HT!, AT!, PT!).

Distribution. Laos.

Heterobrenthus Sharp, 1895

Heterobrenthus Sharp, 1895a: 42.

Type species: *Heterobrenthus distans* Sharp, 1895.

Distribution. USA (Texas); Guatemala, Mexico, Nicaragua.

distans Sharp, 1895

distans Sharp, 1895a: 42.

Type depository. NHM (LT!, PLT!).

Distribution. Guatemala, Mexico, Nicaragua.

Notes. Lectotype of *H. distans* (NHM) designated on a male specimen with the following labels: *Heterobrenthus distans* ♂ type D. S., El Reposo, 800 ft., Champion/ syntype/ type/ B. C. A., Col., IV, 6. *Heterobrenthus distans* Sharp. Paralectotypes: 1 female specimen (NHM), *Heterobrenthus distans* type D. S., Panzos Vera Paz, Champion/ syntype/ B. C. A., Col., IV, 6. *Heterobrenthus distans* Sharp; 1 female specimen (NHM), *Heterobrenthus distans* type D. S., Chontales, Janson/ syntype/ B. C. A., Col., IV, 6. *Heterobrenthus distans* Sharp/ Sharp Coll. 1905-313; 1 female specimen (NHM), *Heterobrenthus distans* D. S./ syntype/ Temax, N Yucatan, Gaumer/ B. C. A., Col., IV, 6. *Heterobrenthus distans* Sharp; 1 female specimen (NHM), *Heterobrenthus distans* var? Temax, Gaumer/ B. C. A., Col., IV, 6. *Heterobrenthus distans* Sharp/ Sharp Coll. 1905-313; 1 female specimen (NHM), *Heterobrenthus distans* var. D. S., Juquila, Mexico, Salle Coll./ syntype/ B. C. A., Col., IV, 6. *Heterobrenthus distans* Sharp.

lacordairei (Power, 1878)

lacordairei Power, 1878b: 241 (*Episphales*).

Type depositories. IRSNB (LT!), MNHN (PLT!).

Distribution. Mexico.

Notes. Damoiseau (1963f: 16) reported that the specimen held in IRSNB is the holotype, but in his original description Power did not designate any of the syntypes as holotype; however, as there is only one syntype in IRSNB, this must be considered as lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999). The lectotype has the following labels: Mexique/ Type/ Coll. Lacordaire/ Type/ *Episphales Lacordairei* Pow./ Prépar. génit. No. 766 R. Damoiseau.

texanus Schaeffer, 1915

texanus Schaeffer, 1915: 51.

Type depository. NMNH (ST).

Distribution. USA (Texas).

***Heterorrhynchus* Calabresi, 1921**

Heterorrhynchus Calabresi, 1921c: 62.

Heterorrhynchus: Kleine, 1922m: 162 (non Lafresnaye, 1839, nec Mandelli, 1873) [lapsus].

Type species: *Heterorrhynchus ornatus* Calabresi, 1921.

Distribution. Borneo, Vietnam.

***ornatus* Calabresi, 1921**

ornatus Calabresi, 1921c: 63.

Type depositories. MZUF (LT!), SMTD (PLT).

Distribution. Borneo, Vietnam.

Notes. Lectotype designated by Bartolozzi *et al.* (1985: 99) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

***Holobrenthus* Kleine, 1923**

Holobrenthus Kleine, 1923h: 52.

Type species: *Holobrenthus aptus* Kleine, 1923.

Distribution. Mexico.

***aptus* Kleine, 1923**

aptus Kleine, 1923h: 53.

optus: Kleine, 1937h: 318 (*Holobrenthus*) [lapsus].

Type depository. NMPC (HT).

Distribution. Mexico.

Hopliterrhynchus Senna, 1892

Hopliterrhynchus Senna, 1892b: 26.

Type species: *Hopliterrhynchus emmae* Senna, 1892.

Distribution. Borneo, Malaysia, Philippines (Luzon, Mindanao).

bicolor Damoiseau, 1989

bicolor Damoiseau, 1989b: 60.

Type depository. IRSNB (HT!).

Distribution. Philippines (Mindanao).

Notes. Damoiseau (1989b: 60) reported that the holotype is in BPBM, but actually it is in IRSNB.

emmae Senna, 1892

emmae Senna, 1892b: 27.

Type depository. MZUF (HT!).

Distribution. Borneo, Malaysia.

magnificus Damoiseau, 1989

magnificus Damoiseau, 1989b: 60.

Type depository. IRSNB (HT!).

Distribution. Philippines (Luzon).

Notes. Damoiseau (1989b: 60) reported that the holotype is in BPBM, but actually it is in IRSNB.

Hyposphales Kleine, 1927

Hyposphales Kleine, 1927b: 434.

Type species: *Hyposphales factus* Kleine, 1927.

Distribution. Brazil.

factus Kleine, 1927

factus Kleine, 1927b: 436.

Type depository. NHM (LT!, PLT!).

Distribution. Brazil.

Notes. Lectotype of *H. factus* (NHM) designated on a male specimen with the following labels: syntype/ Diversip, Bras./ Bowring 63-47/ *Hyposphales factus* Kln. ♂, Type 1923, R. Kleine det. Paralectotypes: 1 female specimen (NHM), syntype/ typus "*posticus*" ♀ Bras./ Bowring 63-47/ *Hyposphales factus* Kln ♀ 1923, R. Kleine det.; 1 male specimen (NHM), syntype/ 169/ "*posticus*" ♀ Bras./ Bowring 63-47/ *Hyposphales factus* Kln., A. W. Donoghue det. 1983; 1 female specimen (NHM), Brasil/ syntype.

gratiosus Kleine, 1941

gratiosus Kleine, 1941c: 192.

Type depository. MZPW (HT).

Distribution. Brazil.

Megateras Kleine, 1921

Megateras Kleine, 1921l: 62.

Type species: *Megateras horribile* Kleine, 1921.

Distribution. Borneo, Malaysia.

***horribile* Kleine, 1921**

horribile Kleine, 1921l: 64.

Type depository. SMFD (HT).

Distribution. Borneo, Malaysia.

***Mesitogenus* Kleine, 1919**

Mesitogenus Kleine, 1919b: 7.

Type species: *Mesitogenus amorphocephaloides* Kleine, 1919.

Distribution. Indonesia (Sumatra).

***amorphocephaloides* Kleine, 1919**

amorphocephaloides Kleine, 1919b: 9.

Type depository. MZPW (HT).

Distribution. Indonesia (Sumatra).

***Orfilaia* Haedo Rossi, 1955**

Orfilaia Haedo Rossi, 1955: 63.

Evpsalis: Fåhræus, 1871: 435 [lapsus].

Eupsalis Kleine, 1927a: 38 [not available, misidentification].

Orphilaia: Commonwealth Institute of Entomology, 1957: 432 [lapsus].

Type species: *Arrhenodes vulsellatus* Gyllenhal, 1833.

Distribution. Cyprus, Greece (Crete), Israel, S Spain, Syria, Tunisia, Yemen; Afrotropical from Ivory Coast to Ethiopia and to South Africa, Madagascar; India, Indonesia (Moluccas: Ambon, Bacan [= Batjan], Buru, Seram [= Ceram]; Sulawesi), Myanmar [= Burma], Philippines (Mindanao), Sri Lanka; New Guinea, Polynesia, Seychelles Is; Italy (I).

amitina (Kolbe, 1910)

amitina Kolbe, 1910: 39 (*Eupsalis*).

Type depository. MNHUB (HT!).

Distribution. Seychelles Is.

anthracina (Klug, 1833)

anthracinus Klug, 1833: 106 (*Arrhenodes*).

bipunctatus Gory, 1839: 328 (*Arrhenodes*); syn. Gemminger & Harold, 1872: 2709.

coracina Kolbe, 1883c: 76 (*Eupsalis*); syn. Kleine, 1917g: 69.

Type depositories. *A. anthracinus*: MNHUB (HT!); *A. bipunctatus*: unknown; *E. coracina*: MNHUB (LT!, PLT!).

Distribution. Madagascar.

Notes. Lectotype of *E. coracina* (MNHUB) designated on a male specimen with the following labels: Madagascar int. austr./ Hildebrandt/ 380/ Type/ *coracina* Kolbe = *anthracina* Kl. Kleine. Paralectotype: 1 male specimen (MNHUB), Madagascar int. austr./ Hildebrandt/ Type/ 34486.

arabica (De Muizon, 1954)

arabica De Muizon, 1954: 327 (*Eupsalis*).

Type depositories. NHM (HT!, AT!, PT!), MNHN (PT!).

Distribution. Yemen.

bifalcata (Fairmaire, 1884)

bifalcatus Fairmaire, 1884: CXLVII (*Eupsalis*).

Type depository. MNHN (HT!).

Distribution. Burundi, Kenya, Somalia, Tanzania; Italy (I).

Notes. Fairmaire redescribed the species in 1887 (p. 325).

***brevirostris* (Kolbe, 1897)**

brevirostris Kolbe, 1897: 286 (*Eupsalis*).

kolbei Kleine, 1917g: 98 (*Eupsalis*); syn. De Muizon, 1955b: 897.

Type depositories. *E. brevirostris*: MNHUB (HT!); *E. kolbei*: MNHUB (LT!, PLT!), DEI (PLT!), ZMUH (PLT).

Distribution. Benin, Cameroon, Central African Republic, Democratic Republic of the Congo (ex Zaire), Eritrea, Ethiopia, Gabon, Ivory Coast, Sudan, Tanzania, Uganda, Zimbabwe.

Notes. Lectotype of *E. kolbei* (MNHUB) designated on a male specimen with the following labels: 831/ Kamerun, Yaunde-Stat. v. Carnap S. G./ Type/ *Eupsalis kolbei* Kl. Paralectotypes: 2 female specimens (MNHUB), with the same locality as the lectotype; 1 female specimen (DEI), Swakopmund/ coll. Kraatz/ Kleine det./ Syntypus/ *Eupsalis kolbei* Kl., Cotypus.

Damoiseau (1967d: 361) did not mention Zimbabwe (Kleine, 1917g: 89) and Sudan (Heller, 1923: 193) when he gave the species distribution.

***erythrea* (De Muizon, 1960)**

erythrea De Muizon, 1960: 180 (*Eupsalis*).

bifalcata De Muizon, 1954: 330 (non Fairmaire, 1884) (*Eupsalis*); syn. Damoiseau, 1967a: 367.

Type depository. MSNG (?) (HT, AT).

Distribution. Eritrea, Somalia, Sudan.

Notes. Although De Muizon (1960: 180) states that the types are in MSNG, we did not find them; the only specimens there come from different localities.

***falsotaruiensis* (Kleine, 1947)**

falsotaruiensis Kleine, 1947: 143 (*Eupsalis*).

Type depository. MCST (HT).

Distribution. Somalia.

***glabrata* (Kleine, 1917)**

glabrata Kleine, 1917g: 102 (*Eupsalis*).

Type depositories. DEI (ST!), SMTD (ST).

Distribution. Ecuador (?), Mediterranean (?).

Notes. In his original description Kleine (1917g: 105) gives Ecuador as type locality, but later Kleine (1923b: 169) states that this locality is not correct and even later (1938d: 94) he writes «unsicher, Mittelmeer» [= not sure, Mediterranean] as the species distribution.

***kleinei* (Heller, 1921)**

kleinei Heller, 1921: 624 (*Schizoeupsalis*).

Type depository. SMTD (HT).

Distribution. Philippines (Mindanao).

***nigeriensis* Damoiseau, 1963**

nigeriensis Damoiseau, 1963f: 11.

Type depository. IRSNB (HT!).

Distribution. Nigeria.

***nigra* Damoiseau, 1963**

nigra Damoiseau, 1963f: 12.

brevirostris Kleine, 1917g: 86 (non Kolbe, 1897) (*Eupsalis*); syn. Damoiseau, 1967a: 362.

Type depository. IRSNB (HT!).

Distribution. Senegal, Tanzania.

reichei (Fairmaire, 1859)

reichei Fairmaire, 1859: CLXIV (*Arrhenodes*).

reichi: Stierlin, 1893: 518 (*Amorphocephalus*) [lapsus].

Type depository. MNHN (?) (ST).

Distribution. Cyprus, Greece (Crete), Israel, S Spain (?), Syria, Tunisia.

Notes. Damoiseau (1967a: 367) states that the holotype is held in the MNHN, but we did not find it there.

somalica (Senna, 1895)

somalica Senna, 1895d: 57 (*Eupsalis*).

Type depository. RMNH (HT).

Distribution. Ethiopia, Kenya, Somalia, Tanzania.

Notes. Damoiseau (1967a: 364) did not mention Tanzania (Kleine, 1917g: 93) when he gave the species distribution.

taruensis (Kolbe, 1897)

taruensis Kolbe, 1897: 286 (*Eupsalis*).

Type depository. MNHUB (HT!).

Distribution. Ethiopia, Kenya, Somalia.

testacea (Kleine, 1917)

testacea Kleine, 1917g: 131 (*Eupsalis*).

Type depositories. MNHUB (LT!), DEI (PLT!), MZPW (PLT), ZMUH (PLT).

Distribution. Indonesia (Moluccas: Ambon, Bacan [= Batjan], Buru, Seram [= Ceram]; Sulawesi); New Guinea (including New Britain), Polynesia.

Notes. Lectotype of *E. testacea* (MNHUB) designated on a male specimen with the following labels: Ceram, Jlo, C. Ribbe, 1884/ 814/ *Eupsalis testacea* Kl., cotypus. Paralectotype: 1 female specimen (DEI), D. N. Guinea/ col. Bennigsen/ Kleine det./ Dtsch. Entomol. Institut Berlin/ Syntype/ *Eupsalis testacea* Kl., Cotype.

truncata (Boheman, 1829)

truncatus Boheman, 1829: 27 (*Brentus*).

Type depository. NHRS (ST).

Distribution. India, Sri Lanka.

vulsellata (Gyllenhal, 1833)

vulsellatus Gyllenhal (in Schoenherr, 1833: 325) (*Arrhenodes*).

gentilis J. Thomson, 1858: 117 (*Arrhenodes*); syn. Damoiseau, 1967a: 359.

vulsatellus: Kolbe, 1883b: 239 (*Eupsalis*) [lapsus].

vulneratus: Quedenfeldt, 1888a: 195 (*Arrhenodes*) [lapsus].

callosoguttis Kolbe, 1897: 286 (*Eupsalis*); syn. Kleine, 1917g: 136.

parviornata Kleine, 1917g: 115 (*Eupsalis*); syn. De Muizon, 1955b: 897.

vulsellata: Uhlig & Jaeger, 1995: 231 (*Orfilaia*) [lapsus].

Type depositories. *A. vulsellatus*: NHRS (HT); *A. gentilis*: MNHN (LT!, PLT!), IRSNB (PLT!); *E. callosoguttis*: MNHUB (HT!); *E. parviornata*: MNHUB (LT!), ZMUH (PLT).

Distribution. Tropical Africa to South Africa; Italy (I).

Notes. Lectotype of *A. gentilis* (MNHN) designated on a male specimen with the following labels: Gabon/ Thomson ♂/ type 42/ ex Musaeo Power. Paralectotypes: 1 specimen (MNHN), Gabon/ Mniszech/ Museum Paris 1952, coll. R. Oberthür; 1 specimen (MNHN), Gabon/ Mniszech 31/ ex Musaeo Power; 2 specimens (MNHN), Gabon/ Chevrolat 130/ ex Musaeo G. Power; 1 specimen (IRSNB), Gabon/ Type/ ♂/ M. R. Belg./ Coll. Roelofs/ nov. sp. *vulsellato proximal* *Eupsalis gentilis* Thoms./ *Orfilaia vulsellata* (Gyllenhal), R. Damoiseau det., 1961.

We also found a specimen of *A. gentilis* (MNHN) labelled "Guinée/ Thomson ♂ 50/ type/ ex Musaeo Power", which cannot belong to the type series because it comes from Guinea and not from Gabon as stated in the description.

Lectotype of *E. parviornata* (MNHUB) designated on a male specimen with the following labels: Damara bis Ngami-See, Fleck S./ 99/ *parviornata* Kl.

Orychodes Pascoe, 1862

Orychodes Pascoe, 1862: 389.

Caenorychodes Kleine, 1921b: 87; syn. Alonso-Zarazaga & Lyal, 1999: 48.

Caenorychydes: Kleine, 1921b: 70 [lapsus].

Caenorhychodes: Zimmerman, 1994: 116 [lapsus].

Orichodes: Zimmerman, 1994: 121 [lapsus].

Chenorhychodes: Li-zhong, 2002: 296 [lapsus]

Type species: *Brentus serrirostris* Lund, 1800.

Distribution. China, Japan, Taiwan; Bangladesh, Borneo, India (including Andaman Is., Assam, Darjeeling, Sikkim), Indonesia (Bali; Batu; Java; Moluccas: Ambon, Aru, Damar, Obi, Seram [= Ceram]; Nias; Sulawesi; Sumatra; Sumba; Ternate), Laos, Myanmar [= Burma], Philippines (Basilan, Leyte, Luzon, Masbate, Mindanao, Negros, Palawan, Samar, Siargao, Sibuyan, Tawi Tawi), Singapore, Sri Lanka, Thailand, Vietnam; Australia (including Christmas Is.), New Guinea (including Bismarck Archipelago: New Britain; Yule Is.), Polynesia, Solomon Is.

abnormis (Kleine, 1921)

abnormis Kleine, 1921b: 100 (*Caenorhychodes*).

Type depository. MZPW (HT).

Distribution. Indonesia (Sumbawa).

andrewsi Gahan, 1900

andrewsi Gahan, 1900: 117.

andrewsii: Schönfeldt, 1908: 41 (*Orychodes*) [lapsus].

andrewesii: Kleine, 1921b: 76 (*Caenorychodes*) [lapsus].

andreuesi: Kleine, 1928a: 83 (*Caenorychodes*) [lapsus].

andrewesi: Kleine, 1938d: 101 (*Caenorychodes*) [lapsus].

Type depositories. NHM (LT!, PLT!), MZUF (PLT!), DEI (PLT!).

Distribution. Australia (Christmas Is.).

Notes. Lectotype of *O. andrewsi* (NHM) designated on a male specimen with the following labels: syntype/ Christmas I., C. W. Andrews, 1898, 98-20/ *Orychodes Andrews* Gahan, Type. Paralectotypes: 3 male and 5 female specimens (NHM), syntype/ Christmas I., C. W. Andrews, 1898, 98-20/ *Orychodes Andrews* Gahan; 1 male and 1 female specimen (DEI), Christmas I., C. W. Andrews, 1898, 98-20/ *Orychodes Andrews* Gahan; 1 male specimen (MZUF), Christmas I., C. W. Andrews, 1898/ *Caenorychodes andrewsi* Gahan, cotypi!/ ex-coll. Senna/ "La Specola" Firenze, 7009; 1 female specimen (MZUF), Christmas I., C. W. Andrews, 1898/ ex-coll. Senna/ "La Specola" Firenze, 7009; 1 male specimen (MZUF), Christmas I., C. W. Andrews, 1898/ *Caenorychodes andrewsi* Gahan, R. Damoiseau det./ Prépar. génit. N° 75 E 29, R. Damoiseau/ ex-coll. Senna/ "La Specola" Firenze, 7009; 1 female specimen (MZUF), Christmas I., C. W. Andrews, 1898/ *Caenorychodes andrewsi* Gahan, R. Damoiseau det./ ex-coll. Senna/ "La Specola" Firenze, 7009.

digramma (Boisduval, 1835)

digramma Boisduval, 1835b: 310 (*Arrhenodes*).

digramma: Masters, 1888: 955 (*Orychodes*) [lapsus].

Type depository. IRSNB (?) (HT).

Distribution. Indonesia (Moluccas: Aru); Australia, New Guinea (including Bismarck Archipelago: New Britain; Yule Is.), Polynesia.

Notes. According to Kleine (1928a: 84) the holotype was held in IRSNB but we did not find it there.

fasciatus (Kleine, 1921)

fasciatus Kleine, 1921b: 97 (*Caenorychodes*).

Type depositories. DEI (LT!), MNHUB (PLT!), MZPW (PLT), SMTD (PLT).

Distribution. India (including Andaman Is.).

Notes. Lectotype of *C. fasciatus* (DEI) designated on a male specimen with the following labels: Andamanen/ Kleine vidit/ Kleine vidit et det./ syntypus/ *Caenorychodes fasciatus* Kleine, patria Andamanen. Paralectotypes: 2

male specimens (MNHUB), Andamanen, de Roepstorff/ Typus; 2 female specimens (MNHUB), Iles Andaman/ Coll. H. C. Vogel.

***indus* Kirsch, 1875**

indus Kirsch, 1875b: 51.

Type depository. SMTD (?) (HT).

Distribution. Taiwan; India (including Assam), Laos, Myanmar [= Burma], Sri Lanka, Thailand, Vietnam.

Notes. According to Kleine (1923f: 90 and 1938d: 101) and Li-zhong (2002: 296) this species is a synonym of *O. planicollis* but Damoiseau (1966) considers it as a *bona species* as we do here.

Kleine (1928a: 84) reported the type for this species to be in SMTD, but according to O. Jaeger (SMTD) (2002, personal communication) it is now missing from the SMTD collections.

***insulanus* (Kleine, 1921)**

insulanus Kleine, 1921b: 96 (*Caenorhynchodes*).

Type depositories. DEI (LT!, PLT!), MNHUB (PLT!), SMTD (PLT).

Distribution. Indonesia (Moluccas: Ambon, Seram [= Ceram]).

Notes. Lectotype of *C. insulanus* (DEI) designated on a male specimen with the following labels: Ceram, Jlo, C. Ribbe, 1884/ col. Kraatz/ syntypus/ *Caenorychodes insulanus* det. 1921. Paralectotypes: 1 specimen (DEI), Ceram, Jlo, C. Ribbe, 1884/ col. Kraatz/ Kleine det./ syntypus/ *Caenorychodes insulanus* det. 1921; 1 specimen (MNHUB), Ceram, Jlo, C. Ribbe, 1884/ Typus/ *Caenorychodes insulanus* Kl., det. Aut., 1920.

***maassi* (Kleine, 1926)**

maassi Kleine, 1926f: 369 (*Caenorhynchodes*).

maasii: Kleine, 1935a: 7 (*Caenorhynchodes*) [lapsus].

Type depositories. NME (HT), SMTD (PT).

Distribution. Solomon Is.

nigerrimus (Kleine, 1921)

nigerrimus Kleine, 1921b: 95 (*Caenorhychodes*).

Type depositories. MNHUB (?) (ST), SMTD (ST).

Distribution. Indonesia (Moluccas: Ambon); New Guinea.

Notes. Kleine (1921b: 95) reported that syntypes were in MNHUB, but we did not find them there.

octoguttatus (Nakane, 1963)

octoguttatus Nakane, 1963: 34 (*Cerobates*).

Type depository. SEHU (HT, AT, PT).

Distribution. Japan.

planicollis (F. Walker, 1859)

planicollis F. Walker, 1859: 262 (*Nemocephalus*).

Type depository. NHM (LT!, PLT!).

Distribution. China, Japan, Taiwan; India (including Andaman Is., Assam, Darjeeling, Sikkim), Indonesia (Java, Moluccas: Ambon), Laos, Myanmar [= Burma], Sri Lanka, Thailand, Vietnam.

Notes. Lectotype of *N. planicollis* (NHM) designated on a male specimen with the following labels: syntype/ Ceylon/ 59-106/ *Nemocephalus planicollis* Walker a. n. H. (Type)/ *planicollis*. Paralectotypes: 1 male specimen (NHM), syntype/ Ceylon/ 59-106/ *Nemocephalus planicollis* Walker a. n. H. (Type)/ Gattung: *Caenorychodes* Kln. syn. *indus* Kirsch, R. Kleine det.; 1 female specimen (NHM), syntype/ Ceylon/ 53-23.

rubrosignatus (Kleine, 1921)

rubrosignatus Kleine, 1921b: 95 (*Caenorhynchodes*).

subrosignatus: Kleine, 1921b: 71 (*Caenorhynchodes*) [lapsus].

Type depositories. MZUF (LT!, PLT!), MZPW (PLT), SMTD (PLT).

Distribution. Indonesia (Moluccas: Ternate).

Notes. Lectotype of *C. rubrosignatus* (MZUF) designated on a male specimen with the following labels: Molucche, Ternate, Coll. Bruijn, 1875/ *Caenorhynchodes rubrosignatus* Kleine, cotypi!/ ex-coll. Senna/ "La Specola" Firenze, 7010. Paralectotype: 1 female (MZUF), Molucche, Ternate, Coll. Bruijn, 1875/ ex-coll. Senna/ "La Specola" Firenze, 7010.

serrirostris Lund, 1800

serrirostris Lund, 1800: 86 (*Brentus*).

striolatus Kirsch, 1875b: 50 (*Orychodes*); syn. Kleine, 1921b: 102.

femoratus Schaufuss, 1885: 206 (*Orychodes*); syn. Kleine, 1921b: 102.

serrirostri: Kleine, 1921b: 99 (*Caenorhynchodes*) [lapsus].

serriroastris: Kano, 1930: 153 (*Caenorhynchodes*) [lapsus].

Type depositories. *B. serrirostris*: ZMUC (HT); *O. striolatus*: SMTD (?) (HT); *O. femoratus*: MNHUB (LT!, PLT!), IRSNB (PLT!), NMPC (Salvator coll.) (PLT).

Distribution. S China (Hainan), Taiwan; Bangladesh, Borneo, India, Indonesia (Bali; Batu; Java; Mentawai; Moluccas: Obi; Nias; Sulawesi; Sumatra; Sumba), Malaysia, Myanmar [= Burma], Philippines (Basilan, Leyte, Luzon, Masbate, Mindanao, Palawan, Samar, Siargao, Tawi Tawi), Singapore, Vietnam.

Notes. Lectotype of *O. femoratus* (MNHUB) designated on a male specimen with the following labels: Type/ *Orychodes femoratus* Schauf., Macassar. Paralectotypes: 1 male and 1 female specimen (MNHUB), Coll. L. W. Schaufuss/ *Orychodes femoratus* m., S. Celebes; 1 male specimen (IRSNB), Celebes. merid. Makassar Ujung Padang, ex coll. P de Moffarts/ Paratype/ *Orychodes femoratus* Schauf. ex typ. Macassar Cel. mer./ Sp. dist.? antenna gracilior, caput angus., spina brevior.

Kleine (1928a: 84) reported the type for *O. striolatus* to be in SMTD, but according to O. Jaeger (SMTD) (2002, personal communication) it is not in the SMTD collections.

***sinensis* Fairmaire, 1888**

sinensis Fairmaire, 1888: 138.

sienensis: Kleine, 1921b: 64 (*Orychodes*) [lapsus].

siennensis: Kleine, 1927f: 343 (*Orychodes*) [lapsus].

Type depository. MNHN (?) (ST).

Distribution. China (Tianjin).

Notes. We did not find any syntype of this species in MNHN, where many types of Fairmaire are held. Kleine (1938d) did not mention this species among the species listed in his *Genera Insectorum*.

***splendens* Kirsch, 1875**

splendens Kirsch, 1875b: 50.

Type depository. SMTD (HT).

Distribution. Philippines (Luzon, Masbate, Negros, Palawan, Sibuyan).

***versicolor* (Kleine, 1921)**

versicolor Kleine, 1921b: 93 (*Caenorhychodes*).

Type depositories. MNHUB (LT!, PLT!), DEI (PLT!), SMTD (PLT), MZPW (PLT).

Distribution. Indonesia (Moluccas: Ambon, Damar).

Notes. Lectotype of *C. versicolor* (MNHUB) designated on a specimen with the following labels: Insel Dammer/ Type/ *Caenorhychodes versicolor* Kleine, det. Aut. 1920. Paralectotype: 1 specimen (MNHUB), with the same locality as the lectotype [head missing].

***Parorychodes* Kleine, 1921**

Parorychodes Kleine, 1921b: 80.

Parabletus Calabresi, 1921a: 8; syn. Kleine, 1927a: 45.

Type species: *Orychodes degener* Senna, 1892.

Distribution. S China; Bangladesh, India (Andaman Is., Assam), Laos, Myanmar [= Burma], Vietnam.

cereus Kleine, 1925

cereus Kleine, 1925b: 151.

Type depository. NHM (LT!, PLT!).

Distribution. S China; India (Assam), Vietnam.

Notes. Lectotype of *P. cereus* (NHM) designated on a female specimen with the following labels: syntype/ type ♀/ ex under bark of *Nagakola*/ Namber Res., Sibsagar, Assam, 29.IV.1921, C. F. C. Beeson/ *Parorychodes* Kleine, R. Kleine det. Paralectotype: 1 female specimen (NHM), with the same labels as the lectotype [head and prothorax missing].

comes Kleine, 1921

comes Kleine, 1921b: 82.

Type depositories. MNHUB (LT!, PLT!), MZPW (PLT).

Distribution. Vietnam.

Notes. Lectotype of *P. comes* (MNHUB) designated on a male specimen with the following labels: Tonkin, Monte Mauson, April-Mai, 2-3000', H. Fruhstorfer/ Typus/ *Parorychodes comes* Kleine, det. aut. 1921. Paralectotypes: 2 specimens (MNHUB), with the same locality as the lectotype [head and prothorax missing in both specimens].

degener (Senna, 1892)

degener Senna, 1892e: 474 (*Orychodes*).

Type depository. MSNG (HT!).

Distribution. Bangladesh, India (Andaman Is., Assam), Laos, Myanmar [= Burma], Vietnam.

***Perorychodes* Kleine, 1925**

Perorychodes Kleine, 1925b: 145.

Type species: *Perorychodes arrowi* Kleine, 1925.

Distribution. S. China; Borneo, India, N Vietnam.

Notes. Kabakov (2001: 213) quotes two indetermined species of this genus from S China (Yunnan) and N Vietnam.

***arrowi* Kleine, 1925**

arrowi Kleine, 1925b: 146.

Type depository. NHM (HT!).

Distribution. India.

***dayakorum* Damoiseau, 1989**

dayakorum Damoiseau, 1989b: 62.

Type depository. MZUF (HT!).

Distribution. Borneo.

***Phymechus* Senna, 1895**

Phymechus Senna, 1895b: 219.

Type species: *Phymechus jordani* Senna, 1895.

Distribution. Mexico.

***jordani* Senna, 1895**

jordani Senna, 1895b: 220.

Type depository. MNHN (HT!).

Distribution. Mexico.

Proepisphales Kleine, 1922

Proepisphales Kleine, 1922f: 213.

Propisphales: Kleine, 1927b: 435 [lapsus].

Proepishales: Döbler & Gaedike, 1995: 447 [lapsus].

Type species: *Episphales rugicollis* Sharp, 1895.

Distribution. Costa Rica, El Salvador, Guatemala, Mexico.

championi (Sharp, 1895)

championi Sharp, 1895a: 39 (*Episphales*).

Type depository. NHM (LT!, PLT!).

Distribution. Guatemala.

Notes. Lectotype of *E. championi* (NHM) designated on a male specimen with the following labels: *Episphales similis* Type D. S., Calderas, Guatemala, Champion/ syntype/ B. C. A., Col., IV, 6. *Episphales facilis*, Sharp/ Sharp Coll. 1905-313. Paralectotypes: 2 male specimens (NHM), with the same label data as the lectotype.

elegans (Sharp, 1895)

elegans Sharp, 1895a: 38 (*Episphales*).

Type depository. NHM (LT!, PLT!).

Distribution. El Salvador, Mexico.

Notes. Lectotype of *E. elegans* (NHM) designated on a male specimen with the following labels: *Episphales elegans* Types D. S., Omilteme, Guerrero, 800 ft., July, H. H. Smith/ syntype/ B. C. A., Col., IV, 6. *Episphales elegans*, Sharp/ Sharp Coll. 1905-313. Paralectotypes: 3 male and 3 female specimens (NHM), with the same label data as the lectotype; 2 male and 1 female specimens (NHM), Mochitlan, Guerrero/ coll. Harford/ B. C. A., Col., IV, 6. *Episphales elegans*, Sharp/ Sharp Coll. 1905-313.

***facilis* (Sharp, 1895)**

facilis Sharp, 1895a: 37 (*Episphales*).

Type depository. NHM (LT!, PLT!).

Distribution. Mexico.

Notes. Lectotype of *E. facilis* (NHM) designated on a female specimen with the following labels: *Episphales facilis* Type D. S., Omilteme, Guerrero, 8000 ft., Aug., H. H. Smith/ syntype/ B. C. A., Col., IV, 6. *Episphales facilis*, Sharp. Paralectotype: 1 female specimen (NHM), with the same label data as the lectotype.

***horni* Kleine, 1922**

horni Kleine, 1922f: 213 .

Type depository. DEI (HT!).

Distribution. Mexico.

***interruptolineatus* (Gyllenhal, 1840)**

interruptolineatus Gyllenhal (in Schoenherr, 1840: 477) (*Arrhenodes*).

Type depositories. MNHN (ST!), NHRS (ST!).

Distribution. Mexico.

***minor* (Sharp, 1895)**

minor Sharp, 1895a: 38 (*Episphales*).

Type depository. NHM (HT!).

Distribution. Mexico.

***optatus* (Sharp, 1895)**

optatus Sharp, 1895a: 37 (*Episphales*).

Type depository. NHM (HT!).

Distribution. Costa Rica.

***rugicollis* (Sharp, 1895)**

rugicollis Sharp, 1895a: 36 (*Episphales*).

Type depository. NHM (HT!).

Distribution. Mexico.

***similis* (Sharp, 1895)**

similis Sharp, 1895a: 37 (*Episphales*).

Type depository. NHM (LT!, PLT!).

Distribution. Guatemala.

Notes. Lectotype of *E. similis* (NHM) designated on a female specimen with the following labels: *Episphales similis* Type D. S., Perula, Vera Paz, Champion/ syntype/ B. C. A., Col., IV, 6. *Episphales facilis*, Sharp/ Sharp Coll. 1905-313. Paralectotype: 1 female specimen (NHM), *Episphales similis* D. S., S. Geronimo, Guatemala, Champion/ syntype/ B. C. A., Col., IV, 6. *Episphales facilis* Sharp.

***Prophthalmus* Lacordaire, 1866**

Prophthalmus Lacordaire, 1866: 427.

Type species: *Brentus tridentatus* Lund, 1800.

Distribution. S China, Taiwan; Borneo, India (including Andaman Is., Assam, Darjeeling, Sikkim), Indonesia (Engano; Java; Mentawai; Moluccas: Bacan [= Batjan], Halmahera, Ternate; Nias; Sulawesi; Sumatra; Sumbawa), Kampuchea [= Cambodia], Laos, Malaysia, Myanmar [= Burma], Nepal, Philippines (Catanduanes, Leyte, Luzon, Mindanao, Negros, Palawan, Samar, Siargao, Sibuyan), Sri Lanka, Thailand, Vietnam; New Guinea.

apparatus Kleine, 1923*apparatus* Kleine, 1923h: 48.**Type depository.** NMPC (?) (ST).**Distribution.** E India.

Notes. Kleine (1923h: 48) described the species on a small series of specimens from the NMPC collection; according to J. Jelinek (NMPC) (2002, personal communication) the NMPC syntypes are missing.

bourgeoisi Power, 1878*bourgeoisi* Power, 1878a: XXXVIII.**Type depositories.** MNHN (LT!, PLT!), IRSNB (PLT!).**Distribution.** E India, Sri Lanka.

Notes. Lectotype of *P. bourgeoisi* (MNHN) designated on a male specimen with the following labels: Ceylon/ Montchicourt/ type/ ex Musaeo Power/ Museum Paris 1952, coll. R. Oberthür. Paralectotypes: 1 specimen (MNHN), Ceylon/ Chevrolat 70/ type/ ex Musaeo Power/ Museum Paris 1952, coll. R. Oberthür; 1 male specimen (IRSNB), Ceylon/ Coll. Roelofs/ M. R. Belg./ *Prophthalmus Bourgeoisi* Pow./ Paratype/ *Prophthalmus bourgeoisi* Power, R. Damoiseau det.

brevis Power, 1878*brevis* Power, 1878a: XLV.**Type depositories.** MNHN (LT!, PLT!), IRSNB (PLT!).**Distribution.** Indonesia (Sumatra), Malaysia, Vietnam.

Notes. Lectotype of *P. brevis* (MNHN) designated on a specimen with the following labels: Malacca/ Roelofs 2/ paratype/ Museum Paris 1952, coll. R. Oberthür. Paralectotypes: 1 specimen (MNHN), Sumatra/ ex Musaeo Power/ Museum Paris 1952, coll. R. Oberthür; 1 male specimen (IRSNB), Malacca/ Coll. Castelnau, Coll. Roelofs/ *Prophthalmus brevis* Pow./ Prépar, génit. No. 75 D 19 R. Damoiseau/ *Prophthalmus brevis* Power, Holotype, R. Damoiseau

rev. 1975/ Lectotype [*in litteris*]; 1 female specimen (IRSNB), Malacca/ ex Coll. Castelnau, Coll. Roelofs/ *Prophthalmus brevis* Power, Allotype R. Damoiseau rev. 1975/ Allotype.

heikertingeri Kleine, 1916

heikertingeri Kleine, 1916b: 255.

Type depositories. MNHUB (LT!, PLT!), MZPW (PLT).

Distribution. India (including Andaman Is., Darjeeling), Malaysia, Myanmar [= Burma].

Notes. Lectotype of *P. heikertingeri* (MNHUB) designated on a male specimen with the following labels: Andaman, de Roepstorff/ Typus/ Lectotype [*in litteris*]/ *Prophthalmus heikertingeri* Kl. n. sp. Paralectotypes: 3 male and 3 female specimens (MNHUB), Andaman, de Roepstorff; 1 male specimen (MNHUB), Andaman, de Roepstorff/ Prépar. génit. No 75D 13, R. Damoiseau; 1 male specimen (MNHUB), Iles Andaman/ 82221; 1 female specimen (MNHUB), 82220/ Typus/ Allotype; 1 male specimen (MNHUB), Iles Mersi/ 82220.

longirostris (Gyllenhal, 1833)

longirostris Gyllenhal (in Schoenherr, 1833: 323) (*Arrhenodes*).

assimilis Kleine, 1916b: 248 (*Prophthalmus*); syn. Damoiseau, 1989b: 52.

Type depositories. *P. longirostris*: NHRS (ST), IRSNB (ST!); *assimilis*: DEI (LT!), SMTD (PLT).

Distribution. S China (Hainan); Borneo, Indonesia (Java, Sulawesi, Sumatra), Laos, Malaysia, Philippines (Luzon, Mindanao), Thailand, Vietnam.

Notes. Lectotype of *P. assimilis* (DEI) designated on a female specimen with the following labels: Laos, Tonkin/ *Prophthalmus assimilis*, cotype Kl./ syntype.

mutabilis* Senna, 1893mutabilis* Senna, 1893b: 274.*mentaweicus* Senna, 1898a: 238 (*Prophthalmus mutabilis* var.); syn. Kleine, 1916b: 283.

Type depositories. *P. mutabilis*: MSNG (LT!), DEI (PLT!), IRSNB (PLT!), MNHN (PLT!), MNHUB (PLT!), MZUF (PLT!), NHM (PLT!), MZPW (PLT), SMTD (PLT); *P. mutabilis* var. *mentaweicus*: MSNG (LT!).

Distribution. Indonesia (Engano, Mentawai, Sumatra).

Notes. Lectotype of *P. mutabilis* (MSNG) designated on a male specimen with the following labels: Engano, Bua Bua, V-VI.1891, Modigliani/ *Prophthalmus mutabilis* Senna typi! Paralectotypes: 8 male and 10 female specimens (MSNG), 1 male specimen (MNHN), 2 male and 2 female specimens (NHM), 2 male and 1 female specimens (DEI), 1 male and 1 female specimen (MNHUB), Engano, Bua Bua, V-VI.1891, Modigliani; 1 male specimen (MNHUB), Engano, Bua Bua, V-VI.1891, Modigliani/ 66699/ *Prophthalmus mutabilis* Senna/ 55/ Paratypus/ Prépar. génit. No 75D 18, R. Damoiseau; 1 male specimen (MZUF), Engano, Bua Bua, V-VI.1891/ *Prophthalmus mutabilis* Senna, typus!/ *Prophthalmus mutabilis* Senna, cotypi!/ Ann. Mus. Genova, ser. 2^a, XIII, 1893, cotypi ♂ ♀ / ex-coll. Senna/ "La Specola" Firenze, 6971; 1 male and 1 female specimen (MZUF), Engano, Bua Bua, V-VI.1891/ *Prophthalmus mutabilis* Senna, R. Damoiseau det./ ex-coll. Senna/ "La Specola" Firenze, 6971; 4 male and 6 female specimen (MZUF), Engano, Bua Bua, V-VI.1891/ ex-coll. Senna/ "La Specola" Firenze, 6971; 3 male and 3 female specimens (IRSNB), Engano, Bua Bua, V-VI.1891, Modigliani/ *Prophthalmus mutabilis* Senna, R. Damoiseau rev./ Paratype.

Lectotype of *P. mutabilis* var. *mentaweicus* (MSNG) designated on a male specimen with the following labels: Mentawai, Si Oban, IV-VIII.94, Modigliani/ *Prophthalmus mutabilis mentaweicus* Senna. Paralectotypes: 4 male and 2 female specimens (MSNG), Mentawai, Si Oban, IV-VIII.94, Modigliani.

nigropunctatus* Damoiseau, 1962nigropunctatus* Damoiseau, 1962e: 113.

Type depositories. MSNG (HT!, AT!, PT!), IRSNB (PT!).

Distribution. Indonesia (Sumatra), Thailand.

obscurus* Power, 1878obscurus* Power, 1878a: XLV.

Type depository. MNHN (HT!).

Distribution. India, Sri Lanka, Vietnam.

planipennis Pascoe, 1872

planipennis Pascoe, 1872: 322.

Type depository. NHM (LT!, PLT!).

Distribution. Indonesia (Moluccas: Bacan [= Batjan], Halmahera, Ternate; Sulawesi), Malaysia, New Guinea.

Notes. Lectotype of *P. planipennis* (NHM) designated on a male specimen with the following labels: syntype/ Manado, Celebes/ Pascoe Coll. 93-60/ *Prophthalmus planipennis* type Pasc./ *Prophthalmus planipennis* Pasc. Paralectotype: 1 male specimen (NHM), syntype/ N. Celebes/ Pascoe Coll. 93-60.

potens potens Lacordaire, 1866

potens Lacordaire, 1866: 428.

sanguinalis Pascoe, 1872: 322 (*Prophthalmus*); syn. Senna, 1892c: 46.

proteus: Desbrochers des Loges, 1891: 4 (*Prophthalmus*) [lapsus].

amethystina Kleine, 1916b: 242 (*Prophthalmus potens* var.); **syn. n.**

Type depositories. *P. potens potens*: IRSNB (LT!, PLT!); *P. sanguinalis*: NHM (?) (HT); *P. potens* var. *amethystina*: SMTD (ST).

Distribution. S China; N India (including Assam, Darjeeling, Sikkim), Kampuchea [= Cambodia], Malaysia, Myanmar [= Burma], Nepal, Vietnam.

Notes. Lectotype of *P. potens* (IRSNB) designated on a male specimen with the following labels: ♂/ A. n. sp. Assam/ Coll. Lacordaire/ Type/ Type/ *Prophthalmus potens* Lac./ *Prophthalmus potens* Lacordaire, R. Damoiseau det. Paralectotypes: 1 female specimen (IRSNB), Assam/ Coll. Lacordaire/ Type/ *Prophthalmus potens* Lacordaire, R. Damoiseau det./ Allotype; 1 female specimen (IRSNB), Assam/ Coll. Lacordaire/ Type/ *Prophthalmus potens* Lacordaire, R. Damoiseau det./ Paratype.

Kleine (1928a: 81) reported that the holotype of *P. sanguinalis* was in NHM, but we did not find it there.

potens delesserti Power, 1878

delesserti Power, 1878a: XXXVII.

Type depository. *P. potens delesserti*: MNHN (HT!).

Distribution. S and E India.

***pugnator* Power, 1878**

pugnator Power, 1878a: XLIV.

Type depositories. MNHN (LT!, PLT!), IRSNB (PLT!), MHNG (PLT).

Distribution. Taiwan; Borneo, India (Andaman Is.), Indonesia (Java, Nias, Sumatra), Malaysia, Myanmar [= Burma].

Notes. Lectotype of *P. pugnator* (MNHN) designated on a male specimen with the following labels: Indes orientales/ Chevrolat 68/ type/ ex Musaeo Power/ Museum Paris 1952, coll. R. Oberthür. Paralectotypes: 1 female specimen (MNHN), Java/ type/ ex Musaeo Power/ Museum Paris 1952, coll. R. Oberthür; 1 male specimen (IRSNB), ♂/ Java/ M. R. Belg./ *Prophthalmus pugnator* Pow./ Lectotype [*in litteris*]/ *Prophthalmus pugnator* Power R. Damoiseau rev.; 1 male specimen (IRSNB), ♂/ Java/ M. R. Belg./ Paratype/ *Prophthalmus pugnator* Power R. Damoiseau rev.

***ruficornis* Senna, 1903**

ruficornis Senna, 1903: 174.

Type depositories. MSNG (LT!, PLT!), MZUF (PLT!).

Distribution. Indonesia (Sumatra, Sumbawa).

Notes. Lectotype of *P. ruficornis* (MSNG) designated on a male specimen with the following labels: Sumatra, Sirambè, XII.90-III.91, E. Modigliani/ *Prophthalmus ruficornis* Senna, cotypi. Paralectotypes: 5 specimens (MSNG), Sumatra, Sirambè, XII.90-III.91, E. Modigliani; 1 male specimen (MZUF), Sumatra, Sirambè, XII.90-III.91, E. Modigliani/ *Prophthalmus ruficornis* Senna, cotypus/ Bull. Soc. Ent. It. XXXIV, 1902, cotypus! ♂/ ex-coll. Senna/ "La Specola" Firenze, 6973.

***rugosicollis* (Power, 1879)**

rugosicollis Power, 1879b: 298 (*Baryrrhynchus*).

Type depository. IRSNB (LT!).

Distribution. India, "Indochina", Malaysia.

Notes. Lectotype of *B. rugosicollis* (IRSNB) designated on a male specimen with the following labels: Malacca/ Coll. Castelnau, Coll. Roelofs/ *Baryrrhynchus rugosicollis* Pow. ♂/ Type/ *Baryrrhynchus rugosicollis* Pow. ♂ [handwritten by Senna]/ *Baryrrhynchus rugosicollis* Pow. ♂ R. Damoiseau det., 1967/ Lectotype [*in litteris*].

tricolor Power, 1878

tricolor Power, 1878a: XXXVIII.

philippinensis Kleine, 1916b: 276 (*Prophthalmus tricolor* f.); **syn. n.**

celebensis Kleine, 1916b: 276 (*Prophthalmus tricolor* f.); **syn. n.**

ceramensis Kleine, 1916b: 276 (*Prophthalmus tricolor* f.); **syn. n.**

philippensis: Kleine, 1925f: 24 (*Prophthalmus tricolor* f.) [*lapsus*].

Type depositories. MNHN (LT!, PLT!), MCZ (PLT).

Distribution. Indonesia (Moluccas: Ambon, Buru, Seram [= Ceram]; Sulawesi), Philippines (Catanduanes, Leyte, Luzon, Mindanao, Negros, Samar, Siargao, Sibuyan).

Notes. Lectotype of *P. tricolor* (MNHN) designated on a male specimen with the following labels: Philippines/ H. Deyrolle ♂/ type/ ex Musaeo G. Power/ Museum Paris 1952, coll. R. Oberthür. Paralectotype: 1 specimen (MNHN), Moluques/ Chevrolat 72/ type/ ex Musaeo G. Power/ Museum Paris 1952, coll. R. Oberthür.

Kleine (1916b: 276) states that three geographical races exist for *tricolor*: *philippinensis*, *celebensis* and *ceramensis*, but does not define the type specimens. We list these forms as synonyms of the species.

tridentatus (Lund, 1800)

tridentatus Lund, 1800: 90 (*Brentus*).

tridentus: Beeson, 1925: 12 (*Prophthalmus*) [*lapsus*].

Type depository. ZMUC (HT).

Distribution. Taiwan; Borneo, India (Andaman Is., Assam), Indonesia (Java, Moluccas, Sulawesi, Sumatra), Malaysia, Philippines (Palawan), Vietnam; New Guinea.

***versicolor* Senna, 1892**

versicolor Senna, 1892e: 466.

Type depositories. MSNG (LT!, PLT!), MZUF (PLT!).

Distribution. Indonesia (Java), Myanmar [= Burma].

Notes. Lectotype of *P. versicolor* (MSNG) designated on a male specimen with the following labels: Carin Chebà, 900-1100 m, L. Fea, V-XII.1888/ *Prophthalmus versicolor* Senna typus! Paralectotypes: 2 specimens (MSNG), Carin Chebà, 900-1100 m, L. Fea, V-XII.1888; 1 male specimen (MZUF), Carin Chebà, 900-1100 m, L. Fea, V-XII.1888/ *Prophthalmus versicolor* Senna, typus!/ Ann. Mus. Genova, s. 2^a XII, 1892, cotypi! ♂ ♀/ ex-coll. Senna/ "La Specola" Firenze, 6972; 1 female specimen (MZUF), Carin Chebà, 900-1100 m, L. Fea, V-XII.1888/ ex-coll. Senna/ "La Specola" Firenze, 6972.

***wichmanni* Kleine, 1916**

wichmanni Kleine, 1916b: 263.

Type depositories. DEI (LT!, PLT!), MNHUB (PLT!), SMTD (PLT).

Distribution. S China, Taiwan; India (Assam, Darjeeling), Laos, Vietnam.

Notes. Lectotype of *P. wichmanni* (DEI) designated on a male specimen with the following labels: Tonkin, Than-Moi, Juni-Juli, H. Fruhstorfer/ *Prophthalmus wichmanni* Kl./ Syntypus. Paralectotypes: 2 female specimens (DEI), Tonkin, Than-Moi, Juni-Juli, H. Fruhstorfer/ col. Kraatz/ Kleine det.; 1 male specimen (MNHUB), Tonkin, Than-Moi, Juni-Juli, H. Fruhstorfer/ Lectotype [*in litteris*]/ *Prophthalmus wichmanni* Kl., det. Kleine; 1 male and 2 female specimens (MNHUB), Tonkin, Than-Moi, Juni-Juli, H. Fruhstorfer/ Paralec-
totype [*in litteris*].

***Pseudobelopherus* Calabresi, 1920**

Pseudobelopherus Calabresi, 1920b: 59.

Type species: *Pseudobelopherus orientalis* Calabresi, 1920.

Distribution. Malaysia.

deductus Kleine, 1925

deductus Kleine, 1925b: 157.

Type depository. NHM (HT!).

Distribution. Malaysia.

orientalis Calabresi, 1920

orientalis Calabresi, 1920b: 61.

Type depository. MZUF (LT!, PLT!).

Distribution. Malaysia.

Notes. Lectotype designated by Bartolozzi *et al.* (1985: 99) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Pseudomiolispa Kleine, 1926

Pseudomiolispa Kleine, 1926c: 169.

Type species: *Pseudomiolispa pendleburyi* Kleine, 1926.

Distribution. Indonesia (Java), Malaysia.

luchti Kleine, 1938

luchti Kleine, 1938f: 289.

Type depository. DEI (HT!).

Distribution. Indonesia (Java).

pendleburyi Kleine, 1926

pendleburyi Kleine, 1926c: 170.

Type depository. NHM (HT!).

Distribution. Malaysia.

Pseudorychodes Senna, 1894

Pseudorychodes Senna, 1894b: 375.

Carausius Senna, 1898e: 71 (non Stål, 1879) [see Alonso-Zarazaga & Lyal, 1999: 48].

Pseudorhychodes: R. Lucas, 1920: 552 [lapsus].

Type species: *Orychodes insignis* Lewis, 1883.

Distribution. China, Japan, Russia (E Siberia), Taiwan; Borneo, Indonesia (Java, Sulawesi, Sumatra), India (Assam, Sikkim), Malaysia, Myanmar [= Burma], Philippines (Mindanao), Thailand, Vietnam.

cochlaearis Damoiseau, 1989

cochlaearis Damoiseau, 1989b: 58.

Type depository. MZUF (HT!).

Distribution. Borneo.

crassus Senna, 1894

crassus Senna, 1894b: 378.

Type depositories. MZUF (LT!), MNHN (PLT!).

Distribution. India (Assam, Sikkim), Myanmar [= Burma].

Notes. Lectotypus of *P. crassus* (MZUF) designated on a male specimen with the following labels: Min. Rubini, Alta Birmania, H. Doherty/ *Pseudorychodes crassus* Senna, cotypus!/ Ann. Soc. Ent. Belg., XXXVIII, 1894, cotypus!/ ex-coll. Senna/ "La Specola" Firenze, 7013. Paralectotype: 1 male specimen (MNHN), Hte Birmanie, Mines des rubis, 1200-2300 m, Doherty

1890/ Dr. Senna vidit 1803/ Museum Paris 1952, coll. R. Oberthür/ *Pseudorychodes crassus* typus! ♂.

***cruentatus* Senna, 1898**

cruentatus Senna, 1898e: 71.

Type depository. IRSNB (HT!).

Distribution. Indonesia (Java, Sumatra), Malaysia, Myanmar [= Burma], Vietnam.

***damnosus* Kleine, 1925**

damnosus Kleine, 1925b: 149.

Type depository. NHM (LT!, PLT!).

Distribution. Malaysia.

Notes. Lectotype of *P. damnosus* (NHM) designated on a male specimen with the following labels: syntype/ Doherty/ Perak/ Fry Coll. 1905.100/ *Pseudorychodes damnosus* Kln. ♂ typus, R. Kleine det, 1922. Paralectotype: 1 female specimen (NHM), with the same label data as the lectotype.

***fruhstorferi* Senna, 1895**

fruhstorferi Senna, 1895c: 53.

Type depositories. MZUF (ST!), RMNH (ST!).

Distribution. Indonesia (Java).

***helleri* Senna, 1899**

helleri Senna, 1899: 304.

helteri: Kleine, 1932e: 287 (*Caenorychodes*) [lapsus].

Type depository. NHMB (LT!, PLT!).

Distribution. Indonesia (Sulawesi).

Notes. Lectotype of *P. helleri* (NHMB) designated on a female specimen with the following labels: typus/ Drs. Sarasin, N. Celebes, Masarang/ O. 12/ *Pseudorychodes helleri* n. Senna, ♀ typus. Paralectotype: 1 female specimen (NHMB), cotypus/ Drs. Sarasin, N. Celebes, Masarang/ O. 12.

honorus Kleine, 1926

honorus Kleine, 1926c: 168.

Type depositories. NHM (LT!, PLT!), SMTD (PLT).

Distribution. Malaysia, Thailand.

Notes. Lectotype of *P. honorus* (NHM) designated on a male specimen with the following labels: syntype/ Pahang, F. M. S. "Cameron's Highlands" N 04 Camp, 4800 ft., 17 Oct. 1923, H. M. Pendlebury/ Malay Pen. B. M. 1927-37/ *Pseudorychodes honorus* Kln., 1924, typus. Paralectotype: 1 female specimen (NHM), syntype/ Malay Pen. B. M. 1927-37/ Peninsular Siam, Nakon Sri Tamarat, Khao Luang, 2000 ft., 27 March 1922, H. M. Pendlebury/ *Pseudorychodes honorus* Kln. 1924, typus! R. Kleine det.

insignis (Lewis, 1883)

insignis Lewis, 1883: 301 (*Orychodes*).

Type depository. NHM (LT!, PLT!).

Distribution. Japan, Russia (E Siberia), Taiwan.

Notes. Lectotype of *P. insignis* (NHM) designated on a male specimen with the following labels: syntype/ Japan, J. Lewis, 1910-320/ Miyanoshita, 11.V.-14.V.1980/ *Orychodes insignis* Lewis, type ♂. Paralectotypes: 1 female specimen (NHM), syntype/ Nikko, 10.VIII.-18.VIII.81/ Japan, G. Lewis, 1910-320/ *Orychodes insignis* Lewis type ♀; 1 male and 2 female specimens (NHM), syntype/ Junsai Lake/ Japan, G. Lewis, 1910-320; 1 male specimen (NHM), syntype/ Junsai 1880/ Junsai Lake/ Japan, G. Lewis, 1910-320; 3 male and 4 female specimens (NHM), syntype/ Sapporo, 5.VIII.-16.VIII.80/ Japan, G. Lewis, 1910-320; 1 female specimen (NHM), syntype/ Japan, G. Lewis, 1910-320/ *Orychodes insignis* Lewis; 1 male specimen (NHM), syntype/ Awomori, 2.IX.-9.IX.80/ Japan, G. Lewis, 1910-320; 1 male specimen (NHM), syntype/ Hakodate/ Hakodate/ Japan, G. Lewis, 1910-320; 1 male specimen (NHM), syntype/ *Orychodes? insignis* Lewis Japan/ Sharp Coll.

1905-313/ *Pseudorychodes insignis* Lewis, 1923, R. Kleine det.; 1 female specimen (NHM), syntype/ paratype/ Japan, G. Lewis, 1910-320; 1 male specimen (NHM), syntype/ paratype/ Yuyama, 10.V.-14.V.81/ Japan, G. Lewis, 1910-320; 1 female specimen (NHM), syntype/ Japan, G. Lewis, 1910-320; *Pseudorychodes insignis* Lewis, co-type.

lineolatus (Kirsch, 1875)

lineolatus Kirsch, 1875b: 49 (*Orychodes*).

Type depository. SMTD (ST).

Distribution. Borneo, Indonesia (Sumatra), Malaysia, Thailand.

mandschuricus Kleine, 1939

mandschuricus Kleine, 1939b: 318.

Type depository. DEI (HT!).

Distribution. China (Heilongjiang).

piliferus (Senna, 1892)

piliferus Senna, 1892: 177 (*Orychodes*).

Type depository. RMNH (HT).

Distribution. Borneo, Indonesia (Java).

praeclarus Kleine, 1925

praeclarus Kleine, 1925c: 600.

Type depository. SMTD (HT).

Distribution. Philippines (Mindanao).

ritsemae (Senna, 1891)

ritsemae Senna, 1891: 161 (*Orychodes*).

Type depository. MZUF (HT!).

Distribution. Indonesia (Sumatra), Malaysia, Myanmar [= Burma].

sauteri Kleine, 1928

sauteri Kleine, 1928e: 100.

Type depository. SMFD (HT, PT).

Distribution. Taiwan.

spinitibia Damoiseau, 1989

spinitibia Damoiseau, 1989b: 59.

Type depositories. IRSNB (HT!), MZUF (AT!, PT!).

Distribution. Indonesia (Java).

tenuirostris Senna, 1894

tenuirostris Senna, 1894b: 376.

Type depositories. MNHN (LT!), SMTD (PLT).

Distribution. Borneo, Malaysia, Myanmar [= Burma].

Notes. An 'Holotype' was quoted by Bartolozzi *et al.* (1985: 101) in MZUF, but this specimen is not present in the collection and it seems to be lost; we designate the Lectotype of *P. tenuirostris* on a male specimen (MNHN) with the following labels: Hte Birmanie, Mines des rubis, 1200-2300 m, Doherty 1890/ Museum Paris 1952, coll. R. Oberthür/ *Pseudo-orychodes tenuirostris* typus! ♂.

Raphirhynchidus Kleine, 1927

Raphirhynchidus Kleine, 1927b: 441.

Rhaphirhynchus: Kleine, 1926a: 177 [lapsus].

Rhaphirhynchidus: Kleine, 1927b: 504 [lapsus].

Rhaphirhynchidus: Kleine, 1938d: 120 [lapsus].

Type species: *Raphirhynchidus excitatus* Kleine, 1927.

Distribution. Brazil.

eliasi Soares, 1970

eliasi Soares, 1970a: 201.

Type depository. MNRJ (Campos Seabra coll.) (HT, PT).

Distribution. Brazil.

excitatus Kleine, 1927

excitatus Kleine, 1927b: 442.

Type depository. NHM (HT!).

Distribution. Brazil.

Raphirhynchus Dejean, 1834

Raphirhynchus Dejean, 1834: 243.

Rhaphirhynchus Schoenherr, 1840: 504 [unjustified emendation].

Raphidorhynchus Agassiz, 1846b: 321 [unjustified emendation].

Rhaphidorhynchus: Agassiz, 1846b: 323 [lapsus].

Rhaphidorrhynchus Gemminger & Harold, 1872: 2711 [unjustified emendation].

Rhaphidorrhynchus: Kleine, 1921a: 42 [lapsus].

Type species: *Arrhenodes nitidicollis* Gyllenhal, 1833 = *Brentus cylindricornis* Fabricius, 1787.

Distribution. Costa Rica (including Coco Is.), Guadeloupe Is., Guatemala, Mexico, Nicaragua, Panama; Brazil, French Guyana.

amazonicus (Senna, 1894)

amazonicus Senna, 1894d: 598 (*Raphidorrhynchus*).

Type depository. MZUF (HT!).

Distribution. Brazil.

amplicollis (Senna, 1894)

amplicollis Senna, 1894d: 597 (*Raphidorrhynchus*).

Type depositories. MZUF (LT!); IRSNB (PLT!).

Distribution. Mexico; Brazil, French Guyana.

Notes. Lectotype designated by Bartolozzi *et al.* (1985: 94) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

We did not find any syntype from Mexico, one of the type localities (Senna, 1894d: 597), either in IRSNB or in MZUF.

anomalus Sharp, 1895

anomalus Sharp, 1895a: 59 (*Rhaphirhynchus*).

Type depository. NHM (HT!).

Distribution. Panama.

auricollis Sharp, 1895

auricollis Sharp, 1895a: 58 (*Rhaphirhynchus*).

Type depository. NHM (LT!, PLT!).

Distribution. Guatemala, Mexico, Nicaragua.

Notes. Lectotype of *R. auricollis* (NHM) designated on a female specimen with the following labels: *Rhaphirhynchus auricollis* Type D. S., Chontales, Nicaragua, T. Belt/ syntype/ type/ B. C. A., Col., IV, 6. *Rhaphirhynchus auricollis* Sharp/ Sharp Coll. 1905-313. Paralectotype: 1 male specimen (NHM),

Rhaphirhynchus auricollis Type D. S., Toxpam, Mexico, Salle Coll./ syntype/ type/ B. C. A., Col., IV, 6. *Rhaphirhynchus auricollis* Sharp/ Sharp Coll. 1905-313.

***chiriquensis* Sharp, 1895**

chiriquensis Sharp, 1895a: 60 (*Rhaphirhynchus*).

Type depository. NHM (LT!, PLT!).

Distribution. Costa Rica, Panama.

Notes. Lectotype of *R. chiriquensis* (NHM) designated on a male specimen with the following labels: *Rhaphirhynchus chiriquensis* ♂ Type, V. de Chiriqui, 2-3000 ft., Champion/ syntype/ B. C. A., Col., IV, 6. *Rhaphirhynchus chiriquensis*, Sharp/ Sharp Coll. 1905-313. Paralectotypes: 1 male specimen (NHM), *Rhaphirhynchus chiriquensis* var. D. S., V. de Chiriqui, 2-3000 ft., Champion/ syntype/ B. C. A., Col., IV, 6. *Rhaphirhynchus chiriquensis*, Sharp/ Sharp Coll. 1905-313; 2 male specimens (NHM), V. de Chiriqui, 2-3000 ft., Champion/ syntype/ B. C. A., Col., IV, 6. *Rhaphirhynchus chiriquensis*, Sharp/ Sharp Coll. 1905-313; 1 male specimen (NHM), *Rhaphirhynchus chiriquensis* D. S., Bugaba, 800-1500 ft., Champion/ syntype/ B. C. A., Col., IV, 6. *Rhaphirhynchus chiriquensis*, Sharp/ Sharp Coll. 1905-313.

***cocoensis* (Calabresi, 1920)**

cocoensis Calabresi, 1920b: 71 (*Rhaphidorrhynchus*).

Type depository. MZUF (HT!).

Distribution. Costa Rica (Coco Is.).

***consanguineus* (Senna, 1895)**

consanguineus Senna, 1895b: 217 (*Rhaphidorrhynchus*).

Type depositories. MNHN (LT!, PLT!), MZUF (PLT!).

Distribution. Venezuela.

Notes. Lectotype of *R. consanguineus* (MNHN) designated on a male specimen with the following labels: Merida, Venezuela/ ♂/*Raphidorhynchus consanguineus* n. sp. Senna, cotypi!/ Senna vidit 1895/ ex Musaeo G. Power/ Museum Paris 1952, coll. R. Oberthür. Paralectotypes: 1 female specimen (MNHN), Merida, Venezuela/ ♀/ Senna vidit 1895/ ex Musaeo G. Power/ Museum Paris 1952, coll. R. Oberthür; 1 specimen (MZUF), Merida, Venezuela, Tring Mus./ *Raphidorhynchus consanguineus* Senna, cotypi!/ Not. Leyden Mus. XVI, 1895, cotypi!/ ex-coll. Senna/ "La Specola" Firenze, 7022; 2 specimens (MZUF), Merida, Venezuela, Tring Mus./ ex-coll. Senna/ "La Specola" Firenze, 7022; 1 specimen (MZUF), Merida, Venezuela, Tring Mus./ cotypi, ex Mus. Tring/ ex-coll. Senna/ "La Specola" Firenze, 7022.

cylindricornis (Fabricius, 1787)

cylindricornis Fabricius, 1787: 96 (*Brentus*).

nitidicollis Gyllenhal (in Schoenherr, 1833: 328) (*Arrhenodes*); syn. Gemminger & Harold, 1872: 2711.

Type depositories. *B. cylindricornis*: ZMUK (HT); *A. nitidicollis*: NHRS (HT).

Distribution. Costa Rica, Dominican Republic, Guadeloupe Is.; Brazil, Colombia, Venezuela.

deceptor Sharp, 1895

deceptor Sharp, 1895a: 62 (*Rhaphirhynchus*).

Type depository. NHM (HT!).

Distribution. Costa Rica, Nicaragua.

duplicatus (Germar, 1824)

duplicatus Germar, 1824: 189 (*Brentus*).

Type depository. MLUH (HT).

Distribution. Brazil.

Notes. According to Schoenherr (1840: 505), Senna (1894e: 602) and Haedo Rossi (1961: 170) this species is a synonym of *R. longimanus*, but Damoiseau (1966f: 305) after examining its genitalia, proved it was a *bona species*.

***excellens* Kleine, 1927**

excellens Kleine, 1927b: 440.

Type depository. NHM (LT!, PLT!).

Distribution. Ecuador.

Notes. Lectotype of *R. excellens* (NHM) designated on a male specimen with the following labels: syntype/ Ecuador/ Buckley/ *Rhaphirhynchus excellens* ♂ Type R. Kleine det. 1923. Paralectotype: 1 female specimen (NHM), with the same label data as the lectotype.

***excelsus* Kleine, 1927**

excelsus Kleine, 1927b: 437.

Type depository. NHM (HT!).

Distribution. Brazil.

***favorabilis* Kleine, 1927**

favorabilis Kleine, 1927b: 439.

Type depository. NHM (HT!).

Distribution. Brazil.

***fecundus* Kleine, 1927**

fecundus Kleine, 1927b: 438.

Type depository. NHM (HT!).

Distribution. Brazil, French Guyana, Peru.

***filicornis* Sharp, 1895**

filicornis Sharp, 1895a: 54 (*Rhaphirhynchus*).

Type depository. NHM (HT!).

Distribution. Panama.

***insculptus* (Senna, 1889)**

insculptus Senna, 1889: 36 (*Raphidorrhynchus*).

Type depository. MZUF (HT!).

Distribution. Brazil, Ecuador, Peru.

***jansoni* Sharp, 1895**

jansoni Sharp, 1895a: 59 (*Rhaphirhynchus*).

Type depository. NHM (HT!).

Distribution. Nicaragua.

***linearis* (Senna, 1894)**

linearis Senna, 1894d: 607 (*Raphidorrhynchus*).

Type depository. IRSNB (HT!).

Distribution. Guatemala, Mexico, Nicaragua; Bolivia.

Notes. Bartolozzi *et al.* (2002: 99) reported a male syntype in MZUF, but the specimen is actually a female and does not belong to the type material.

***longimanus* (Lund, 1800)**

longimanus Lund, 1800: 85 (*Brentus*).

rhinoprion Perty, 1832: 69 (*Arrhenodes*); syn. Gemminger & Harold, 1872: 2711.

indicatus Gyllenhal (in Schoenherr, 1833: 330) (*Arrhenodes*); syn. Schoenherr, 1840: 505.

Type depositories. *B. longimanus*: ZMUC (ST); *A. rhinoprion*: ZSM (?) (HT); *A. indicatus*: NHRS (HT).

Distribution. Guatemala, Mexico, Puerto Rico; Argentina, Brazil, Colombia, Venezuela.

Notes. Kleine (1928a: 85) reported that the type for *A. rhinoprion* was in ZSM, but according to M. Baehr (ZSM) (2001, personal communication) this type is missing; it was probably lost during World War II like many other Perity types.

longulus Sharp, 1895

longulus Sharp, 1895a: 54 (*Rhaphirhynchus*).

Type depository. NHM (LT!, PLT!).

Distribution. Costa Rica, Nicaragua, Panama.

Notes. Lectotype of *R. longulus* (NHM) designated on a male specimen with the following labels: *Rhaphirhynchus longulus* Type D. S., Chontales, Jonson/ syntype/ *Rhaphirhynchus longulus* Sharp. Paralectotypes: 1 male specimen (NHM), *Rhaphirhynchus longulus*/ syntype/ Chontales, Jonson/ Sharp Coll. 1905-313/ B. C. A., Col., IV, 6. *Rhaphirhynchus longulus* Sharp; 1 male specimen (NHM), *Rhaphirhynchus longulus*/ syntype/ Chontales, Jonson/ Sharp Coll. 1905-313/ B. C. A., Col., IV, 6. *Rhaphirhynchus* D. S., V. de Chiriqui, 25-4000 ft., Champion; 1 female specimen (NHM), *Rhaphirhynchus longulus* ♀ D. S., Bugaba, 800-1,500 ft., Champion/ syntype/ B. C. A., Col., IV, 6. *Rhaphirhynchus longulus* Sharp.

mexicanus (Senna, 1894)

mexicanus Senna, 1894d: 608 (*Raphidorrhynchus*).

Type depository. IRSNB (HT!).

Distribution. Guatemala, Mexico.

obenbergeri Kleine, 1926

obenbergeri Kleine, 1926e: 177 (*Rhaphirhynchus*).

Type depository. NMPC (HT).

Distribution. Brazil.

***obscurus* Sharp, 1895**

obscurus Sharp, 1895a: 56 (*Rhaphirhynchus*).

Type depository. NHM (LT!, PLT!).

Distribution. Guatemala.

Notes. Lectotype of *R. obscurus* (NHM) designated on a female specimen with the following labels: *Rhaphirhynchus obscurus* Type D. S., Cerro Zunil, 4000 ft., Champion/ syntype/ type/ B. C. A., Col., IV, 6. *Rhaphirhynchus obscurus* Sharp. Paralectotype: 1 female specimen (NHM), with the same label data as the lectotype.

***panamensis* (Senna, 1894)**

panamensis Senna, 1894d: 609 (*Raphidorrhynchus*).

panamanensis: Kleine, 1938d: 114 (*Raphirhynchus*) [lapsus].

Type depository. MZUF (HT!).

Distribution. Mexico, Nicaragua, Panama; Venezuela.

***politus* (Senna, 1894)**

politus Senna, 1894d: 605 (*Rhaphidorrhynchus*).

Type depository. IRSNB (HT!).

Distribution. Costa Rica, Mexico, Nicaragua.

***principalis* Sharp, 1895**

principalis Sharp, 1895a: 55 (*Rhaphirhynchus*).

Type depository. NHM (HT!).

Distribution. Costa Rica, Guatemala.

***prosperus* Kleine, 1943**

prosperus Kleine, 1943b: 139 (*Raphirrhynchus*).

Type depository. HNHM (HT).

Distribution. Peru.

***pulcherrimus* Soares, 1966**

pulcherrimus Soares, 1966b: 319.

Type depository. MNRJ (Campos Seabra coll.) (HT).

Distribution. Peru.

***rothschildi* (Senna, 1895)**

rothschildi Senna, 1895b: 214 (*Rhaphidorrhynchus*).

Type depository. MNHN (LT!, PLT!).

Distribution. Nicaragua; Brazil.

Notes. Lectotype of *R. rothschildi* (MNHN) designated on a male specimen with the following labels: Nicaragua/ *Raphidorhynchus rothschildi* Senna ♂ typus! n. sp/ Senna vidit 1895/ ex Musaeo G. Power/ Museum Paris 1952, coll. R. Oberthür. Paralectotype: 1 female specimen (MNHN), Nicaragua/ Senna vidit 1895/ ex Musaeo G. Power/ Museum Paris 1952, coll. R. Oberthür.

***scobinirostris* (Gyllenhal, 1833)**

scobinirostris Gyllenhal (in Schoenherr, 1833: 328) (*Arrhenodes*).

Type depository. NHRS (HT).

Distribution. Brazil.

sennai Sharp, 1895

sennai Sharp, 1895a: 61 (*Rhaphirhynchus*).

Type depository. NHM (LT!, PLT!).

Distribution. Guatemala.

Notes. Lectotype of *R. sennai* (NHM) designated on a male specimen with the following labels: *Rhaphirhynchus sennai* ♂ Type D. S., Perula, Vera Paz, Champion/ syntype/ B. C. A., Col., IV, 6. *Rhaphirhynchus sennai* Sharp/ Sharp Coll. 1905-313. Paralectotypes: 4 male and 1 female specimens (NHM), with the same label data as the lectotype; 1 male and 1 female specimen (NHM), Perula, Vera Paz, Champion/ syntype/ B. C. A., Col., IV, 6. *Rhaphirhynchus sennai* Sharp/ Sharp Coll. 1905-313.

severini (Senna, 1894)

severini Senna, 1894d: 602 (*Raphidorrhynchus*).

Type depository. MZUF (HT!).

Distribution. Mexico.

sexvittatus (Senna, 1894)

sexvittatus Senna, 1894d: 596 (*Raphidorrhynchus*).

Type depository. MZUF (LT!, PLT!).

Distribution. Costa Rica, Guatemala, Mexico, Nicaragua, Panama; Colombia, Brazil.

Notes. Lectotype of *R. sexvittatus* (MZUF) designated on a male specimen with the following labels: Panama, A. Boucard/ *Rhaphidorr. sexvittatus* Senna ♂ ♀ typi!/ Ann. Soc. Ent. Belg., XXXVIII, 1894 typi! ♂ ♀/ ex coll. Senna/ "La Specola" Firenze, 7027. Paralectotype: 1 female specimen (MZUF), Chiriqui, Americ. centr., d. Staudinger/ ex coll. Senna/ "La Specola" Firenze, 7028.

***signifer* Boheman, 1840**

signifer Boheman (in Schoenherr, 1840: 505) (*Rhaphirhynchus*).

Type depository. NHRS (HT).

Distribution. Mexico.

***variabilis* (Senna, 1894)**

variabilis Senna, 1894d: 606 (*Raphidorrhynchus*).

Type depositories. MZUF (LT!, PLT!), IRSNB (PLT!).

Distribution. Mexico; Brazil, Peru.

Notes. Lectotype of *R. variabilis* (MZUF) designated on a male specimen with the following labels: Oajaca, Messico, ex Mus. Belg./ *Raphidorrhynchus* 4-lineatus (Jek.) marginistriatus (Chevr.) Power ♂ / *Raphidorrhynchus variabilis* typus, dono Severin, Mus. Belgique/ Ann. Soc. Ent. Belg. XXXVIII, 1894, cotypi ♂ ♀ / *R. variabilis* Senna, cotypus ♂ / ex-coll. Senna/ "La Specola" Firenze, 7029. Paralectotypes: 1 female specimen (MZUF), Messico, ex coll. Chevrolat, d. Mus. Belg./ *Claeoderes marginistriatus* Chev. Mexique/ ex-coll. Senna/ "La Specola" Firenze, 7029; 1 female specimen (MZUF), Oajaca, Messico, ex Mus. Belg./ ex-coll. Senna/ "La Specola" Firenze, 7029; 1 male specimen (IRSNB), Oajaca/ Coll. Lacordaire/ ♂ / M. R. Belg./ Type/ *Raphidorrhynchus lineatus* Jek. in litt./ *Raphidorrhynchus variabilis* Senna typus!; 1 male specimen (IRSNB), Mexique/ Coll. Roelofs/ M. R. Belg./ *Claeodores biserrirostris* Schoenherr/ Paratype; 1 female specimen (IRSNB), Mexique/ Coll. Roelofs/ M. R. Belg./ Paratype; 1 female specimen (IRSNB), Coll. Roelofs/ ♀ / M. R. Belg./ Paratype.

***vicinus* (Senna, 1894)**

vicinus Senna, 1894d: 599 (*Raphidorrhynchus*).

Type depositories. MZUF (LT!, PLT!), IRSNB (PLT!).

Distribution. Brazil.

Notes. Lectotype of *R. vicinus* (MZUF) designated on a specimen with the following labels: Petropolis, Brasile, A. Boucard/ *Rhaphirhynchus vicinus* Senna, olotypi! [sic!] ♂ ♀ / Ann. Soc. Ent. Belg., XXXVIII, 1894, olotypi!

[sic!] ♂ ♀/ ex-coll. Senna/ "La Specola" Firenze, 7030; 1 specimen (MZUF), Petropolis, Brasile, A. Boucard/ ex-coll. Senna/ "La Specola" Firenze, 7030; 2 specimens (MZUF), Rio Janeiro, Brasile, ex Staudinger/ ex-coll. Senna/ "La Specola" Firenze, 7031; 1 male specimen (IRSNB), Brésil/ Coll. Roelofs/ M. R. Belg./ Type/ *Raphirhynchus vicinus* Dej. Brasil/ *Rhaphid. vicinus* Senna ♂ typus!; 1 female specimen (IRSNB), ♀/ Brésil/ Coll. Dejean, Coll. Roelofs/ M. R. Belg./ *Vicinus* Dej./ *Raphirhynchus vicinus* mihi, h. in Brasilia, D. Gory/ Paratype.

***Rhynchoneus* Sharp, 1895**

Rhynchoneus Sharp, 1895a: 47.

Type species: *Rhynchoneus belti* Sharp, 1895.

Distribution. Nicaragua; Brazil; Paraguay, Venezuela.

***belti* Sharp, 1895**

belti Sharp, 1895a: 47.

vampyrus Senna, 1897a: 230 (*Arrhenodes*); syn. Soares, 1966a: 276.

Type depositories. *R. belti*: NHM (LT!, PLT!); *A. vampyrus*: IRSNB (HT!).

Distribution. Nicaragua; Brazil, Paraguay, Venezuela.

Notes. Lectotype of *R. belti* (NHM) designated on a female specimen with the following labels: *Rhynchoneus belti* Type D. S./ Chontales, Nicaragua, T. Belt/ syntype/ Sharp Coll. 1905-313/ B. C. A., Col., IV, 6. *Rhynchoneus belti* Sharp. Paralectotypes (NHM): 3 female specimens with the same label data as the lectotype.

***Schizoeupsalis* Kleine, 1917**

Schizoeupsalis Kleine, 1917g: 127.

Type species: *Eupsalis promissa* Pascoe, 1872.

Distribution. Indonesia (Mofuccas: Aru, Bacan [= Batjan], Ceram, Damar [= Dammer], Halmahera [= Gilolo], Kisar [= Kisser], Ternate; Sulawesi, Sumba, Sumbawa); Australia, New Guinea.

promissa (Pascoe, 1872)

promissus Pascoe, 1872: 323 (*Eupsalis*).

Type depository. NHM (LT!, PLT!).

Distribution. Indonesia (Moluccas: Aru, Bacan [= Batjan], Ceram, Damar [= Dammer], Halmahera [= Gilolo], Kisar [= Kisser], Ternate; Sulawesi, Sumba, Sumbawa); Australia, New Guinea.

Notes. Lectotype of *E. promissa* (NHM) designated on a male specimen with the following labels: syntype/ *Eupsalis promissa* Type Pascoe/ Batchian/ Pascoe, Coll. 93-60/ *Eupsalis promissus* Pasc. Paralectotypes: 2 male and 2 female specimens (NHM), syntype/ Batchian/ Pascoe, Coll. 93-60.

Schoenfeldtia Senna, 1893

Schoenfeldtia Senna, 1893d: 120.

Type species: *Schoenfeldtia impressicollis* Senna, 1893.

Distribution. Bolivia, Brazil.

impressicollis Senna, 1893

impressicollis Senna, 1893c: 121.

Type depository. SMFD (von Schönfeldt coll.) (HT).

Distribution. Brazil.

sennai Haedo Rossi, 1965

sennai Haedo Rossi, 1965: 318.

Type depository. CSMBA (HT, AT).

Distribution. Bolivia.

***Schoenfeldtiopsis* Soares & Scivittaro, 1977**

Schoenfeldtiopsis Soares & Scivittaro, 1977: 96.

Type species: *Schoenfeldtiopsis zikani* Soares & Scivittaro, 1977.

Distribution. Brazil.

***zikani* Soares & Scivittaro, 1977**

zikani Soares & Scivittaro, 1977: 96.

Type depositories. FIOC (HT), MZSP (Diringshofen coll.) (PT).

Distribution. Brazil.

***Spatherrhinus* Power, 1879**

Spatherrhinus Power, 1879a: 493.

Spatherrhinus: Power, 1879a: 491 [lapsus].

Spatherrhinus Rye, 1880: 105 [unjustified emendation].

Eupsalithopsis Kleine, 1914a: 196; syn. Kleine, 1927a: 37.

Eupralithopsis: Sharp, 1916: 166 [lapsus].

Spatherrhinus: Kleine, 1921a: 35 [lapsus].

Type species: *Arrhenodes gabonicus* J. Thomson, 1858.

Distribution. Angola, Cameroon, Central African Republic, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea, Gabon, Ghana, Ivory Coast, Liberia, Malawi, Nigeria, Republic of the Congo, Tanzania, Togo; Italy (I).

***gabonicus* (J. Thomson, 1858)**

gabonicus J. Thomson, 1858: 116 (*Arrhenodes*).

submaculatus Kolbe, 1883b: 238 (*Eupsalis*); syn. Kleine, 1927a: 38.

juvenilis Kolbe, 1888: 307 (*Spatherrhinus*); syn. Schönfeldt, 1908: 40.

stuhlmanni Kolbe, 1897: 286 (*Spatherrhinus*); syn. De Muizon, 1955b: 892.

curiosus Kleine, 1926f: 368 (*Spatherrhinus*); syn. De Muizon, 1955b: 892.

Type depositories. *A. gabonicus*: MNHN (HT!); *E. submaculatus*: MNHUB (HT!); *S. juvenilis*: MNHUB (HT!); *S. stuhlmanni*: MNHUB (HT!); *S. curiosus*: NHM (LT!, PLT!).

Distribution. Angola, Cameroon, Central African Republic, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea, Gabon, Ghana, Ivory Coast, Liberia, Nigeria, Republic of the Congo, Tanzania.

Notes. In MNHN we found a female specimen of *A. gabonicus* with the following labels “Type 45/ Gabon/ type/ ex Musaeo Power/ Museum Paris 1952 coll. R. Oberthür”; we are not sure if this specimen belongs to the type series.

Lectotype of *S. curiosus* (NHM) designated on a male specimen with the following labels: syntype/ Gold Coast/ G. A. Higlett., 1903-13, Tamsoo near Tarkwa/ *Spatherrhinus curiosus* Kln. R. Kleine det, 1922. Paralectotype: 1 female specimen (NHM) with the same label data as the lectotype.

Damoiseau (1967a: 350) did not mention Nigeria (Senna, 1894a: 411), Liberia (Senna, 1896b: 254), Ghana (Kleine, 1926f: 369), and Tanzania (Kleine, 1936a: 33) when he gave the species distribution.

grandis Kleine, 1914

grandis Kleine, 1914a: 191.

Type depository. MZPW (ST).

Distribution. Democratic Republic of the Congo (ex Zaire), Tanzania.

Notes. According to Damoiseau (1967a: 356) Kleine's syntypes were destroyed during the World War II, so he designated a neotype in IRSNB. T. Huflejt (MZPW) (2002, personal communication) informed us that a syntype for *S. grandis* does exist in MZPW, thus Damoiseau's neotype is not valid.

longiceps Kolbe, 1888

longiceps Kolbe, 1888: 308.

longuiceps: Mayné & Donis, 1962: 100 (*Spatherrhinus*) [lapsus].

Type depository. MNHUB (LT!, PLT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Gabon, Guinea Bissau, Ivory Coast, Liberia, Nigeria, Principe, Republic of the Congo, Tanzania, Togo; Italy (I).

Notes. Lectotype of *S. longiceps* (MNHUB) designated on a male specimen with the following labels: Gabon, dr. Büttner/ Type/ *Spatherrhinus longiceps* n. sp. Kolbe/ 65862. Paralectotype: 1 male specimen (MNHUB), Gabon, dr. Büttner/ Type/ *Buttneri*.

Damoiseau (1967a: 355) did not mention Nigeria (Senna, 1894a: 411) and Liberia (Senna, 1896b: 254) when he gave the species distribution.

medioximus (J. Thomson, 1858)

medioximus J. Thomson, 1858: 117 (*Arrhenodes*).

eupsaloides Kolbe, 1888: 306 (*Spatherrhinus*); syn. Kleine, 1927a: 38.

picturatus Kolbe, 1888: 306 (*Spatherrhinus*); syn. Kleine, 1927a: 38.

Type depositories. *A. medioximus*: MNHN (HT!); *S. eupsaloides*: MNHUB (HT!); *S. picturatus*: MNHUB (HT!).

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Gabon, Ivory Coast, Liberia (?), Malawi, Nigeria, Republic of the Congo, Togo.

Notes. We also found a female specimen of *A. medioximus* in MNHN with the following labels "Type 40/ type/ ex Musaeo Power"; this can not be a type because the species was described on a single male specimen.

Damoiseau (1967a: 347) did not mention Angola (Senna, 1892c: 47), Nigeria (Senna, 1894a: 411), and Liberia (Senna, 1896b: 254) when he gave the species distribution.

opacus (J. Thomson, 1858)

opacus J. Thomson, 1858: 116 (*Arrhenodes*).

directus Kleine, 1924a: 106 (*Spatherrhinus*); syn. De Muizon, 1955b: 893.

Type depositories. *A. opacus*: MNHN (HT!); *S. directus*: MRAC (HT!).

Distribution. Democratic Republic of the Congo (ex Zaire), Gabon, Ivory Coast.

Notes. In MNHN we also found two other specimens of *A. opacus* with the following labels “ Gabon/ type/ Mniszech 9/ ex Musaeo Power/ Museum Paris 1952, coll. R. Oberthür ” and “ Gabon/ type/ Mniszech 7/ ex Musaeo Power/ Museum Paris 1952, coll. R. Oberthür” which can not be types because the species was described on a single specimen.

Kleine (1938d) did not mention *S. directus* among the species listed in the Genera Insectorum; Damoiseau (1967a) did not mention this species either.

***ophthalmicus* Kolbe, 1888**

ophthalmicus Kolbe, 1888: 307.

spatherinoides Kleine, 1914a: 196 (*Eupsalithopsis*); syn. De Muizon, 1955b: 893.

Type depositories. *S. ophthalmicus*: MNHUB (HT!); *E. spatherinoides*: MZPW (ST).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Gabon.

Notes. The quotation for the People's Republic of Congo (Sforzi, 1998b: 268) is incorrect and should be referred to as the Democratic Republic of the Congo.

***Stratiorrhina* Pascoe, 1872**

Stratiorrhina Pascoe, 1872: 322.

Stratiorrhina: Kleine, 1932d: 279 [lapsus].

Type species: *Arrhenodes xiphias* Westwood, 1848.

Distribution. Borneo, Indonesia (Java; Moluccas: Aru; Sumatra), Malaysia, Thailand; Australia (Prince of Wales Is.), New Guinea.

***amatoria* Kleine, 1923**

amatoria Kleine, 1923c: 128.

Type depository. CUMZ (HT!).

Distribution. Borneo.

***concors* Kleine, 1923**

concors Kleine, 1923i: 272.

Type depository. NHM (LT!, PLT!).

Distribution. Malaysia.

Notes. Lectotype of *S. concors* (NHM) designated on a male specimen with the following labels: syntype/ typus/ Maxwell's Hill, Perak, 4076', 19 Aug. 1908/ *Stratiorrhina concors* Kleine, det. Aut., 1922, Paralectotypes: 1 female specimen (NHM), Maxwell's Hill, Perak, 3000', 24 Aug. 1908/ ex F.M.S. Museum B.M. 1955-354/ *Stratiorrhina concors* Kleine, det. by Kleine 1922, Type sent to Brit. Mus.; 1 female specimen (NHM), Maxwell's Hill, Perak, 4078', 16 Aug. 1908/ ex F.M.S. Museum B.M. 1955-354; 1 female specimen (NHM), Maxwell's Hill, Perak, 3758', Aug. 1908/ ex F.M.S. Museum B.M. 1955-354; 1 male specimen (NHM), syntype/ Typus!/ 87-4/ Perak/ *Stratiorrhina concors* Kleine, R. Kleine det., 1922; 1 female specimen (NHM), syntype/ 87-4.

***femoralis* Ritsema, 1882**

femoralis Ritsema, 1882a: 188.

Type depository. RMNH (ST).

Distribution. Indonesia (Sumatra).

***inermis* Calabresi, 1921**

inermis Calabresi, 1921b: 51.

Type depository. MSNG (HT!).

Distribution. Borneo.

***major* Calabresi, 1921**

major Calabresi, 1921b: 47.

Type depository. MZUF (LT!, PLT!).

Distribution. Borneo, Indonesia (Sumatra), Malaysia, Thailand.

Notes. Lectotype of *S. major* (MZUF) designated on a male specimen with the following labels: Borneo, A. Boucard/ *Stratiorrhina major* Calabresi, typus!/ Bull. Soc. Ent. It., LIII, 1921, cotypi!/ ex-coll. Senna/ "La Specola" Firenze, 7005. Paralectotypes: 1 male specimen (MZUF) Sumatra, Dohrn, don. Dr. Faust/ ex-coll. Senna/ "La Specola" Firenze, 7007; 1 female specimen (MZUF), Borneo, A. Boucard/ *Stratiorrhina major* Calabresi ♀/ ex-coll. Senna/ "La Specola" Firenze, 7005; 1 female specimen (MZUF), Coll. Nonfried, Siam/ ex-coll. Senna/ "La Specola" Firenze, 7006.

mirabilis Kleine, 1926

mirabilis Kleine, 1926a: 43.

Type depository. NHM (HT!).

Distribution. Indonesia (Moluccas: Aru); New Guinea.

pascoei Kirsch, 1875

pascoei Kirsch, 1875b: 48.

Type depository. SMTD (ST!).

Distribution. Borneo, Malaysia.

pulla Kleine, 1936

pulla Kleine, 1936b: 121.

Type depositories. NHM (LT!, PLT!), DEI (PLT!).

Distribution. Malaysia.

Notes. Lectotype of *S. pulla* (NHM) designated on a male specimen with the following labels: syntype/ typus ♂/ F. M. S., Cameron's Highlands, 5000-5500 ft., 12.6.1935, H. M. Pendlebury/ Brit. Mus. 1937-97/ *Stratiorrhina pulla* Kln., R. Kleine det., 1935. Paralectotypes: 1 female (NHM), Syntype/ typus ♀/ Pahang F. M. S., Cameron's Highlands, 5000 ft., 11.6.1935, H.M. Pendlebury/ ex F. M. S. Museum B.M. 1955-354; 1 female specimen (DEI), Pahang,

F. M. S., Cameron's Highlands, 5000 ft., 9.6.1935, H. M. Pendlebury/ ex F. M. S. Museums/ *Stratiorrhina pulla* Kln., R. Kleine det., 1935.

***tarsalis* Sforzi & Bartolozzi, 1993**

tarsalis Sforzi & Bartolozzi, 1993: 293.

Type depositories. MZUF (HT!), NHM (PT!).

Distribution. Borneo, Malaysia.

***xiphias* (Westwood, 1848)**

xiphias Westwood, 1848: 31 (*Arrhenodes*).

Type depository. OXUM (HT).

Distribution. Borneo, Indonesia (Java), Malaysia; Australia (Prince of Wales Is.).

***Stratiorrhynchus* Damoiseau, 1962**

Stratiorrhynchus Damoiseau, 1962b: 32.

Type species: *Belopherus strigicollis* Lacordaire, 1866.

Distribution. Cuba, Dominican Republic.

***strigicollis* (Lacordaire, 1866)**

strigicollis Lacordaire, 1866: 435 (*Belopherus*).

Type depository. IRSNB (HT!).

Distribution. Cuba, Dominican Republic.

Suborychodes Kleine, 1917

Suborychodes Kleine, 1917f: 328.

Subdorychodes: Calabresi, 1921a: 9 [lapsus].

Surboychodes: Kleine, 1921b: 73 [lapsus].

Type species: *Suborychodes intermedius* Kleine, 1917.

Distribution. Borneo, India (including Assam), Indonesia (Sumatra), Malaysia, Vietnam.

delectabilis Kleine, 1925

delectabilis Kleine, 1925b: 150.

Type depository. NHM (HT!).

Distribution. India.

intermedius Kleine, 1917

intermedius Kleine, 1917f: 330.

Type depositories. DEI (LT!), MZPW (PLT).

Distribution. Borneo, India (including Assam), Indonesia (Sumatra), Malaysia, Vietnam.

Notes. Lectotype of *S. intermedius* (DEI) designated on a male specimen with the following labels: Sumatra/ syntypus/ coll. Kraatz/ Kleine det./ *Suborychodes intermedius* Kleine.

Synorychodes Kleine, 1921

Synorychodes Kleine, 1921n: 9.

Syormychodes: Kleine, 1925b: 46 [lapsus].

Type species: *Synorychodes opacus* Kleine, 1921.

Distribution. Borneo, India, Sri Lanka, Vietnam.

***formidabilis* Kleine, 1923**

formidabilis Kleine, 1923e: 163.

Type depository. SMTD (?) (HT).

Distribution. Borneo.

Notes. Kleine (1923e: 164) reported the type for this species to be in SMTD, but according to O. Jaeger (SMTD) (2002, personal communication) it is missing from the collections.

***glaber* Kleine, 1937**

glaber Kleine, 1937i: 204.

Type depositories. NHM (LT!, PLT!), DEI (PLT!).

Distribution. India, Vietnam.

Notes. Lectotype of *S. glaber* (NHM) designated on a male specimen with the following labels: syntype/ Sukna, Kurseong, Bengal, C. F. C. Beeson, 15.V.1936/ Brit. Mus. 1946-11/ ex unknown wood/ R. R. D. 899 B. C. R. 23 cage 813/ *Synorychodes glaber* Kln., R. Kleine det. 1936. Paralectotypes: 3 female specimens (NHM), with the same label data as the lectotype; 1 female specimen (DEI), Bengalen, Kurseong, R. Kleine det. 1936/ *Synorychodes glaber* Kln.

***opacus* Kleine, 1921**

opacus Kleine, 1921n: 10.

Type depository. MZHF (HT).

Distribution. Sri Lanka.

***Teraticorhynchus* Kleine, 1925**

Teraticorhynchus Kleine, 1925b: 155.

Teraticorrhynchus: Kleine, 1938d: 118 [lapsus].

Type species: *Teraticorhynchus defectus* Kleine, 1925.

Distribution. Malaysia, Myanmar [= Burma].

defectus Kleine, 1925

defectus Kleine, 1925b: 157.

Type depository. NHM (HT!).

Distribution. Myanmar [= Burma].

serratus Kleine, 1936

serratus Kleine, 1936b: 122.

Type depository. NHM (HT!).

Distribution. Malaysia.

Tmetogonus Senna, 1895

Tmetogonus Senna, 1895b: 224.

Type species: *Tmetogonus chiriquensis* Senna, 1895.

Distribution. Panama.

chiriquensis Senna, 1895

chiriquensis Senna, 1895b: 225.

Type depository. MZUF (HT!).

Distribution. Panama.

***Toxobrentus* Damoiseau, 1965**

Toxobrentus Damoiseau, 1965b: 1.

Type species: *Toxobrentus expalliatu*s Damoiseau, 1965.

Distribution. Laos.

***expalliatu*s Damoiseau, 1965**

*expalliatu*s Damoiseau, 1965b: 2.

Type depository. IRSNB (HT!).

Distribution. Laos.

***Ubaniopsis* Soares, 1970**

Ubaniopsis Soares, 1970b: 205.

Type species: *Ubaniopsis magna* Soares, 1970.

Distribution. Bolivia, Brazil.

***confirmata* Soares, 1970**

confirmatus Soares, 1970b: 208.

Type depository. APC (HT).

Distribution. Bolivia.

***magna* Soares, 1970**

magnus Soares, 1970b: 206.

Type depository. MNRJ (Campos Seabra coll.) (HT, AT, PT).

Distribution. Brazil.

Ubanus Senna, 1895

Ubanus Senna, 1895b: 221.

Psalobrenthus Sharp, 1895a: 41; syn. Sharp, 1895b: 165.

Type species: *Ubanus aeneus* Senna, 1895.

Distribution. Belize, Costa Rica, Mexico, Panama; Brazil, Peru.

aeneus Senna, 1895

aeneus Senna, 1895b: 222 [partim: ♂].

solitarius Sharp, 1895a: 41 (*Psalobrenthus*); syn. Sharp, 1895b: 165.

Type depositories. *U. aeneus*: IRSNB (LT!), MZUF (PLT!), NHM (ex coll. Rothschild, Tring) (PLT!); *P. solitarius*: NHM (HT!).

Distribution. Costa Rica, Mexico, Panama.

Notes. According to Sharp (1895d: 164) only the male syntype specimen of *U. aeneus* (IRSNB) belongs to this species, whilst the two female paralectotypes (NHM and MZUF) are *Ubanus potens* Sharp, 1895.

Lectotype of *U. aeneus* (IRSNB) designated on a male specimen with the following labels: ♂ / coll. Roelofs / *Episphales* ? S. G. ? *aeneus* Pow. ♂ / *Ubanus* n. g. (Arrhenodinae) *aeneus* Senna typus!! = *Psalobrenthus* n. *solitarius* n. Sharp in Biolog. Centr. Amer. [handwritten by Senna] / Type / Prépar. génit. No. 768 R. Damoiseau. Paralectotypes: 1 female specimen (NHM), Mexico; 1 female specimen (MZUF), Chiriqui.

perlaetus (Gyllenhal, 1840)

perlaetus Gyllenhal (in Schoenherr, 1840: 468) (*Arrhenodes*).

Type depository. NHRS (HT).

Distribution. Brazil, Peru.

potens Sharp, 1895

potens Sharp, 1895b: 164.

aeneus Senna, 1895b: 222 [partim: ♀] (*Ubanus*); syn. Sharp, 1895b: 165.

minor Sharp, 1895b: 165 (*Ubanus potens* var.); syn. Sharp, 1895b: 165.

Type depositories. *U. potens*: NHM (HT!); *U. potens* var. *minor*: NHM (HT!).

Distribution. Belize, Mexico, Panama.

Notes. Regarding the status of the *U. aeneus* syntypes, see the note on this species.

EREMOXENINI Semenov-Tian-Shanskij, 1892

Eremoxenidae Semenov-Tian-Shanskij, 1892: 440 [see Alonso-Zarazaga & Lyal, 1999: 49].

Amorphocephalides Power, 1879a: 478.

Amorphocephalidae Gozis, 1882: 58.

Amorphocephalini Kolbe, 1883b: 237.

Amorphocephalinae Senna, 1892c: 46.

Amorphocephalina Sharp, 1895a: 32.

Eremoxenini Schönfeldt, 1910a: 46.

Paussobrenthini Gestro, 1919: 270.

Acramorphocephalus Kleine, 1918

Acramorphocephalus Kleine, 1918b: 118.

Type species: *Acramorphocephalus gebieni* Kleine, 1918.

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea (Bioko Is. = [Fernando Poo]), Gabon, Ivory Coast, Kenya, Malawi, Republic of the Congo, Tanzania, Zambia.

costipennis Calabresi, 1920

costipennis Calabresi, 1920c: 66.

Type depository. MZUF (LT!, PLT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Gabon, Ivory Coast, Republic of the Congo.

Notes. Lectotype of *A. costipennis* (MZUF) designated on a male speci-

men with the following labels: Stat. J. Albrechts Höhe, N. Kamerun, L. Conradt d. Kraatz/ *Acramorphocephalus costipennis* ♂ typus! n. sp./ Bull. Soc. Ent. It., LII, 1920, typus! ♂ 1898/ ex-coll. Senna/ "La Specola" Firenze, 6981. Paralectotype: 1 female specimen (MZUF), Bénito, Congo Franc./ typus ♀/ *Acramorphocephalus costipennis* Calabresi sp. n./ ex-coll. Senna/ "La Specola" Firenze, 6982). According to De Muizon (1955b: 890) this female paralectotype belongs to a different species.

gebieni Kleine, 1918

gebieni Kleine, 1918b: 120.

robustus Kleine, 1922h: 141 (*Acramorphocephalus*); syn. Damoiseau, 1967a: 413.

Type depositories. *A. gebieni*: MNHUB (?) (NT); *A. robustus*: MNHUB (?) (HT).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea (Bioko Is. = [Fernando Poo]), Ivory Coast, Tanzania.

Notes. Damoiseau (1967a: 416) designated the neotype for *A. gebieni* on the holotype specimen of *A. robustus* in MNHUB, but we did not find it there.

schoutedeni Kleine, 1918

schoutedeni Kleine, 1918b: 122.

schoutodeni Kleine, 1924d: 213 (*Acramorphocephalus*) [lapsus].

Type depository. MRAC (HT!).

Distribution. Angola, Democratic Republic of the Congo (ex Zaire).

stabilis Kleine, 1918

stabilis Kleine, 1918b: 122.

Type depositories. MNHUB (?) (LT, PLT), DEI (PLT!), SMTD (PLT), MZPW (PLT).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea, Gabon, Kenya, Malawi, Republic of the Congo, Tanzania, Zambia.

Notes. Lectotype and paralectotypes were designated by Damoiseau (1967a: 419) on the specimens held in MNHUB, DEI, and SMTD. We found neither lectotype nor allolectotype in MNHUB.

Afrocordus Damoiseau, 1980

Afrocordus Damoiseau, 1980: 30.

Type species: *Cordus vicarius* Kleine, 1922.

Distribution. Angola, Benin, Cameroon, Democratic Republic of the Congo (ex Zaire), Gabon, Ivory Coast, Malawi, Republic of the Congo, Republic of Guinea, Senegal, Sierra Leone, Tanzania.

asper (Calabresi, 1920)

asper Calabresi, 1920d: 30 (*Cordus*).

Type depository. MSNG (LT!, PLT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Gabon, Republic of the Congo.

Notes. Damoiseau (1967a: 396) reported that the specimens held in MSNG are holotype and allotype, but in her original description Calabresi did not designate either of the syntypes as holotype; however, as there is only one male syntype in MSNG, this must be considered as lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999). The lectotype has the following labels: Congo francese, Nkogo', XII.1902, L. Feal *Cordus asper* n.sp., ♂ ♀ Typi!

vicarius (Kleine, 1922)

vicarius Kleine, 1922e: 154 (*Cordus*).

gigas De Muizon, 1955b: 886 (*Cordus*); syn. Damoiseau, 1967a: 396.

Type depositories. *A. vicarius*: MNHUB (?) (HT, AT); *C. gigas*: NHM (HT!).

Distribution. Angola, Benin, Democratic Republic of the Congo (ex Zaire), Gabon, Ivory Coast, Malawi, Republic of the Congo, Republic of Guinea, Senegal, Sierra Leone, Tanzania.

Notes. According to Damoiseau (1967a: 397) the types for *A. vicarius* are in MNHUB, but we did not find them there.

***Amorphocephala* Damoiseau, 1966**

Amorphocephala Damoiseau, 1966f: 307 [replacement name for *Amorphocephalus*].

Amorphocephalus Schoenherr, 1840: 485 (non Bowdich, 1825).

Type species: *Brentus coronatus* Germar, 1817.

Distribution. Albania, Algeria, Bosnia Hercegovina, Bulgaria, Croatia, Cyprus, Egypt, France, Greece, Italy, Libya, Macedonia, Morocco (including Spanish possessions), Portugal, Spain, Turkey, Yugoslavia (Montenegro); Iran, Israel, Jordan, Saudi Arabia, Syria, Yemen; Afrotropical Region; India, Pakistan.

***aethiopica* Bartolozzi, 1991**

aethiopica Bartolozzi, 1991: 20.

Type depository. MRSN (HT).

Distribution. Eritrea

***angusta* Damoiseau, 1967**

angusta Damoiseau, 1967a: 425.

Type depository. TMSA (HT!).

Distribution. South Africa.

***arabica* (Damoiseau, 1964)**

arabicus Damoiseau, 1964b: 428 (*Amorphocephalus*).

Type depository. HNHM (HT!).

Distribution. Saudi Arabia.

Notes. According to W. Schedl (1975a: 33) the species is a synonym of *A. sulcata* Calabresi, but Damoiseau (1979b: 10, footnote) states that the synonymy is possible but not certain.

***basilewskyi* Damoiseau, 1967**

basilewskyi Damoiseau, 1967a: 434.

Type depositories. MRAC (HT!, AT!), IRSNB (PT!).

Distribution. Tanzania.

***calcarata* Damoiseau, 1967**

calcarata Damoiseau, 1967a: 432.

Type depositories. TMSA (HT, AT), IRSNB (PT!).

Distribution. Ethiopia, Kenya, Somalia, Tanzania.

***capensis* Damoiseau, 1979**

capensis Damoiseau, 1979b: 5.

Type depositories. MRAC (HT!, AT!, PT!), IRSNB (PT!).

Distribution. South Africa.

***coronata* (Germar, 1817)**

coronatus Germar, 1817: 247 (*Brentus*).

italicus Guérin-Ménéville, 1833: pl. 36, figs. 7, 7a, 7b (*Brentus*); syn. Guérin-Ménéville, 1844: 139.

Type depository. *B. coronatus*: MLUH (HT).

Distribution. Albania, Algeria, Bosnia Hercegovina, Bulgaria, Croatia, Cyprus, France, Greece, Italy (including Sardinia and Sicily), Macedonia, Morocco, Portugal, Spain, Turkey, Yugoslavia (Montenegro); Iran, Israel, Syria.

Notes. Guérin-Méneville illustrated this species in 1833 (pl. 36, figs 7, 7a, 7b) under the name of *Brentus italicus* Latr. In the text explaining the plate, published in 1844 (p. 139), he wrote that the correct name should be *Brentus coronatus*, as stated by Schoenherr (1833: 331). We consider Guérin-Méneville as the author of *B. italicus* because he illustrated a specimen (♀), whilst Latreille (1828: 593) used the name but did not supply any description.

According to Kleine (1918b: 92) localities also include Russia and the Caucasus.

dahomeensis (Senna, 1894)

dahomeensis Senna, 1894a: 407 (*Amorphocephalus*).

intermedius Kleine, 1918b: 94 (*Amorphocephalus*); syn. De Muizon, 1960: 167.

parasitus Kleine, 1938d: 86 (*Amorphocephalus*); syn. Damoiseau, 1967a: 428.

Type depositories. *A. dahomeensis*: unknown; *A. intermedius*: MZPW (ST), MNHUB (?) (ST), ZSM (ST); *A. parasitus*: MRAC (HT!).

Distribution. Central African Republic, Democratic Republic of the Congo (ex Zaire), Ethiopia, Ghana, Nigeria, Republic of Guinea, Senegal, Somalia, Sudan.

Notes. According to Kleine (1928a: 80) the type for *A. dahomeensis* was in the Alluaud collection, but we did not find it in MNHN. Damoiseau (1967a: 431) erroneously wrote that the holotype is in MZUF, but this is not so.

According to Kleine (1928a: 81) syntypes for *A. intermedius* are in MNHUB, but we did not find them there.

Damoiseau (1967a: 431) did not mention Ethiopia (Kleine, 1937: 580) and Somalia (Kleine, 1918b: 96) when he gave the species distribution.

delicata (Kleine, 1925)

delicatus Kleine, 1925b: 144 (*Amorphocephalus*).

delicatulus: Kleine, 1926h: 24 (*Amorphocephalus*) [lapsus].

deltcatus: Kleine, 1938a: 16 (*Amorphocephalus*) [lapsus].

Type depository. NHM (HT!).

Distribution. Pakistan.

deserticola (Kleine, 1944)

deserticolus Kleine, 1944: 153 (*Amorphocephalus*).

Type depository. MNHN (?) (HT).

Distribution. Morocco (including Spanish possessions).

Notes. According to Kleine (1944: 153) the holotype is in MNHN, but neither Damoiseau (1967a: 437) nor ourselves found it there.

diademata (Power, 1879)

diadematus Power, 1879a: 486 (*Amorphocephalus*).

Type depository. MNHN (HT!).

Distribution. Democratic Republic of the Congo (ex Zaire), Ethiopia, Guinea Bissau, Kenya, Senegal, Somalia, Sudan, Tanzania, Togo.

Notes. Damoiseau (1967a: 431) did not mention Senegal (Power, 1879a: 486), Togo (Kleine, 1918b: 108) and the Democratic Republic of the Congo (ex Zaire) (Kleine, 1936a: 32) when he gave the species distribution.

excantator (Damoiseau, 1964)

excantator Damoiseau, 1964b: 425 (*Amorphocephalus*).

Type depository. HNHM (HT, AT!).

Distribution. Jordan.

imitator (Fåhræus, 1871)

imitator Fåhræus, 1871: 434 (*Amorphocephalus*).

imitador: Schaufuss, 1876: 402 (*Amorphocephalus*) [lapsus].

hospes Kolbe, 1885: 188 (*Amorphocephalus*); syn. Damoiseau, 1979b: 4.

Type depositories. *A. imitator*: NHRS (ST); *A. hospes*: MNHUB (HT!).

Distribution. Tropical Africa; Italy (I).

***jickelii* (Schaufuss, 1876)**

jickelii Schaufuss, 1876: 402 (*Amorphocephalus*).

jickeli: Schönfeldt, 1908: 29 (*Amorphocephalus*) [lapsus].

jikelii: Kleine, 1938d: 85 (*Amorphocephalus*) [lapsus].

jikeli: Kleine, 1938d: 188 (*Amorphocephalus*) [lapsus].

Type depository. Unknown.

Distribution. Egypt; Sudan.

Notes. Damoiseau (1979b: 11) listed this species in the genus *Amorphocephala*, adding a question mark because he was unable to study the type specimen. We did not find the type in MNHUB, where Schaufuss types are held.

***mandibularis* Damoiseau, 1967**

mandibularis Damoiseau, 1967a: 436.

princeps De Muizon, 1960: 168 (non Kleine, 1918) (*Amorphocephalus*); syn. Damoiseau, 1967a: 436.

Type depository. *A. mandibularis*: MNHN (?) (HT, AT).

Distribution. Ethiopia, Eritrea, Kenya, Niger, Tanzania.

Notes. According to Damoiseau (1967a: 436) the types of *A. mandibularis* are in MNHN, but we did not find them there.

***poweri* (De Muizon, 1960)**

poweri De Muizon, 1960: 161 (*Amorphocephalus*).

diadematus Kleine, 1918b: 106 (non Power, 1879) (*Amorphocephalus*); syn. De Muizon, 1960: 161.

Type depository. MNHN (HT!, AT!).

Distribution. Benin, Central African Republic, Democratic Republic of the Congo (ex Zaire), Ghana, Guinea Bissau, Ivory Coast, Nigeria, Republic of Guinea, Senegal, Sierra Leone, Sudan, Togo.

pubescens (Damoiseau, 1961)

pubescens Damoiseau, 1961a: 286 (*Amorphocephalus*).

Type depository. MRAC (HT!).

Distribution. Democratic Republic of the Congo (ex Zaire).

scotti (De Muizon, 1954)

scotti De Muizon, 1954: 325 (*Amorphocephalus*).

Type depository. NHM (HT!, AT!).

Distribution. Yemen.

senegalensis (Power, 1879)

senegalensis Power, 1879a: 486 (*Amorphocephalus*).

princeps Kleine, 1918b: 101 (*Amorphocephalus*); syn. Damoiseau, 1967a: 435.

Type depositories. *A. senegalensis*: IRSNB (LT!), MNHN (PLT!); *A. princeps*: MNHUB (?) (HT), NMW (Werner coll.) (AT).

Distribution. Algeria, Egypt; Cameroon, Chad, Ethiopia, Nigeria, Senegal, Sudan.

Notes. Lectotype and paralectotype for *A. senegalensis* designated by Damoiseau (1967a: 435).

According to Kleine (1928a: 81) the holotype of *A. princeps* is in MNHUB, but we did not find it there.

Damoiseau (1967a: 435) did not mention Cameroon (Kleine, 1918b: 99), Algeria and Egypt (Roudier, 1954: 135) when he gave the species distribution.

sulcata (Calabresi, 1920)

sulcatus Calabresi, 1920c: 64 (*Amorphocephalus*).

Type depositories. MSNG (LT!), MZUF (PLT!).

Distribution. Egypt, Libya; Eritrea, Ethiopia, Niger, Sudan; Saudi Arabia.

Notes. Damoiseau (1967a: 428) reported that the specimen held in MSNG is the holotype, and that the allotype is held in MZUF, but in her original description Calabresi did not designate either of the syntypes as holotype; however, as there is only one male syntype in MSNG, this must be considered as lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999). The lectotype has the following labels: Agordat 1896, F. Derchi, *Amorphocephalus sulcatus* n. sp. ♂ typus!

Ankleineella Zimmerman, 1994

Ankleineella Zimmerman, 1994: 43.

Type species: *Amorphocephalus sulcicollis* Pascoe, 1872.

Distribution. Indonesia (Moluccas: Halmahera, Waigeo); Australia, New Guinea, Solomon Is.

australis (Lacordaire, 1866)

australis Lacordaire, 1866: 423 (*Amorphocephalus*).

Type depository. IRSNB (NT!).

Distribution. Australia.

Notes. Neotype designated by Damoiseau (1979b: 25).

mniszechi (Power, 1879)

mniszechi Power, 1879a: 487 (*Amorphocephalus*).

Type depository. MNHN (HT!).

Distribution. Australia.

subscripta (Damoiseau, 1979)

subscripta Damoiseau, 1979b: 25 (*Kleineella*).

Type depositories. SAMA (Lea coll.) (HT, AT, PT), IRSNB (PT!), QMA (PT).

Distribution. Australia.

sulcicollis (Pascoe, 1872)

sulcicollis Pascoe, 1872: 321 (*Amorphocephalus*).

Type depository. NHM (HT!).

Distribution. Australia.

Cobalocephalus Morimoto, 1982

Cobalocephalus Morimoto, 1982: 33.

Type species: *Amorphocephalus gyotokui* Nakane, 1963.

Distribution. Japan.

gyotokui (Nakane, 1963)

gyotokui Nakane, 1963: 35 (*Amorphocephalus*).

Type depository. SEHU (HT, AT).

Distribution. Japan.

Cordus Schoenherr, 1847

Cordus Schoenherr, 1847: 10.

Type species: *Cordus hospes* Schoenherr, 1847.

Distribution. Australia, New Guinea, Solomon Is.

acutipennis Power, 1879

acutipennis Power, 1879a: 483.

Type depository. MNHN (LT!, PLT!).

Distribution. Australia.

Notes. Lectotype designated by Damoiseau (1840: 14).

archboldi Damoiseau, 1980

archboldi Damoiseau, 1980: 10.

Type depository. NMNH (HT, AT).

Distribution. New Guinea.

armaticeps Senna, 1894

armaticeps Senna, 1894e: 559.

Type depositories. MSNG, (LT!, PLT!), MZUF (PLT!).

Distribution. New Guinea.

Notes. Lectotype designated by Damoiseau (1980: 7).

cheesmanae (Kleine, 1939)

cheesmanae Kleine, 1939c: 108 (*Kleineella*).

Type depository. NHM (HT!).

Distribution. New Guinea.

demarzi Damoiseau, 1980

demarzi Damoiseau, 1980: 18.

Type depositories. NHMB (HT!), SAMA (AT, PT), IRSNB (PT!).

Distribution. Australia.

***elytrostriatus* (Kleine, 1939)**

elytrostriatus Kleine, 1939c: 108 (*Kleineella*).

Type depository. NHM (HT!).

Distribution. New Guinea.

***festus* Damoiseau, 1980**

festus Damoiseau, 1980: 11.

Type depository. SAMA (HT).

Distribution. Australia.

***firmus* Damoiseau, 1980**

firmus Damoiseau, 1980: 16.

Type depositories. NMNH (HT), SAMA (AT), IRSNB (PT!), NHMB (PT!).

Distribution. Australia.

***ganglbaueri* Senna, 1919**

ganglbaueri Senna, 1919: 81.

Type depository. NMW (HT).

Distribution. Australia.

***hospes* Schoenherr, 1847**

hospes Schoenherr, 1847: 11.

Type depository. MLUH (HT).

Distribution. Australia (including Tasmania).

laevis Damoiseau, 1980

laevis Damoiseau, 1980: 12.

Type depositories. SAMA (HT, AT, PT), IRSNB (PT!).

Distribution. Australia.

occidentalis Damoiseau, 1980

occidentalis Damoiseau, 1980: 21.

Type depositories. NHMB (HT!), IRSNB (PT!).

Distribution. Australia.

pascoei Power, 1879

pascoei Power, 1879a: 484.

Type depository. MNHN (HT!).

Distribution. Australia.

plumipennis Damoiseau, 1980

plumipennis Damoiseau, 1980: 15.

Type depositories. SAMA (Lea coll.) (HT), IRSNB (AT!).

Distribution. Australia.

queenslandicus Senna, 1919

queenslandicus Senna, 1919: 80.

Type depository. MZUF (HT!).

Distribution. Australia.

recticornis Damoiseau, 1980

recticornis Damoiseau, 1980: 20.

Type depositories. SAMA (HT, AT), IRSNB (PT!).

Distribution. Australia.

rostralis Damoiseau, 1980

rostralis Damoiseau, 1980: 24.

Type depositories. NHMB (HT!, AT!, PT!), IRSNB (PT!).

Distribution. Australia.

salomonensis Damoiseau, 1980

salomonensis Damoiseau, 1980: 9.

Type depository. NHM (HT!).

Distribution. Solomon Is.

schoenherri Power, 1879

schoenherri Power, 1879a: 483.

Type depositories. IRSNB (LT!, PLT!), MNHN (PLT!), MCZ (PLT), NHRS (PLT).

Distribution. Australia.

Notes. Lectotype designated by Damoiseau (1980: 22).

ultimus Damoiseau, 1980

ultimus Damoiseau, 1980: 23.

Type depository. UQDZ (HT).

Distribution. Australia.

Notes. Damoiseau (1980: 23) wrote that the holotype is in IRSNB, but according to M. Cludts (IRSNB) (2002, personal communication) it is actually held in the UQDZ collections.

***vermiculatus* Damoiseau, 1980**

vermiculatus Damoiseau, 1980: 19.

Type depositories. NHMB (HT!, AT!), IRSNB (PT!), NMNH (PT), SAMA (PT).

Distribution. Australia.

***vicinus* Senna, 1919**

vicinus Senna, 1919: 82.

Type depository. MZUF (HT!).

Distribution. Australia.

Notes. Zimmerman (1994: 109) states that the holotype is not in MZUF. This is incorrect, the holotype is held there and is also listed in Bartolozzi *et al.* (1984: 102).

***Eremoxenus* Semenov-Tian-Schanskij, 1892**

Eremoxenus Semenov-Tian-Schanskij, 1892: 438.

Acramorphocephaloides Bartolozzi, 1988b: 155; syn. Bartolozzi & Sforzi (in Alonso-Zarazaga & Lyal, 1999: 49).

Type species: *Eremoxenus chan* Semenov-Tian-Schanskij, 1892.

Distribution. Algeria; Iran, Kazakhstan, Turkmenistan, Uzbekistan.

***algirus* (Bartolozzi, 1988)**

algirus Bartolozzi, 1988b: 157 (*Acramorphocephaloides*).

Type depository. ZMUC (HT!).

Distribution. Algeria.

chan Semenov-Tian-Schanskij, 1892

chan Semenov-Tian-Schanskij, 1892: 440.

Type depository. ZIN (ST).

Distribution. Iran, Kazakhstan, Turkmenistan, Uzbekistan.

Hadramorphocephalus Kleine, 1918

Hadramorphocephalus Kleine, 1918b: 114.

Type species: *Amorphocephalus calvei* Power, 1879.

Distribution. Chad, Mali, Senegal.

calvei (Power, 1879)

calvei Power, 1879a: 485 (*Amorphocephalus*).

calwei: Kleine, 1938a: 14 (*Hadramorphocephalus*) [lapsus].

Type depositories. MNHN (LT!, PLT!), IRSNB (PLT!).

Distribution. Chad, Mali, Senegal.

Notes. Damoiseau (1967a: 412) reported that the specimens held in IRSNB are holotype and allotype, but in his original description Power did not designate any of the syntypes as holotype; however, as there is more than one syntype in MNHN, and as Damoiseau did not specifically select one as holotype, we designate the lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Lectotype of *A. calvei* (MNHN) designated on a specimen with the following labels: Senegal/ Thomson 137/ type/ ex Musaeo G. Power/ Museum Paris 1952, coll. R. Oberthür. Paralectotypes: 1 specimen (MNHN), Senegal/ Chevrolat 62/ ex Musaeo G. Power/ Museum Paris 1952, coll. R. Oberthür; 1 specimen (MNHN), Senegal/ Thomson 136/ ex Musaeo G. Power/ Museum Paris, 1952 coll. R. Oberthür; 1 specimen (IRSNB), Sénégal/ Coll. Dejean, Coll. Roelofs/ M. R. Belg./ Type/ *Calvei* Petit/ *Arrhenodes Calvei* Petit, h. in Senegal, D. Petit/ *Amorphocephalus Calvei* Pow./ *Hadramorphocephalus calvei* Kleine, R. Damoiseau det.; 1 specimen (IRSNB), Sénégal/ Coll. Dejean, Coll. Roelofs/ M. R. Belg./ *Hanetii* Petit/ Paratype/ *Arrhenodes Hanetii*

Petit, h. in Senegal, D. Petit/ *Hadramorphocephalus calvei* Kleine, R. Damoiseau det.; 1 specimen (IRSNB), Sénégal/ Coll. Dejean, Coll. Roelofs/ M. R. Belg./ *Amorphocephalus Calvei* Pow. ♂/ Allotype/ *Hadramorphocephalus calvei* Kleine, R. Damoiseau det.

Hemicordus Kleine, 1922

Hemicordus Kleine, 1922r: 159.

Cordomorphus Damoiseau, 1971b: 271; syn. Damoiseau, 1980: 26.

Type species: *Hemicordus minax* Kleine, 1922 = *Cordus peguanus* Senna, 1892.

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea, Gabon, Ghana, Guinea Bissau, Ivory Coast, Republic of the Congo, Sierra Leone, South Africa, Togo; Borneo, Indonesia (Sumatra), Malaysia, Myanmar [= Burma], Philippines (Basilan, Negros), Sri Lanka.

bellicosus (Kleine, 1926)

bellicosus Kleine, 1926f: 366 (*Cordus*).

bellicosus: Kleine, 1938a: 11 (*Cordus*) [lapsus].

Type depository. MNHUB (?) (HT).

Distribution. Cameroon, Republic of the Congo.

Notes. According to Kleine (1928a: 79) the holotype is in MNHUB, but neither Damoiseau (1967a: 399) nor ourselves found it there.

guineensis (Calabresi, 1920)

guineensis Calabresi, 1920d: 29 (*Cordus*).

Type depository. MSNG (HT!).

Distribution. Guinea Bissau.

***kraatzii* (Senna, 1898)**

kraatzii Senna, 1898c: 375 (*Cordus*).

myrmecophilus Kleine, 1921i: 111 (*Cordus*); syn. Damoiseau 1967a: 392.

bellatorius Kleine, 1924a: 105 (*Cordus*); syn. Damoiseau 1967a: 392.

Type depositories. *C. kraatzii*: MZUF (HT!); *C. myrmecophilus*: MWIC (HT); *C. bellatorius*: MRAC (HT!), MNHUB (?) (AT).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Gabon, Ghana, Ivory Coast, Republic of the Congo.

Notes. According to Kleine (1924a: 106) the allotype of *C. bellatorius* is in MNHUB, but we did not find it there.

***ivorensis* Damoiseau, 1980**

ivorensis Damoiseau, 1980: 28.

Type depository. MRAC (HT!).

Distribution. Cameroon, Ivory Coast, Sierra Leone, Togo.

***operosus* (Damoiseau, 1971)**

operosus Damoiseau, 1971b: 271 (*Cordomorphus*).

Type depository. MZLU (HT, AT).

Distribution. Sri Lanka.

***peguanus* (Senna, 1892)**

peguanus Senna, 1892e: 463 (*Cordus*).

minax Kleine, 1922r: 160 (*Hemicordus*); syn. Damoiseau, 1980: 26.

Type depositories. *C. peguanus*: MSNG (HT!); *H. minax*: SMTD (HT).

Distribution. Borneo, Indonesia (Sumatra), Malaysia, Myanmar [= Burma], Philippines (Basilan, Negros).

plagiator (Kleine, 1922)

plagiator Kleine, 1922h: 142 (*Cordus*).

Type depository. MNHUB (?) (HT).

Distribution. Democratic Republic of the Congo (ex Zaire), Equatorial Guinea, Republic of the Congo.

Notes. According to Damoiseau (1967a: 399) the holotype is in MNHUB, but we did not find it there.

puncticollis (Power, 1878)

puncticollis Power, 1878a: 484 (*Cordus*).

Type depository. MNHN (HT!).

Distribution. South Africa.

sculpturatus (Senna, 1903)

sculpturatus Senna, 1903: 173 (*Cordus*).

Type depository. MSNG (HT!).

Distribution. Indonesia (Sumatra).

Kleinella Kleine, 1917

Kleinella Kleine, 1917h: 276.

Mastax Kleine, 1918c: 163 (non Fischer de Waldheim, 1828, nec Perty, 1832).

Kleineella Strand, 1918: 167 [replacement name for *Mastax*].

Kleineela: Zimmerman, 1994: 115 [lapsus].

Type species: *Kleinella compressicornis* Kleine, 1917.

Distribution. Indonesia (Moluccas: Halmahera, Waigeo); Australia, New Guinea, Polynesia (Tahiti) (I?), Solomon Is.

Notes. Kleine (1917h: 276) described the new species *Kleinella compressicornis*, specifying that the genus *Kleinella* Strand was in publication. Actu-

ally, the genus of Strand was not published until 1918 under the name *Kleineella*.

***barbata* (Kleine, 1918)**

barbatus Kleine, 1918c: 164 (*Mastax*).

piceonitens Kleine, 1918b: 150 (*Kleineella*); syn. Zimmerman, 1994: 115.

piceoniteus: Kleine, 1924d: 210 (*Kleineella*) [lapsus].

Type depositories. *M. barbata*: SMTD (HT); *K. piceonitens*: MNHUB (?) (HT).

Distribution. Australia, Polynesia (Tahiti) (I?).

Notes. According to Kleine (1918b: 152) the holotype of *K. piceonitens* was held in MNHUB; Zimmerman (1994: 115) was unable to find it in MNHUB (we did not find it either). Zimmerman also considers that the holotype was either a mislabelled specimen or a specimen that had been introduced to Tahiti through trade with Australia.

***bougainvillei* Damoiseau, 1966**

bougainvillei Damoiseau, 1966c: 150.

Type depository. MZPW (HT).

Distribution. Solomon Is.

***compressicornis* Kleine, 1917**

compressicornis Kleine, 1917h: 276.

Type depository. RMNH (HT).

Distribution. Indonesia (Moluccas: Waigeo).

Notes. Kleine (1917h: 276) described this species under the genus *Kleinella* [sic!] even though the description of the new genus *Kleineella* by Strand was only still in press. Thus *K. compressicornis* is the type species of the genus for monotypy and combined description.

fraterna Kleine, 1924*fraterna* Kleine, 1924b: 33.**Type depository.** MNHUB (HT!).**Distribution.** New Guinea.*moluccana* Kleine, 1944*moluccana* Kleine, 1944: 153.**Type depository.** MNHN (HT!).**Distribution.** Indonesia (Moluccas: Halmahera).*novaeguineae* (Senna, 1894)*novaeguineae* Senna, 1894e: 560 (*Amorphocephalus*).**Type depository.** MSNG (HT!).**Distribution.** New Guinea.*Leptamorphocephalus* Kleine, 1918*Leptamorphocephalus* Kleine, 1918b: 132.*Leptamorphocephalus*: Kleine, 1927a: 33 [lapsus].**Type species:** *Leptamorphocephalus laborator* Kleine, 1918.**Distribution.** Taiwan; Borneo, India (including Andaman Is., Assam, Darjeeling), Indonesia (Java, Nias, Sumatra), Laos, Malaysia, Myanmar [= Burma], Philippines (Negros, Palawan), Singapore, Thailand, Vietnam.*acutipennis* Damoiseau, 1979*acutipennis* Damoiseau, 1979b: 15.**Type depositories.** IRSNB (HT!), NHM (AT!).**Distribution.** India.

cuneatus Kleine, 1925*cuneatus* Kleine, 1925b: 142.**Type depository.** NHM (HT!).**Distribution.** India (Assam).*dissentaneus* Kleine, 1922*dissentaneus* Kleine, 1922r: 160.**Type depository.** SMTD (HT).**Distribution.** Borneo.*foederatus* Kleine, 1923*foederatus* Kleine, 1923e: 163.**Type depository.** SMTD (HT).**Distribution.** Indonesia (Sumatra), Malaysia, Philippines (Negros).*laborator* Kleine, 1918*laborator* Kleine, 1918b: 133.*labrator*: Kano, 1930: 153 (*Leptamorphocephalus*) [lapsus].**Type depository.** DEI (HT!).**Distribution.** Taiwan; Indonesia (Sumatra), Malaysia, Singapore, Vietnam.*laevis* (Power, 1879)*laevis* Power, 1879a: 486 (*Amorphocephalus*).*cupidus* Kleine, 1925b: 143 (*Leptamorphocephalus*); syn. Damoiseau, 1979b: 12.**Type depositories.** *A. laevis*: IRSNB (LT!), MNHN (PLT!); *L. cupidus*: NHM (HT!).

Distribution. India (including Assam, Darjeeling), Indonesia (Java), Myanmar [= Burma].

Notes. Damoiseau (1979b: 15) reported that the specimen of *A. laevis* held in IRSNB is the holotype, but in his original description Power did not designate any of the syntypes as holotype; however, as there is only one syntype in IRSNB, this must be considered as lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

monstratus (Kleine, 1933)

monstratus Kleine, 1933e: 324 (*Paramorphocephalus*).

Type depository. NHRS (HT).

Distribution. Thailand, Vietnam.

sumatranus (Senna, 1895)

sumatranus Senna, 1895a: 195 (*Amorphocephalus*).

Type depository. RMNH (HT).

Distribution. Borneo, India (Andaman Is.), Indonesia (Java, Nias, Sumatra), Laos, Malaysia, Philippines (Palawan), Vietnam.

Micramorphocephalus Kleine, 1918

Micramorphocephalus Kleine, 1918b: 125.

Micramorcephalus: Kleine, 1924d: 224 [lapsus].

Type species: *Micramorphocephalus frater* Kleine, 1918.

Distribution. Benin, Central African Republic, Democratic Republic of the Congo (ex Zaire), Ethiopia, Ghana, Ivory Coast, Kenya, Republic of the Congo, Republic of Guinea, Senegal, Sierra Leone, South Africa, Tanzania, Togo, Zambia.

consobrinus Kleine, 1918

consobrinus Kleine, 1918b: 131.

Type depository. MRAC (HT!).

Distribution. Central African Republic, Democratic Republic of the Congo (ex Zaire), Ghana, Ivory Coast, Republic of Guinea, Republic of the Congo, Senegal, Sierra Leone, Tanzania.

frater Kleine, 1918

frater Kleine, 1918b: 128.

Type depositories. MNHUB (?) (LT), TMSA (PLT).

Distribution. Ethiopia, Kenya, South Africa, Tanzania, Zambia.

Notes. Damoiseau (1967a: 409) reported that the specimen held in MNHUB is the holotype and the one held in TMSA is the allotype, but in his original description Kleine did not designate either of the two syntypes as holotype; however, as there is only one syntype in MNHUB, this must be considered as lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999). We did not find this lectotype in MNHUB.

soror Kleine, 1918

soror Kleine, 1918b: 130.

Type depository. MNHUB (?) (LT, PLT).

Distribution. Benin, Sierra Leone, Togo.

Notes. Damoiseau (1967a: 412) reported that the specimens held in MNHUB are holotype and allotype, but in his original description Kleine did not designate either of the syntypes as holotype. However on the basis of article 74.5 of the International Commission of Zoological Nomenclature (1999) the specimen that Damoiseau gives as holotype must be considered as the lectotype. We did not find the lectotype and paralectotype in MNHUB.

Myrmecobrenthus Kleine, 1920

Myrmecobrenthus Kleine, 1920i: 25.

Myrmecobrenthus: Kleine, 1926f: 367 [lapsus].

Type species: *Myrmecobrenthus wasmanni* Kleine, 1920.

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Gabon, Ghana, Ivory Coast.

ater Damoiseau, 1961

ater Damoiseau, 1961b: 21.

Type depository. IRSNB (HT!).

Distribution. Gabon, Ivory Coast.

wasmanni Kleine, 1920

wasmanni Kleine, 1920i: 27.

Type depository. MRAC (NT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Ghana, Ivory Coast.

Paramorphocephalus Kleine, 1920

Paramorphocephalus Kleine, 1920j: 236.

Type species: *Paramorphocephalus diabolus* Kleine, 1920.

Distribution. Japan; Indonesia (Java, Mentawai, Sumatra), Malaysia, Philippines (Samar, Tawi Tawi), Vietnam.

binotatus Calabresi, 1922

binotatus Calabresi, 1922: 109.

Type depository. MNHN (LT!, PLT!).

Distribution. Vietnam.

Notes. Lectotype of *P. binotatus* (MNHN) designated on a male specimen with the following labels: Long Xuyen, Cochinchine Doit/ collection E. Fleutiaux/ *Paramorphocephalus binotatus* n. sp. ♂ ♀ Typi!/ Calabresi, Bull. Soc. ent. Ital. 1922 p. 107, collection Fleutiaux/ Museum Paris/ *Paramorphocephalus binotatus* Calabresi R. Damoiseau det., Prép. Génit. 78 R 03 R. Damoiseau. Paralectotype: 1 female specimen (MNHN), Long Xuyen, Cochinchine Doit/ collection E. Fleutiaux/ *Paramorphocephalus binotatus* Calabresi, R. Damoiseau det.

diabolus Kleine, 1920

diabolus Kleine, 1920j: 240.

Type depository. RMNH (ST).

Distribution. Indonesia (Java, Sumatra).

fumosus (Morimoto, 1976)

fumosus Morimoto, 1976a: 279 (*Leptamorphocephalus*).

Type depository. ELKU (HT, PT).

Distribution. Japan.

loricatus Kleine, 1920

loricatus Kleine, 1920j: 241.

Type depository. RMNH (HT).

Distribution. Indonesia (Sumatra).

mentaweicus (Senna, 1898)

mentaweicus Senna, 1898a: 237 (*Amorphocephalus*).

Type depository. MSNG (HT!).

Distribution. Indonesia (Mentawai).

nodosifer (Kleine, 1918)

nodosifer Kleine, 1918b: 143 (*Leptamorphocephalus*).

Type depository. MZPW (HT).

Distribution. Indonesia (Sumatra).

setosus Kleine, 1925

setosus Kleine, 1925c: 597.

Type depository. SMTD (HT).

Distribution. Philippines (Samar).

tawitawi (Damoiseau, 1966)

tawitawi Damoiseau, 1966e: 448 (*Leptamorphocephalus*).

Type depositories. ZMUC (HT), IRSNB (PT!).

Distribution. Philippines (Tawi Tawi).

variolosus (Power, 1879)

variolosus Power, 1879a: 485 (*Amorphocephalus*).

Type depository. IRSNB (HT!).

Distribution. Malaysia.

Paussobrenthus Gestro, 1919

Paussobrenthus Gestro, 1919: 271.

Paussobrenthis: Imperial Bureau of Entomology, 1924: 235 [lapsus].

Type species: *Paussobrenthus bakeri* Gestro, 1919.

Distribution. India (Andaman Is.), Kampuchea (= Cambodia), Singapore, Vietnam.

***bakeri* Gestro, 1919**

bakeri Gestro, 1919: 273.

Type depository. MSNG (HT!).

Distribution. Singapore, Vietnam.

***concitatus* Kleine, 1925**

concitatus Kleine, 1925b: 141.

Type depository. NHM (HT!).

Distribution. India (Andaman Is.).

***conterminus* Kleine, 1925**

conterminus Kleine, 1925b: 141.

Type depository. NHM (HT!).

Distribution. Kampuchea (= Cambodia), Vietnam.

Notes. Head and thorax missing in the type specimen.

***Pericordus* Kolbe, 1883**

Pericordus Kolbe, 1883b: 237.

Thoracantharus Kleine, 1926f: 366; syn. De Muizon, 1960: 136.

Type species: *Pericordus latipes* Kolbe, 1883.

Distribution. Angola, Ivory Coast, Kenya, Mozambique, Namibia, Republic of Guinea, Senegal, Somalia, South Africa, Tanzania, Uganda, Zambia.

***curiatus* (Kleine, 1926)**

curiatus Kleine, 1926f: 368 (*Thoracantharus*).

Type depository. NHM (LT!).

Distribution. Kenya, Tanzania, Uganda.

Notes. Damoiseau (1967a: 377) reported that the specimen held in NHM is the holotype, but in his original description Kleine did not designate any of the syntypes as holotype; however, as there is only one syntype in NHM, this must be considered as lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

freyi Damoiseau, 1966

freyi Damoiseau, 1966d: 16.

australis Damoiseau, 1967e: 214 (*Pericordus*); **syn. n.**

Type depositories. *P. freyi*: NHMB (HT!), IRSNB (PT!); *P. australis*: MZLU (HT!).

Distribution. Mozambique, South Africa, Zambia.

Notes. After studying the type material, we consider *P. australis* to be a new synonym of *P. freyi*.

latipes Kolbe, 1883

latipes Kolbe, 1883b: 238.

Type depository. MNHUB (HT!).

Distribution. Angola, Senegal.

occidentalis Damoiseau, 1964

occidentalis Damoiseau, 1964b: 420.

Type depositories. MRAC (HT!, AT!, PT!), IRSNB (PT!), MNHN (PT!).

Distribution. Ivory Coast, Republic of Guinea, Tanzania.

scutellaris Kleine, 1922

scutellaris Kleine, 1922e: 155.

Type depository. MNHUB (?) (HT!).

Distribution. Kenya, Namibia, Somalia, South Africa (?), Tanzania.

Notes. According to Damoiseau (1967a: 377) the holotype is in MNHUB, but we did not find it there.

Damoiseau (1967a: 377) did not mention Namibia (K. E. Schedl, 1965: 111) when he gave the species distribution.

Perisymphocerus Kleine, 1919

Perisymphocerus Kleine, 1919c: 13.

Type species: *Perisymphocerus gracilis* Kleine, 1919 = *Cordus latirostris* Power, 1879.

Distribution. Benin, Central African Republic, Eritrea, Ethiopia, Gambia, Ghana, Guinea Bissau, Ivory Coast, Kenya, Nigeria, Republic of Guinea, Senegal, Somalia, Sudan, Tanzania, Togo.

compactus Kleine, 1919

compactus Kleine, 1919c: 17.

Type depository. MNHUB (?) (LT), MSNG (?) (PLT).

Distribution. Eritrea, Kenya, Somalia, Tanzania.

Notes. Damoiseau (1967a: 404) reported that the specimen held in MNHUB is the holotype and the one held in MSNG is the allotype, but in his original description Kleine did not designate any of the syntypes as holotype; however, as there is only one syntype in MNHUB, this must be considered as lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999). We did not find the lectotype in MNHUB and the paralectotype in MSNG.

elongatus (Power, 1879)

elongatus Power, 1879a: 484 (*Cordus*).

Type depository. MNHN (HT!).

Distribution. Eritrea, Ghana, Senegal, Sudan.

Notes. There are two specimens labelled as types in MNHN (1 ♀ Senegal/ Thomson 99/ type/ ex Musaeo G. Power/ Museum Paris 1952, coll. R. Oberthür, holotype; 1 ♂/ [Sudan]: Ht. Niger Koulouba, J. Vuillet, VII.1908/ type ♂/ Museum Paris 1952, coll. R. Oberthür). In his original paper, Power only described the female from Senegal which is therefore the true holotype. De Muizon mistakenly labelled the male specimen from Sudan as type. Damoiseau (1967a: 403) reports that there are the male holotype and the female allotype in MNHN, but the male was collected 10 years after the species was first described and therefore can not be a type.

latirostris (Power, 1879)

latirostris Power, 1879a: 484 (*Cordus*).

gracilis Kleine, 1919c: 15 (*Perisymphorocerus*); syn. Calabresi, 1920d: 32.

Type depositories. *C. latirostris*: MZUF (LT!); *P. gracilis*: DEI (LT!), SMTD (?) (PLT), ZSM (Ertl coll.) (PLT).

Distribution. Benin, Central African Republic, Ethiopia, Gambia, Ghana, Guinea Bissau, Ivory Coast, Kenya, Nigeria, Republic of Guinea, Senegal, Sudan, Tanzania, Togo.

Notes. Power (1879a: 484) described *C. latirostris* on males from Senegal. The MNHN holds two specimens (1 female: Senegal/ type/ ex Musaeo G. Power/ Museum Paris 1952 coll. R. Oberthür; 1 male: Tererè Dahomey/ type ♂/ Museum Paris 1952 coll. R. Oberthür). De Muizon (1960: 142) defines the specimen from Senegal as "type unique", deducing that Power had described the female as a male. Damoiseau (1967a: 402) defines the female from Senegal as the holotype and the male from Dahomey as allotype; therefore the only specimen that definitely belongs to the type series is the one in MZUF. In his original description Power, when he mentions the rostrum, explicitly writes "*anteriori parte fere quadrata*", from which it is clear that he is describing a male because in the female the prorostrum is narrower and more elongated. The only male specimen that we could find from the type series of Power, is the one in MZUF and which we define as lectotype.

Lectotype of *C. latirostris* (MZUF) designated on a male specimen with the following labels: Bogandé, Africa Occ. franc., ex coll. Power/ type du ♂/ Bocandé/ *Cordus latirostris* ♂/ *Perisymphorocerus latirostris* (Power) ♂ typus!/ ex-coll. Senna/ "La Specola" Firenze, 12258.

Lectotype of *P. gracilis* (DEI) designated on a male specimen with the following labels: *Perisymphorocerus gracilis* Kln, det. Kleine, 1917/ Kleine det.

Damoiseau (1967a: 402) reported that syntypes of *P. gracilis* are in

SMTD, but according to O. Jaeger (SMTD) (2002, personal communication) no syntype of this species is in the SMTD collections.

Damoiseau (1967a: 402) did not mention Tanzania (Kleine, 1922e: 155) and Kenya (Kleine, 1935d: 113) when he gave the species distribution. The quotation for Equatorial Guinea (Sforzi, 1998b: 271) is incorrect.

sulcifrons Kleine, 1919

sulcifrons Kleine, 1919c: 16.

Type depository. MNHUB (?) (HT).

Distribution. Ethiopia, Kenya, Somalia.

Notes. According to Damoiseau (1967a: 403) the holotype is in MNHUB, but we did not find it there.

Damoiseau (1967a: 403) only reports East African localities for the species while Kleine (1938d: 77) also quotes Guinea, Senegal and Sudan; these quotations must be verified.

tenuis Kleine, 1919

tenuis Kleine, 1919c: 16.

Type depository. MNHUB (?) (HT).

Distribution. Kenya, Somalia, Sudan, Tanzania.

Notes. According to Damoiseau (1967a: 406) the holotype is in MNHUB, but we did not find it there.

Symmorphocerus Schoenherr, 1847

Symmorphocerus Schoenherr, 1847: 8.

Scymmorphocerus: De Muizon, 1960: 139 [lapsus].

Type species: *Symmorphocerus monticola* Schoenherr, 1847.

Distribution. Greece (Crete); Angola, Benin, Cameroon, Central African Republic, Democratic Republic of the Congo (ex Zaire), Eritrea, Ethiopia, Gabon, Guinea Bissau, Ivory Coast, Kenya, Liberia, Nigeria, Republic of the

Congo, Republic of Guinea, Senegal, Somalia, South Africa, Sudan, Tanzania, Zambia; India, Iraq, Jordan, Kazakhstan, Lebanon, Pakistan, Syria, Turkey.

***alluaudi* Senna, 1894**

alluaudi Senna, 1894a: 409.

Type depository. Unknown.

Distribution. Benin, Cameroon, Democratic Republic of the Congo (ex Zaire), Guinea Bissau, Ivory Coast, Kenya, Nigeria, Republic of the Congo, Republic of Guinea, Senegal, Sudan, Tanzania.

Notes. We did not find the holotype in MNHN or MZUF. De Muizon (1960: 140) states that there is a cotype in MZUF but this specimen does not exist. Damoiseau (1967a: 385) states that he was not able to find the species holotype and defines some plesiotypes in IRSNB.

Damoiseau (1967a: 385) did not mention Cameroon, Congo and Tanzania (Kleine, 1920h: 7) when he gave the species distribution.

***approximatus* Damoiseau, 1964**

approximatus Damoiseau, 1964b: 421.

Type depository. MRAC (HT!), MNHN (PT!).

Distribution. Angola, Democratic Republic of the Congo (ex Zaire), Gabon, Republic of Guinea.

Notes. The quotation for the People's Republic of Congo (Sforzi, 1998b: 268) is incorrect and should be referred to as the Democratic Republic of the Congo (ex Zaire).

***beloni* Power, 1879**

beloni Power, 1879a: 488.

Type depository. MNHN (LT!, PLT!).

Distribution. Iraq, Turkey.

Notes. Lectotype of *S. beloni* (MNHN) designated on a male specimen with the following labels: Mesopotamie/ Type/ ex Musaeo G. Power/ Museum Paris 1952, coll. R. Oberthür. Paralectotypes: 1 female specimen (MNHN), without locality/ Type/ ex Musaeo G. Power/ Museum Paris 1952 coll. R. Oberthür; 1 male specimen (MNHN), Mossoul, Mesopotamie/ *beloni*/ Type/ ex Musaeo G. Power/ Museum Paris 1952, coll. R. Oberthür.

cardoni Senna, 1894

cardoni Senna, 1894c: 385.

Type depositories. IRSNB (LT!, PLT!), MZUF (PLT!).

Distribution. India, Pakistan.

Notes. Lectotype of *S. cardoni* (IRSNB) designated on a male specimen with the following labels: Barway, P. Cardon/ *Symmorphocerus cardoni* Senna Typus!/ Prépar. génit. No. 74 K 22 R Damoiseau/ *Symmorphocerus cardoni* Senna, R. Damoiseau det./ Lectotype [*in litteris*]. Paralectotypes: 1 female specimen (MZUF), Barway, P. Cardon/ *Symmorphocerus cardoni* Senna, ♀ typi/ Ann. Soc. Ent. Belg., XXXVIII, 1894, ♀ ♀ cotypi!/ ex-coll. Senna/ "La Specola" Firenze, 6976; 2 female specimens (MZUF), Barway, P. Cardon/ ex-coll. Senna/ "La Specola" Firenze, 6976; 1 female specimen (MZUF), India, ♀, ex coll. Boucard, sub *Am. aestivus*/ ex-coll. Senna/ "La Specola" Firenze, 6976; 1 male and 5 female specimens (IRSNB), Barway, P. Cardon/ *Symmorphocerus cardoni* Senna R. Damoiseau det./ Paralectotype [*in litteris*].

consequens Kleine, 1926

consequens Kleine, 1926f: 366.

consequena: Shalaby *et al.*, 1968: 4 (*Symmorphocerus*) [lapsus].

Type depository. NHM (LT!, PLT!).

Distribution. Iraq, Kazakhstan.

Notes. W. Schedl (1975a: 31) reported that a specimen held in NHM is the holotype, but in his original description Kleine did not designate any of the syntypes as holotype; however, as there is more than one syntype in NHM, and as W. Schedl did not specifically select one as holotype, we designate the lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Lectotype of *S. consequens* (NHM) designated on a male specimen with the following labels: Syntype/ 40179/ Millingen/ Turcom^a Kurdⁿ/ Fry Coll. 1905-100. Paralectotype: 1 male specimen (NHM), Syntype/ Typus!/ Turcom^a Kurdⁿ/ Millingen/ Fry Coll. 1905-100/ *Symmorphocerus consequens* Kln., R. Kleine det. 1922/ Prépar. génit. No. BM 5, R. Damoiseau.

frontalis (Olivier, 1807)

frontalis Olivier, 1807: 434 (*Brentus*).

lottoi De Muizon, 1960: 139 (*Symmorphocerus*); syn. Damoiseau, 1989b: 64.

Type depositories. *B. frontalis*: unknown; *S. lottoi*: NHM (LT!, PLT!).

Distribution. Angola, Cameroon, Central African Republic, Democratic Republic of the Congo (ex Zaire), Eritrea, Ethiopia, "Guinea", Ivory Coast, Kenya, Senegal, South Africa, Tanzania, Zambia.

Notes. Damoiseau (1967a: 385) writes that he was unable to find the holotype of *B. frontalis* and designates plesiotypes in IRSNB.

Lectotype of *S. lottoi* (NHM) designated on a male specimen [incorrectly labelled as holotype] with the following labels: Holotype/ Type ♂/ Eritrea, Ghinda, River Ghinda, 9.I.1940, G. de Lotto, 900 m. a.s.l. 90100/ Pres. by Com. Inst. ent. B. M. 1948-142/ *Symmorphocerus* sp., D.J. Atkinson det. 1950/ *Symmorphocerus lottoi* n. sp. Type ♂/ J. De Muizon det. Paralectotype: 1 female specimen (NHM), Paratype/ Type ♀/ Eritrea, Ghinda, River Ghinda, 9.I.1940, G. de Lotto, 900 m. a.s.l. 90100/ Imp. Inst. Ent. Coll. No. 10811/ Pres. by Com. Inst. ent. B. M. 1948-142/ *Symmorphocerus lottoi* n. sp. Type ♀/ J. De Muizon det.

Damoiseau (1967a: 385) did not mention South Africa and Guinea (Power, 1879: 488) and Senegal (Kleine, 1935d: 113) when he gave the species distribution.

minutus Power, 1879

minutus Power, 1879a: 488.

Type depository. MNHN (HT!).

Distribution. Eritrea, Ethiopia, Sudan.

***monticola* Schoenherr, 1847 stat. res.**

monticola Schoenherr, 1847: 8.

semipunctatus Pascoe, 1872: 321 (*Cordus*); syn. Damoiseau, 1967a: 389.

Type depositories. *S. monticola*: NHRS (?) (HT); *C. semipunctatus*: NHM (HT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Liberia, South Africa, Tanzania, Zambia.

Notes. Kleine (1928a: 79) reported that the type for *S. monticola* is in NHRS but the species is not listed in the NHRS web site.

Kleine (1927a: 30; 1938d: 77) mistakenly attributes the species to Fåhræus (1871: 434) and considers it a synonym of *B. frontalis* Olivier. Damoiseau (1967a: 389) also mistakenly attributes the species to Fåhræus but considers it as "*bona species*". According to Alonso-Zarazaga & Lyal (1999: 49) *S. monticola* Schoenherr is a synonym of *B. frontalis* Olivier, but in our opinion the two species are distinct.

Damoiseau (1967a: 389) did not mention Liberia (Senna, 1896b: 253) and Cameroon (Kleine, 1936a: 29) when he gave the species distribution.

***nagporensis* Kleine, 1922**

nagporensis Kleine, 1922f: 211.

Type depository. SMTD (ST).

Distribution. India.

***piochardi* (Bedel, 1877)**

piochardi Bedel, 1877: CLXXXIV (*Amorphocephalus*).

piliger Desbrocher des Loges, 1895: 99 (*Amorphocephalus*); syn. Pic, 1905: 280.

saalasi Sahlberg, 1913a: 61 (*Symmorphocerus*); syn. W. Schedl, 1970: 102.

unionis Sahlberg, 1913b: 201 (*Amorphocephalus*); syn. W. Schedl, 1970: 102.

muchi Hertel, 1961: 117 (*Amorphocephalus*); syn. W. Schedl, 1970: 102.

cretensis Hertel, 1964: 167 (*Amorphocephalus muchei* ssp.); syn. W. Schedl, 1970: 102.

Type depositories. *S. piochardi*: MNHN (HT!); *A. piliger*: unknown; *S. saalasi*: unknown; *A. muchei*: SMTD (HT); *A. muchei* ssp. *cretensis*: SMTD (HT, PT).

Distribution. Greece (Crete); Sudan (?); Jordan, Lebanon, Syria, Turkey.

Notes. Sahlberg (1913a: 61) described *Symmorphocerus saalasi*; in the same year (1913b: 201) he referred to the same species, but curiously he called it *Amorphocephalus unionis*.

supercillatus Damoiseau, 1967

supercillatus Damoiseau, 1967a: 388.

Type depositories. MNHUB (HT!), MRAC (AT!).

Distribution. Democratic Republic of the Congo (ex Zaire), Kenya, Somalia, Tanzania.

Systellus Kleine, 1917

Systellus Kleine, 1917d: 175.

Eusystellus Kleine, 1917e: 316 [unjustified replacement name].

Eurystellus: Kleine, 1931i: 307 [lapsus].

Type species: *Systellus rex* Kleine, 1917.

Distribution. Borneo, Indonesia (Sumatra).

rex Kleine, 1917

rex Kleine, 1917d: 177.

Type depositories. DEI (HT!), RMNH (PT).

Distribution. Borneo, Indonesia (Sumatra).

CYPHAGOGINAE Kolbe, 1892

CYPHAGOGINI Kolbe, 1892

- Cyphagoginae Kolbe, 1892: 162.
 Cyphagogi Senna, 1896a: 210.
 Calodrominen Kolbe, 1916: 50.
 Calodromini Kleine, 1922q: 148.
 Calodermini: Vitzthum, 1921b: 5 [lapsus].
 Calodrominidae: Kleine, 1922h: 133 [lapsus].
 Calodrominae Kleine, 1925b: 43.
 Caladromini: Kleine, 1938d: 176 [lapsus].
 Zemiosinae Damoiseau, 1965a: 6.
 Cyphagogidae Ienistea, 1986: 32.
 Cyphagogina Zimmerman, 1994: 130.
 Cyphagogini Morrone, 1998: 132.

Adidactus Senna, 1894

Adidactus Senna, 1894a: 406.

Type species: *Zemioses cancellatus* Lacordaire, 1866.

Distribution. Angola, Cameroon, Comores Is., Democratic Republic of the Congo (ex Zaire), Equatorial Guinea (including Bioko Is. [= Fernando Poo]), Gabon, Ghana, Ivory Coast, Kenya, Madagascar, Namibia, Principe, Republic of the Congo, Republic of Guinea, South Africa, Tanzania, Togo, Uganda, Zambia.

cancellatus (Lacordaire, 1866)

cancellatus Lacordaire, 1866: 409 (*Zemioses*).

conradsi Kleine, 1934a: 304 (*Genogogus*); syn. De Muizon, 1955a: 464.

Type depositories. *Z. cancellatus*: MCZ (HT); *G. conradsi*: DEI (HT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea (including Bioko Is. [= Fernando Poo]), Gabon, Ghana, Ivory Coast, Kenya, Namibia, Natal, Republic of the Congo, Republic of Guinea, South Africa, Tanzania, Togo.

Notes. Kleine (1934a: 304) described *G. conradsi* on a series of five specimens but only gave the locality for the holotype; it was not possible to discover where the paratypes are held.

Damoiseau (1967a: 68) did not mention South Africa (Natal) (Senna, 1892c: 40) and Togo (Kolbe, 1916: 62) when he gave the species distribution.

contumax (Kleine, 1924)

contumax Kleine, 1924a: 98 (*Schizoadidactus*).

Type depository. MRAC (HT!).

Distribution. Democratic Republic of the Congo (ex Zaire), Gabon, Republic of the Congo.

Notes. According to Damoiseau (1967a: 71) the status of this species is doubtful because it was described on a very small specimen which is very similar to *A. striolatus* Fairmaire.

madagascariensis De Muizon, 1955

madagascariensis De Muizon, 1955a: 466.

Type depository. MNHN (HT!).

Distribution. Madagascar.

striolatus (Fairmaire, 1897)

striolatus Fairmaire, 1897: 187 (*Zemioses*).

infantulus Kolbe, 1916: 62 (*Adidactus*); syn. Damoiseau, 1967a: 69.

dispositus Kleine, 1926f: 359 (*Schizoadidactus*); syn. De Muizon, 1955a: 470.

tibialis K. E. Schedl, 1961a: 10 (non Kleine, 1916) (*Adidactus*); syn. Damoiseau, 1967a: 69.

Type depositories. *Z. striolatus*: MNHN (LT!, PLT!); *A. infantulus*: MNHUB (ST!), IRSNB (ST!); *S. dispositus*: NHM (HT!).

Distribution. Angola, Cameroon, Comores Is., Democratic Republic of the Congo (ex Zaire), Equatorial Guinea, Gabon, Ghana, Ivory Coast, Kenya, Madagascar, Principe, Republic of the Congo, Tanzania, Uganda, Zambia.

Notes. Damoiseau (1967a: 71) reported that the specimen of *Z. striolatus* held in MNHN is the holotype, but in his original description Fairmaire did not designate any of the syntypes as holotype; however, as there is more than

one syntype in MNHN, and as Damoiseau did not specifically select one as holotype, we designate the lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Lectotype of *Zemioses striolatus* (MNHN) designated on a specimen with the following labels: Madagascar, Tsarasoatra, 2e semestre 1895, Lieut. Scalabre/ Museum Paris, Madagascar Scalabre 1897/ *Zemioses striolatus* n. sp./ *Zemioses striolatus* Fairmaire, 1897 type. Paralectotypes: 1 specimen (MNHN), Madagr., L. Scalabre/ Museum Paris 1952, coll. R. Oberthür/ *Zemioses striolatus* n. sp.; 3 specimens (MNHN), Madagascar, Tsarasoatra, 2^e semestre 1895, Lieut. Scalabre/ Collection A. Lajoie/ Museum Paris 1952, coll. R. Oberthür; 1 specimen (MNHN), Madagr. L. Scalabre/ Museum Paris 1952, coll. R. Oberthür/ *Zemioses striolatus* n. sp.; 1 specimen (MNHN), ex Musaeo L. Fairmaire 1893/ Museum Paris 1952, coll. R. Oberthür/ *Zemioses striolatus* Faurmaire, 1896/ Kleine vidit V.1934; 5 specimens (MNHN), Madagr., L. Scalabre/ Museum Paris 1952, coll. R. Oberthür.

According to Quentin (1966: 1633) *A. infantulus* syntypes are also held in MZPW.

Damoiseau (1967a: 71) did not mention Uganda (Calabresi, 1920d: 18) when he gave the species distribution.

Allaeometrus Senna, 1903

Allaeometrus Senna, 1903: 157.

Subdysmorphorhynchus Kleine, 1917f: 323; syn. Damoiseau, 1964a: 395.

Subdismorphorhynchus: Kleine, 1922m: 161 [lapsus].

Allacometrus: Döbler & Gaedike, 1995: 443 [lapsus].

Type species: *Allaeometrus breviceps* Senna, 1903.

Distribution. China (Jiangxi), Taiwan; Borneo, India (including Assam), Indonesia (Java, Sumatra), "Indochina", Laos, Malaysia, Thailand, Vietnam; Australia, New Guinea.

breviceps Senna, 1903

breviceps Senna, 1903: 158.

carinatus Kleine, 1917f: 326 (*Subdysmorphorhynchus*); syn. Damoiseau, 1964a: 396.

Type depositories. *A. breviceps*: MSNG (LT!, PLT!), SMTD (PLT); *S. carinatus*: DEI (LT!).

Distribution. China (Jiangxi), Taiwan; Borneo, India (including Assam), Indonesia (Java, Sumatra), "Indochina", Laos, Malaysia, Thailand, Vietnam; Australia, New Guinea.

Notes. Damoiseau (1964a: 398) reported that the specimens of *A. breviceps* held in MSNG are holotype and a paratype, but in his original description Senna did not designate either of the syntypes as holotype; however, as there is more than one syntype in MSNG, and as Damoiseau did not specifically select one as holotype, we designate the lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Lectotype of *A. breviceps* (MSNG) designated on a specimen with the following labels: Sumatra: Penang, 600-800 m, II.1889, leg. Loria e Fea/ *Allaeometrus breviceps* typus! Senna. Paralectotype: 1 specimen (MSNG), Sumatra, Si Rambé, XII.1890-III.1891, leg. Modigliani.

Lectotype of *S. carinatus* (DEI) designated on a specimen with the following labels: Coen District, Cape York, Queensland, H. Hacker/ Dtsch. Entomol. Institut Berlin/ Syntypus/ Kleine det./ coll. Hacker/ Comp. au Type/ *Subdysmorphorhynchus carinatus* Kleine, det. Kleine, 1917/ *Allaeometrus breviceps* Senna, R. Damoiseau det., 1962. We were not able to find the other syntype (species description was based on two specimens).

setosus (Schönfeldt, 1906)

setosus Schönfeldt, 1906: 35 (*Zemioses*).

Type depository. RMNH (HT).

Distribution. New Guinea.

Notes. Damoiseau (1964a: 396) places this species in *Allaeometrus* as *incertae sedis* and writes that he was unable to find the type specimen. Damoiseau (1989a: 106) however states that the genus only includes the type species.

Allagogus Gahan, 1909

Allagogus Gahan, 1909: 225.

Oxybasius Kolbe, 1916: 61; syn. Damoiseau, 1962b: 6.

Phobetrum Kleine, 1916c: 31; syn. Damoiseau, 1962b: 6.

Diplohoplizes Kleine, 1916c: 37; syn. Damoiseau, 1962b: 6.

Allogogus: Damoiseau, 1962b: 4 [lapsus].

Type species: *Allagogus brunneus* Gahan, 1909.

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea (including Bioko Is. [= Fernando Poo]), Ethiopia, Gabon, Ghana, Ivory Coast, Kenya, Senegal, Tanzania, Uganda, Zambia, Zimbabwe.

***brunneus* Gahan, 1909**

brunneus Gahan, 1909: 225.

brevithorax Kolbe, 1916: 61 (*Oxybasius*); syn. Damoiseau, 1962b: 6.

laticornis Kolbe, 1916: 61 (*Oxybasius*); syn. Kleine, 1927a: 9.

pectinicornis Kolbe, 1916: 61 (*Oxybasius*); syn. Kleine, 1927a: 9.

sulcirostris Kolbe, 1916: 61 (*Oxybasius*); syn. Kleine, 1927a: 9.

paradoxum Kleine, 1916c: 33 (*Phobetrum*); syn. De Muizon, 1955a: 489.

armatus Kleine, 1916c: 38 (*Diplohoplizes*); syn. De Muizon, 1955a: 491.

burgeoni De Muizon, 1955a: 492 (*Diplohoplizes*); syn. Damoiseau, 1962b: 6.

Type depositories. *A. brunneus*: NHM (HT!); *O. brevithorax*: MNHUB (?) (HT); *O. laticornis*: MNHUB (?) (ST); *O. pectinicornis*: MNHUB (?) (HT); *O. sulcirostris*: MNHUB (?) (HT); *P. paradoxum*: DEI (LT!, PLT!), ZMUH (PLT); *D. armatus*: DEI (LT!, PLT!); *D. burgeoni*: MNHN (HT).

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea (including Bioko Is. [= Fernando Poo]), Ethiopia, Gabon, Ghana, Ivory Coast, Kenya, Senegal, Tanzania, Uganda, Zambia, Zimbabwe.

Notes. Lectotype of *P. paradoxum* (DEI) designated on a male specimen with the following labels: N Kamerun, L. Conradt, 96/ Syntypus/ Dtsch. Entomol. Institut Berlin/ Kleine det./ *Allagogus brunneus* Gahan ♂, R. Damoiseau, det., 1961. Paralectotypes: 4 female specimens (DEI), N Kamerun, L. Conradt, 96/ Syntypus/ coll. Kraatz/ Kleine det./ Dtsch. Entomol. Institut Berlin/ *Allagogus brunneus* ♀, R. Damoiseau, det., 1961.

Lectotype of *D. armatus* (DEI) designated on a male specimen with the following labels: N Kamerun, L. Conradt, 96/ Syntypus/ ♂/coll. Kraatz/ *Diplo. armatus* Kl./ Kleine det./ *Diplohoplizes armatus* n. g. n. sp./ *Allagogus brunneus* Gahan ♂, R. Damoiseau, det., 1961. Paralectotype: 1 specimen (DEI), N Kamerun, L. Conradt, 96/ coll. Kraatz/ Kleine det./ Syntypus/ *Allagogus brunneus* Gahan ♂, R. Damoiseau, det., 1961.

We did not find any types of *O. brevithorax*, *O. laticornis*, *O. pectinicornis*, *O. sulcirostris* in MNHUB where they should be held.

***yangambiensis* Damoiseau, 1961**

yangambiensis Damoiseau, 1961a: 279.

Type depository. MRAC (HT!, AT!, PT!).

Distribution. Angola, Democratic Republic of the Congo (ex Zaire), Ghana.

Amobaeus Kleine, 1925

Amobaeus Kleine, 1925a: 192.

Type species: *Cyphagogus costipennis* Fairmaire, 1889 *sensu* Kleine, 1925 = *Amobaeus costipennis* Quentin, 1961.

Distribution. Madagascar

costipennis Quentin, 1961

costipennis Quentin, 1961: 206.

Type depository. MNHN (?) (HT).

Distribution. Madagascar.

Notes. We did not find the holotype in MNHN. See Quentin (1961) for the complex taxonomical problem related to this species.

Amphithetobrentus Damoiseau, 1963

Amphithetobrentus Damoiseau, 1963c: 236.

Type species: *Schizoadidactus concolor* Kleine, 1916.

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Gabon, Ghana, Republic of the Congo, Uganda.

concolor (Kleine, 1916)

concolor Kleine, 1916c: 22 (*Schizoadidactus*).

congoensis Kleine, 1926f: 357 (*Genogogus*); syn. De Muizon, 1955a: 469.

Type depositories. *S. concolor*: DEI (HT!); *G. congoensis*: MRAC (HT!).

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Gabon, Ghana, Republic of the Congo, Uganda.

Notes. Damoiseau (1967a: 108) did not mention Uganda (Kleine, 1936a: 14) when he gave the species distribution.

Ancylobrentus Damoiseau, 1965

Ancylobrentus Damoiseau, 1965a: 10.

Type species: *Ancylobrentus australicola* Damoiseau, 1965.

Distribution. Australia.

australicola Damoiseau, 1965

australicola Damoiseau, 1965a: 11.

Type depository. SAMA (HT).

Distribution. Australia.

Autometrus Kleine, 1922

Autometrus Kleine, 1922p: 130.

Type species: *Autometrus punctulatus* Kleine, 1922.

Distribution. Borneo, Singapore, Vietnam; Solomon Is.

papuensis Damoiseau, 1979

papuensis Damoiseau, 1979a: 24.

Type depository. NHM (HT!).

Distribution. Solomon Is.

punctulatus Kleine, 1922

punctulatus Kleine, 1922p: 131.

Type depository. SMTD (HT).

Distribution. Borneo, Singapore, Vietnam.

Autosebus Kolbe, 1916

Autosebus Kolbe, 1916: 57.

Autosebius: Döbler & Gaedike, 1995: 444 [lapsus].

Type species: *Autosebus setosellus* Kolbe, 1916.

Distribution. Angola, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea, Gabon, Ghana, Ivory Coast, Nigeria, Republic of the Congo, Rwanda, Sierra Leone, Tanzania, Togo, Uganda.

burgeoni (De Muizon, 1959)

burgeoni De Muizon, 1959: 74 (*Microsebus*).

Type depository. MNHN (HT!).

Distribution. Democratic Republic of the Congo (ex Zaire), Ivory Coast, Rwanda.

donisi Damoiseau, 1962

donisi Damoiseau, 1962f: 334.

Type depositories. MRAC (HT!, AT!), DEI (PT!), IRSNB (PT!).

Distribution. Democratic Republic of the Congo (ex Zaire).

kolbei (De Muizon, 1960)

kolbei De Muizon, 1960: 88 (non Kleine, 1916) (*Microsebus*).

Type depository. MNHN (?) (HT).

Distribution. Ivory Coast, Sierra Leone.

Notes. According to Damoiseau (1967a: 131) the holotype is in MNHN, but we did not find it there.

magnus Damoiseau, 1967

magnus Damoiseau, 1967a: 133.

Type depository. MRAC (HT!).

Distribution. Ivory Coast.

mucronatus Damoiseau, 1972

mucronatus Damoiseau, 1972a: 263.

Type depository. HNHM (HT).

Distribution. Ghana.

setosellus Kolbe, 1916

setosellus Kolbe, 1916: 57.

calcaratus Calabresi, 1920d: 20 (*Microsebus*); syn. Damoiseau, 1962f: 333.

Type depositories. *A. setosellus*: MNHUB (LT!, PLT!); *M. calcaratus*: MSNG (HT!).

Distribution. Angola, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea, Gabon, Ghana, Ivory Coast, Nigeria, Republic of the Congo, Tanzania, Togo, Uganda.

Notes. According to Quentin (1966: 1635), the type for *A. setosellus* is in the Museum of Stettin (now in MZPW), but it is not there [T. Huflejt (MZPW) (2002, personal communication)]. According to Damoiseau (1967a: 130) the specimen held in MNHUB is the holotype, but in his original description Kolbe did not designate any of the syntypes as holotype; however, as there is more than one syntype in MNHUB, and as Kolbe did not specifically select one as holotype, we designate the lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Lectotype of *A. setosellus* (MNHUB) designated on a specimen with the following labels: Togo, Bismarckburg, L. Conradt S./ Type/ *Autosebus setosellus* n. sp. Paralectotypes: 1 specimen (MNHUB), with the same label data as the lectotype; 1 specimen (MNHUB), Span. Guinea, Uellebg. Benitogbt., 15-31.07/ G. Tessmann/ S. G./ 3132-220/ *Autosebus setosellus*/ Type/ *Autosebus setosellus* Kolbe, R. Damoiseau det., 1962; 1 specimen (MNHUB), D.-Ostafri-

ka, Tendaguru, Lindi, Mitte VIII-Mitte XI. 09, Janensch S. G./ *Autosebus setosellus*/ Paratype/ *Autosebus setosellus* Kolbe, R. Damoiseau det., 1962.

Azemius Damoiseau, 1961

Azemius Damoiseau, 1961b: 6.

Type species: *Acidotus vaneyeni* De Muizon, 1959.

Distribution. Democratic Republic of the Congo (ex Zaire), Ghana, Republic of the Congo.

vaneyeni (De Muizon, 1959)

vaneyeni De Muizon, 1959: 75 (*Acidotus*).

Type depository. MRAC (HT!).

Distribution. Democratic Republic of the Congo (ex Zaire), Ghana, Republic of the Congo.

Basenius Kolbe, 1892

Basenius Kolbe, 1892: 170.

Pachydactylus Kleine, 1926f: 358 (non Wiegmann, 1834, nec Menge, 1868); syn. De Muizon, 1955a: 485.

Neopachydactylus Hedicke, 1927: 145; syn. De Muizon, 1955a: 485.

Type species: *Basenius laticornis* Kolbe, 1892.

Distribution. Tanzania, Mozambique.

laticornis Kolbe, 1892

laticornis Kolbe, 1892: 170.

marshalli Kleine, 1926f: 359 (*Pachydactylus*); syn. De Muizon, 1955a: 485.

Type depositories. *B. laticornis*: MNHUB (HT!); *P. marshalli*: NHM (HT!).

Distribution. Tanzania, Mozambique.

Caenosebus Kleine, 1916

Caenosebus Kleine, 1916c: 42.

Gaenosebus: Kleine, 1937d: 23 [lapsus].

Type species: *Caenosebus fraudator* Kleine, 1916.

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire).

fraudator Kleine, 1916

fraudator Kleine, 1916c: 45.

Type depositary. DEI (HT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire).

Callipareius Senna, 1892

Callipareius Senna, 1892e: 444.

Type species: *Callipareius feae* Senna, 1892.

Distribution. China, Japan, Taiwan; Democratic Republic of the Congo (ex Zaire), Ghana; Bhutan, Borneo, Indonesia (Java), India (including Darjeeling), Myanmar [= Burma], Philippines (Luzon), Vietnam; Australia, New Caledonia, New Guinea, Vanuatu [= New Hebrides].

subgenus *Callipareius* s. s.

Diastrophus Perroud & Montrouzier, 1865: 141 (non Hartig, 1840); syn. Damoiseau, 1965a: 19.

Asaphepterum Kleine, 1916c: 85; syn. Calabresi, 1922: 107.

Calliparius: Calabresi, 1922: 107 [lapsus].

Pseudosebasius Kleine, 1944: 150; syn. Damoiseau, 1989a: 108.

Asaphoptorum: Döbler & Gaedike, 1995: 445 [lapsus].

Notes. Alonso-Zarazaga & Lyal (1999: 50 and 248) give Perroud as author of the genus, but we believe that Perroud and Montrouzier should be considered as co-authors.

bicolor Damoiseau, 1965

bicolor Damoiseau, 1965a: 20.

Type depository. NHM (HT!).

Distribution. Myanmar [= Burma].

bistriatus (Kleine, 1935)

bistriatus Kleine, 1935c: 73 (*Opysthenoxys*).

lineatosignatus Kleine, 1939d: 1 (*Eterozemus*); syn. Damoiseau, 1989a: 111.

Type depositories. *O. bistriatus*: NHM (HT!, PT!), DEI (PT!); *E. lineatosignatus*: NHRS (?) (HT).

Distribution. China (Fujian); Bhutan, India (Darjeeling), Myanmar [= Burma].

Notes. Kleine (1939d: 1) reported that the type for *E. lineatosignatus* is in NHRS but the species is not listed in the NHRS web site.

feae Senna, 1892

feae Senna, 1892e: 445.

feai: Damoiseau, 1989a: 109 (*Callipareius*) [unjustified emendation].

Type depositories. MSNG (LT!, PLT!), MZUF (PLT!).

Distribution. S China; India (including Darjeeling), Myanmar [= Burma], Vietnam.

Notes. Damoiseau (1989a: 109) reported that the specimens held in MSNG are holotype and paratypes, but in his original description Senna did not designate any of the syntypes as holotype; however, as there is more than one syntype in MSNG, and as Damoiseau did not specifically select one as holotype, we designate the lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Lectotype of *C. feae* (MSNG) designated on a specimen with the following labels: Birmania, Carin Ghecù, 1300-1400 m, L. Fea, VI. 1888/ typus/ fam. Taphroderinae. *Callipareius* n. gen. *Feae* n. sp. typus! Paralectotypes: 2 specimens (MSNG), Birmania, Carin Ghecù, 1300-1400 m, L. Fea, V. 1888; 1

specimen (MZUF), Carin Ghecù, 1300-1400 m, L. Fea, V. 1888/ *Callipareius* ♂ n. g. *feae* Senna n. sp., Typus!/ Ann. Mus. civ. St. nat. Genova, XXXII, 1892, Typus!/ *Callipareius* (s. s.) *feai* Senna, nom. emend., R. Damoiseau det., 1966/ ex-coll. Senna/ "La Specola" Firenze, 6919.

formosanus (Kleine, 1916)

formosanus Kleine, 1916c: 87 (*Asaphepterum*).

Type depository. DEI (HT!).

Distribution. Japan, Taiwan; Borneo, Indonesia (Java), Philippines (Luzon), Vietnam.

foveatus Senna, 1894

foveatus Senna, 1894c: 384.

mediocris Kleine, 1935c: 74 (*Pittodes*); syn. Damoiseau, 1965a: 19.

Type depositories. *C. foveatus*: MZUF (LT!), IRSNB (PLT!), MNHN (?) (PLT); *P. mediocris*: NHM (HT!).

Distribution. India (including Darjeeling), Myanmar [= Burma].

Notes. According to Damoiseau (1965a: 20) the syntypes of *C. foveatus* are held in MNHN, but we did not find them there. The original description mentions 3 male specimens in IRSNB and a female in Oberthür's collection. Type specimens are present in MZUF and in IRSNB.

Lectotype of *C. foveatus* (MZUF) designated on a specimen with the following labels: Kurseong, P. Braet/ *Callipareius foveatus* Senna, typus/ Ann. Soc. Ent. Belg., XXXVIII, 1894, Typus!/ *Callipareius* (s. s.) *foveatus* Senna, R. Damoiseau det., 1966/ ex-coll. Senna/ "La Specola" Firenze, 6920. Paralectotypes: 1 female specimen [incorrectly labelled as holotype] (IRSNB), Kurseong, R. P. Cardon/ *Callipareius foveatus*, typus Senna/ Holotype/ *Callipareius foveatus* Senna Type ♀ Dam. det.; 1 specimen (IRSNB), Barway, P. Cardon/ *Callipareius foveatus* n. sp. det. Senna 1894/ Paratype/ *Callipareius* (*Callipareius*) *foveatus* Senna, R. Damoiseau vid. 1965.

japonicus (Nakane, 1963)

japonicum Nakane, 1963: 34 (*Asaphepterum*).

Type depository. SEHU (HT).

Distribution. Japan.

miyakawai Morimoto, 1979

miyakawai Morimoto, 1979: 25.

Type depository. ELKU (HT, PT).

Distribution. Japan.

planitarsus (Perroud & Montrouzier, 1865)

planitarsus Perroud & Montrouzier, 1865: 142 (*Diastrophus*).

flavolineatus Calabresi, 1922: 107 (*Callipareius*); syn. Damoiseau, 1965a: 19.

planitarsis: Alonso-Zarazaga & Lyal, 1999: 50 (*Diastrophus*) [lapsus].

Type depositories. *D. planitarsus*: MNHN (HT!); *C. flavolineatus*: MNHN (HT!).

Distribution. Australia, New Caledonia, New Guinea, Vanuatu [= New Hebrides].

Notes. Damoiseau (1965a: 19; 1989a: 108) considered *D. planitarsus* as a synonym of *C. flavolineatus* but this is not possible as *planitarsus* has priority (Zimmerman, 1994: 133).

Alonso-Zarazaga & Lyal (1999: 50 and 248) give Perroud as author of the species *planitarsus*, but we believe that Perroud and Montrouzier should be considered as co-authors.

preangeri Damoiseau, 1965

preangeri Damoiseau, 1965a: 22.

Type depository. MNHUB (HT!).

Distribution. Indonesia (Java).

***rubrocastaneus* (Kleine, 1944)**

rubrocastaneus Kleine, 1944: 150 (*Pseudosebasius*).

Type depository. MNHN (HT!).

Distribution. India.

subgenus *Metacidotes* K. E. Schedl, 1961

Metacidotes K. E. Schedl, 1961c: 193.

Metacidotus: K. E. Schedl, 1961c: 194 [lapsus].

Coomania Kleine, 1941b: 226 (non Cameron, 1939); syn. Damoiseau, 1961b: 15.

Coomanisia Gomy, 1987: 27; syn. Alonso-Zarazaga & Lyal, 1999: 50.

***kojimai* Morimoto, 1982**

kojimai Morimoto, 1982: 31.

Type depository. ELKU (HT, PT).

Distribution. Japan, Taiwan.

Notes. Li-zhong (2002) also quotes the species for Borneo, Indonesia, Malaysia, Myanmar, and India.

***ponderosus* (Kleine, 1941)**

ponderosus Kleine, 1941b: 227 (*Coomania*).

Type depository. MNHN (HT!).

Distribution. S China (Hainan); Vietnam.

***projectus* Damoiseau, 1961**

projectus Damoiseau, 1961b: 17.

Type depository. IRSNB (HT!).

Distribution. Vietnam.

***pulvifrons* (K. E. Schedl, 1961)**

pulvifrons K. E. Schedl, 1961c: 194 (*Metacidotes*).

Type depository. MRAC (HT!).

Distribution. Democratic Republic of the Congo (ex Zaire), Ghana.

Notes. K. E. Schedl (1961c: 194) reported that one paratype was in his collection but W. Schedl (2001, personal communication) did not find it in his father's collection.

***Calodromus* Guérin-Méneville, 1832**

Calodromus Guérin-Méneville, 1832: Cl. IX, Pl. 34.

Caladromus: Castelnau, 1840: 295 [lapsus].

Amorphopus Schoenherr, 1840: 577 (non Audinet-Serville, 1838) syn. Alonso-Zarazaga & Lyal, 1999: 50.

Callidromus Agassiz, 1846b: 60 [unjustified replacement name].

Alloeodromus Senna, 1895a: 179; syn. Damoiseau, 1964c: 464.

Allaeodromus: Schönfeldt, 1910a: 4 [lapsus].

Calochromus: Commonwealth Institute of Entomology, 1950: 76 [lapsus].

Type species: *Calodromus mellyi* Guérin-Méneville & Gory, 1832.

Distribution. S China, Taiwan; Borneo, Kampuchea [= Cambodia], India (including Andaman Is., Assam, Nicobares Is.), Indonesia (Java, Sumatra), Laos, Malaysia, Myanmar [= Burma], Philippines (Luzon, Mindanao), Sri Lanka, Thailand, Vietnam.

***amabilis* Kleine, 1916**

amabilis Kleine, 1916d: 111.

Type depository. ZMAN (HT!).

Distribution. Indonesia (Java).

crinitus Kleine, 1922

crinitus Kleine, 1922b: 24.

Type depository. SMTD (HT).

Distribution. Philippines (Luzon).

douglasi Kleine, 1931

douglasi Kleine, 1931g: 184.

Type depository. ZMAN (HT!).

Distribution. Indonesia (Sumatra).

horishanus Kano, 1930

horishanus Kano, 1930: 153.

Type depository. TARI (?) (HT).

Distribution. Taiwan.

insignis (Senna, 1895)

insignis Senna, 1895a: 180 (*Allaeodromus*).

Type depository. RMNH (HT).

Distribution. Indonesia (Sumatra).

integer Damoiseau, 1971

integer Damoiseau, 1971a: 17.

Type depository. IRSNB (HT!).

Distribution. "Indonesia".

ito Heller, 1910

ito Heller, 1910: 180.

Type depositories. NHM (ST!), SMTD (ST), RMNH (ST).

Distribution. Taiwan.

Notes. Damoiseau (1971a: 11) reported that the holotype and allotype are at DEI and paratypes are in DEI, RMNH, and SMTD, but species description was based on several specimens from SMTD. We did not find any syntypes in DEI, neither are they mentioned by Döbler & Gaedike (1995).

kalabakani Damoiseau, 1971

kalabakani Damoiseau, 1971a: 16.

Type depository. BPBM (HT).

Distribution. N Borneo.

luzonicus Damoiseau, 1964

luzonicus Damoiseau, 1964c: 465.

Type depository. MNHUB (HT!).

Distribution. Philippines (Luzon).

mellyi Guérin-Méneville & Gory, 1832

mellyi Guérin-Méneville & Gory, 1832: Cl. IX, Pl. 34.

mellesi: Boheman, 1837: 221 (*Calodromus*) [lapsus].

mellii: Westwood, 1848: 32 (*Calodromus*) [lapsus].

sarawakensis Heller, 1910: 179 (*Calodromus mellyi* ssp.); syn. Damoiseau, 1964c: 465.

Type depositories. *C. mellyi*: unknown; *C. mellyi* ssp. *sarawakensis*: NHM (HT!).

Distribution. S China; Borneo, Kampuchea [= Cambodia], India (including Andaman Is., Assam, Nicobares Is.), Indonesia (Sumatra), Laos, Malaysia, Myanmar [= Burma], Philippines (Luzon, Mindanao), Sri Lanka, Thailand, Vietnam.

simplex Heller, 1910

simplex Heller, 1910: 179.

Type depository. RMNH (HT).

Distribution. Indonesia (Sumatra), Vietnam.

vethi Kleine, 1915

vethi Kleine, 1915: 120.

Type depository. RMNH (HT).

Distribution. Indonesia (Java).

Catagogus Kleine, 1926

Catagogus Kleine, 1926f: 354.

Catagonus: Kleine, 1938d: 18 (non Ameghino, 1904) [lapsus].

Type species: *Catagogus desidiosus* Kleine, 1926 = *Cyphagogus diorymerus* Lea, 1898.

Distribution. Australia.

diorymerus (Lea, 1898)

diorymerus Lea, 1898: 633 (*Cyphagogus*).

desidiosus Kleine, 1926f: 355 (*Catagogus*); syn. Damoiseau, 1965a: 18.

Type depositories. *C. diorymerus*: SAMA (Lea coll.) (ST); *C. desidiosus*: NHM (HT!).

Distribution. Australia.

Notes. According to Damoiseau (1989a: 112) the holotype of *C. diorymerus* is held in the Macleay Museum. Zimmerman (1994: 135) states the holotype as being in SAMA. In the Lea collection there are 2 specimens glued to a card with Lea's typical red "Type" writing on the label. These specimens should be the types, but since Lea did not designate one of them as holotype in the original description, they are syntypes (R. Oberprieler, 2001, personal communication).

Ceragogus Kleine, 1925

Ceragogus Kleine, 1925a: 190.

Type species: *Cyphagogus appendiculatus* Fairmaire, 1889.

Distribution. Madagascar.

appendiculatus (Fairmaire, 1889)

appendiculatus Fairmaire, 1889: XCII (*Cyphagogus*).

Type depository. MNHN (LT!, PLT!).

Distribution. Madagascar.

Notes. Lectotype of *C. appendiculatus* (MNHN) designated on a specimen with the following labels: Madagascar, Tenerive, E. Perrot/ Museum Paris 1952, coll. R. Oberthür/ *Cyphagogus appendiculatus*. Paralectotypes: 2 specimens (MNHN) with the same label data as the lectotype.

Chelorhinus Kleine, 1922

Chelorhinus Kleine, 1922o: 92.

Epigogus Kleine, 1923e: 159; syn. Damoiseau 1965a: 16.

Pseudopisthenoxys Kleine, 1942: 130; syn. Damoiseau, 1979a: 5.

Type species: *Chelorhinus propheti* Kleine, 1922.

Distribution. Taiwan; Myanmar [= Burma], Philippines (Basilan, Mindanao, Negros).

flexibilis (Kleine, 1923)

flexibilis Kleine, 1923e: 159 (*Epigogus*).

insculptus Kleine, 1942: 130 (*Pseudopisthenoxys*); syn. Damoiseau, 1979a: 5.

Type depositories. *E. flexibilis*: SMTD (ST); *P. insculptus*: NHM (HT!), unknown (PT).

Distribution. Myanmar [= Burma], Philippines (Basilan, Mindanao, Negros).

Notes. Damoiseau (1989a: 115) reported that the specimens of *E. flexibilis* held in SMTD are holotype, allotype and paratypes, but in his original description Kleine did not designate any of the syntypes as holotype.

propheti Kleine, 1922

propheti Kleine, 1922a: 93.

Type depository. DEI (HT!).

Distribution. Taiwan.

Cormopus Kolbe, 1892

Cormopus Kolbe, 1892: 166.

Type species: *Cormopus penicillifer* Kolbe, 1892.

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Ethiopia, Gabon, Ghana, Ivory Coast, Nigeria, Republic of the Congo, Republic of Guinea, Rwanda, Tanzania, Togo, Zambia.

aberrans Kolbe, 1916

aberrans Kolbe, 1916: 63.

Type depositories. MNHUB (LT!), IRSNB (?) (PLT).

Distribution. Cameroon.

Notes. Damoiseau (1967a: 31) reported that the specimen held in MNHUB is the holotype and the one held in IRSNB is a paratype, but in his original description Kolbe did not designate any of the syntypes as holotype; however, as there is only one syntype in MNHUB, this must be considered as lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999). We did not find any paralectotype in IRSNB. The lectotype has the following labels: Kamerun, Joh.-Albrechshöhe, 21.XI-28.XII.98, L. Conradt/ Type/ *penicillifer* var. *aberrans* Kolbe.

distinctus Kleine, 1916

distinctus Kleine, 1916c: 17.

clavatus Kleine, 1921l: 59 (*Cormopus*); syn. Damoiseau, 1962b: 11.

Type depositories. *C. distinctus*: DEI (LT!, PLT!); MRAC (PLT!); *C. clavatus*: SMFD (HT).

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Ivory Coast, Zambia.

Nota: According to Damoiseau (1962b: 11) *C. clavatus* is a synonym of *C. distinctus*, but according to Quentin (1970: 207) *C. clavatus* is a synonym of *C. simplex*; here we follow Damoiseau's opinion.

Lectotype of *C. distinctus* (DEI) designated on a specimen with the following labels: N Kamerun/ L. Conradt, 96/ ♂/ Coll. Kraatz/ Kleine det./ *Cormopus distinctus* n. sp./ Dtsch. Entomol. Institut Berlin/ *Cormopus distinctus* ♂ Kleine, R. Damoiseau vid., 1961. Paralectotypes: 1 specimen (MRAC), Paratypus/ Musée du Congo, N Kamerun (Conradt), D. Ent. Nat. Mus., ex Coll. Kraatz/ Kleine det./ Coll. Kraatz/ N Kamerun, L. Conradt, '96/ Paratyp./ R. det. D. 1331/ *Cormopus distinctus* Kleine cotype!; 2 male and 1 female specimens (DEI), with the same label data as the lectotype. A further paralectotype (DEI) is actually a female of *Cormopus penicillifer*.

diversitarsis Kleine, 1916

diversitarsis Kleine, 1916c: 16.

diversitarisis: Damoiseau, 1962b: 11 (*Cormopus*) [lapsus].

Type depository. DEI (LT!, PLT!).

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Republic of the Congo, Republic of Guinea, Tanzania, Zambia.

Notes. Damoiseau (1967a: 35) reported that the holotype is in MNHUB and the allotype in MNHN, but Kleine did not designate any of the syntypes, which are held in DEI, as holotype.

Lectotype of *C. diversitarsis* (DEI) designated on a specimen with the following labels: N Kamerun/ L. Conradt, 96/ ♂/ Coll. Kraatz/ Dtsch. Entomol. Institut Berlin/ Kleine det./ Syntypus/ siehe mikroskop. Präparate/ *Cormopus diversitarsis* Kl./ *Cormopus diversitarsis* ♂ Kleine R., Damoiseau vid., 1961. Paralectotype: 1 female specimen (DEI), with the same label data as the lectotype.

diversus Kleine, 1926

diversus Kleine, 1926f: 360.

Type depositories. NHM (LT!, PLT!), IRSNB (PLT!), SMTD (?) (PLT).

Distribution. Tanzania, Zambia.

Notes. Damoiseau (1967a: 31) reported that the specimens held in NHM are holotype and allotype, and that paratypes are in NHM and SMTD, but in his original description Kleine did not designate any of the 11 syntypes as holotype; however, as there is more than one syntype in NHM, and as Damoiseau did not specifically select one as holotype, we designate the lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Lectotype of *C. diversus* (NHM) designated on a specimen with the following labels: Syntype/ N. W. Rhodesia: Kashitu [actually in Zambia], I.1915, H. C. Dollman. 1919-79/ *Cormopus diversus* Kleine Typus ♂ R. Kleine det. 1923. Paralectotypes: 1 female specimen (NHM), Syntype/ N. W. Rhodesia, Kashitu, XI.1914, H. C. Dollman. 1919-79/ *Cormopus diversus* Kleine Typus ♀ R. Kleine det. 1923; 1 specimen (NHM), Syntype/ N. W. Rhodesia, Kashitu, I.1915, H. C. Dollman. 1919-79/ R. Damoiseau det., 1964 *Cormopus diversus* Kleine; 5 specimens (NHM), Syntype/ N. W. Rhodesia, Kashitu, I.1915, H. C. Dollman. 1919-79; 1 male specimen (IRSNB), N. Rhodesia, Kashitu, I.1915, H. C. Dollman/ H. C. Dollman Coll. 1919-79/ Staatl. Museum für Tierkunde Dresden/ R. Damoiseau det. 1961 *Cormopus diversus* Kleine/ R. I. Sc. N. B., I. G. 23.643/ Paratype.

Damoiseau (1967a: 31) reported paratypes of *C. diversus* in SMTD; according to O. Jaeger (SMTD) (2002, personal communication) two specimens of the species are present in the SMTD collection, but they are not labelled as types.

filum Kolbe, 1916 stat. res.

filum Kolbe, 1916: 64.

simplex Kleine, 1916c: 17 (*Cormopus*); syn. De Muizon, 1960: 37.

Type depositories. *C. filum*: MNHUB (HT!); *C. simplex*: DEI (LT!, PLT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Gabon, Ivory Coast, Tanzania, Zambia.

Notes. Damoiseau (1967a: 40) reported that the specimens of *C. simplex*

held in DEI are holotype and allotype, but in his originale description Kleine did not designate any of the syntypes as holotype; however, as there is only one male syntype in DEI, this must be considered as lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999). The lectotype has the following labels: Kamerun Conradt/ Syntypus/ Type/ ♂/ Coll. Kraatz/ Kleine det./ Dtsch. Entomol. Institut Berlin/ *Cormopus simplex* n. sp/ *Cormopus simplex* ♂ Kleine, R. Damoiseau vid., 1961.

***guineensis* Damoiseau, 1967**

guineensis Damoiseau, 1967a: 38.

Type depository. MNHN (?) (HT).

Distribution. Cameroon, Republic of Guinea.

Notes. According to Damoiseau (1967a: 39) the holotype is in MNHN, but we did not find it there.

***mimus* Kleine, 1921**

mimus Kleine, 1921: 59.

Type depository. MNHUB (HT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire).

***penicillifer* Kolbe, 1892**

penicillifer Kolbe, 1892: 167.

edentatus Kleine, 1914b: 170 (*Cormopus*); syn. Kleine, 1916c: 19.

pennicillifer: Kleine, 1914b: 170 (*Cormopus*) [lapsus].

dilutus Kleine, 1944: 149 (*Cormopus*); syn. De Muizon, 1955a: 458.

Type depositories. *C. penicillifer*: MNHUB (?) (HT); *C. edentatus*: MZPW (ST); *C. dilutus*: MNHN (HT!, PT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Ethiopia, Gabon, Ghana, Ivory Coast, Nigeria, Republic of Guinea, Rwanda, Tanzania, Zimbabwe.

Notes. Damoiseau (1967a: 29) incorrectly reported that holotype and allotype of *C. penicillifer* are in MNHUB and paratypes in MNHUB and IRSNB, but the species description was based on a single specimen. We did not find it in MNHUB.

Damoiseau (1967a: 29) did not mention S Rhodesia [= Zimbabwe] (Kleine, 1936a: 12) when he gave the species distribution.

pumilio Kolbe, 1916

pumilio Kolbe, 1916: 65.

Type depository. MNHUB (HT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Nigeria.

Cyphagogus Parry, 1849

Cyphagogus Parry, 1849: 182.

Nardodes Motschulsky, 1866: 430; syn. Alonso-Zarazaga & Lyal, 1999: 51.

Cyphagonus: Desbrochers des Loges, 1890: 221 [lapsus].

Chypagogus: Kleine, 1914b: 162 [lapsus].

Xyphagogus: Kleine, 1933c: 289 [lapsus].

Arthrepigogus Damoiseau, 1964c: 491; syn. Alonso-Zarazaga & Lyal, 1999: 51.

Type species: *Cyphagogus westwoodii* Parry, 1849.

Distribution. S China, Japan, Taiwan; Madagascar; Bangladesh, Borneo, Bhutan, India (including Andaman Is., Assam, Darjeeling, Meghalaya), Indonesia (Java; Engano; Mentawai; Moluccas: Aru, Bacan [= Batjan], Seram [= Ceram], Ternate; Nias; Sangihe [= Sangir]; Sulawesi; Sumatra), Laos, Malaysia, Myanmar [= Burma], Philippines (Alabat, Basilan, Iligan, Luzon, Mindanao, Mindoro, Negros, Palawan, Panay, Samar, Sibuyan), Singapore, Sri Lanka, Thailand, Vietnam; Australia (including Tasmania), Fiji Is., New Guinea (including Bismarck Archipelago: New Britain), New Hebrids, Samoa, Solomon Is.; Europe (I).

Notes. According to Zimmerman (1994: 139) all species of *Cyphagogus* known in Australia and Tasmania are believed to be immigrants or introductions through the aid of commerce.

***angusticeps* Senna, 1893**

angusticeps Senna, 1893e: 296.

Type depositories. MZUF (LT!), SMTD (PLT).

Distribution. India, Indonesia (Java, Sumatra), Malaysia; Europe (I).

Notes. Lectotype designated by Bartolozzi *et al.* (1985: 94) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Following Damoiseau (1989a: 148) the paralectotype in SMTD is actually a specimen of *C. tabacicola*.

***angustirostris* Damoiseau, 1964**

angustirostris Damoiseau, 1964c: 494.

Type depository. RMNH (HT!).

Distribution. Indonesia (Sulawesi).

***aulacothorax* Damoiseau, 1989**

aulacothorax Damoiseau, 1989a: 154.

Type depository. MNHN (HT!).

Distribution. Indonesia (Java).

***bakeri* Damoiseau, 1989**

bakeri Damoiseau, 1989a: 152.

Type depositories. NMNH (HT, PT), IRSNB (PT!).

Distribution. Philippines (Luzon, Negros).

***beesoni* Damoiseau, 1964**

beesoni Damoiseau, 1964c: 484.

Type depository. SMTD (HT, AT).

Distribution. Borneo (?), India.

bimaculatus Damoiseau, 1989

bimaculatus Damoiseau, 1989a: 134.

Type depository. MONZ (HT).

Distribution. New Guinea.

bipunctatus Senna, 1897

bipunctatus Senna, 1897a: 227.

Type depository. IRSNB (LT!, PLT!).

Distribution. Australia.

Notes. Lectotype of *C. bipunctatus* (IRSNB) designated on a specimen with the following labels: Australia – N. S. W., Kyama/ Coll. Castelnau, Coll. Roelofs/ *Cyphagogus bipunctatus* Senna Cotypi! ♂ ♀/ Prépar. génit. No. 63 A 01 R. Damoiseau/ R. Damoiseau det., *Cyphagogus bipunctatus* Senna/ Lectotype [*in litteris*]. Paralectotypes: 1 male specimen (IRSNB), Australia – N. S. W., Kyama/ Coll. Castelnau, Coll. Roelofs/ *Cyphagogus bipunctatus* Pow./ Prépar. génit. No. 63 A 02 R. Damoiseau/ R. Damoiseau det., *Cyphagogus bipunctatus* Senna/ Paralectotype [*in litteris*]; 2 specimens (IRSNB), Australia – N. S. W., Kyama/ Coll. Castelnau, Coll. Roelofs/ R. Damoiseau det., *Cyphagogus bipunctatus* Senna/ Paralectotype [*in litteris*]. [According to Damoiseau (1964c: 477) the fifth paralectotype is actually a specimen of *C. delicatus*].

bishopi Damoiseau, 1989

bishopi Damoiseau, 1989a: 136.

Type depository. BPBM (HT).

Distribution. Borneo.

brownei Kleine, 1936

brownei Kleine, 1936c: 134.

Type depository. NHM (HT!).

Distribution. Malaysia, Myanmar [= Burma].

Notes. According to Damoiseau (1979a: 5; 1989a: 114) this species is a synonym of *Chelorhinus flexibilis*, but it is also considered as valid in the same paper (1989a: 133)!

***cheesmanae* Damoiseau, 1989**

cheesmanae Damoiseau, 1989a: 135.

Type depository. NHM (HT!).

Distribution. New Guinea.

***conciliatus* Kleine, 1926**

conciliatus Kleine, 1926a: 8.

Type depository. NHM (HT!).

Distribution. Borneo, Malaysia.

***confertulus* Kleine, 1925**

confertulus Kleine, 1925b: 125.

fragosus Kleine, 1925b: 126 (*Cyphagogus*); syn. Damoiseau, 1989a: 151.

Type depositories. *C. confertulus*: NHM (LT!), NME (PLT), NZSI (PLT), SMTD (PLT); *C. fragosus*: NZSI (ST).

Distribution. S China; Bhutan, India (Andaman Is., Assam), Myanmar [= Burma], Vietnam.

Notes. Damoiseau (1989a: 152) reported that the specimen of *C. confertulus* held in NHM is the holotype, but in his original description Kleine did not designate any of the syntypes as holotype; however, as there is only one syntype in NHM, this must be considered as lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999). The lectotype has the following labels: Type/ Typus! 64414/ Fry Coll. 1905.100/ Doherty/ *Cyphagogus confertulus* Kln., R. Kleine det. 1922.

***confidens* Kleine, 1925**

confidens Kleine, 1925b: 126.

Type depository. NHM (HT!).

Distribution. Thailand, Vietnam.

***corporaali* Kleine, 1918**

corporaali Kleine, 1918i: 177.

Type depository. ZMAN (HT!).

Distribution. Borneo, India, Indonesia (Java), Malaysia, Myanmar [= Burma], Sri Lanka.

***crassicollis* Damoiseau, 1989**

crassicollis Damoiseau, 1989a: 137.

Type depositories. MSNG (HT!, AT!), IRSNB (PT!), SAMA (PT).

Distribution. New Guinea.

***crassitarsus* Damoiseau, 1964**

crassitarsus Damoiseau, 1964c: 480.

Type depository. MMA (HT).

Distribution. Australia.

Notes. The IRSNB holds a specimen (labelled: C. G. Oke collection/ [no locality]/ MMA) labelled as paratype, but this paratype is not mentioned in the original description.

***curvitibia* Damoiseau, 1989**

curvitibia Damoiseau, 1989a: 139.

Type depository. MNHN (HT!).

Distribution. Borneo.

cyrtotrachelus (J. Thomson, 1857)

cyrtotrachelus J. Thomson, 1857: 119 (*Caladromus*) (sic!).

buccatus Kleine, 1916c: 9 (*Cyphagogus*); syn. Damoiseau, 1964c: 496.

Type depositories. *C. cyrtotrachelus*: MNHN (LT!, PLT!); *C. buccatus*: DEI (LT!, PLT!), NHRS (PLT).

Distribution. Borneo, India (including Andaman Is., Assam), Indonesia (Java, Sumatra), Malaysia, Myanmar [= Burma], Philippines (Mindanao, Samar), Singapore, Sri Lanka.

Notes. *C. cyrtotrachelus* was described on two specimens. We found both syntypes in MNHN, but they belong to two different species. Damoiseau (1964c: 498) incorrectly reported that the holotype is in MNHN and the allotype in DEI; however, as there are two syntypes in MNHN, and as Damoiseau did not specifically select one as holotype, we designate the lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Lectotype of *C. cyrtotrachelus* (MNHN) designated on a specimen with the following labels: Borneo/ Thomson type 113/ ex Musaeo G. Power/ Museum Paris 1952 coll. R. Oberthür. Paralectotype: 1 specimen (MNHN), with the same labels.

Lectotype of *C. buccatus* (DEI) designated on a specimen with the following labels: N.O. Sumatra, Tebing-tinggi, Coll. Schultheiss/ Coll. Schultheiss/ Coll. Kraatz/ Syntype/ Dtsch. Entomol. Institut Berlin/ *Cyphagogus buccatus* Kln., R. Damoiseau det., 1962. Paralectotype: 1 specimen (DEI), with the same label data as the lectotype.

delicatus Lea, 1899

delicatus Lea, 1899: 634.

hauseri Kleine, 1924b: 33 (*Cyphagogus*); syn. Damoiseau, 1989a: 122.

Type depositories. *C. delicatus*: SAMA (Lea coll.) (ST); *C. hauseri*: MNHUB (HT!).

Distribution. Australia (including Tasmania), New Guinea.

Notes. Damoiseau (1989a: 123) reported that the specimens of *C. delicatus* held in SAMA are holotype, allotype and paratypes, but in his original description Lea did not designate any of the syntypes as holotype.

***densepunctatus* Kleine, 1916**

densepunctatus Kleine, 1916c: 4.

Type depository. DEI (HT!).

Distribution. Borneo, Malaysia, Singapore.

***dieckmanni* Damoiseau, 1967**

dieckmanni Damoiseau, 1967c: 1.

Type depositories. DEI (HT!), IRSNB (PT!), MNHUB (PT!), MSNG (PT!), NHM (PT!), SMTD (PT), ZSM (PT).

Distribution. Borneo, Indonesia (Sumatra), Malaysia, Sri Lanka.

***eggersi* Kleine, 1921**

eggersi Kleine, 1921e: 324.

raptor Kleine, 1936c: 136 (*Cyphagogus*); syn. Damoiseau, 1989a: 145.

Type depositories. *C. eggersi*: MNHUB (HT!); *C. raptor*: NHM (HT!).

Distribution. Borneo, Indonesia (Sumatra), Laos, Malaysia; Australia (I).

***eichhorni* Kirsch, 1875**

eichhorni Kirsch, 1875b: 45.

erichsoni: Lewis, 1883: 297 (*Cyphagogus*) [lapsus].

longisetosus Kleine, 1916c: 11 (*Cyphagogus*); syn. Damoiseau, 1964c: 498.

humilis Kleine, 1925c: 590 (*Cyphagogus*); syn. Damoiseau, 1989a: 142.

dissociabilis Kleine, 1926a: 8 (*Cyphagogus*); syn. Damoiseau, 1989a: 142.

Type depositories. *C. eichhorni*: SMTD (ST); *C. longisetosus*: DEI (HT!); *C. humilis*: MPMP (HT); *C. dissociabilis*: NHM (ST!).

Distribution. Borneo, India (including Andaman Is., Assam), Indonesia (Mentawai, Moluccas, Sumatra), Malaysia, Myanmar [= Burma], Philippines (Iligan, Luzon, Mindanao, Negros, Panay, Sibuyan), Thailand, Vietnam; New Guinea.

elongatus Kleine, 1916

elongatus Kleine, 1916c: 5.

Type depository. DEI (LT!, PLT!).

Distribution. Borneo, Indonesia (Sumatra?); New Guinea.

Notes. According to Damoiseau (1964c: 498) one of the two syntypes belongs to a different species; later Damoiseau (1989a: 139) stated that the holotype is in DEI. Even if Kleine did not designate any of the syntypes as holotype, on the basis of article 74.5 of the International Commission of Zoological Nomenclature (1999) the specimen that Damoiseau gives as holotype must be considered as the lectotype.

According to Damoiseau (1989a: 139) the presence of this species in Sumatra is extremely doubtful.

fijianus Kleine, 1928

fijianus Kleine, 1928b: 48.

Type depositories. BPBM (HT), SMTD (PT).

Distribution. Fiji Is:

gemellus Kleine, 1931

gemellus Kleine, 1931a: 93.

Type depository. IFRI (HT, PT).

Distribution. India.

gracilirostris Damoiseau, 1964

gracilirostris Damoiseau, 1964c: 473.

Type depositories. MNHUB (HT!), IRSNB (PT!).

Distribution. Borneo, Philippines (Mindanao, Palawan).

***greensladi* Damoiseau, 1989**

greensladi Damoiseau, 1989a: 154.

Type depository. NHM (HT!).

Distribution. Solomon Is.

***hornabrooki* Damoiseau, 1989**

hornabrooki Damoiseau, 1989a: 153.

Type depository. MONZ (HT).

Distribution. New Guinea.

***incisus* Senna, 1903**

incisus Senna, 1903: 153.

obconiceps Senna, 1903: 154 (*Cyphagogus*); syn. Damoiseau, 1989a: 144.

Type depositories. *C. incisus*: MSNG (HT!); *C. obconiceps*: MSNG (HT!).

Distribution. Borneo, India (including Andaman Is., Assam, Darjeeling), Indonesia (Java, Sumatra), Malaysia, Vietnam.

***indicus* Damoiseau, 1964**

indicus Damoiseau, 1964c: 486.

Type depository. SMTD (HT).

Distribution. India.

iwatensis Morimoto, 1976

iwatensis Morimoto, 1976a: 268.

Type depository. ELKU (HT, PT).

Distribution. Japan.

javanus Kleine, 1916

javanus Kleine, 1916c: 13.

Type depositories. RMNH (LT, PLT), MZPW (PLT), DEI (PLT!).

Distribution. Indonesia (Java).

Notes. Lectotype and paralectotypes designated by Damoiseau (1989a: 151). According to Damoiseau (1964c: 494) the paralectotype held in DEI is actually *C. longulus*.

longulus Senna, 1898

longulus Senna, 1898d: 52.

simulator Senna 1903: 155 (*Cyphagogus*); syn. Damoiseau, 1964c: 494.

Type depositories. *C. longulus*: RMNH (HT); *C. simulator*: MSNG (LT!, PLT!).

Distribution. Bhutan, Borneo, India (Andaman Is., Assam), Indonesia (Java; Moluccas: Bacan [= Batjan], Seram [= Ceram]; Sumatra), Malaysia, Philippines (Basilan, Luzon, Mindanao), Thailand, Vietnam.

Notes. Lectotype of *C. simulator* (MSNG) designated on a specimen with the following labels: Sumatra, Si Rambé, XII.90-III.1991, E. Modigliani/*Cyphagogus simulator* cotypi! Senna. Paralectotypes: 3 specimens (MSNG), Sumatra, Si Rambé, XII.90-III.1991, E. Modigliani.

madagascariensis Damoiseau, 1964

madagascariensis Damoiseau, 1964c: 489.

Type depository. MRAC (HT!).

Distribution. Madagascar.

***malabaricus* Damoiseau, 1964**

malabaricus Damoiseau, 1964c: 488.

Type depository. SMFD (HT).

Distribution. India.

***malaysianus* Damoiseau, 1989**

malaysianus Damoiseau, 1989a: 134.

Type depository. MHNG (HT).

Distribution. Malaysia.

***modiglianii* Senna, 1893**

modiglianii Senna, 1893b: 258.

modigliani: Kleine, 1914b: 163 (*Cyphagogus*) [lapsus].

Type depositories. MSNG (LT!, PLT!), MNHN (PLT!), MZUF (PLT!).

Distribution. Taiwan; Borneo, Indonesia (Engano, Java, Sumatra), Malaysia, Philippines (Basilan); Australia, New Guinea.

Notes. Damoiseau (1989a: 127) reported that the specimens held in MSNG are holotype and allotype and those held in MZUF as paratypes, but in his original description Senna did not designate any of the syntypes as holotype; however, as there is more than one syntype in MSNG, and as Damoiseau did not specifically select one as holotype, we designate the lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Lectotype of *C. modiglianii* (MSNG) designated on a male specimen with the following labels: Engano, Bua Bua V.VI.1891 Modigliani/ *Cyphagogus Modiglianii* Senna typus! Paralectotypes: 2 specimens (MSNG), Engano, Malaconni, VI.1891, Modigliani; Engano, Bua Bua, V.VI.1891, Modigliani; 1 female specimen (MZUF), Bua Bua, V-VI.1891, Modigliani/ *Cyphagogus modiglianii* Senna/ Ann. Mus. Genova, XXXIII, 1893, Typi!! *Cyphagogus modiglianii* Senna ♀, R. Damoiseau, det., 1966/ ex-coll. Senna/ "La Specola" Firenze, 6912; 1 male specimen (MZUF), Bua Bua, V-VI.1891, Modigliani/ *Cyphagogus modiglianii* Senna ♂/ Prépar. génit. n° Senna 1, R. Damoiseau/

ex-coll. Senna/ "La Specola" Firenze, 6912; 1 male specimen (MNHN), Engano, Bua Bua, V-VI.1891, Modigliani.

***moluccanus* Damoiseau, 1989**

moluccanus Damoiseau, 1989a: 154.

Type depository. MNHN (HT!).

Distribution. Indonesia (Moluccas: Ternate).

***nigraustralis* Damoiseau, 1966**

nigraustralis Damoiseau, 1966d: 8.

Type depositories. NHMB (HT!), MMA (AT, PT), IRSNB (PT!), SAMA (PT).

Distribution. Australia, New Guinea.

***nodosus* Damoiseau, 1964**

nodosus Damoiseau, 1964c: 474.

Type depository. IRSNB (HT!, PT!).

Distribution. Laos, Vietnam.

***odewahnii* Pascoe, 1864**

odewahnii Pascoe, 1864: 46.

suspendiosus Lea, 1899: 635 (*Cyphagogus*); syn. Zimmerman, 1994: 147.

odewahni: Schönfeldt, 1908: 6 (*Cyphagogus*) [lapsus].

praecipuus Kleine, 1943b: 138 (*Cyphagogus*); syn. Damoiseau, 1964c: 478.

Type depositories. *C. suspendiosus*: SAMA (Lea coll.) (ST); *C. odewahnii*: NHM (HT!); *C. praecipuus*: HNHM (HT).

Distribution. Indonesia (Moluccas, Sulawesi), Philippines; Australia, New Guinea.

palawani Damoiseau, 1964

palawani Damoiseau, 1964c: 491.

Type depository. MNHUB (HT!).

Distribution. Philippines (Palawan).

Notes. The head is missing from the holotype specimen.

papuanus Damoiseau, 1989

papuanus Damoiseau, 1989a: 135.

Type depository. AMNH (HT).

Distribution. New Guinea.

pasteuri Damoiseau, 1989

pasteuri Damoiseau, 1989a: 155.

Type depositories. MZUF (HT!, AT!), IRSNB (PT!).

Distribution. Indonesia (Java).

pilosus Damoiseau, 1989

pilosus Damoiseau, 1989a: 136.

Type depository. IRSNB (HT!, AT!).

Distribution. New Guinea.

Notes. Damoiseau (1989a: 137) reported that the holotype is in CKS, but actually it is in IRSNB.

planifrons Kirsch, 1875

planifrons Kirsch, 1875b: 46.

gladiator Kleine, 1921e: 307 (*Cyphagogus*); syn. Damoiseau, 1989a: 120.

Type depositories. *C. planifrons*: SMTD (HT); *C. gladiator*: SMTD (HT, PT), NHRS (PT).

Distribution. S China; Bhutan, Borneo, India (including Assam), Indonesia (Java, Mentawai, Nias, Sumatra), Laos, Malaysia, Philippines (Basilan, Luzon, Mindanao, Negros, Samar), Thailand, Vietnam.

rubidus Damoiseau, 1967

rubidus Damoiseau, 1967c: 3.

Type depository. NHM (HT!).

Distribution. Myanmar [= Burma], Vietnam (?).

rufirostris Kleine, 1914

rufirostris Kleine, 1914b: 162.

rufifrons: Kleine, 1922c: 41 (*Cyphagogus*) [lapsus].

Type depository. MZPW (ST).

Distribution. Philippines, Sri Lanka.

Notes. Damoiseau (1989a: 147) reported that the specimens held in MZPW are holotype and paratypes, but in his original description Kleine did not designate any of the syntypes as holotype.

rugaticollis Damoiseau, 1989

rugaticollis Damoiseau, 1989a: 155.

Type depository. MNHN (HT!, AT!).

Distribution. New Guinea.

samoanus Kleine, 1928

samoanus Kleine, 1928c: 157.

Type depositories. NHM (LT!), SMTD (PLT).

Distribution. Samoa.

Notes. Damoiseau (1989a: 127) reported that the holotype is in NHM and the allotype in SMTD, but in his original description Kleine did not designate one of the syntypes as holotype; however, as there is only one syntype in NHM, this must be considered as lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999). The lectotype has the following labels: Syntype/ Type/ Lectotype [*in litteris*]/ ♂/ Samoa Is., Apia/ III.1924, P. A. Buxton and G. H. Hopkins/ pres. by Imp. Bur. Ent. Brit. Mus. 1927-294/ see slide Coll./ *Cyphagogus samoanus* Kln. Typus! R. Kleine det. 1926/ *Cyphagogus samoanus* Kleine lect. ♂, R. Damoiseau rev. 1965.

According to Damoiseau (1989a: 128) and Zimmerman (1994: 146) this species could be synonymous of *C. modiglianii*.

sarasini Senna, 1899

sarasini Senna, 1899: 302.

frugalis Kleine, 1925f: 16 (*Cyphagogus*); syn. Damoiseau, 1964c: 493.

Type depositories. *C. sarasini*: NHMB (HT!); *C. frugalis*: ZMAN (HT!).

Distribution. Indonesia (Moluccas: Buru, Ternate; Misool; Sangihe [=Sangir]; Sulawesi); New Guinea.

scissicollis Damoiseau, 1989

scissicollis Damoiseau, 1989a: 139.

Type depositories. NMNH (HT, AT), IRSNB (PT!).

Distribution. Borneo, Philippines (Mindanao).

scutellatus Damoiseau, 1964

scutellatus Damoiseau, 1964c: 471.

Type depositories. SMTD (HT, PT), IRSNB (PT!).

Distribution. Philippines (Alabat, Luzon, Mindoro).

signipes Lewis, 1883

signipes Lewis, 1883: 297.

suaviter Kleine, 1916c: 2 (*Cyphagogus*); syn. Damoiseau, 1989a: 140.

segnipes: Sharp, 1936: 296 (*Cyphagogus*) [lapsus].

suavifer: Li-zhong, 2002: 296 [lapsus].

Type depositories. *C. signipes*: NHM (LT!, PLT!), MNHN (PLT!); *C. suaviter*: DEI (HT!).

Distribution. China (Yunnan), Japan, Taiwan.

Notes. Damoiseau (1989a: 141) reported that the specimens of *C. signipes* held in NHM are holotype and allotype, but in his original description Lewis did not designate any of the syntypes as holotype; however, as there is more than one syntype in NHM, and as Damoiseau did not specifically select one as holotype, we designate the lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Lectotype of *C. signipes* (NHM) designated on a specimen with the following labels: syntype/ type/ Japan, G. Lewis, 1910-320/ Nagasaki, 22.V.-3.VI.81/ *Cyphagogus signipes* Lewis ♂ Type/ R. Damoiseau rev., 1965 *Cyphagogus signipes* Lewis. Paralectotypes: 1 female specimen (NHM), syntype/ Japan, G. Lewis, 1910-320/ Nagasaki, 22.V.-3.VI.81/ *Cyphagogus signipes* Lewis ♀ Type; 10 specimens (NHM), syntype/ Japan, G. Lewis, 1910-320; 1 specimen (NHM), syntype/ Japan, G. Lewis, 1910-320/ Japan/ Andrewes Bequest. B. M. 1922-221/ *Cyphagogus signipes* Lewis; 1 specimen (NHM), *Cyphagogus signipes* Lewis, syntype, Japan, 1.VI.1881, G. Lewis/ syntype/ Japan/ *Cyphagogus signipes* Lewis/ Sharp Coll. 1905-313; 3 specimens (NHM), syntype/ Japan, G. Lewis, 1910-320/ Kashiwagi, 15.VI.-24.VI.81; 1 specimen (NHM), syntype/ Japan, G. Lewis, 1910-320/ Kashiwagi, 15.VI.-24.VI.81/ *Cyphagogus signipes* Lewis; 1 specimen (NHM), syntype/ Higo, 1882/ Japan, G. Lewis, 1910-320.

silvanus Senna, 1903

silvanus Senna, 1903: 154.

Type depository. MSNG (LT!, PLT!).

Distribution. Borneo, India, Indonesia (Java; Moluccas: Buru; Mentawai; Sumatra), Malaysia, Philippines (Basilan, Luzon); New Guinea.

Notes. Damoiseau (1989a: 149) reported that a specimen held in MSNG is the holotype, but in his original description Senna did not designate any of the

syntypes as holotype; however, as there is more than one syntype in MSNG, and as Damoiseau did not specifically select one as holotype, we designate the lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Lectotype of *C. silvanus* (MSNG) designated on a specimen with the following labels: Sumatra, Si Rambé, XII.90-III.1991, E. Modigliani/ *Cyphagogus silvanus* cotypi! Senna. Paralectotypes: 3 specimens (MSNG), Sumatra, Si Rambé, XII.90-III.1991, E. Modigliani.

splendens Kleine, 1916

splendens Kleine, 1916c: 7.

mundus Kleine, 1941a: 131 (*Cyphagogus*); syn. Damoiseau, 1989a: 141.

Type depositories. *C. splendens*: MZPW (LT, PLT), DEI (PLT!), SAMA (PLT), SMTD (PLT); *C. mundus*: MNHN (HT!).

Distribution. Indonesia (Moluccas: Aru, Bacan [= Batjan], Seram [= Ceram]; Sulawesi), Philippines (Mindanao); New Guinea (including Bismarck Archipelago: New Britain), New Hebrids, Solomon Is.

Notes. Lectotype and paralectotypes designated by Damoiseau (1989a: 142).

tabacicola Senna, 1893

tabacicola Senna, 1893e: 294.

Type depositories. MZUF (LT!), IRSNB (PLT!), MSNG (PLT!), RMNH (PLT), SMTD (PLT).

Distribution. Bangladesh, Borneo, India (including Andaman Is., Assam), Indonesia (Java, Nias, Sumatra), Malaysia, Myanmar [= Burma], Philippines (Basilan, Luzon, Mindanao), Vietnam; Europe (I).

Notes. Lectotype designated by Bartolozzi *et al.* (1985: 101) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

thoracicus Kleine, 1933

thoracicus Kleine, 1933d: 299.

Type depository. ZMAN (?) (HT).

Distribution. Borneo, Indonesia (Java).

Notes. Damoiseau (1989a: 146) reported that the holotype could be in ZMAN, but we did not find it there. According to H. E. Bomans (1995, personal communication) and B. Brugge (ZMAN) (2002, personal communication) some types of Drescher's collection could be in MZB.

tristriatus Kleine, 1916

tristriatus Kleine, 1916c: 16 (*Cyphagogus delicatus* f.).

Type depositories. DEI (LT!, PLT!), ZMAN (PLT!).

Distribution. Australia, New Guinea.

Notes. Damoiseau (1989a: 124) reported that the holotype and the allotype are in DEI and paratypes are in IRSNB and SAMA. Kleine (1916c: 16) described the species on three syntypes, two in DEI and one in ZMAN. Döbler & Gaedike (1995) forgot to mention these syntypes in the DEI type catalogue. As there are two syntypes in DEI, and as Damoiseau did not specifically select one as holotype, we designate the lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Lectotype of *C. delicatus* f. *tristriatus* (DEI) designated on a specimen with the following labels: Coen, 26.01.06/ Coen District, Cape York, Queensland, H. Hacker/ Type/ coll. Hacker/ Kleine det./ Dtsch. Entomol. Institut Berlin/ forma *tristriata* Kleine/ *Cyphagogus delicatus* Lea, det. Kleine., 1915/ *Cyphagogus tristriatus* Kln, R. Damoiseau det., 1962. Paralectotypes: 1 specimen (DEI), Coen District, Cape York, Queensland, H. Hacker/ Paratype/ coll. Hacker/ Kleine det./ Dtsch. Entomol. Institut Berlin/ *Cyphagogus delicatus* f. *tristriata* Kl./ *Cyphagogus tristriatus* Kln, R. Damoiseau det., 1962; 1 specimen (ZMAN), Z. Nieuw Guinea, Versteeg, 1912.13, 27.II.12, cotypus.

undulatus Damoiseau, 1964

undulatus Damoiseau, 1964c: 487.

Type depository. SMTD (HT).

Distribution. India.

***westwoodii* Parry, 1849**

westwoodii Parry, 1849: 182.

westwoodi: Schönfeldt, 1908: 6 (*Cyphagogus*) [lapsus].

Type depository. NHM (HT!).

Distribution. Bangladesh, Bhutan, Borneo, India (including Assam), Indonesia (Java, Mentawai, Nias, Sumatra), Laos, Malaysia, Myanmar [= Burma], Philippines (Negros), Sri Lanka, Thailand, Vietnam.

***whitii* (Westwood, 1848)**

whitii Westwood, 1848: 32 (*Taphroderes*).

whitei: Senna, 1898a: 224 (*Cyphagogus*) [lapsus].

Type depository. NHM (HT!).

Distribution. Philippines (Luzon, Mindanao).

Notes. The head is missing from the holotype specimen. Senna (1898a: 224) mentioned the locality Mentawai, but his quotation was based on a misidentification as the species was *C. westwoodi* (Senna, 1898, manuscript correction on the reprint).

***wittmeri* Damoiseau, 1989**

wittmeri Damoiseau, 1989a: 137.

Type depository. NHMB (HT!).

Distribution. India (Meghalaya).

***Dentisebus* De Muizon, 1960**

Dentisebus De Muizon, 1960: 72.

Microsebus Kleine, 1938d: 35 (non Kolbe, 1892); syn. Alonso-Zarazaga & Lyal, 1999: 51.

Type species: *Microsebus dentipes* Kleine, 1922 = *Microsebus pusio* Kolbe, 1892.

Distribution. Democratic Republic of the Congo (ex Zaire), Equatorial Guinea, Gabon, Madagascar, Tanzania.

***fossicollis* (Kolbe, 1916)**

fossicollis Kolbe, 1916: 57 (*Autosebus*).

fossicollis: Kleine, 1936a: 18 (*Microsebus*) [lapsus].

Type depository. MNHUB (HT!).

Distribution. Democratic Republic of the Congo (ex Zaire), Equatorial Guinea.

***malgasicus* (Senna, 1895) comb. n.**

malgasicus Senna, 1895e: 290 (*Microsebus*).

Type depository. MNHN (HT!).

Distribution. Madagascar.

***pusio* (Kolbe, 1892)**

pusio Kolbe, 1892: 169 (*Microsebus*).

dentipes Kleine, 1922k: 172 (*Microsebus*); syn. Damoiseau, 1962f: 333.

Type depositories. *M. pusio*: MNHUB (?) (HT); *M. dentipes*: MNHUB (HT!).

Distribution. Democratic Republic of the Congo (ex Zaire), Equatorial Guinea, Gabon, Tanzania.

Notes. According to Damoiseau (1967a: 126) the holotype of *M. pusio* is in MNHUB, but we did not find it there.

Damoiseau (1967a: 126) quotes the species for Kenya (Usambara), but these mountains are actually in Tanzania.

***Diastrophocoleps* Damoiseau, 1967**

Diastrophocoleps Damoiseau, 1967a: 42.

Type species: *Cormopus femoralis* Damoiseau, 1961.

Distribution. Democratic Republic of the Congo (ex Zaire).

***femoralis* (Damoiseau, 1961)**

femoralis Damoiseau, 1961a: 278 (*Cormopus*).

Type depository. MRAC (HT!, AT!).

Distribution. Democratic Republic of the Congo (ex Zaire).

Notes. Damoiseau (1987a: 43) quotes some paratypes in IRSNB, but in his original description he only designated holotype and allotype in MRAC.

***Dyscheromorphus* Kleine, 1916**

Dyscheromorphus Kleine, 1916c: 60.

Atopomorphus Kleine, 1925c: 593; syn. Damoiseau, 1979a: 34.

Type species: *Dyscheromorphus helleri* Kleine, 1916.

Distribution. Borneo, Indonesia (Sumatra), Malaysia, Philippines (Negros).

***helleri* Kleine, 1916**

helleri Kleine, 1916c: 62.

Type depository. DEI (HT!).

Distribution. Borneo, Indonesia (Sumatra), Malaysia.

Notes. Damoiseau (1979a: 35) incorrectly reported that a paratype is in SMTD but in his original description Kleine only designated the holotype.

***schultzei* (Kleine, 1925)**

schultzei Kleine, 1925c: 594 (*Atopomorphus*).

Type depository. MPMP (HT).

Distribution. Philippines (Negros).

Ecnomobrentus Damoiseau, 1965

Ecnomobrentus Damoiseau, 1965a: 6.

Type species: *Ecnomobrentus afrosimulans* Damoiseau, 1965.

Distribution. Borneo, Indonesia (Java), Myanmar [= Burma], Vietnam.

afrosimulans Damoiseau, 1965

afrosimulans Damoiseau, 1965a: 7.

Type depositories. MNHN (HT!), MZPW (PT).

Distribution. Indonesia (Java), Vietnam.

bidens Kabakov, 2001

bidens Kabakov, 2001: 218.

Type depository. ZIN (HT, PT).

Distribution. Vietnam.

gracilis Damoiseau, 1979

gracilis Damoiseau, 1979a: 13.

Type depository. NHM (HT!).

Distribution. Borneo.

uncus Damoiseau, 1965

uncus Damoiseau, 1965a: 9.

Type depository. NHM (HT!).

Distribution. Myanmar [= Burma].

Eterozemus Senna, 1903

Eterozemus Senna, 1903: 160.

Dictyotopterus Kleine, 1916c: 75; syn. Damoiseau, 1963g: 37.

Dictiotopterus: Kleine, 1922b: 26 [lapsus].

Dictyotpterus: Kleine, 1925b: 44 [lapsus].

Dictyopterus: Kleine, 1937d: 24 (non Mulsant, 1838) [lapsus].

Orthopareia Kleine, 1925c: 591; syn. Damoiseau, 1963g: 37.

Sparganophasma Kleine, 1926a: 11; syn. Damoiseau, 1963g: 37.

Dictyoperus: Kano, 1930: 152 [lapsus].

Eterocemus: Kleine, 1940b: 90 [lapsus].

Distyotopterus: Damoiseau, 1963g: 6 [lapsus].

Heterozemus: Li-zhong, 2002: 296 [lapsus].

Type species: *Zemioses pubens* Senna, 1892.

Distribution. Japan, Taiwan; Borneo, India (including Assam), Indonesia (Java; Moluccas: Buru; Sumatra), Malaysia, Myanmar [= Burma], Philippines (Luzon, Mindanao, Tawi Tawi), Sri Lanka, Thailand, Vietnam; New Guinea, Solomon Is.

Notes. Kleine (1922m: 161) mentioned *Sparganophasma* in a list of genera, but he did not describe it until four years later (1926a: 11).

biguttatus (Kleine, 1921)

biguttatus Kleine, 1921m: 58 (*Neosebus*).

Type depository. RMNH (HT).

Distribution. Borneo.

carus (Kleine, 1926)

cara Kleine, 1926a: 12 (*Sparganophasma*).

Type depositories. ZMAN (LT!), NHRS (PLT).

Distribution. Borneo, India, Indonesia (Sumatra), Myanmar [= Burma], Sri Lanka, Vietnam.

Notes. Lectotypes designated by Damoiseau (1963g: 34). According to Kleine (1926a: 12) some syntypes are in NHM but we did not find them there.

Kleine in his original description quotes Borneo, Sumatra, Burma and Tonkin as species locality, but Damoiseau (1963g: 34; 1979a: 19) quotes only Sumatra.

celtis (Lewis, 1883)

celtis Lewis, 1883: 296 (*Zemioses*).

Type depository. NHM (LT!, PLT!).

Distribution. Japan.

Notes. Lectotype of *Z. celtis* (NHM) designated on a specimen with the following labels: Syntype/ type H. T./ Japan, G. Lewis, 1910-320/ Hitoyoshi, 15.V.-17.V.81/ *Zemioses celtis* Lewis type. Paralectotypes: 1 specimen (NHM), *Zemioses celtis* Lewis, Japan, 17.V.1881, Lewis/ Syntype/ Sharp Coll. 1905-313/ *Zemioses celtis* Lewis/ R. Damoiseau rev., 1967 *Eterozemus celtis* Lewis; 8 specimens (NHM), Syntype/ Japan, G. Lewis, 1910-320/ Hitoyoshi, 15.V.-17.V.81; 1 specimen (NHM), syntype/ Japan/ Pascoe coll. 93-60/ *Zemioses celtis* co-typi; 1 specimen (NHM), syntype/ Japan, G. Lewis, 1910-320/ Hitoyoshi, 15.V.-17.V.81/ *Zemioses celtis* Lewis/ R. Damoiseau rev., 1967 *Eterozemus celtis* Lewis.

deformis (Kleine, 1925)

deformis Kleine, 1925b: 127 (*Allaeometrus*).

coomani Kleine, 1941b: 226 (*Sparganophasma*); syn. Damoiseau, 1965a: 14.

Type depositories. *A. deformis*: NHM (LT!, PLT!), SMTD (PLT); *S. coomani*: MNHN (HT!).

Distribution. India (Assam), Vietnam.

Notes. Lectotype of *A. deformis* (NHM) designated on a male specimen with the following labels: Syntype/ type/ Assam, 15.V.1906/ Kachugaon/ 233/ India B. M. 1931-84/ *Allaeometrus deformis* ♂ Type, R. Kleine det., 1922. Paralectotype: 1 specimen (NHM), syntype/ type/ India, B. M. 1931-84/ *Allaeometrus deformis* ♀ Type, R. Kleine det., 1922.

fucatus (Kleine, 1925)

fucatus Kleine, 1925f: 17 (*Mesoderes*).

rufomaculatus Damoiseau, 1963g: 29 (*Eterozemus*); syn. Damoiseau, 1979a: 18.

Type depositories. *M. fucatus*: ZMAN (HT!, AT!); *E. rufomaculatus*: MSNG (HT!).

Distribution. Indonesia (Moluccas: Buru); New Guinea.

javanus Damoiseau, 1963

javanus Damoiseau, 1963g: 12.

Type depositories. RMNH (HT, AT, PT), IRSNB (PT!).

Distribution. Indonesia (Java).

laetus (Senna, 1892)

laetus Senna, 1892e: 441 (*Zemioses*).

Type depository. MSNG (LT!, PLT!).

Distribution. India, Indonesia (Java, Sumatra), Malaysia, Myanmar [= Burma], Philippines (Luzon).

Notes. Damoiseau (1963g: 20) reported that a specimen held in MSNG is the holotype, but in his original description Senna did not designate any of the syntypes as holotype; however, as there are two syntypes in MSNG, and as Damoiseau did not specifically select one as holotype, we designate the lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Lectotype of *Z. laetus* (MSNG) designated on a male specimen with the following labels: Birmania, Carin Ghecù, 1300-1400 m, L. Fea, II.III. 1888/ ♂/ R. Damoiseau det., 1963 *Eterozemus laetus* Senna Allotype ♂/ R. Damoiseau det., 1963 *Eterozemus laetus* (Senna). Paralectotype: 1 specimen (MSNG), Birmania, Carin Ghecù, 1300-1400 m, L. Fea, IV. 1888/ typus / *Zemioses laetus* Senna, typus! / *Eterozemus laetus* (Senna) ♀ R. Damoiseau det., 1963.

Kleine (1938d: 43) quoted Sumatra and Philippines, but these localities are not given by Damoiseau (1963g: 20; 1979a: 19).

papuanus Damoiseau, 1962

papuanus Damoiseau, 1962e: 109.

Type depository. MSNG (HT!).

Distribution. New Guinea

Notes. In MSNG there is a specimen labelled as paratype (N. Guinea S. E., Moroka, 1300 m, VII.11.1893, Loria) but the species was described on a single specimen.

philippinensis (Kleine, 1922)

philippinensis Kleine, 1922b: 25 (*Dictyopterus*).

idonea Kleine, 1925c: 592 (*Orthopareia*); syn. Damoiseau, 1963g: 10.

Type depositories. *D. philippinensis*: SMTD (HT); *O. idonea*: MPMP (HT).

Distribution. Philippines (Luzon, Mindanao, Tawi Tawi), Vietnam.

pretiosus (Kleine, 1921)

pretiosus Kleine, 1921m: 60 (*Neosebus*).

concitata Kleine, 1926a: 12 (*Sparganophasma*); syn. Damoiseau 1965a: 14.

Type depositories. *N. pretiosus*: RMNH (HT); *S. concitata*: NHM (HT?)

Distribution. Borneo, Indonesia (Java).

Notes. In NHM there is a specimen of *S. concitata* from Borneo labelled as holotype but it can not be the true holotype because the type locality is Java.

pubens (Senna, 1892)

pubens Senna, 1892e: 439 (*Zemioses*).

coomani Kleine, 1944: 151 (*Allaeometrus*); syn. Damoiseau, 1964a: 417.

Type depositories. *Z. pubens*: MSNG (HT!); *A. coomani*: MNHN (HT!), DEI (PT!).

Distribution. Taiwan; India, Myanmar [= Burma], Philippines (Luzon), Thailand, Vietnam.

Notes. Damoiseau (1963g: 10; 1979a: 20) did not mention Malaysia, Java and the Philippines (Kleine, 1938d: 43) as species localities because he doubted the identifications made by Kleine and consequently the corresponding distributions (Damoiseau, 1963g: 10).

***pulcherrimus* (Kleine, 1922)**

pulcherrimus Kleine, 1922b: 26 (*Dictiotopterus*).

Type depository. SMTD (HT).

Distribution. Philippines (Luzon).

***rubicundus* Damoiseau, 1967**

rubicundus Damoiseau, 1967c: 8.

rubicundus: Bartolozzi *et al.*, 1985: 100 (*Eterozemus*) [lapsus].

Type depository. MZUF (HT!).

Distribution. India (Assam).

***salomonensis* Damoiseau, 1979**

salomonensis Damoiseau, 1979a: 16.

Type depository. NHM (HT!).

Distribution. Solomon Is.

***splendens* (Kleine, 1916)**

splendens Kleine, 1916c: 77 (*Dictyotopterus*).

Type depositories. SMTD (LT), DEI (PLT!), RMNH (?) (PLT).

Distribution. Taiwan.

Notes. Damoiseau (1963g: 33) reported the specimens held in SMTD as holotype and allotype, and those held in DEI and in RMNH as paratypes, but

in his original description Kleine did not designate any of the syntypes as holotype. On the basis of article 74.5 of the International Commission of Zoological Nomenclature (1999) the specimen that Damoiseau gives as holotype must be considered as the lectotype.

Kleine (1938d: 44) quotes Andaman Is., but this locality is not given by Damoiseau (1963g: 33; 1979a: 20).

[*Eugeneiobrentus* Damoiseau, 1963]

Eugeneiobrentus Damoiseau, 1963b: 200 [no description].

Eugeneiobrentus Damoiseau, 1989a: 104 [no type species designation].

Notes. Invalid genus. Damoiseau (1963b: 200) quoted it for the first time but he did not describe it, later (1989a: 104) he included the genus in a key but he did not designate the type species. The genus is listed by Alonso-Zarazaga & Lyal (1999: 51) as invalid genus.

Eurorhinus Damoiseau, 1961

Eurorhinus Damoiseau, 1961a: 282.

Type species: *Eurorhinus maynei* Damoiseau, 1961.

Distribution. Democratic Republic of the Congo (ex Zaire).

maynei Damoiseau, 1961

maynei Damoiseau, 1961a: 283.

Type depository. MRAC (HT!).

Distribution. Democratic Republic of the Congo (ex Zaire).

Genogogus Kleine, 1925

Genogogus Kleine, 1925a: 193.

Type species: *Genogogus famosus* Kleine, 1925 = *Cyphagogus sculptilis* Fairmaire, 1889.

Distribution. Madagascar.

***olsufieffi* Kleine, 1934**

olsufieffi Kleine, 1934a: 304.

olsufiewi: Kleine, 1938d: 22 (*Genogogus*) [lapsus].

Type depository. DEI (HT!).

Distribution. Madagascar.

Notes. In 1934 Kleine included this species in a key with the name “*olsufieffi*”, but gave a full description only four years later (1938d) using the name “*olsufiewi*”. The name “*olsufieffi*” has priority because Kleine (1934a: 304) gave enough diagnostic characters in the key.

***sculptilis* (Fairmaire, 1889)**

sculptilis Fairmaire, 1889: XCIII (*Cyphagogus*).

famosus Kleine, 1925a: 193 (*Genogogus*); syn. Damoiseau, 1963c: 240.

Type depositories. *C. sculptilis*: MNHN (HT!); *G. famosus*: NHM (HT!).

Distribution. Madagascar.

***Isomorphus* Kleine, 1916**

Isomorphus Kleine, 1916c: 55.

Isosomus: Kleine, 1920a: 3 [lapsus].

Type species: *Isomorphus unicolor* Kleine, 1916 = *Microsebus kerimi* Senna, 1903.

Distribution. Taiwan; Indonesia (Java, Sumatra); Australia, New Caledonia, New Guinea.

Notes. According to Zimmerman (1994: 147) this genus occurs in Australia (Queensland and Northern Territories) with two indetermined species.

***fauveli* Damoiseau, 1962**

fauveli Damoiseau, 1962c: 283.

Type depository. IRSNB (HT!).

Distribution. New Caledonia.

kerimi (Senna, 1903)

kerimi Senna, 1903: 162 (*Microsebus*).

unicolor Kleine, 1916c: 56 (*Isomorphus*); syn. Damoiseau, 1964a: 401.

Type depositories. *M. kerimi*: MSNG (HT!); *I. unicolor*: IRSNB (ST!), MZPW (ST), RMNH (ST).

Distribution. Indonesia (Java, Sumatra).

minutus Kleine, 1916

minutus Kleine, 1916c: 59.

Type depository. SMTD (HT).

Distribution. Indonesia (Sumatra).

papuanus Damoiseau, 1964

papuanus Damoiseau, 1964a: 407.

Type depository. HNHM (HT).

Distribution. New Guinea.

sennai Damoiseau, 1979

sennai Damoiseau, 1979a: 25.

Type depository. MSNG (HT!).

Distribution. New Guinea.

Notes. See Damoiseau (1979a: 25) for the taxonomical problem related to this species.

Macropareia Kleine, 1932

Macropareia Kleine, 1932a: 571.

Type species: *Macropareia schultzei* Kleine, 1932.

Distribution. Philippines (Panay).

schultzei Kleine, 1932

schultzei Kleine, 1932a: 572.

Type depository. SMTD (HT).

Distribution. Philippines (Panay).

Notes. Damoiseau (1964a: 394; 1989a: 158) reported that the present type depository of the species was unknown; but according to O. Jaeger (SMTD) (2002, personal communication) the type is held in SMTD.

Megalosebus Kolbe, 1916

Megalosebus Kolbe, 1916: 57.

Anomalopleura Kleine, 1916c: 70; syn. De Muizon, 1960: 245.

Anomalopleurus: Kleine, 1920a: 31 [lapsus].

Type species: *Megalosebus separabilis* Kolbe, 1916.

Distribution. Angola, Benin, Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea (Bioko Is. [= Fernando Poo]), Gabon, Ivory Coast, Kenya, Madagascar, Republic of the Congo, Republic of Guinea, Tanzania, Togo, Uganda.

babaulti De Muizon, 1955

babaulti De Muizon, 1955a: 518.

Type depository. MNHN (HT!).

Distribution. Democratic Republic of the Congo (ex Zaire).

bidenticulatus Kolbe, 1916

bidenticulatus Kolbe, 1916: 58.

Type depository. MNHUB (HT!).

Distribution. Benin, Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea (Bioko Is. [= Fernando Poo]), Ivory Coast, Republic of the Congo, Republic of Guinea.

***fallaciosus* Kolbe, 1916 stat. res.**

fallaciosus Kolbe, 1916: 58.

hagedorni Kleine, 1916c: 71 (*Anomalopleura*); syn. De Muizon, 1955a: 521.

feae Calabresi, 1920d: 22 (*Megalosebus*); syn. Kleine, 1927a: 11.

Type depositories. *M. fallaciosus*: MNHUB (HT!); *A. hagedorni*: MZPW (HT); *M. feae*: MSNG (HT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Gabon, Kenya, Republic of the Congo, Tanzania, Uganda.

***perrieri* (Fairmaire, 1899)**

perrieri Fairmaire, 1899: 546 (*Cyphagogus*).

Type depository. MNHN (LT!, PLT!).

Distribution. Madagascar.

Notes. Lectotype of *C. perrieri* (MNHN) designated on a specimen with the following labels: Madag., Perrier/ type/ Museum Paris, Madagascar, coll. Perrier de la Bathie 1906. Paralectotypes: 2 specimens (MNHN) with the same label data as the lectotype.

***separabilis* Kolbe, 1916**

separabilis Kolbe, 1916: 57.

maynei De Muizon, 1959: 72 (*Megalosebus*); syn. Damoiseau, 1962f: 330.

Type depositories. *M. separabilis*: MNHUB (HT!); *M. maynei*: MRAC (HT!, AT!), MNHN (PT!).

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire).

Notes. Damoiseau (1967a: 122) incorrectly reported that the specimens of *M. maynei* held in MRAC are syntypes, but in his original description De Muizon designated holotype, allotype and one paratype.

***testaceus* (Kleine, 1916)**

testaceus Kleine, 1916c: 72 (*Anomalopleura*).

Type depository. DEI (HT!).

Distribution. Cameroon.

***togonicus* Kolbe, 1916**

togonicus Kolbe, 1916: 58.

Type depository. MNHUB (HT!).

Distribution. Republic of the Congo, Togo.

***Mesoderes* Senna, 1898**

Mesoderes Senna, 1898e: 65.

Hellodius: Senna, 1898e: 67 [see Alonso-Zarazaga & Lyal, 1999: 51].

Dysmorphorhynchus Kleine, 1916c: 51; syn. Kleine, 1926a: 13.

Thrasycephalus Kleine, 1916c: 53; syn. Zimmerman, 1994: 148.

Thrasycephalus: Kleine, 1920a: 18 [lapsus].

Trasycephalus: Kabakov, 2001: 209 [lapsus].

Type species: *Mesoderes sexnotatus* Senna, 1898.

Distribution. India to New Guinea, Australia.

Notes. As Damoiseau (1989a: 159) already mentioned, this genus requires in-depth revision because of the systematic confusion Kleine created. Zimmerman (1994: 148) puts the genus *Thrasicephalus* Kleine, 1916 as synonym for *Mesoderes* Senna, 1898 so the species Damoiseau (1989a: 168) included in the genus *Thrasicephalus* must be shifted to *Mesoderes* again.

***argutus* Kleine, 1923**

argutus Kleine, 1923d: 132.

Type depository. NHM (HT!).

Distribution. India.

Notes. Damoiseau (1965, manuscript label) doubts that the specimen from London is the real type. Indeed, Kleine (1923d: 132) writes "keine Makel auf den Elytren", whilst the specimen in question is clearly blotched. On the other hand Kleine compares *M. argutus* with *M. maculatus* Senna, 1898 (a blotched species) and writes that the two species are very similar and only differ in the shape of the antennae. Therefore the specimen could be the true type and that the error lies in the diagnosis furnished by Kleine.

***boettcheri* (Kleine, 1925)**

boettcheri Kleine, 1925c: 593 (*Opistenoxys*).

Type depositories. SMTD (HT).

Distribution. Philippines (Mindanao).

Notes. In his original description Kleine (1925c: 593) wrote that his collection included a paratype, but according to T. Huflejt (MZPW) (2002, personal communication) the specimen is not in MZPW, where Kleine's collection is now held.

***fessus* Kleine, 1923**

fessus Kleine, 1923e: 160.

Type depository. SMTD (?) (HT).

Distribution. Philippines (Negros).

Notes. Kleine (1928a: 73) reported the type for this species to be in SMTD; according to O. Jaeger (SMTD) (2002, personal communication) it is now missing from the SMTD collections.

***guttatus* (Kleine, 1916)**

guttatus Kleine, 1916c: 54 (*Thrasycephalus*).

aberrans Kleine, 1922f: 201 (*Mesoderes*); syn. Damoiseau, 1989a: 168.

gratus Kleine, 1926f: 361 (*Mesoderes*); syn. Damoiseau, 1989a: 168.

Type depositories. *T. guttatus*: RMNH (LT, PLT), MZPW (PLT); *M. aberrans*: SMTD (HT); *M. gratus*: NHM (HT!).

Distribution. India, Indonesia (Java, Timor), Malaysia, Singapore, Vietnam; Australia.

Notes. Damoiseau (1989a: 168) reported that the specimens of *T. guttatus* held in RMNH are holotype and allotype, but in his original description Kleine did not designate any of the syntypes as holotype. However, on the basis of article 74.5 of the International Commission of Zoological Nomenclature (1999) the specimen that Damoiseau gives as holotype must be considered as the lectotype.

latecostatus (Kleine, 1916)

latecostatus Kleine, 1916c: 58 (*Isomorphus*).

latecortatus: Kano, 1930: 152 (*Isomorphus*) [lapsus].

Type depository. DEI (HT!).

Distribution. Taiwan; Vietnam.

maculatus Senna, 1898

maculatus Senna, 1898e: 67.

Type depository. IRSNB (HT!).

Distribution. Borneo, Indonesia (Sumatra), Malaysia, Vietnam; Australia, New Guinea.

sexnotatus Senna, 1898

sexnotatus Senna, 1898e: 66.

amabilis Kleine, 1916c 52 (*Dysmorphorhynchus*); syn. Damoiseau, 1989a: 160.

Type depositories. *M. sexnotatus*: MZUF (LT!, PLT!); *D. amabilis*: DEI (LT!, PLT!).

Distribution. Borneo, India (Andaman Is., Assam), Indonesia (Java, Sumatra), Malaysia, Singapore, Vietnam; Australia, New Guinea.

Notes. Lectotype of *M. sexnotatus* (MZUF) designated on a male specimen with the following labels: Humboldt Bay, Sept.-Oct. 1893, W. Doherty/ *Mesoderes b-notatus* typus! Senna ♂ ♀ / Notes Leyd. Mus., XX, 1898/ ex-

coll. Senna/ "La Specola" Firenze, 6917. Paralectotype: 1 female specimen (MZUF), Humboldt Bay, N. Guinea, leg. Doherty/ Notes Leyd. Mus., XX, 1898/ ex-coll. Senna/ "La Specola" Firenze, 6922.

Lectotype of *D. amabilis* (DEI) designated on a specimen with the following labels: Coen District, Cape York, Queensland, H. Hacker/ coll. Hacker/ Siehe mikroskop. Präparate/ Kleine det./ Dtsch. Entomol. Institut Berlin/ Syntypus/ *Dysmorphorhynchus amabilis* n. g., n. sp./ *Mesoderes sexnotatus* Senna, R. Damoiseau det., 1962. Paralectotypes: 3 specimens (DEI), Coen District, Cape York, Queensland, Hacker/ Syntypus/ coll. Hacker/ Kleine det./ Dtsch. Entomol. Institut Berlin/ Syntypus/ *Dysmorphorhynchus amabilis* Kl./ *Mesoderes sexnotatus* Senna, R. Damoiseau det., 1962.

sphaeroidalis Damoiseau, 1967

sphaeroidalis Damoiseau, 1967c: 12.

shpaeroidalis: Damoiseau, 1989a: 160 (*Mesoderes*) [lapsus].

Type depository. SMTD (HT).

Distribution. Borneo.

Metusambius Kolbe, 1916

Metusambius Kolbe, 1916: 62.

Prosogogus Kleine, 1938e: 102 ; syn. De Muizon, 1955a: 472.

Prosogus: De Muizon, 1955a: 472 [lapsus].

Type species: *Metusambius insularis* Kolbe, 1916.

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea (including Bioko Is. [= Fernando Poo]), Gabon, Ghana, Ivory Coast, Togo.

insularis Kolbe, 1916

insularis Kolbe, 1916: 62.

susplicax Kolbe, 1916: 62 (*Metusambius*); syn. Damoiseau, 1963c: 239.

primoti Kleine, 1938e: 103 (*Prosogogus*); syn. De Muizon, 1955a: 472.

Type depositories. *M. insularis*: MNHUB (LT!), IRSNB (?) (PLT); *M. susplicax*: MNHUB (ST!); *P. primoti*: MNHN (HT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea (including Bioko Is. [= Fernando Poo]), Gabon, Ghana, Ivory Coast, Togo.

Notes. Lectotypes of *M. insularis* designated by Damoiseau (1967a: 78). Damoiseau (1967a: 77) reported that “paratypes” for *M. insularis* are in IRSNB, but we did not find them.

Microsebus Kolbe, 1892

Microsebus Kolbe, 1892: 168.

Eusebus Kleine, 1922k: 173; syn. Alonso-Zarazaga & Lyal, 1999: 51.

Type species: *Microsebus adelphus* Kolbe, 1892.

Distribution. S China, Taiwan; Bhutan, Borneo, India, Indonesia (Java, Sumatra, Sumba), Sri Lanka, Vietnam; Fiji Is., New Guinea.

adelphus Kolbe, 1892

adelphus Kolbe, 1892: 169.

Type depository. MNHUB (HT!).

Distribution. India, Sri Lanka.

bhutanicola (Damoiseau, 1979)

bhutanicola Damoiseau, 1979a: 30 (*Eusebus*).

Type depository. MNHN (HT!).

Distribution. Bhutan.

biakianus (Damoiseau, 1989) **n. comb.**

biakianus Damoiseau, 1989a: 156 (*Eusebus*).

Type depository. NHM (HT!).

Distribution. New Guinea.

***fijianus* (Damoiseau, 1989) n. comb.**

fijianus Damoiseau, 1989a: 156 (*Eusebus*).

Type depository. DSIRN (HT!).

Distribution. Fiji Is.

***globithorax* (Damoiseau, 1979)**

globithorax Damoiseau, 1979a: 29 (*Eusebus*).

Type depository. IRSNB (HT!, AT!, PT!).

Distribution. Borneo, Vietnam.

***hirsutus* (Shaw, 1953)**

hirsutus Shaw, 1953: 328 (*Mesoderes*).

Type depository. NHMB (HT!).

Distribution. Indonesia (Java, Sumatra, Sumba).

***loriae* Senna, 1894**

loriae Senna, 1894e: 554.

Type depository. MSNG (LT!, PLT!).

Distribution. New Guinea.

Notes. Damoiseau (1979a: 28) reported that a specimen held in MSNG is the holotype, but in his original description Senna did not designate any of the syntypes as holotype; however, as there are two syntypes in MSNG, and as Damoiseau did not specifically select one as holotype, we designate the lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Lectotype of *M. loriae* (MSNG) designated on a specimen with the following labels: N Guinea, Ighibirei, Loria, VII-VIII.1890/ *Microsebus loriae* typus! Senna. Paralectotype: 1 specimen (MSNG), N Guinea, Ighibirei, Loria, VII-VI-II.1890.

***trifasciatus* (Kleine, 1916)**

trifasciatus Kleine, 1916c: 57 (*Isomorphus*).

Type depositories. DEI (LT!, PLT!), SMTD (?) (PLT).

Distribution. S China, Taiwan; Indonesia (Sumatra), Vietnam.

Notes. Lectotype of *I. trifasciatus* (DEI) designated on a specimen with the following labels: Formosa, Taihorin, 1911, H. Sauter/ Syntypus/ Kleine det./ *Isomorphus trifasciatus* n. sp/ 7.VIII/ Dtsch. Entomol. Institut Berlin/ *Eusebus trifasciatus* (Kleine) comb. nov., R. Damoiseau det., 1962. Paralectotype: 1 specimen (DEI), Formosa, Taihorin, 1911, H. Sauter/ Syntypus/ Kleine det./ Dtsch. Entomol. Institut Berlin/ *Eusebus trifasciatus* (Kln.), R. Damoiseau det., 1966.

Kleine (1916c: 58) reported four syntypes for this species in SMTD; according to O. Jaeger (SMTD) (2002, personal communication) two specimens of the species are present in the SMTD collection, but they are not labelled as types.

Kabakov (2001: 209) states that the species distribution is: "Ceylon, India, Indochina".

***Nannobrenthus* Kolbe, 1916**

Nannobrenthus Kolbe, 1916: 60.

Rhytidopterus Kleine, 1916c: 46; syn. Kleine, 1927a: 13.

Type species: *Nannobrenthus infantulus* Kolbe, 1916 = *Microsebus compressithorax* Senna, 1898.

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea, Ivory Coast, Republic of the Congo, Rwanda, Sudan, Togo, Zambia.

***compressithorax* (Senna, 1898)**

compressithorax Senna, 1898c: 370 (*Microsebus*).

infantulus Kolbe, 1916: 60 (*Nannobrenthus*); syn. Kleine, 1938d: 41.

variabilis Kleine, 1916c: 47 (*Rhytidopterus*); syn. De Muizon, 1955a: 495.

Type depositories. *M. compressithorax*: MZUF (LT!), DEI (PLT!), MRAC (?) (PLT), SMTD (PLT); *N. infantulus*: MNHUB (?) (ST); *R. variabilis*: DEI (LT!, PLT!); MRAC (PLT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea, Ivory Coast, Republic of the Congo, Rwanda, Sudan, Togo, Zambia.

Notes. Lectotype of *M. compressithorax* designated by Bartolozzi *et al.* (1985: 95) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999). A paralectotype is in DEI, but according to Damoiseau (1967a: 113) it is actually a specimen of *Podozemius kolbei*.

Lectotype of *R. variabilis* (DEI) designated on a specimen with the following labels: N Kamerun, L. Conradt, 96/ Syntypus/ Kleine det./ Dtsch. Entomol. Institut Berlin/ coll. Kraatz/ *Rhytidopterus variabilis* n. g. n. sp./ *Rhytidopterus compressithorax* Senna, R. Damoiseau det., 1961. Paralectotypes: 4 specimens (DEI), N Kamerun, L. Conradt, 96/ Syntypus/ coll. Kraatz/ Kleine det./ Dtsch. Entomol. Institut Berlin/ *Rhytidopterus variabilis* Kleine/ *Rhytidopterus compressithorax* Senna, R. Damoiseau det., 1961; 4 specimens (DEI), Kamerun, Conradt/ coll. Kraatz/ Kleine det./ Syntypus/ *Rhytidopterus variabilis* Kleine/ Dtsch. Entomol. Institut Berlin/ *Rhytidopterus compressithorax* Senna, R. Damoiseau det., 1961; 1 specimen (NHM), Syntype/ cotype/ N Kamerun, L. Conradt, 96/ coll. Kraatz/ Brit. Mus. 1987-250/ *Rhytidopterus variabilis* Kl./ Kleine det.; 1 specimen (MRAC), Paratypus/ Musée du Congo, N Kamerun, L. Conradt, '96/ R. det. C 1331/ *Rhytidopterus variabilis* Kleine, cotype!/ *Rhytidopterus compressithorax* Senna, R. Damoiseau det., 1961.

According to Damoiseau (1967a: 104) syntypes of *N. infantulus* are in MNHUB, but we did not find them there.

Damoiseau (1967a: 104) did not mention Togo (Kolbe, 1916: 57) when he gave the species distribution.

Neocyphagogus Damoiseau, 1979

Neocyphagogus Damoiseau, 1979a: 6.

Type species: *Cyphagogus concavus* Kleine, 1925.

Distribution. Malaysia.

concavus (Kleine, 1925)

concavus Kleine, 1925b: 124 (*Cyphagogus*).

Type depository. NHM (HT!).

Distribution. Malaysia.

***reductus* Damoiseau, 1979**

reductus Damoiseau, 1979a: 8.

Type depositories. IRSNB (HT!), MNHN (AT!).

Distribution. Malaysia.

***violaceonitens* (Kleine, 1931)**

violaceonitens Kleine, 1931a: 94 (*Cyphagogus*).

Type depository. NHM (HT!).

Distribution. Malaysia.

***Neosebus* Senna, 1903**

Neosebus Senna, 1903: 156.

Exostenus Kleine, 1916c: 88; syn. Damoiseau, 1967c: 15.

Type species: *Neosebus bidentatus* Senna, 1903.

Distribution. Borneo, Indonesia (Sumatra); Australia (?).

Notes. Zimmerman (1994: 152) states that he examined an unidentified specimen collected in Australia (Queensland) that could belong to the genus *Neosebus*.

***bidentatus* Senna, 1903**

bidentatus Senna, 1903: 156.

Type depository. MSNG (HT!).

Distribution. Indonesia (Sumatra).

***hospiton* (Kleine, 1916)**

hospiton Kleine, 1916c: 90 (*Exostenus*).

Type depository. DEI (HT!).

Distribution. Borneo.

***sulcirostris* Damoiseau, 1967**

sulcirostris Damoiseau, 1967c: 15.

Type depository. SMTD (HT).

Distribution. Borneo.

***Neoxybasius* Kolbe, 1916**

Neoxybasius Kolbe, 1916: 61.

Phobetromimus Kleine, 1916c: 33; syn. De Muizon, 1955a: 491.

Type species: *Oxybasius fatuelus* Kolbe, 1916.

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea (including Bioko Is. [= Fernando Poo]), Ghana, Ivory Coast, Rwanda, Sierra Leone, Tanzania.

***fatuelus* Kolbe, 1916**

fatuelus Kolbe, 1916: 61.

simulans Kleine, 1916c: 35 (*Phobetromimus*); syn. Alonso-Zarazaga & Lyal, 1999: 51.

exiguus Kleine, 1916c: 36 (*Phobetromimus*); syn. Damoiseau, 1962b: 7.

unicolor Kleine, 1918d: 56 (*Diplohoplizes*); syn. Damoiseau, 1962b: 7.

Type depositories. *N. fatuelus*: DEI (HT!, PT!); *P. simulans*: DEI (LT!, PLT!), ZMUH (PLT); *P. exiguus*: DEI (LT!, PLT!); *D. unicolor*: MRAC (HT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea (including Bioko Is. [= Fernando Poo]), Ghana, Ivory Coast, Rwanda, Sierra Leone, Tanzania.

Notes. Damoiseau (1967a: 60) reported that the specimens of *P. simulans* and *P. exiguus* held in DEI are holotype and paratypes, but in his original description Kleine did not designate any of the syntypes as holotype; however, as there is more than one syntype in DEI, and as Damoiseau did not specifically select one as holotype of each species, we designate the lectotypes (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Lectotype of *P. simulans* (DEI) designated on a specimen with the following labels: ♂ / N Kamerun, L. Conradt, 96/ Syntypus/ coll. Kraatz/ Dtsch. Entomol. Institut Berlin/ *Phobetromimus simulans* ♂ Kleine. Paralectotypes: 2 female specimens (DEI), N Kamerun, L. Conradt, 96/ Syntypus/ *Phobetromimus simulans* n. g. n. sp./ ♀ / coll. Kraatz/ Kleine det./ Dtsch. Entomol. Institut Berlin/ *Phobetromimus simulans* ♀, Kleine Type.

Lectotype of *P. exiguus* (DEI) designated on a male specimen with the following labels: N. Kamerun, L. Conradt, 96/ syntypus/ ♂ / coll. Kraatz/ Deutsche Entomologische Institut/ *Phobetromimus exiguus* n. sp./ *Phobetromimus simulans* Kleine R. Damoiseau det. 1961. Paralectotype: 1 male specimen (DEI), N. Kamerun, L. Conradt, 96/ syntypus/ ♂ / Kleine det./ coll. Kraatz/ Deutsche Entomologische Institut/ *exiguus*/ *Phobetromimus simulans* Kleine, R. Damoiseau det. 1961.

The quotations for the People's Republic of Congo and Guinea (Sforzi, 1998b: 271) are incorrect.

pugionatus Kolbe, 1916

pugionatus Kolbe, 1916: 61.

Type depository. MNHUB (?) (ST).

Distribution. Equatorial Guinea (Bioko Is. [= Fernando Poo]).

Notes. According to Damoiseau (1967a: 61) the status of this species is doubtful. It could be a synonym of *N. fatuelus* Kolbe but he was unable to find the type series in MNHUB. We did not find the syntypes of *N. pugionatus* in MNHUB either.

Neozemioses Damoiseau, 1989

Neozemioses Damoiseau, 1989a: 162.

Type species: *Neozemioses humboldti* Damoiseau, 1989.

Distribution. New Guinea.

humboldti Damoiseau, 1989

humboldti Damoiseau, 1989a: 163.

Type depository. NHM (?) (HT).

Distribution. New Guinea.

Notes. According to Damoiseau (1989a: 163) the holotype is in NHM, but we did not find it there.

Nesidiobrentus Damoiseau, 1964

Nesidiobrentus Damoiseau, 1964a: 415.

Type species: *Opisthenoxys samoanus* Kleine, 1944.

Distribution. New Guinea, Samoa.

bacchusi Damoiseau, 1989

bacchusi Damoiseau, 1989a: 163.

Type depository. NHM (HT!).

Distribution. New Guinea.

samoanus (Kleine, 1944)

samoanus Kleine, 1944: 150 (*Opisthenoxys*).

Type depository. MNHN (HT!).

Distribution. Samoa.

Odontopareius Damoiseau, 1964

Odontopareius Damoiseau, 1964a: 412.

Odontopareius Damoiseau, 1963b: 200 [no description].

Odontopareius Damoiseau, 1963g: 4 [no type species designation].

Type species: *Mesoderes aridus* Kleine, 1923.

Distribution. India.

Notes. Damoiseau (1963b: 200; 1963g: 4) mentioned *Odontopareius* in a list of genera, but he described it in another paper (1964a: 412).

aridus (Kleine, 1923)

aridus Kleine, 1923d: 132 (*Mesoderes*).

Type depositories. NHM (LT!, PLT!), DEI (PLT!), IRSNB (PLT!), SMTD (?) (PLT).

Distribution. India.

Notes. Lectotype of *M. aridus* (NHM) designated on a specimen with the following labels: Syntype/ holotype ♂/ typus!/ 1610/ Nilgiri Hills/ Andrewes Bequest. B. M. 1922-221/ *Mesoderes aridus* Kleine det. 22/ R. Damoiseau det., 1965, *Ondontopareius aridus* (Kleine) Ht. Paralectotypes: 1 specimen (NHM), Syntype/ Allotype/ Nilgiri Hills/ Andrewes/ R. Damoiseau det., 1965, *Ondontopareius aridus* (Kleine) At.; 1 specimen (NHM), Syntype/ cotype/ H. L. Andrewes, Nilgiri Hills/ H. E. Andrewes Bequest. B. M. 1922-221/ *Mesoderes aridus* Kleine; 1 specimen (NHM), Syntype/ cotype/ H. L. Andrewes, Nilgiri Hills/ *Mesoderes aridus* Kleine/ H. E. Andrewes Bequest. B. M. 1922-221/ R. Damoiseau det., 1965, *Ondontopareius aridus*; 3 specimens (NHM), Syntype/ cotype/ Nilgiri Hills/ 1610/ H. E. Andrewes Bequest. B. M. 1922-221; 17 specimens (NHM), Syntype/ cotype/ H. L. Andrews Nilgiri Hills/ Andrewes Bequest. B. M. 1922-221; 5 specimens (DEI), H. L. Andrewes, Nilgiri Hills/ Syntypus/ *Mesoderes aridus* Kleine, H. E. Andrewes det./ *Ondontopareius aridus* (Kleine), R. Damoiseau rev., 1966; 2 specimens (IRSNB), Inde: Nilgiri Hills/ 1922 8/ Staatl. Museum für Tierkunde Dresden/ R. Damoiseau det. 1963, *Ondontopareius aridus* (Kleine)/ Paratype.

Damoiseau (1964a: 414) reported syntypes of this species in SMTD; according to O. Jaeger (SMTD) (2002, personal communication) six specimens of the species are present in the SMTD collection, but they are not labelled as types.

Oncodemerus Senna, 1892

Oncodemerus Senna, 1892d: 157.

Oncodemerus: De Muizon, 1955a: 486 [lapsus].

Type species: *Oncodemerus costipennis* Senna, 1892 (non Fairmaire, 1889) = *Oncodemerus sennai* Quentin, 1961.

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea (including Bioko Is. [= Fernando Poo]), Gabon, Ghana, Ivory Coast, Madagascar, Republic of the Congo, Sierra Leone, Togo.

***costipennis* (Fairmaire, 1889)**

costipennis Fairmaire, 1889: XCIII (*Cyphagogus*).

fairmairei De Muizon, 1955a: 487 (*Oncodermes*) (sic!); syn. Quentin, 1961: 206.

Type depositories. *C. costipennis*: MNHN (LT!, PLT!); *O. fairmairei*: MNHN (?) (ST).

Distribution. Madagascar.

Notes. Damoiseau (1989a: 165) reported that the specimens of *C. costipennis* held in MNHN are holotype, allotype and paratypes, but in his original description Fairmaire did not designate any of the syntypes as holotype; however, as there is more than one syntype in MNHN, and as Damoiseau did not specifically select one as holotype, we designate the lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Lectotype of *C. costipennis* (MNHN) designated on a specimen with the following labels: Madagascar, Tenerive, E. Perrot/ Museum Paris 1952, coll. R. Oberthür. Paralectotypes: 5 specimens (MNHN) with the same label data as the lectotype.

We did not find the syntypes of *O. fairmairei* in MNHN.

See Quentin (1961) for the complex taxonomical problem related to this species.

***sennai* Quentin, 1961**

sennai Quentin, 1961: 206 [nom. nov. for *costipennis* Senna].

costipennis Senna, 1892d: 160 (non Fairmaire, 1889) (*Oncodermes*); syn. Quentin, 1961: 206.

Type depositories. *O. sennai* (= *O. costipennis* Senna): MZUF (LT!), MNHN (PLT!), SMTD (PLT).

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea (including Bioko Is. [= Fernando Poo]), Gabon, Ghana, Ivory Coast, Republic of the Congo, Republic of Guinea, Sierra Leone, Togo.

Notes. Lectotype of *O. sennai* (sub *O. costipennis*) designated by Bartolozzi *et al.* (1985: 95) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

See Quentin (1961) for the complex taxonomical problem related to this species.

Opisthenoxys Kleine, 1922

Opisthenoxys Kleine, 1922b: 26.

Opistenoxys: Kleine, 1937d: 23 [lapsus].

Barathrodes Damoiseau, 1961b: 11; syn. Damoiseau, 1964a: 408.

Type species: *Opisthenoxys ochraceus* Kleine, 1922.

Distribution. Borneo, India (including Assam, Darjeeling), Indonesia (Java, Sumatra), Malaysia, Philippines (Basilan, Mindanao, Negros), Thailand, Vietnam; Australia (I), New Guinea.

arrogans Kleine, 1923

arrogans Kleine, 1923d: 133.

Type depository. NHM (HT!).

Distribution. India.

confirmatus (Kleine, 1926)

confirmatus Kleine, 1926a: 15 (*Allaeometrus*).

Type depository. NHM (HT!).

Distribution. "Indonesia".

Notes. According to Damoiseau (1979a: 23) this species might be a synonym of *O. famulus*.

famulus Kleine, 1925

famulus Kleine, 1925b: 128.

Type depository. NZSI (HT).

Distribution. India (including Darjeeling), Malaysia, Vietnam.

ochraceus Kleine, 1922

ochraceus Kleine, 1922b: 28.

binotatus Damoiseau, 1961b: 12 (*Barathrodes*); syn. Damoiseau, 1964a: 409.

Type depositories. *O. ochraceus*: SMTD (HT); *B. binotatus*: IRSNB (HT!).

Distribution. Borneo, India (including Assam), Indonesia (Java, Sumatra), Malaysia, Philippines (Basilan, Mindanao, Negros), Thailand, Vietnam; Australia (I), New Guinea.

Opisthozemius Kolbe, 1916

Opisthozemius Kolbe, 1916: 58.

Acidotus De Muizon, 1955a: 506; syn. Damoiseau, 1961b: 2.

Type species: *Opisthozemius appendiculatus* Kolbe, 1916.

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Ivory Coast, Republic of the Congo, Zambia; Indonesia (Java, Sumatra).

appendiculatus Kolbe, 1916

appendiculatus Kolbe, 1916: 58.

honestus Kleine, 1936a: 22 (*Opisthozemius*); syn. Damoiseau, 1961b: 5.

Type depositories. *O. appendiculatus*: MNHUB (HT!); *O. honestus*: MRAC (HT!).

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Republic of the Congo.

sulcithorax Damoiseau, 1967

sulcithorax Damoiseau, 1967c: 5.

Type depository. MZUF (HT!).

Distribution. Indonesia (Java, Sumatra).

***villiersi* (De Muizon, 1955)**

villiersi De Muizon, 1955a: 506 (*Acidotus*).

Type depository. MNHN (HT!).

Distribution. Ivory Coast.

***vittatus* Kolbe, 1916**

vittatus Kolbe, 1916: 58.

hautmanni De Muizon, 1959: 76 (*Acidotus*); syn. Damoiseau, 1961b: 5.

Type depositories. *O. vittatus*: MNHUB (HT!); *A. hautmanni*: MRAC (HT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Ivory Coast, Zambia.

***Palaeoparagogus* Damoiseau, 1979**

Palaeoparagogus Damoiseau, 1979a: 12.

Type species: *Palaeoparagogus dentatus* Damoiseau, 1979.

Distribution. Borneo, Vietnam (?).

***dentatus* Damoiseau, 1979**

dentatus Damoiseau, 1979a: 13.

Type depository. BPBM (HT).

Distribution. Borneo, Vietnam (?).

Nota: According to Kabakov (2001: 209) this species is known from Java and Vietnam (?), but actually this species was described from Borneo.

Paraclidorhinus Senna, 1903

Paraclidorhinus Senna, 1903: 159.

Paraclidorrhinus: Kleine, 1927a: 15 [lapsus].

Type species: *Paraclidorhinus modiglianii* Senna, 1903.

Distribution. Borneo, Indonesia (Sumatra), Laos, Malaysia, Vietnam.

modiglianii Senna, 1903

modiglianii Senna, 1903: 160.

Type depository. MSNG (HT).

Distribution. Borneo, Indonesia (Sumatra), Malaysia.

rondoni Damoiseau, 1964

rondoni Damoiseau, 1964a: 388.

Type depository. IRSNB (HT!, PT).

Distribution. Laos.

Notes. Damoiseau (1964a: 389) reported that the holotype and a paratype are held in IRSNB, but we did not find the paratype there.

vietnamicus Kabakov, 2001

vietnamicus Kabakov, 2001: 215.

Type depository. ZIN (HT).

Distribution. Vietnam.

Paradidactus Quentin, 1966

Paradidactus Quentin, 1966: 1650.

Type species: *Paradidactus gibbosus* Quentin, 1966.

Distribution. Republic of the Congo.

Notes. Damoiseau (1967a) did not mention this genus in his Monograph on African Brentidae.

gibbosus Quentin, 1966

gibbosus Quentin, 1966: 1651.

Type depository. MNHN (HT!).

Distribution. Republic of the Congo.

Notes. Damoiseau (1967a) did not mention this species in his Monograph on African Brentidae.

Paragogus Kleine, 1926

Paragogus Kleine, 1926f: 356.

Type species: *Paragogus definitus* Kleine, 1926.

Distribution. Tanzania.

definitus Kleine, 1926

definitus Kleine, 1926f: 357.

Type depository. NHM (HT!).

Distribution. Tanzania.

Paramicrosebus Damoiseau, 1962

Paramicrosebus Damoiseau, 1962f: 335.

Type species: *Paramicrosebus mucronatus* Damoiseau, 1962.

Distribution. Democratic Republic of the Congo (ex Zaire), Ghana, Ivory Coast, Togo.

***angustirostris* Damoiseau, 1962**

angustirostris Damoiseau, 1962f: 338.

Type depository. MNHUB (HT!).

Distribution. Ghana, Ivory Coast, Togo.

***mucronatus* Damoiseau, 1962**

mucronatus Damoiseau, 1962f: 336.

Type depository. MRAC (HT!, AT!).

Distribution. Democratic Republic of the Congo (ex Zaire), Ivory Coast.

Notes. In DEI, MSNG and IRSNB there are some specimens labelled as paratypes, but Damoiseau in his original description only designated holotype and allotype.

***Parasebasius* De Muizon, 1955**

Parasebasius De Muizon, 1955a: 504.

Type species: *Sebasius meticulosus* Kolbe, 1916.

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Ghana, Ivory Coast, Namibia, South Africa, Tanzania, Zambia.

***methneri* (Kleine, 1922)**

methneri Kleine, 1922e: 153 (*Sebasius*).

controversus Kleine, 1924a: 98 (*Glaucocephalus*); syn. De Muizon, 1955a: 499.

Type depositories. *S. methneri*: SMTD (HT); *G. controversus*: MRAC (HT!).

Distribution. Democratic Republic of the Congo (ex Zaire), Namibia, Tanzania, Zambia.

Notes. According to Damoiseau (1967a: 156) the syntypes of *S. methneri* should be in MNHUB but he did not find them there. Kleine based his description of the species on a single specimen which consequently is the holotype; later (1928a: 72) he reported the type to be in MNHUB (Methner coll.). According to O. Jaeger (SMTD) (2002, personal communication) the type is in SMTD.

meticulosus (Kolbe, 1916)

meticulosus Kolbe, 1916: 60 (*Sebasius*).

densepunctatus Kleine, 1936a: 17 (*Glaucocephalus*); syn. De Muizon, 1955a: 499.

versicolor Kleine, 1940a: 245 (*Glaucocephalus*); syn. De Muizon, 1955a: 499.

Type depositories. *S.eticulosus*: MNHUB (HT!); *G. densepunctatus*: MRAC (HT!); *G. versicolor*: MRAC (HT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Ghana, Ivory Coast, South Africa.

Parusambius Kleine, 1925

Parusambius Kleine, 1925b: 129.

Type species: *Parusambius fraudulentus* Kleine, 1925.

Distribution. India (Assam).

fraudulentus Kleine, 1925

fraudulentus Kleine, 1925b: 129.

Type depository. NZSI (?) (HT).

Distribution. India (Assam).

Notes. According to Damoiseau (1989a: 165) the holotype has disappeared.

Pittodes Kleine, 1922

Pittodes Kleine, 1922f: 201.

Type species: *Pittodes aculeatus* Kleine, 1922.

Distribution. Borneo, Malaysia.

aculeatus Kleine, 1922

aculeatus Kleine, 1922f: 203.

Type depository. SMTD (HT).

Distribution. Borneo.

selangorensis Kleine, 1932

selangorensis Kleine, 1932b: 115.

Type depository. NHM (HT!).

Distribution. Malaysia.

Podozemius Kolbe, 1916

Podozemius Kolbe, 1916: 57.

Paramegalosebus De Muizon, 1955a: 522; syn. Damoiseau, 1962f: 322.

Type species: *Podozemius conradti* Kolbe, 1916.

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea, Ethiopia, Gabon, Gambia, Ghana, Ivory Coast, Nigeria, Republic of the Congo, Republic of Guinea, Rwanda, South Africa, Sudan, Tanzania, Togo, Uganda.

comparabilis (Kolbe, 1916) **stat. res.**

comparabilis Kolbe, 1916: 58 (*Megalosebus*).

kolbei Kleine, 1916c: 41 (*Microsebus*); syn. Damoiseau, 1967a: 113.

dilucidus Kleine, 1924a: 99 (*Rhytidopterus*); syn. Damoiseau, 1962f: 322.

mustus De Muizon, 1955a: 510 (non Kolbe, 1916) (*Podozemius*); syn. Damoiseau, 1962f: 323.

Type depositories. *M. comparabilis*: MNHUB (HT!); *M. kolbei*: DEI (LT!, PLT!), MZPW (PLT); *R. dilucidus*: MRAC (LT!), IRSNB (PLT!).

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea, Ethiopia, Gabon, Gambia, Ghana, Ivory Coast, Nigeria, Republic of the Congo, Rwanda, Sudan, Tanzania, Togo, Uganda.

Notes. Damoiseau (1962f: 322) reported that the specimens of *M. kolbei* held in DEI are holotype and allotype, but in his original description Kleine did not designate any of the syntypes as holotype. In the DEI collection we found three syntypes of *M. kolbei*: one of which we designate as lectotype. The other two syntypes belong to the species *Oncodemerus costipennis*.

Lectotype of *M. kolbei* (DEI) designated on a specimen with the following labels: N. Kamerun, L. Conradt, 96/ syntypus/ *Microsebus kolbei* n. sp./ *Paramegalosebus kolbei* Kleine R. Damoiseau det. 1961.

Lectotype of *R. dilucidus* (MRAC) designated on a male specimen with the following labels: Holotypus [incorrectly labelled as holotype]/ Musée du Congo, Haut Uele, Watsa, XI.19919, L. Burgeon/ R. det. C 609/ R. det. F 748/ Typus!/ *Rhytidopterus dilucidus* Kln. typus!, 1922/ Comp. au Type/ *Podozemius kolbei* (Kleine), det. R. Damoiseau, 1962. Paralectotype: 1 specimen (IRSNB), West Africa, Belg. Congo/ 1925-9/ Musée du Congo, Congo da Lemba, II-III.1913, R. Mayné/ *Rhytidopterus dilucidus* Kln. 1922 Cotypus!/ Paratype/ *dilucidus* Kln./ *Podozemius kolbei* (Kleine) R. Damoiseau det., 1962.

conradti Kolbe, 1916

conradti Kolbe, 1916: 57.

gigas Kleine, 1940a: 245 (*Anomalopleura*); syn. Damoiseau, 1962f: 322.

separabilis De Muizon, 1960: 78 (non Kolbe, 1916) (*Paramegalosebus*); syn. Damoiseau, 1962f: 322.

Type depositories. *P. conradti*: MNHUB (LT!); *A. gigas*: MRAC (HT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Ghana, Ivory Coast, Republic of Guinea.

Notes. Damoiseau (1967a: 111) incorrectly reported one paratype of *A. gigas* (MRAC) from Congo (Eala, J. Ghesquière, 1935), but Kleine (1940a: 246) quoted only "Belgischer Congo: Maniema (P. Henrard)" as type locality.

mustus Kolbe, 1916

mustus Kolbe, 1916: 57.

barbatus Calabresi, 1920d: 19 (*Podozemius*); syn. Kleine, 1927a: 11.

dilectus Kleine, 1924a: 99 (*Rhytidopterus*); syn. Damoiseau, 1962f: 323.

gabonicus De Muizon, 1955a: 512 (*Podozemius*); syn. Damoiseau, 1967a: 111.

Type depositories. *P. mustus*: MNHUB (LT!, PLT!); *P. barbatus*: MSNG (HT!); *R. dilectus*: MRAC (HT!); *P. gabonicus*: MNHN (HT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea, Gabon, Ivory Coast, Republic of the Congo, South Africa.

Notes. Lectotype and paralectotype of *P. mustus* designated by Damoiseau (1967a: 112). Damoiseau (1967a: 112) did not mention Congo (Calabresi, 1920d: 19) when he gave the species distribution.

Protoproctus Kolbe, 1916

Protoproctus Kolbe, 1916: 59.

Anablyzostoma Kleine, 1916c: 48; syn. Kleine, 1927a: 12.

Discothorax Damoiseau, 1961b: 10; syn. Damoiseau, 1962f: 340.

Type species: *Protoproctus debilis* Kolbe, 1916.

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Ghana, Ivory Coast, Nigeria, Republic of the Congo, Sierra Leone, Tanzania.

debilis Kolbe, 1916

debilis Kolbe, 1916: 59.

ferrugineum Kleine, 1916c: 50 (*Anablyzostoma*); syn. Kleine, 1927a: 12.

angustirostris K. E. Schedl, 1961c: 193 (*Sebasius*); syn. Damoiseau, 1962f: 340.

herrugineum: Damoiseau, 1962f: 339 (*Anablyzostoma*) [lapsus].

Type depositories. *P. debilis*: MNHUB (?) (HT); *A. ferrugineum*: DEI (HT!); *S. angustirostris*: MRAC (HT!, PT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Ghana, Ivory Coast, Sierra Leone.

Notes. According to Kleine (1928a: 73) the holotype of *P. debilis* was in

MNHUB, but we did not find it there. Quentin (1970: 211) states that it is held in the Museum of Stettin (now in MZPW), but according to T. Huflejt (MZPW) (2002, personal communication) it is not held there.

K. E. Schedl (1961c: 193) reported that two paratypes of *S. angustirostris* are in his collection but W. Schedl (2001, personal communication) did not find them in his father's collection.

***minor* (K. E. Schedl, 1961) comb. n.**

minor K. E. Schedl, 1961c: 192 (*Sebasius*).

ferrugineum De Muizon, 1960: 71 (non Kleine, 1916) (*Anablyzostoma*); syn. Damoiseau, 1962f: 341.

Type depository. MRAC (HT!, PT!).

Distribution. Democratic Republic of the Congo (ex Zaire), Ghana, Ivory Coast, Republic of the Congo, Nigeria, Tanzania.

Notes. K. E. Schedl (1961c: 192) reported that some paratypes of *S. minor* are in his collection but W. Schedl (2001, personal communication) did not find them in his father's collection.

***Protusambius* Kolbe, 1916**

Protusambius Kolbe, 1916: 62.

Type species: *Protusambius praecursor* Kolbe, 1916.

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Ivory Coast.

***praecursor* Kolbe, 1916**

praecursor Kolbe, 1916: 62.

tibialis Kleine, 1916c: 23 (*Schizoadidactus*); syn. De Muizon, 1955a: 470.

proecursor: De Muizon, 1955a: 470 (*Protusambius*) [lapsus].

pracensor: Mayné & Donis, 1962: 95 (*Protusambius*) [lapsus].

Type depositories. *P. praecursor*: MNHUB (LT!, PLT!); *S. tibialis*: DEI (LT!, PLT!), MZPW (PLT).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Ivory Coast.

Notes. Lectotype of *P. praecursor* (MNHUB) designated on a specimen with the following labels: Kamerun, Joh.-Albrechtshöhe, 28.V.-12.VI.98, L. Conradt S/ Type/ *Protusambius praecursor* n. sp. Kolbel/ 5/ *Protusambius tibialis* (Kleine) R. Damoiseau det., 1962/ Type de *Protusambius tibialis praecursor* Kolbe, R. Damoiseau det., 1963. Paralectotype: 1 specimen (MNHUB), Kamerun, Conradt/ *praecursor* n. sp. Kolbel/ *Protusambius tibialis* (Kleine) R. Damoiseau det., 1962.

Damoiseau (1967a: 81) reported that the specimens of *S. tibialis* held in DEI are holotype and allotype, but in his original description Kleine did not designate either of the syntypes as holotype; however, as there are two syntypes in DEI, and as Damoiseau did not specifically select one as holotype, we designate the lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Lectotype of *S. tibialis* (DEI) designated on a specimen with the following labels: N Kamerun, Johan Albrechtshöhe, L. Conradt, 96/ Typus/ Syntypus/ Coll. Kraatz/ Kleine det./ Dtsch. Entomol. Institut Berlin/ *Schizoadidactus tibialis* Kl., det. Kleine, 1915/ *Protusambius tibialis* (Kleine), R. Damoiseau vid., 1961. Paralectotype: 1 specimen (DEI), with the same label data as the lectotype.

Pseudamobaeus Damoiseau, 1967

Pseudamobaeus Damoiseau, 1967b: 2.

Type species: *Pseudamobaeus procax* Damoiseau, 1967.

Distribution. Republic of the Congo.

procax Damoiseau, 1967

procax Damoiseau, 1967b: 3.

Type depository. HNHM (HT).

Distribution. Republic of the Congo.

Pseudoadidactus De Muizon, 1955

Pseudoadidactus De Muizon, 1955a: 471.

Type species: *Schizoadidactus famatus* Kleine, 1925.

Distribution. Madagascar.

famatus (Kleine, 1925)

famatus Kleine, 1925a: 191 (*Schizoadidactus*).

advena K. E. Schedl, 1961c: 187 (non Pascoe, 1866) (*Usambius*); syn. Damoiseau, 1989a: 166.

Type depository. NHM (HT!).

Distribution. Madagascar.

Pseudocyphagonus Desbrochers des Loges, 1890

Pseudocyphagonus Desbrochers des Loges, 1890: 221.

Pseudocyphagogus Senna, 1892c: 41 [unjustified emendation].

Homoiozemius Damoiseau, 1963g: 38; syn. Damoiseau, 1989a: 166.

Type species: *Pseudocyphagonus squamifer* Desbrochers des Loges, 1890.

Distribution. Borneo, India (Andaman Is., Assam), Indonesia (Sumatra), Laos, Malaysia, Philippines (Mindanao, Negros, Samar), Thailand, Vietnam.

Notes. Damoiseau (1963b: 200) mentioned *Homoiozemius* in a list of genera, but he described it in another paper (1963g: 38).

reticulatus (Damoiseau, 1963)

reticulatus Damoiseau, 1963g: 39 (*Homoiozemius*).

Type depository. MNHUB (HT!, AT!, PT!).

Distribution. Philippines (Mindanao).

Notes. Damoiseau (1963g: 43) reported that one paratype is in IRSNB but actually it is in MNHUB.

squamifer Desbrochers des Loges, 1890

squamifer Desbrochers des Loges, 1890: 222.

Type depository. MNHN (ST!).

Distribution. Borneo, India (Andaman Is., Assam), Indonesia (Sumatra), Laos, Malaysia, Philippines (Negros, Samar), Thailand, Vietnam.

Notes. Bartolozzi *et al.* (1984: 101) quoted that Damoiseau [*in litteris*] designated the neotype of this species on a specimen of MZUF but this neotype designation was never published by Damoiseau.

According to Damoiseau (1964a: 399) the distribution given by Kleine (1938d: 39) for this species is doubtful.

Pseudoparagogus De Muizon, 1955

Pseudoparagogus De Muizon, 1955a: 458.

Type species: *Pseudoparagogus turneri* De Muizon, 1955.

Distribution. Angola, Democratic Republic of the Congo (ex Zaire), Gabon, Ghana, Republic of the Congo, South Africa, Swaziland.

ensifer Quentin, 1966

ensifer Quentin, 1966: 1647.

Type depository. MNHN (HT!).

Distribution. Republic of the Congo.

Notes. Damoiseau (1967a) did not mention this species in his Monograph on African Brentidae.

foveatus Damoiseau, 1972

foveatus Damoiseau, 1972a: 261.

Type depository. HNHN (HT, AT).

Distribution. Angola, Ghana.

longitarsis Damoiseau, 1961

longitarsis Damoiseau, 1961a: 281.

Type depository. MRAC (HT!), AT!).

Distribution. Democratic Republic of the Congo (ex Zaire).

parvulus Damoiseau, 1963

parvulus Damoiseau, 1963a: 119.

Type depository. MRAC (HT!).

Distribution. Democratic Republic of the Congo (ex Zaire).

turneri De Muizon, 1955

turneri De Muizon, 1955a: 459.

Type depositories. NHM (HT!), MNHN (PT!).

Distribution. Democratic Republic of the Congo (ex Zaire), Gabon, South Africa, Swaziland.

Pseudousambius De Muizon, 1955

Pseudousambius De Muizon, 1955a: 482.

Type species: *Pseudousambius overlaeti* De Muizon, 1955 = *Metusambius silvanus* Kolbe, 1916.

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Ivory Coast, Republic of the Congo.

silvanus (Kolbe, 1916)

silvanus Kolbe, 1916: 62 (*Metusambius*).

overlaeti De Muizon, 1955a: 483 (*Pseudousambius*); syn. Damoiseau, 1963a: 117.

Type depositories. *M. silvanus*: MNHUB (HT!); *P. overlaeti*: MRAC (HT!, PT!), MNHN (PT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Ivory Coast, Republic of the Congo.

Schizoadidactus Kleine, 1916

Schizoadidactus Kleine, 1916c: 19.

Type species: *Schizoadidactus aethiops* Kleine, 1916 = *Adidactus napaeus* Kolbe, 1916.

Distribution. Angola, Cameroon, Central African Republic, Comores Is., Democratic Republic of the Congo (ex Zaire), Equatorial Guinea, Ethiopia, Ghana, Ivory Coast, Kenya, Republic of the Congo, Rwanda, Sierra Leone, Tanzania, Togo, Zimbabwe.

humbloti Quentin, 1978

humbloti Quentin, 1978: 133.

Type depository. MNHN (HT!, PT!).

Distribution. Comores Is.

napaeus (Kolbe, 1916) **stat. res.**

napaeus Kolbe, 1916: 62 (*Adidactus*).

aethiops Kleine, 1916c: 21 (*Schizoadidactus*); syn. Damoiseau, 1963c: 233.

Type depositories. *A. napaeus*: MNHUB (?) (ST); *S. aethiops*: SMTD (ST), ZMUH (ST).

Distribution. Angola, Cameroon, Central African Republic, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea, Ethiopia, Ghana, Ivory Coast, Kenya, Republic of the Congo, Rwanda, Sierra Leone, Tanzania, Togo, Zimbabwe.

Notes. According to Damoiseau (1967a: 74) the syntypes of *A. napaesus* are in MNHUB, but we did not find them there.

Damoiseau (1967a: 74) reported that the specimen of *S. aethiops* held in SMTD is the holotype, but in his original description Kleine did not designate any of the three syntypes as holotype.

Damoiseau (1967a: 74) did not mention Zimbabwe (Kleine, 1936a: 13) when he gave the species distribution.

***Sebasius* Lacordaire, 1866**

Sebasius Lacordaire, 1866: 409.

Sebarius: Kolbe, 1883b: 237 [lapsus].

Stilbonotus Kleine, 1916c: 78; syn. De Muizon, 1955a: 497.

Type species: *Sebasius deyrollei* Lacordaire, 1866.

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Ivory Coast, Madagascar, Nigeria, South Africa; Brazil.

***brasiliensis* (Soares, 1954)**

brasiliensis Soares, 1954: 163 (*Stilbonotus*).

Type depository. IPEACS (HT).

Distribution. Brazil.

***deyrollei* Lacordaire, 1866**

deyrollei Lacordaire, 1866: 409.

Type depository. MCZ (HT).

Distribution. Madagascar.

***strohmeyeri* (Kleine, 1916)**

strohmeyeri Kleine, 1916c: 79 (*Stilbonotus*).

Type depositories. SMTD (LT), DEI (PLT!), MZPW (PLT).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Ivory Coast, Madagascar, Nigeria, South Africa (Natal).

Notes. Lectotype designated by Damoiseau (1967a: 158).

Damoiseau (1967a: 157) did not mention South Africa (Natal) and Madagascar (Kleine, 1924a: 109) when he gave the species distribution.

Stibacephalus Kleine, 1916

Stibacephalus Kleine, 1916c: 73.

Protosebus Kleine, 1922h: 133; syn. De Muizon, 1955a: 524.

Type species: *Stibacephalus wagneri* Kleine, 1916.

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Gabon.

wagneri Kleine, 1916

wagneri Kleine, 1916c: 74.

piceus Kleine, 1922h: 135 (*Protosebus*); syn. De Muizon, 1955a: 523.

Type depositories. *S. wagneri*: DEI (HT!); *P. piceus*: MNHUB (LT!, PLT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Gabon.

Notes. Lectotype of *P. piceus* (MNHUB) designated on a specimen with the following labels: Kamerun, Namiong h. Lolodorf a. Lokundjeft., O. Ulbrich V./ Typus/ *Protosebus piceus* Kleine/ *Stibacephalus wagneri* Kln., R. Damoiseau det., 1962. Paralectotype: 1 specimen (MNHUB), Kamerun, Namiong h. Lolodorf a. Lokundjeft., O. Ulbrich V./ Typus.

Synsebasius Kolbe, 1916

Synsebasius Kolbe, 1916: 59.

Glaucocephalus Kleine, 1916c: 82; syn. Kleine, 1927a: 9.

Type species: *Synsebasius imitator* Kolbe, 1916.

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Gabon, Ivory Coast, Madagascar, Republic of the Congo, Togo.

imitator Kolbe, 1916

imitator Kolbe, 1916: 59.

robustus Kleine, 1916c: 84 (*Glaucocephalus*); syn. Kleine, 1927a: 10.

Type depositories. *S. imitator*: MNHUB (?) (ST); *G. robustus*: DEI (LT!, PLT!), ZSM (Ertl coll.) (?) (PLT).

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Gabon, Ivory Coast, Republic of the Congo, Togo, Zimbabwe.

Notes. The syntypes of *S. imitator* should be in MNHUB, but neither Damoiseau (1967a: 168) nor ourselves could find them there.

Lectotype of *G. robustus* (DEI) designated on a specimen with the following labels: N Kamerun, L. Conradt, 96/ Syntypus/ Coll. Kraatz/ Kleine det./ *Glaucocephalus robustus* n. sp./ Dtsch. Entomol. Institut Berlin/ Prépar. génit N. 65/11.25.03/ *Glaucocephalus robustus* Kleine, R. Damoiseau det., 1965. Paralectotype: 1 female specimen (DEI), N Kamerun, L. Conradt, 96/ Syntypus/ Coll. Kraatz/ ♀/ Dtsch. Entomol. Institut Berlin/ *Glaucocephalus robustus* Kleine, R. Damoiseau det., 1965.

In his original description Kleine (1916c: 84) reported that two syntypes for *G. robustus* were in the Ertl collection (Munich), but according to M. Baehr (ZSM) (2001, personal communication) these types are missing.

Damoiseau (1967a: 167) did not mention Zimbabwe (Kleine, 1936a: 17) when he gave the species distribution.

The specimen from Gabon which Bartolozzi & Sforzi (1997: 80) identified as *S. roubaudi* De Muizon actually belongs to this species.

opacus Kolbe, 1916

opacus Kolbe, 1916: 60.

Type depository. MNHUB (?) (HT).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Gabon, Republic of the Congo.

Notes. According to Damoiseau (1967a: 167) the holotype disappeared. We did not find it in MNHUB.

roubaudi (De Muizon, 1955)

roubaudi De Muizon, 1955a: 500 (*Glaucocephalus*).

Type depository. MNHN (HT!).

Distribution. Republic of the Congo.

Notes. The specimen from Gabon which Bartolozzi & Sforzi (1997: 80) identified as this species actually belongs to *S. imitator* Kolbe.

seminitens (Kleine, 1916)

seminitens Kleine, 1916c: 85 (*Glaucocephalus*).

Type depositories. DEI (LT!), unknown (ST).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Gabon, Ivory Coast, Republic of the Congo.

Notes. Lectotype of *G. seminitens* (DEI) designated on a specimen with the following labels: Kamerun, Conradt/ Syntypus/ ♀/Coll. Kraatz/ Kleine det./ Dtsch. Entomol. Institut Berlin/ *Glaucocephalus seminitens* n. sp. ♀/ *Glaucocephalus seminitens* Kleine, R. Damoiseau det., 1965.

sicardi (De Muizon, 1955)

sicardi De Muizon, 1955a: 503 (*Glaucocephalus*).

Type depository. MNHN (HT!).

Distribution. Madagascar.

vadoni (De Muizon, 1955)

vadoni De Muizon, 1955a: 502 (*Glaucocephalus*).

Type depository. MNHN (HT!, PT).

Distribution. Madagascar.

Notes. According to De Muizon (1955a: 503) three paratypes are held in MNHN, but we did not find them there.

Tetanocephalus Kleine, 1916

Tetanocephalus Kleine, 1916c: 66.

Type species: *Tetanocephalus nigronitens* Kleine, 1916.

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire).

nigronitens Kleine, 1916

nigronitens Kleine, 1916c: 68.

Type depository. DEI (LT!, PLT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire).

Notes. Lectotype of *T. nigronitens* (DEI) designated on a specimen with the following labels: N Kamerun, L. Conradt, 96/ Syntypus/ Coll. Kraatz/ *Tetanocephalus nigripes* n. sp./ Kleine det./ *Tetanocephalus nigripes* Kleine, R. Damoiseau det., 1965. Paralectotypes: 2 specimens (DEI), N Kamerun Conradt, 96/ Kleine det./ Coll. Kraatz/ Syntypus/ *Tetanocephalus nigripes* Kleine, R. Damoiseau det., 1965.

rugosus Kleine, 1916

rugosus Kleine, 1916c: 69.

Type depository. DEI (HT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire).

Usambioproctus Kolbe, 1916

Usambioproctus Kolbe, 1916: 62.

Hesperobius Kolbe, 1916: 60; syn. Kleine, 1927a: 8.

Xestocoryphus Kleine, 1916c: 24; syn. Kleine, 1927a: 8.

Xestocryphus: K. E. Schedl, 1964: 68 [lapsus].

Xestocryptus: K. E. Schedl, 1964: 68 [lapsus].

Type species: *Usambioproctus paniscus* Kolbe, 1916.

Distribution. Angola, Cameroon, Central African Republic, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea (including Bioko Is. [= Fernando Poo]), Gabon, Ghana, Ivory Coast, Republic of Guinea, Tanzania, Zambia.

***collarti* (Damoiseau, 1963) comb. n.**

collarti Damoiseau, 1963a: 109 (*Xestocoryphus*).

Type depository. IRSNB (HT!).

Distribution. Democratic Republic of the Congo (ex Zaire).

***compressicornis* (Kleine, 1916) comb. n.**

compressicornis Kleine, 1916c: 28 (*Xestocoryphus*).

Type depository. DEI (HT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Ivory Coast.

Notes. Damoiseau (1967a: 87) did not mention the Democratic Republic of the Congo (ex Zaire) (Kleine, 1936a: 14) when he gave the species distribution.

***congoensis* (De Muizon, 1955) comb. n.**

congoensis De Muizon, 1955a: 475 (*Xestocoryphus*).

Type depository. MRAC (HT!).

Distribution. Democratic Republic of the Congo (ex Zaire).

***conjunctus* (Kleine, 1916) comb. n.**

conjunctus Kleine, 1916c: 30 (*Xestocoryphus*).

dissimilis De Muizon, 1955a: 475 (non Kleine, 1916) (*Xestocoryphus*); syn. Damoiseau, 1963a: 107.

conjonctus De Muizon, 1955a: 475 (*Xestocoryphus*) [lapsus].

Type depository. DEI (HT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Gabon.

***decellei* (Damoiseau, 1967) comb. n.**

decellei Damoiseau, 1967a: 94 (*Xestocoryphus*).

Type depositories. MRAC (HT!, AT!), IRSNB (PT!), MNHN (PT!).

Distribution. Cameroon, Ivory Coast, Republic of Guinea.

***dissimilis* (Kleine, 1916) comb. n.**

dissimilis Kleine, 1916c: 27 (*Xestocoryphus*).

Type depository. DEI (HT!).

Distribution. Cameroon, Central African Republic, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea (Bioko Is. [= Fernando Poo]), Ghana, Ivory Coast, Republic of Guinea, Zambia.

***gracilis* (Kleine, 1916) comb. n.**

gracilis Kleine, 1916c: 26 (*Xestocoryphus*).

Type depositories. DEI (LT!, PLT!), MZPW (PLT).

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea (Bioko Is. [= Fernando Poo]), Ghana, Republic of Guinea, Tanzania.

Notes. Damoiseau (1967a: 85) reported that the specimen held in DEI is the holotype, but in his original description Kleine did not designate any of the syntypes as holotype; however, as there is more than one syntype in DEI, and as Damoiseau did not specifically select one as holotype, we designate the lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Lectotype of *X. gracilis* (DEI) designated on a female specimen with the following labels: N Kamerun L. Conradt, 96/ Coll. Kraatz/ Syntypus/ Kleine

det./ Dtsch. Entomol. Institut Berlin/ Siehe mikroskop. Präparate/ *Xestocoryphus gracilis* ♀ Kleine, Paratype, R. Damoiseau vid., 1961. Paralectotypes: 1 female specimen (DEI), N Kamerun Conradt, 96/ Senna det./ Coll. Kraatz/ Syntypus/ Dtsch. Entomol. Institut Berlin/ Kleine det./ *Xestocoryphus gracilis* Kl. det, 1920/ *Xestocoryphus gracilis* ♀ Kleine, R. Damoiseau vid., 1961; 1 female specimen (DEI), N Kamerun Conradt, 96/ Syntypus/ Coll. Kraatz/ Dtsch. Entomol. Institut Berlin/ Kleine det./ Siehe mikroskop. Präparate/ *Eucinetus* (olim!) *gracilis* n. sp./ *Xestocoryphus*/ *Xestocoryphus gracilis* ♀ Kleine, Type, R. Damoiseau vid., 1961.

Damoiseau (1967a: 84) did not mention Equatorial Guinea (Bioko Is. [= Fernando Poo]) (Kleine, 1936a: 14) when he gave the species distribution.

***intermedius* (Damoiseau, 1961) comb. n.**

intermedius Damoiseau, 1961a: 280 (*Xestocoryphus*).

Type depository. MRAC (HT!, PT!).

Distribution. Democratic Republic of the Congo (ex Zaire), Ghana, Tanzania.

***muizoni* (Damoiseau, 1963) comb. n.**

muizoni Damoiseau, 1963a: 113 (*Xestocoryphus*).

gracilis De Muizon, 1960: 64 (non Kleine, 1916) (*Xestocoryphus*); syn. Damoiseau, 1963a: 113.

Type depositories. MRAC (HT!, PT), IRSNB (PT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Ghana, Ivory Coast.

Notes. Damoiseau (1963a: 115) reported that the holotype and two paratypes are held in MRAC, but we did not find any paratype there.

***paniscus* Kolbe, 1916**

paniscus Kolbe, 1916: 63.

Type depository. MNHUB (?) (ST).

Distribution. Cameroon, Equatorial Guinea.

Notes. According to Kleine (1928a: 71) the types are in MNHUB, but we did not find them there.

In his Monograph on the African Brentidae Damoiseau (1967a: 468) did not redescribe the species but only listed it with a question mark in the genus *Xestocoryphus*.

***pierloti* (Damoiseau, 1967) comb. n.**

pierloti Damoiseau, 1967a: 95 (*Xestocoryphus*).

Type depository. MRAC (HT!).

Distribution. Democratic Republic of the Congo (ex Zaire).

***planistriatus* (Damoiseau, 1961) comb. n.**

planistriatus Damoiseau, 1961a: 279 (*Xestocoryphus*).

Type depository. MRAC (HT!).

Distribution. Democratic Republic of the Congo (ex Zaire), Ivory Coast.

***rufus* (Kolbe, 1916) comb. n.**

rufus Kolbe, 1916: 60 (*Hesperobius*).

Type depository. MNHUB (?) (HT).

Distribution. Cameroon.

Notes. According to Kleine (1928a: 71) the holotype is in MNHUB, but we did not find it there.

In his Monograph on the African Brentidae Damoiseau (1967a: 468) did not redescribe the species but only listed it with a question mark in the genus *Xestocoryphus*.

***tempestivus* (Kolbe, 1916) comb. n.**

tempestivus Kolbe, 1916: 60 (*Hesperobius*).

Type depository. MNHUB (?) (HT).

Distribution. Equatorial Guinea (Bioko Is. [= Fernando Poo]).

Notes. According to Kleine (1928a: 71) the holotype is in MNHUB, but we did not find it there.

In his Monograph on the African Brentidae Damoiseau (1967a: 468) did not redescribe the species but only listed it with a question mark in the genus *Xestocoryphus*.

***vestigialis* (Kolbe, 1916) comb. n.**

vestigialis Kolbe, 1916: 61 (*Hesperobius*).

Type depository. MNHUB (?) (HT).

Distribution. Equatorial Guinea, Ivory Coast.

Notes. According to Kleine (1928a: 71) the holotype is in MNHUB, but we did not find it there.

Damoiseau did not quote this species in his Monograph on the African Brentidae (1967a).

***Usambius* Kolbe, 1892**

Usambius Kolbe, 1892: 167.

Type species: *Cyphagogus advena* Pascoe, 1866.

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea (including Bioko Is. [= Fernando Poo]), Ghana, Ivory Coast, Madagascar, Republic of the Congo, Rwanda, São Thomé and Príncipe, South Africa, Tanzania, Uganda, Zambia; Italy (I).

***advena* (Pascoe, 1866)**

advena Pascoe, 1866: 48 (*Cyphagogus*).

wahlbergii Fähræus, 1871: 433 (*Calodromus*); syn. Kleine, 1927a: 8.

conradti Kolbe, 1892: 168 (*Usambius*); syn. De Muizon, 1955a: 479.

laevis Senna, 1898c: 370 (*Usambius conradti* var.); syn. Calabresi, 1920d: 18.

wahlbergii: Schönfeldt, 1908: 5 (*Calodromus*) [lapsus].

loevis: De Muizon, 1955a: 479 (*Usambius conradti* var.) [lapsus].

laevicollis: Quentin, 1970: 208 (*Usambius*) [lapsus for *laevis* Senna, 1898].

Type depositories. *C. advena*: NHM (HT!); *C. wahlbergi*: NHRS (HT); *U. conradti*: MNHUB (LT!, PLT!), MNHN (PLT!), MSNG (PLT!); *U. conradti* var. *laevis*: MZUF (LT!, PLT!), DEI (PLT!), MRAC (PLT!), MNHUB (PLT!).

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea (including Bioko Is. [= Fernando Poo]), Ghana, Ivory Coast, Madagascar, Republic of the Congo, Rwanda, São Thomé and Príncipe, South Africa, Tanzania, Uganda, Zambia; Italy (I).

Notes. Lectotype of *U. conradti* (MNHUB) designated on a specimen with the following labels: Usambara, Derema, 850 m, 16.9-7.10.91, Conradt S./ Type. Paralectotypes: 1 specimen (MNHN), Usambara, Derema, 850 m, October 91, Conradt S./ aus d. Kgl Museum F. Naturk Berlin/ Museum Paris 1952 coll. R. Oberthür; 2 specimens (MNHN), Usambara, Derema, 850 m, 8-24 Octbr. 91, Conradt S./ Museum Paris 1952, coll. R. Oberthür; 1 specimen (MSNG), Usambara, Derema, 850 m, october 91, Conradt S. (typus); 3 specimens (MNHUB), Usambara, Derema, 850 m, october 91 Conradt S. (typus); 1 specimen (MNHUB), Usambara, Derema, 850 m, dezember 91, Conradt S.

Damoiseau (1967a: 48) reported that syntypes of *U. conradti* var. *laevis* are held in MNHUB, but Senna's description was based on four specimens, all held in MZUF. We found other specimens labelled 'syntype' or 'paratype' in MRAC and DEI, but they cannot belong to the type series.

Lectotype of *U. conradti* var. *laevis* (MZUF) designated on a specimen with the following labels: Station Johann Albrechts Höhe, N Kamerun, L. Conradt/ *Usambius conradti laevis* Senna/ Deutsch. Ent. Z., 1898, Typi!! *Usambius advena* Pascoe, R. Damoiseau det., 1966/ ex-coll. Senna/ "La Specola" Firenze, 6915. Paralectotypes: 2 specimens (MZUF), N. Camerun, Stat. Albrechts, L. Conradt/ ex-coll. Senna/ "La Specola" Firenze, 6915; 1 specimen (MZUF), N. Camerun, leg. Conradt, ex Dr. Kraatz/ *Usambius advena* Pascoe, R. Damoiseau det., 1966.

decellei Damoiseau, 1967

decellei Damoiseau, 1967a: 49.

Type depository. MRAC (HT!, AT!).

Distribution. Angola, Ivory Coast.

madagascariensis De Muizon, 1955

madagascariensis De Muizon, 1955a: 479.

Type depository. MNHN (HT!).

Distribution. Madagascar.

***Vietobrentus* Kabakov, 2001**

Vietobrentus Kabakov, 2001: 214.

Type species: *Vietobrentus napolovi* Kabakov, 2001.

Distribution. Vietnam.

***napolovi* Kabakov, 2001**

napolovi Kabakov, 2001: 215.

Type depository. ZIN (HT).

Distribution. Vietnam.

***Xenadidactus* Damoiseau, 1979**

Xenadidactus Damoiseau, 1979a: 10.

Type species: *Xenadidactus xestocoryphoides* Damoiseau, 1979.

Distribution. Ghana, Republic of the Congo.

Notes. Damoiseau (1972a: 263) mentioned *Xenadidactus xestocoryphoides* in a list of species from Ghana, but he did not describe the genus and the species until six years later (1979a: 10).

***xestocoryphoides* Damoiseau, 1979**

xestocoryphoides Damoiseau, 1979a: 11.

Type depositories. MNHN (HT!, AT!), IRSNB (PT!), HNHM (PT).

Distribution. Ghana, Republic of the Congo.

Xestothorax Damoiseau, 1989

Xestothorax Damoiseau, 1989a: 169.

Type species: *Xestothorax papuanus* Damoiseau, 1989.

Distribution. New Guinea.

papuanus Damoiseau, 1989

papuanus Damoiseau, 1989a: 169.

Type depository. AMNH (HT).

Distribution. New Guinea.

Zemioses Pascoe, 1862

Zemioses Pascoe, 1862: 394.

Dactylobarus Kleine, 1914b: 159; syn. De Muizon, 1955a: 527.

Zemiose: Mayné & Donis, 1960: 56 [lapsus].

Dactylbarus: Döbler & Gaedike, 1995: 446 [lapsus].

Type species: *Zemioses porcatus* Pascoe, 1862.

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea (including Bioko Is. [= Fernando Poo]), Gabon, Ghana, Ivory Coast, Kenya, Liberia, Madagascar, Republic of the Congo, Rwanda, Sierra Leone, South Africa, Swaziland, Tanzania, Togo, Zambia.

crassicollis Fairmaire, 1899

crassicollis Fairmaire, 1899: 546.

Type depository. MNHN (LT!, PLT!).

Distribution. Madagascar.

Notes. Lectotypes of *Zemioses crassicollis* (MNHN) designated on a specimen with the following labels: Madagr., Superb Ile, H. Perrier/ type/ Museum Paris, Madagascar, coll. Perrier de la Bathie 1906/ *Zemioses crassicollis*,

Fairm. Paralectotypes: 3 specimens (MNHN) with the same data as the lectotype; 2 specimens (MNHN), Madagr., Superb Ile, H. Perrier/ type/ Museum Paris, Madagascar, coll. Perrier de la Bathie, 1906.

porcatus Pascoe, 1862

porcatus Pascoe, 1862: 394.

parvatus: Faust, 1895: 260 (*Zemioses*) [lapsus].

camerunus Kleine, 1914b: 164 (*Zemioses*); syn. De Muizon, 1955a: 526.

bicolor Kolbe, 1916: 59 (*Zemioses camerunus* var.); syn. De Muizon, 1955a: 526.

densepunctatus Kolbe, 1916: 59 (*Zemioses camerunus* var.); syn. De Muizon, 1955a: 526.

nigricollis Kolbe, 1916: 59 (*Zemioses camerunus* var.); **syn. n.**

carbonarius Kleine, 1916c: 63 (*Dactylobarus*); syn. De Muizon, 1955a: 528.

fraterculus Kleine, 1916c: 65 (*Dactylobarus*); syn. De Muizon, 1955a: 528.

Type depositories. *Z. porcatus*: NHM (HT!); *Z. camerunus*: unknown; *Z. camerunus* var. *bicolor*: MNHUB (LT!); *Z. camerunus* var. *densepunctatus*: MNHUB (?) (ST); *Z. camerunus* var. *nigricollis*: MNHUB (LT!); *D. carbonarius*: DEI (LT!, PLT!); *D. fraterculus*: DEI (LT!, PLT!).

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea (including Bioko Is. [= Fernando Poo]), Gabon, Ghana, Ivory Coast, Kenya, Liberia, Republic of the Congo, Sierra Leone, South Africa, Swaziland, Tanzania, Togo, Zambia.

Notes. Lectotype of *Z. camerunus* var. *bicolor* (MNHUB) designated on a specimen with the following labels: Togo, Bismarckburg, L. Conradt S./ N. 19 Biol./ Type/ razza *bicolor* Kolbel/ *Zemioses porcatus* Pascoe, R. Damoiseau det., 1962. We were unable to identify other syntypes.

Lectotype of *Z. camerunus* var. *nigricollis* (MNHUB) designated on a specimen with the following labels: Kamar. Lolodorf, Jacob S., Prof. Ziemann G./ Type/ razza *nigricollis* Kolbel/ *Zemioses porcatus* Pascoe, R. Damoiseau det., 1962. We were unable to identify other syntypes.

Lectotype of *D. carbonarius* (DEI) designated on a specimen with the following labels: N Kamerun, L. Conradt, 96/ Syntypus/ Coll. Kraatz/ Kleine det./ *Dactylobarus fraterculus* n. sp./ *Zemioses porcatus* Pascoe, R. Damoiseau det., 1965/. Paralectotypes: 3 specimens (DEI), N Kamerun, L. Conradt, 96/ Syntypus/ Coll. Kraatz/ Kleine det.

Lectotype of *D. fraterculus* (DEI) designated on a specimen with the following labels: N Kamerun, L. Conradt, 96/ Syntypus/ *Dactylobarus fraterculus* n. sp./ Kleine det./ *Zemioses porcatus* Pascoe, R. Damoiseau det., 1965/ Coll. Kraatz. Paralectotypes: 2 specimens (DEI), N Kamerun, L. Conradt, 96/ Syntypus/ Coll. Kraatz/ Kleine det.

According to Kolbe (1916: 59) the types of *Z. camerunus* var. *densepunctatus* are in MNHUB, but neither Damoiseau (1967a: 152) nor ourselves could find them there.

***rufostriatus* (Kleine, 1914)**

rufostriatus Kleine, 1914b: 160 (*Dactylobarus*).

Type depository. MZPW (HT).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea, Ivory Coast, Republic of the Congo, Rwanda.

ATOPOBRENTINI Damoiseau, 1965

Atopobrentini Damoiseau, 1965a: 1.

Atapobrentini: Damoiseau, 1987: 62 [lapsus].

***Anaraiorrhinus* Damoiseau, 1987**

Anaraiorrhinus Damoiseau, 1987: 69.

Type species: *Araiorrhinus conquisitus* Kleine, 1925.

Distribution. India (Assam), Malaysia.

Notes. The position of this genus is still to be clarified and it is provisionally included in the Atopobrentini. Nevertheless since it is very similar to *Araiorrhinus*, which is placed in the Microtrachelizini, perhaps *Anaraiorrhinus* should also be included in this tribe. Zimmerman (1994) does not elucidate the position of this genus because it is not present in Australia.

***conquisitus* (Kleine, 1925)**

conquisitus Kleine, 1925b: 138 (*Araiorrhinus*).

Type depository. NHM (HT!).

Distribution. India (Assam), Malaysia.

Atopobrentus Damoiseau, 1965

Atopobrentus Damoiseau, 1965a: 3.

Type species: *Atopobrentus allardi* Damoiseau, 1965.

Distribution. Democratic Republic of the Congo (ex Zaire).

allardi Damoiseau, 1965

allardi Damoiseau, 1965a: 3.

Type depositories. MRAC (HT!, PT!), IRSNB (PT!).

Distribution. Democratic Republic of the Congo (ex Zaire).

Neoceocephalus Senna, 1898

Neoceocephalus Senna, 1898c: 376.

Type species: *Neoceocephalus rostralis* Senna, 1898.

Distribution. Angola, Benin, Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea, Eritrea, Ethiopia, Gabon, Ghana, Ivory Coast, Kenya, Madagascar, Malawi, Mozambique, Nigeria, Republic of the Congo, Republic of Guinea, Senegal, Sierra Leone, Somalia, South Africa, Tanzania, Uganda, Zambia.

decellei De Muizon, 1959

decellei De Muizon, 1959: 78.

sulcifrons K. E. Schedl, 1961c: 201 (*Araiorrhinus*); syn. Damoiseau, 1963d: 13.

Type depositories. *N. decellei*: MRAC (HT!); *A. sulcifrons*: WSC (HT), IERTA (PT).

Distribution. Democratic Republic of the Congo (ex Zaire), Ivory Coast, Republic of the Congo.

Notes. K. E. Schedl (1961c: 201) reported that the holotype of *A. sulcifrons* is in IERTA and that one paratype is in his collection, but according to W. Schedl (2001, personal communication) the holotype is in his father's collection.

fausti (Senna, 1898)

fausti Senna, 1898c: 374 (*Araeorrhinus*).

Type depositories. MZUF (LT!, PLT!), DEI (PLT!), SMTD (PLT).

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Ivory Coast, Republic of the Congo, Republic of Guinea, South Africa, Tanzania.

Notes. Damoiseau (1967a: 251) reported that the specimen held in MZUF is the holotype, but in his original description Senna did not designate any of the syntypes as holotype; however, as there are two syntypes in MZUF, and as Damoiseau did not specifically select one as holotype, we designate the lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Lectotype of *A. fausti* (MZUF) designated on one specimen with the following labels: Stat. Albrechts-Höhe, N. Camerun, leg. L. Conradt/ Deutsch. Ent. Zeits., 1898, typus! ♂ / ex-coll. Senna/ "La Specola" Firenze, 6915. Paralectotypes: 1 specimen (MZUF), S.O. Kamerun, Stat. Lolodorf, L. Conradt/ *Araeorrhinus fausti* Senna, typus" ♂ / ex-coll. Senna/ "La Specola" Firenze, 6915; 3 specimens (DEI), S.O. Kamerun, Lolodorf, L. Conradt, 1895/ Syntypus/ coll. Kraatz/ Senna det./ Dtsch. Entomol. Institut Berlin; 7 specimens [on two labels but on the same pin] (DEI), S.O. Kamerun, Lolodorf, L. Conradt, 1895/ Syntypus/ coll. Kraatz/ *Araeorrhinus fausti*/ Senna det./ Dtsch. Entomol. Institut Berlin/ *Neoceocephalus fausti* Senna, R. Damoiseau det.

Damoiseau (1967a: 251) did not mention Tanzania (Kleine, 1922e: 154) and the Democratic Republic of the Congo (ex Zaire) (Kleine, 1924a: 111; 1936a: 27) when he gave the species distribution.

freyi De Muizon, 1960

freyi De Muizon, 1960: 214.

Type depository. NHMB (HT!).

Distribution. Democratic Republic of the Congo (ex Zaire), Kenya, Mozambique, South Africa, Tanzania, Zambia.

interrupticostatus (Kleine, 1922)

interrupticostatus Kleine, 1922h: 144 (*Araiorrhinus*).

interruptecostatus: Kleine, 1930f: 101 (*Araiorrhinus*) [lapsus].

interruptus De Muizon, 1955b: 903 (*Neoceocephalus*); syn. Damoiseau, 1963d: 18.

Type depositories. *A. interrupticostatus*: MNHUB (?) (ST); *N. interruptus*: MNHN (HT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Gabon, Ghana.

Notes. Damoiseau (1967a: 248) reported that the specimen of *A. interrupticostatus* held in MNHUB is the holotype, but in his original description Kleine did not designate any of the syntypes as holotype. We did not find any syntype for *A. interrupticostatus* in MNHUB.

malgasicus Damoiseau, 1963

malgasicus Damoiseau, 1963d: 25.

Type depository. IRSNB (HT!).

Distribution. Madagascar.

minutus Damoiseau, 1963

minutus Damoiseau, 1963d: 23.

Type depository. MRAC (HT!).

Distribution. Democratic Republic of the Congo (ex Zaire), Ghana, Uganda.

nasifer Damoiseau, 1987

nasifer Damoiseau, 1987: 70.

Type depositories. MNHN (HT!, AT!, PT!), IRSNB (PT!).

Distribution. Madagascar.

olseni De Muizon, 1960

olseni De Muizon, 1960: 214.

punctifrons K. E. Schedl, 1961c: 201 (*Araiorrhinus*); syn. Damoiseau, 1963d: 19.

Type depositories. *N. olseni*: NHMB (LT!), MNHN (PLT!); *A. punctifrons*: WSC (HT), IERTA (PT).

Distribution. Angola, Democratic Republic of the Congo (ex Zaire), Ghana, Ivory Coast, Republic of the Congo, Republic of Guinea, Sierra Leone.

Notes. Damoiseau (1967a: 249) reported that the specimen of *N. olseni* held in NHMB (Frey coll.) is the holotype, but in his original description De Muizon did not designate either of the syntypes as holotype; however, as there is only one syntype in NHMB, this must be considered as lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999). The lectotype has the following labels: N'Zerekoré Guin. Fr. 14-30.IV.52, S. H. Olsen A. O. F./ type/ Museum Frey Tutzing/ *Neoceocephalus olseni* J. De Muizon det.

K. E. Schedl (1961c: 201) reported that the holotype for *A. punctifrons* is in IERTA and that one paratype is in his collection, but according to W. Schedl (2001, personal communication) the holotype is in his father's collection.

ophthalmicus Calabresi, 1920

ophthalmicus Calabresi, 1920d: 38.

agilis Kleine, 1922g: 223 (*Araiorrhinus*); syn. Damoiseau, 1963d: 14.

Type depositories. *N. ophthalmicus*: MSNG (HT!); *A. agilis*: DEI (LT!).

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Gabon, Ghana, Ivory Coast, Republic of the Congo, Republic of Guinea, Somalia, Zambia.

Notes. Lectotype of *A. agilis* (DEI) designated on a specimen with the following labels: Kamerun, Conradt/ Syntypus/ coll. Kraatz/ Dtsch. Entomol. Institut Berlin/ *Araiorrhinus agilis* Kleine/ *Neoceocephalus sculpturatus* Senna.

We did not find the second syntype mentioned in the original description.

punctatus Damoiseau, 1961

punctatus Damoiseau, 1961a: 287.

Type depository. MRAC (HT!).

Distribution. Democratic Republic of the Congo (ex Zaire), Republic of Guinea.

Notes. The quotations for Cameroon, Republic of the Congo, Gabon and Ghana (Sforzi, 1998b: 276) are incorrect.

punicans Damoiseau, 1963

punicans Damoiseau, 1963d: 21.

Type depositories. NHMB (HT!), IRSNB (PT!).

Distribution. Democratic Republic of the Congo (ex Zaire), Ghana, Ivory Coast, Republic of Guinea.

Notes. The allotype specimen (NHMB) is missing from the card.

rostralis Senna, 1898

rostralis Senna, 1898c: 376.

longisulcatus K. E. Schedl, 1961c: 202 (*Araiorrhinus*); syn. Damoiseau, 1963a: 130.

Type depositories. *N. rostralis*: MZUF (LT!), DEI (PLT!); *A. longisulcatus*: MRAC (HT!).

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Republic of Guinea.

Notes. Lectotype of *N. rostralis* designated by Bartolozzi *et al.* (1985: 100) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

K. E. Schedl (1961c: 203) reported that one paratype for *A. longisulcatus* is in his collection, but according to W. Schedl (2001, personal communication) it is not in his father's collection.

sculpturatus Senna, 1898

sculpturatus Senna, 1898c: 377.

bifurcatus K. E. Schedl, 1961c: 200 (*Araiorrhinus*); syn. Damoiseau, 1963a: 129.

Type depositories. *N. sculpturatus*: MZUF (LT!), DEI (PLT!), SMTD (PLT); *A. bifurcatus*: MRAC (HT!, PT!), IERTA (PT).

Distribution. Benin, Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea, Eritrea, Ethiopia, Ghana, Ivory Coast, Malawi, Mozambique, Nigeria, Republic of the Congo, Senegal, Tanzania.

Notes. Damoiseau (1967a: 240) reported the the specimen of *N. sculpturatus* held in MZUF is the holotype, but in his original description Senna did not designate any of the syntypes as holotype; however, as there is only one syntype in MZUF, this must be considered as lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999). The lectotype has the following labels: Lolodorf, S. O. Camerun, leg. L. Conradt/ *Neoceocephalus sculpturatus* Senna, typus! / Deutsch. Ent. Zeitsch., 1898, H. 2/ typus! ♀ / ex coll. Senna/ "La Specola" Firenze, 6955.

K. E. Schedl (1961c: 200) reported that some paratypes of *A. bifurcatus* are in his collection, but according to W. Schedl (2001, personal communication) they are not in his father's collection.

Damoiseau (1967a: 241) did not mention Mozambique (Kleine, 1936a: 40) when he gave the species distribution.

***thoracicus* Damoiseau, 1963**

thoracicus Damoiseau, 1963d: 12.

Type depository. IRSNB (HT!).

Distribution. Democratic Republic of the Congo (ex Zaire).

HOPLOPISTHIINI Senna & Calabresi, 1919

Hoplopisthi Senna & Calabresi, 1919: 63.

Hoplopisthiini Damoiseau, 1963b: 201.

Hoplopisthiina Zimmerman, 1994: 130.

Hoplopisthinini: Environment Agency, 1995: 315 [lapsus].

Hoplopisthini Morrone, 1998: 132.

***Carcinopisthius* Kolbe, 1892**

Carcinopisthius Kolbe, 1892: 174.

Pseudotaphroderes Bolkay, 1911: 263; syn. Kleine, 1916c: 85.

Stratiopisthius Senna & Calabresi, 1919: 76; syn. Damoiseau, 1987: 32.

Garcinopisthius: Kleine, 1925f: 24 [lapsus].

Carinopisthius: Li-zhong, 2002: 296 [lapsus].

Type species: *Carcinopisthius fruhstorferi* Kolbe, 1892.

Distribution. Taiwan; Borneo, India (including Assam), Indonesia (Java, Mentawai, Moluccas, Sumatra), Malaysia, Myanmar [= Burma], Philippines, Vietnam; Australia, New Guinea, Solomon Is.

doriae (Senna, 1893)

doriae Senna, 1893c: 254 (*Hoplopisthius*).

doriai: Damoiseau, 1987: 43 [unjustified emendation].

Type depository. MSNG (HT!).

Distribution. Australia, New Guinea, Solomon Is.

forcipitiger Damoiseau, 1987

forcipitiger Damoiseau, 1987: 41.

Type depositories. HNHM (HT, AT), IRSNB (?) (PT), NHM (PT!).

Distribution. New Guinea.

Notes. Damoiseau (1987: 42) reported that paratypes are held in IRSNB, but we did not find them there.

fruhstorferi Kolbe, 1892

fruhstorferi Kolbe, 1892: 174.

interrupticosta Senna, 1898a: 224 (*Carcinopisthius*); syn. Damoiseau, 1987: 36.

Type depositories. *C. fruhstorferi*: MNHUB (HT!); *C. interrupticosta*: MSNG (HT!).

Distribution. Borneo, Indonesia (Java, Mentawai, Sumatra), Malaysia, Myanmar [= Burma]; New Guinea.

Notes. The material from the Moluccas Islands which Kleine identified as this species belongs to other taxa (Damoiseau, 1987: 38).

***kolbei* (Senna, 1893)**

kolbei Senna, 1893c: 255 (*Hoplopisthius*).

papuanus Senna, 1894e: 555 (*Carcinopisthius*); syn. Damoiseau, 1987: 39.

forficatus Bolkey, 1911: 264 (*Pseudotaphroderes*); syn. Kleine, 1916c: 85.

papuanus Bolkey, 1911: 265 (*Pseudotaphroderes*); syn. Kleine, 1916c: 85.

forficula Arrow, 1940: 320 (*Stratiopisthius*); syn. Damoiseau, 1987: 39.

Type depositories. *H. kolbei*: MSNG (LT!, PLT!); *C. papuanus*: MSNG (HT!); *P. forficatus*: HNHM (ST); *P. papuanus*: HNHM (ST); *S. forficula*: NHM (?) (LT, PLT).

Distribution. Indonesia (Moluccas); Australia, New Guinea.

Notes. Damoiseau (1987: 40) reported that the specimens for *H. kolbei* held in MSNG are holotype and paratypes, but in his original description Senna did not designate any of the syntypes as holotype; however, as there is more than one syntype in MSNG, and as Damoiseau did not specifically select one as holotype, we designate the lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Lectotype of *H. kolbei* (MSNG) designated on a specimen with the following labels: Nuova Guinea, Salvatti, XI.1875, Beccari/ typus/ *Flaminius spinator* Pow./ *Carcinopisthius kolbei* Senna typus! Paralectotypes: 2 specimens (MSNG), Nuova Guinea, Andai, VIII.72, L.M. D'Albertis; 1 specimen (MSNG), Nuova Guinea, Fly river, 1876/77, L.M. D'Albertis.

Damoiseau (1987: 40) reported that the specimens of *P. forficatus* held in HNHM are holotype and paratypes, but in his original description Bolkey did not designate any of the syntypes as holotype.

We did not find the lectotype and paralectotypes for *S. forficula* designated by Damoiseau (1987: 40) in NHM.

***lamingtoni* Damoiseau, 1987**

lamingtoni Damoiseau, 1987: 42.

Type depository. SAMA (HT).

Distribution. New Guinea.

***loriae* Senna & Calabresi, 1919**

loriae Senna & Calabresi, 1919: 72.

loriai: Damoiseau, 1987: 40 [unjustified emendation].

Type depository. MSNG (LT!, PLT!).

Distribution. New Guinea.

Notes. Damoiseau (1987: 40) reported that the specimens of *C. loriae* held in MSNG are holotype, allotype and paratypes, but in their original description Senna and Calabresi did not designate any of the syntypes as holotype; however, as there is more than one syntype in MSNG, and as Damoiseau did not specifically select one as holotype, we designate the lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Lectotype of *C. loriae* (MSNG) designated on a specimen with the following labels: N. Guinea, S. E. Moroka, 1300 m, VII-XI.93, Loria/ *Carcinopisthius loriae* Senna & Calabresi, typus! Paralectotypes: 3 specimens (MSNG), N. Guinea, S. E. Moroka, 1300 m, VII-XI.93, Loria.

oberthueri (Senna, 1893)

oberthueri Senna, 1893c: 255 (*Hoplopisthius*).

felschei Kolbe, 1892: 175 (*Carcinopisthius*); syn. Senna & Calabresi, 1894b: 359.

maculatus Senna, 1903: 163 (*Carcinopisthius*); syn. Damoiseau, 1966b: 448.

formosanus Bolkey, 1911: 265 (*Pseudotaphroderes*); syn. Senna & Calabresi, 1919: 71.

oberthuri: Kano, 1930: 153 (*Carcinopisthius*) [lapsus].

Type depositories. *H. oberthueri*: MNHN (?) (LT, PLT), MZUF (PLT!); *C. felschei*: MNHUB (LT!); *C. maculatus*: MSNG (HT!); *P. formosanus*: HNHN (HT).

Distribution. Taiwan; Borneo, India (including Assam), Indonesia (Sumatra), Malaysia, Myanmar [= Burma], Philippines, Vietnam.

Notes. We did not find the lectotype and paralectotype of *H. oberthueri* (Damoiseau, 1987: 38) in MNHN.

The material identified by Kleine as *C. oberthueri* from the Moluccas Islands and New Guinea belongs to other taxa (Damoiseau, 1987: 38).

Lectotype of *C. felschei* (MNHUB) designated on a specimen with the following labels: Ober-Burmah, Ruby Mines, 5000-7500 Fuss, Heyne V./ Felsche G./ 106368/ *Carcinop. felschei* n. Type/ *Carcinopisthius oberthueri* Senna. We were unable to find other syntypes of this species.

ocularis Damoiseau, 1987

ocularis Damoiseau, 1987: 42.

Type depository. ZMAN (HT!).

Distribution. New Guinea.

Hoplopisthius Senna, 1892

Hoplopisthius Senna, 1892e: 451.

Holopisthis: Kano, 1930: 153 [lapsus].

Moplopisthius: Li-zhong, 2002: 296 [lapsus].

Type species: *Hoplopisthius trichemerus* Senna, 1892.

Distribution. S China, Taiwan; Borneo, India (Assam), Indonesia (Bali, Java, Mentawai, Nias, Sulawesi, Sumatra), Laos, Malaysia, Myanmar [= Burma], Philippines (Luzon, Palawan), Vietnam; Europe (I).

celebensis Kolbe, 1892

celebensis Kolbe, 1892: 174.

Type depository. MNHUB (LT!, PLT!).

Distribution. Indonesia (Sulawesi).

Notes. Lectotype of *H. celebensis* (MNHUB) designated on a specimen with the following labels: S. Celebes, Bonthain, C. Ribbe, 1882/ 106366/ Type/ *Hoplopisth. celebensis* n. Type. Paralectotype: 1 specimen (MNHUB), with the same locality as the lectotype.

trichemerus Senna, 1892

trichemerus Senna, 1892e: 452.

javanus Kolbe, 1892: 173 (*Hoplopisthius*); syn. Senna, 1893e: 299.

trichimerus: Senna, 1893e: 291 (*Hoplopisthius*) [lapsus].

Type depositories. *H. trichemerus*: MSNG (HT!); *H. javanus*: MNHUB (LT!).

Distribution. S China, Taiwan; Borneo, India (Assam), Indonesia (Bali, Java, Mentawai, Nias, Sumatra), Laos, Malaysia, Myanmar [= Burma], Philippines (Luzon, Palawan), Vietnam; Europe (I).

Notes. Lectotype of *H. javanus* (MNHUB) designated on a specimen with the following labels: Ost Java, Fruhstorfer S./ Felsche G./ 106365/ Type/ *Hoplopisth. javanus* n. Type.

Ipsopisthius Kabakov, 2001

Ipsopisthius Kabakov, 2001: 215.

Type species: *Ipsopisthius hirtus* Kabakov, 2001.

Distribution. Vietnam.

hirtus Kabakov, 2001

hirtus Kabakov, 2001: 215.

Type depository. ZIN (HT).

Distribution. Vietnam.

STEREODERMINI Sharp, 1895

STEREODERMINI Sharp, 1895a: 7.

Stereodermina Sharp, 1895a: 7.

Stereoderminae Senna, 1896a: 210.

Stereodermini Kleine, 1922q: 148.

Stereoderminidae: Kleine, 1922h: 136 [lapsus].

Cerobates Schoenherr, 1840

Cerobates Schoenherr, 1840: 487.

Creobates: Kleine, 1922b: 31 [lapsus].

Cereobates: Kleine, 1922c: 132 [lapsus].

Type species: *Brentus tristriatus* Lund, 1800.

Distribution. Tropical Africa (Ivory Coast to South Africa); China, Japan, Taiwan; Borneo, India (including Andaman Is., Assam, Darjeeling), Indonesia (Batu; Engano; Flores; Java; Moluccas: Ambon, Seram [= Ceram], Halma-hera, Larat, Ternate; Mentawai; Nias; Sangie [= Sangi]; Sulawesi; Sumatra; Sumba; Sumbawa; Timor), Kampuchea [= Cambodia], Laos, Malaysia, Myanmar [= Burma], Philippines (Basilan, Luzon, Masbate, Mindanao, Negros, Palawan, Samar, Siargao), Sri Lanka, Thailand, Vietnam; Australia, Fiji Is., New Guinea, Solomon Is.; Europe (I).

subgenus *Cerobates* s. s.*aemulus* Kleine, 1922

aemulus Kleine, 1922f: 204.

clinatus Kleine, 1923j: 405 (*Cerobates*); syn. Damoiseau, 1987: 71.

Type depositories. *C. aemulus*: MZPW (HT); *C. clinatus*: Buitenzorg Museum (?) (HT).

Distribution. India, Indonesia (Java), Laos, Malaysia, Philippines (Mindanao), Thailand, Vietnam.

angustipennis Senna, 1895

angustipennis Senna, 1895a: 182.

adustus Senna, 1895a: 184 (*Cerobates*); syn. Damoiseau, 1987: 70.

Type depositories. *C. angustipennis*: RMNH (HT); *C. adustus*: RMNH (HT).

Distribution. Borneo, India (including Assam, Darjeeling), Indonesia (Java, Nias, Sumatra), Malaysia, Philippines (Luzon, Mindanao), Sri Lanka, Thailand, Vietnam; Fiji Is, New Guinea.

Notes. Damoiseau (1987: 71) designated the lectotype for *C. angustipennis*, but as the species was described by Senna on a single specimen, it is the holotype.

birmanicus Senna, 1894

birmanicus Senna, 1894b: 364.

birmanicus: Kleine, 1931a: 117 (*Cerobates*) [lapsus].

birmanensis: Kleine, 1933c: 289 (*Cerobates*) [lapsus].

Type depositories. MZUF (LT!, PLT!), MNHN (?) (PLT).

Distribution. Taiwan; India (including Andaman Is., Assam, Darjeeling), Myanmar [= Burma].

Notes. Lectotype *C. birmanicus* (MZUF) designated on a male specimen with the following labels: Rubis Mines, H. Birmanie, M. Doherty/ *Cerobates*

birmanicus Senna, cotypus!// Ann. Soc. Ent. Belg. XXXVIII, 1894, cotypi!// ex-coll. Senna/ "La Specola" Firenze, 6940. Paralectotype: 1 specimen (MZUF), Ruby Mines, H. B./ "La Specola" Firenze, 6925.

According to Kleine (1928a: 75) other syntypes should be in Oberthür's collection (MNHN and IRSNB), but we did not find them there.

***canaliculatus* Motschulsky, 1858**

canaliculatus Motschulsky, 1858: 96.

Type depository. ZMUM (HT).

Distribution. S China, Japan, Taiwan; Borneo, India (Darjeeling), Indonesia (Java, Mentawai, Sumatra), Malaysia, Myanmar [= Burma], Vietnam.

***cingulatus* Kleine, 1939**

cingulatus Kleine, 1939c: 106.

Type depository. NHM (HT!).

Distribution. New Guinea.

***collectivus* Kleine, 1925**

collectivus Kleine, 1925b: 130.

Type depository. NHM (LT!, PLT!).

Distribution. Myanmar [= Burma].

Notes. Lectotype of *C. collectivus* (NHM) designated on a specimen with the following labels: Syntype/ Typus/ Ruby Mines, Burma, 5,500 to 7,500 ft 1904-150 / *Cerobates collectivus* Kln, R. Kleine det. 1922. Paralectotype: 1 specimen (NHM) with the same label data as the lectotype.

***complanatus* Senna, 1896**

complanatus Senna, 1896a: 217.

Type depository. MSNG (LT!, PLT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea, Gabon, Ghana, Ivory Coast, Kenya, Nigeria, Republic of the Congo, Rwanda, Uganda.

Notes. Lectotype designated by Damoiseau (1967a: 203).

conveniens Kleine, 1924

conveniens Kleine, 1924a:100.

Type depositories. MRAC (LT!, PLT!), NHM (PLT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea, Gabon, Ghana, Ivory Coast, Kenya, Republic of the Congo, Senegal.

Notes. Damoiseau (1967a: 206) reported that the specimens held in MRAC are holotype (male) and allotype, but in his original description Kleine (1924a: 100) did not designate any of the syntypes as holotype; however, as there is only one male syntype in MRAC, this must be considered as lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

copiosus copiosus Kleine, 1924

copiosus copiosus Kleine, 1924a: 101.

Type depository. MRAC (LT!, PLT!).

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Gabon, Ghana, Kenya, Nigeria.

Notes. Lectotype and paralectotypes designated by Damoiseau (1967a: 211).

copiosus projectus Kleine, 1934

copiosus projectus Kleine, 1934a: 301.

Type depository. NHM (HT!, PT!).

Distribution. Democratic Republic of the Congo (ex Zaire), Ethiopia, Rwanda.

Notes. Kleine (1934a: 302) quoted a cotype in DEI, but we did not find this specimen neither it is listed in Döbler & Gaedike (1995).

***corruptus* Kleine, 1926**

corruptus Kleine, 1926f: 362.

Type depository. NHM (HT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Gabon, Kenya, Mozambique, Republic of the Congo, Tanzania.

***costatus* Kleine, 1922**

costatus Kleine, 1922r: 153.

Type depository. SMTD (?) (HT).

Distribution. Philippines (Mindanao).

Notes. Kleine (1928a: 75) reported that the type for this species was in SMTD; according to O. Jaeger (SMTD) (2002, personal communication) it is not in the SMTD collections.

***credibilis* Kleine, 1926**

credibilis Kleine, 1926f: 363.

kleinei Müller, 1939: 242 (*Cerobates*); syn. Damoiseau, 1967a: 189.

Type depositories. *C. credibilis*: NHM (HT!); *C. kleinei*: MCST (HT, PT).

Distribution. Ethiopia, Sudan.

Notes. Müller in his original description of *C. kleinei* (1939: 242) designated the holotype and five paratypes. Damoiseau (1967a: 191) incorrectly reported that syntypes of *C. kleinei* are in DEI, but Döbler & Gaedike (1995) did not mention them.

***cruentatus* Senna, 1898**

cruentatus Senna, 1898c: 372.

Type depositories. MZUF (LT!), SMTD (PLT).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea (Bioko Is. [= Fernando Poo]), Gabon, Ghana, Ivory Coast, Kenya, Republic of the Congo, Rwanda, Togo, Uganda.

Notes. Damoiseau (1967a: 192) reported that the specimen held in MZUF is the holotype, but in his original description Senna did not designate any of the syntypes as holotype; however, as there is only one syntype in MZUF, this must be considered as lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999). The lectotype has the following labels: Stat. Joh. Albrecht Höhe, N Kamerun, leg. Conradt, ♂ / *Cerobates cruentatus* Senna, typus! / Deutsch. Ent. Zeit., Heft II, 1898, typus! / ex coll. Senna / "La Specola" Firenze, 7088.

Döbler & Gaedike (1995: 443) quoted four syntypes in DEI, but their localities do not correspond to those of the original description.

Damoiseau (1967a: 192) did not mention Togo (Kleine, 1924a: 110) when he gave the species distribution.

***debilis* J. Thomson, 1858**

debilis J. Thomson, 1858: 118.

sulcata Kolbe, 1897: 286 (*Cerobates debilis* var.); **syn. n.**

Type depositories. *C. debilis*: MNHN (LT, PLT?); *C. debilis* var. *sulcata*: unknown.

Distribution. Tropical Africa (Ivory Coast to South Africa).

Notes. Damoiseau (1967a: 197) designated lectotype and paralectotypes on specimens of MNHN (Deyrolle coll.). We did not find these lectotypes in MNHN, but we found 3 specimens there with the following labels "Gabon/ Thomson Type 122" and "Gabon/ Thomson Type 112" which seem to belong to the original series of Thomson.

***dilutus* Kleine, 1924**

dilutus Kleine, 1924a: 102.

fortunatus Kleine, 1924a: 103 (*Cerobates*); **syn.** De Muizon, 1955b: 877.

Type depositories. *C. dilutus*: MRAC (LT!), SMTD (PLT); *C. fortunatus*: MRAC (HT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea, Ethiopia, Ivory Coast, Sudan, Tanzania, Uganda.

Notes. Damoiseu (1967a: 215) reported that the specimens of *C. dilutus* held in MRAC are holotype and allotype, but in his original description Kleine did not designate any of the syntypes as holotype; however, as there is only one syntype in MRAC, this must be considered as lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999). The lectotype [incorrectly labelled as holotype] has the following labels: Holotypus/ Musée du Congo, Kindu, XI.1913, L. Burgeon/ Typus!/ R. det. L. 747/ *Cerobates dilutus* Kln. typus!, 1922/ Prépar. génit. n° 4/10.28.01 R. Damoiseau/ *Cerobates* (*Cerobates*) *dilutus* Kleine stat. nov., R. Damoiseau det., 1964.

endroedyi Damoiseau, 1972

endroedyi Damoiseau, 1972a: 265.

Type depositories. HNHM (HT, AT, PT), IRSNB (PT!).

Distribution. Ghana.

enganoensis Damoiseau, 1962

enganoensis Damoiseau, 1962e: 111.

Type depository. MSNG (HT!).

Distribution. Indonesia (Engano).

fleutiauxi De Muizon, 1955

fleutiauxi De Muizon, 1955b: 883.

fleutiauxii: Damoiseau, 1963a: 125 (*Cerobates*) [lapsus].

Type depository. MNHN (HT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Ghana.

***formosanus* Schönfeldt, 1911**

formosanus Schönfeldt, 1911: 190.

Type depository. DEI (LT!, PLT!).

Distribution. Japan, Taiwan; Philippines (Luzon, Mindanao, Negros), Vietnam.

Notes. This species was described on a series of three syntypes and Damoiseau (1987: 84) designated lectotype, allolectotype and paralectotype. We only found the lectotype and paralectotype in DEI. In NHM there is a specimen of the species labelled as "cotypus" but it can not belong to the type series because the collecting date is 1912, posterior to the published one.

Damoiseau (1987: 74) did not mention Philippines (Kleine, 1931h: 404) when he gave the species distribution.

***fossulatus* Motschulsky, 1858**

fossulatus Motschulsky, 1858: 96.

articulosus Kleine, 1923d: 134 (*Cerobates*); syn. Damoiseau, 1987: 73.

Type depositories. *C. fossulatus*: SMTD (LT, PLT); *C. articulosus*: NHM (HT!).

Distribution. India (including Assam), Indonesia (Java, Sumatra), Malaysia, Myanmar [= Burma], Vietnam.

Notes. According to Kleine (1929a: 51) the type for *C. fossulatus* was in ZMUM; Damoiseau (1987: 73) designated lectotype, allolectotype and paralectotype on material of SMTD.

***grouvellei* Senna, 1893**

grouvellei Senna, 1893e: 307.

Type depositories. MZUF (LT!), SMTD (PLT).

Distribution. Borneo, Indonesia (Sumatra), Philippines (Mindanao, Palawan); Australia; Europe (I).

Notes. Lectotype designated by Bartolozzi *et al.* (1985: 97) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

hybridus Senna, 1898

hybridus Senna, 1898e: 371.

aduncus Kleine, 1922f: 205 (*Cerobates*); syn. Damoiseau, 1967a: 217.

Type depositories. *C. hybridus*: MZUF (LT!), DEI (PLT!), SMTD (PLT); *C. aduncus*: MZPW (HT).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea (including Bioko Is. [= Fernando Poo]), Gabon, Ghana, Ivory Coast, Kenya, Nigeria, Republic of the Congo, Sudan, Tanzania.

Notes. Lectotype designated by Bartolozzi *et al.* (1985: 97) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

laevipennis Senna, 1896

laevipennis Senna, 1896a: 219.

aequalis Kleine, 1922f: 203 (*Cerobates*); syn. Damoiseau, 1971b: 270.

Type depositories. *C. laevipennis*: MZUF (HT!); *C. aequalis*: MZPW (ST).

Distribution. Borneo, India (including Andaman Is.), Indonesia (Java, Moluccas, Sangi Is., Sulawesi, Sumatra, Sumbawa), Laos, Malaysia, Myanmar [= Burma], Philippines (Luzon, Mindanao, Samar), Sri Lanka, Thailand, Vietnam; Australia, Fiji Is., New Guinea, Solomon Is.

Notes. Damoiseau (1987: 75) incorrectly reported that the holotype for *C. laevipennis* is in NHM and paratypes are in MZUF, but the species description was based on a single specimen which is in MZUF.

matanganus Damoiseau, 1987

matanganus Damoiseau, 1987: 75.

Type depository. NHM (HT!, AT!).

Distribution. Borneo.

pidigala Damoiseau, 1963*pidigala* Damoiseau, 1963i: 38.**Type depository.** MRAC (HT!).**Distribution.** Angola, Democratic Republic of the Congo (ex Zaire), Kenya, Senegal.***planicollis*** Morimoto, 1976*planicollis* Morimoto, 1976a: 276.**Type depository.** ELKU (HT, PT).**Distribution.** Japan.***puerilis*** Kleine, 1936*puerilis* Kleine, 1936c: 137.**Type depository.** NHM (HT!).**Distribution.** Malaysia.***punctulatus*** Senna, 1898*punctulatus* Senna, 1898c: 373.**Type depositories.** MZUF (LT!), DEI (PLT!), MRAC (PLT!), SMTD (PLT).**Distribution.** Cameroon, Democratic Republic of the Congo (ex Zaire), Gabon, Ghana, Ivory Coast, Nigeria, Republic of the Congo, Tanzania.**Notes.** Lectotype designated by Bartolozzi *et al.* (1985: 99) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).***sennae*** Calabresi, 1920*sennae* Calabresi, 1920d: 25.*sennai* Damoiseau, 1967a: 219 [unjustified emendation].**Type depository.** MSNG (LT!, PLT!).

Distribution. Democratic Republic of the Congo (ex Zaire), Equatorial Guinea, Gabon, Ivory Coast, Republic of the Congo, São Thomé and Príncipe.

Notes. Damoiseau (1967a: 220) reported that the specimens held in MSNG are holotype and allotype, but in her original description Calabresi did not designate any of the syntypes as holotype; however, as there is more than one syntype in MSNG, and as Damoiseau did not specifically select one as holotype, we designate the lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Lectotype of *C. sennae* (MSNG) designated on a male specimen with the following labels: Is. S. Thomé, Agua-Ize, XII.1900, 400-700 m, L. Fea/ Lectotype ♂/ *Cerobates sennae* Calabresi lectotype ♂ R. Damoiseau rev. 1962 [*in litteris*]. Paralectotypes: 67 specimens (MSNG), Is. Príncipe, Roca Inf. D. Henrique, I-IV.1901, 100-300 m, L. Fea; 3 specimens (MSNG), Is. S. Thomé, Agua-Ize, XII.1900, 400-700 m, L. Fea; 2 specimens (MSNG), Congo Francese, Ncogo', XII.1902, L. Fea; 1 specimen (MSNG), Is. Príncipe, Bahia do Oeste, V-VI.1901, 100-200 m, L. Fea.

sexsulcatus Motschulsky, 1858

sexsulcatus Motschulsky, 1858: 95.

glaberrima Senna, 1892e: 450 (*Cerobates sexsulcatus* var.); syn. Senna, 1893b: 264.

sexmaculatus: Kleine, 1931a: 117 (*Cerobates*) [lapsus].

Type depositories. *C. sexsulcatus*: ZMUM (ST); *C. sexsulcatus* var. *glaberrima*: MSNG (LT!, PLT!), MNHN (PLT!).

Distribution. China; Borneo, India (including Andaman Is., Assam), Indonesia (Batu; Engano; Java; Moluccas: Seram [= Ceram], Halmahera, Larat, Ternate; Nias; Sulawesi; Sumatra; Sumbawa), Kampuchea [= Cambodia], Laos, Malaysia, Myanmar [= Burma], Philippines (Basilan, Luzon, Masbate, Mindanao, Palawan), Sri Lanka, Thailand, Vietnam; Australia, New Guinea; Europe (I).

Notes. Lectotype of *C. sexsulcatus* v. *glaberrima* (MSNG) designated on a specimen with the following labels: Bhamò, Birmania, VIII.1885, Fea/ typus/ *Cerobates sexsulcatus* var. *glaberrima* Senna. Paralectotypes: 1 specimen (MSNG), Bhamò, Birmania, VI.1886, Fea; 1 specimen (MSNG), Carin Chebà, 900-1100 m, L. Fea, V-XII.1888; 1 specimen (MSNG), Tenasserim, Thagatà, Fea, Apr. 1887; 6 specimens (MNHN), Bhamò, Birmania, VIII.1886, Fea.

***sulcatus sulcatus* Boheman, 1840**

sulcatus sulcatus Boheman (in Schoenherr, 1840: 488).

Type depository. NHRS (HT).

Distribution. South Africa.

***sulcatus sulcirostris* J. Thomson, 1858**

sulcatus sulcirostris J. Thomson, 1858: 119.

Type depository. MNHN (?) (LT, PLT).

Distribution. Tropical Africa.

Notes. Damoiseau (1967a: 199) designated lectotype and paralectotypes in MNHN, but we did not find them there.

***sumatranus* Senna, 1893**

sumatranus Senna, 1893e: 306.

andamanicus Senna, 1896a: 223 (*Cerobates*); syn. Damoiseau, 1987: 72.

Type depositories. *C. sumatranus*: MZUF (LT!), SMTD (PLT), RMNH (PLT); *C. andamanicus*: MZUF (HT!).

Distribution. Taiwan; Borneo, India (Andaman Is., Assam, Darjeeling), Indonesia (Java, Mentawai, Sulawesi, Sumatra, Timor), Malaysia, Philippines (Luzon, Mindanao), Sri Lanka, Thailand, Vietnam; Europe (I).

Notes. Lectotype of *C. sumatranus* designated by Bartolozzi *et al.* (1985: 101) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

***tristriatus* (Lund, 1800)**

tristriatus Lund, 1800: 66 (*Brentus*).

elytralis Senna, 1897c: 545 (*Cerobates tristriatus* ssp.); syn. Schönfeldt, 1908: 21.

longirostris Senna, 1898a: 227 (*Cerobates tristriatus* ssp.); syn. Schönfeldt, 1908: 21.

Type depositories. *B. tristriatus*: ZMUC (HT); *C. tristriatus* ssp. *elytralis*: MSNG (LT!, PLT!); *C. tristriatus* ssp. *longirostris*: MSNG (LT!, PLT!).

Distribution. Borneo, India (Andaman Is., Assam), Indonesia (Batu, Engano, Java, Mentawai, Moluccas, Nias, Sulawesi, Sumatra, Sumba), Malaysia, Myanmar [= Burma], Philippines (Basilan, Luzon, Mindanao, Palawan, Samar, Siargao), Sri Lanka, Thailand, Vietnam; Australia, New Guinea; Europe (I).

Notes. Lectotype of *C. tristriatus elytralis* (MSNG) designated on a specimen with the following labels: Is. Batu, 1896-97, H. Raap/ *Cerobates tristriatus* Lund *elytralis* Senna. Paralectotype: 1 specimen (MSNG), Is. Batu, 1896-97, H. Raap.

Lectotype of *C. tristriatus longirostris* (MSNG) designated on a specimen with the following labels: Mentawai Si Oban, IV-VIII.94, Modigliani/ *tristriatus* (Senna)/ *longirostris* Senna. Paralectotypes: 24 specimens (MSNG), Mentawai, Si Oban, IV-VIII.94, Modigliani; 6 specimens (MSNG), Mentawai, Sipora Sereinu, V-VI.94, Modigliani.

turgidulus Damoiseau, 1987

turgidulus Damoiseau, 1987: 78.

Type depositories. DEI (HT!, AT!, PT!), IRSNB (PT!), SMTD (PT).

Distribution. Taiwan.

Notes. Döbler & Gaedike (1995) did not include these types in their catalogue.

vitiensis Fairmaire, 1881

vitiensis Fairmaire, 1881c: 422.

Type depository. ZMUH (?) (HT).

Distribution. Fiji Is.

Notes. Kleine (1928a: 75) wrote that probably the type for this species is in ZMUH but we were unable to verify this statement.

subgenus *Ionthocerus* Lacordaire, 1866

Ionthocerus Lacordaire, 1866: 415.

Ionthocerus: Gemminger & Harold, 1872: 2705 [lapsus].

Ionthoceras: Gardner, 1935: 143 [lapsus].

Type species: *Ionthocerus crematus* Lacordaire, 1866.

Notes. According to O. N. Kabakov (2001: 209) *Ionthocerus* should be considered as a valid genus and not a subgenus, on account of "...being different both in morphology and mode of life...". Here we consider it as a subgenus, awaiting further studies on this matter.

asiaticus Kleine, 1920

asiaticus Kleine, 1920f: 47 (*Ionthocerus*).

angustifrons Kleine, 1922q: 148 (*Ionthocerus*); syn. Damoiseau, 1987:80.

Type depositories. *J. asiaticus*: DEI (LT!, PLT!), IRSNB (PLT!), NHRS (PLT), SMTD (PLT), ZMUH (PLT); *J. angustifrons*: ZMAN (?) (ST), MZPW (?) (ST).

Distribution. S China, Japan, Taiwan; Borneo, Indonesia (Java, Sumatra), Malaysia, Philippines (Luzon, Mindanao, Palawan), Sri Lanka, Thailand, Vietnam.

Notes. Lectotype of *J. asiaticus* (DEI) designated on a specimen with the following labels: Syntypus/ Formosa, Taihorisho, Sauter, III.09/ Kleine det./ *Ionthocerus asiaticus* Kleine, 1920. Paralectotypes: 1 specimen (DEI), Formosa, Taihorisho, Sauter, IX.09/ v. Schönfeldt det./ Syntypus/ DEI/ *Cerobates* (*Ionthocerus*) *asiaticus* Kleine, R. Damoiseau det., 1967; 1 specimen (IRSNB), Butuan Mindanao Baker/ Staatl. Museum für Tierkunde, Dresden/ Kleine det. 1919 *Ionthocerus asiaticus* n. sp./ Paratype/ *Cerobates* (*Ionthocerus*) *laticostatis* Kleine R. Damoiseau det./ Prépar. génit. No. 67/080301, R. Damoiseau.

According to Kleine (1928a: 74) the types for *J. angustifrons* were in ZMAN and in his collection, but we did not find them in ZMAN, and according to T. Huflejt (MZPW) (2002, personal communication) they are not in MZPW, where Kleine's collection is held.

***bicolor* Heller, 1916**

bicolor Heller, 1916: 297 (*Jonthocerus*).

Type depository. SMTD (HT!).

Distribution. Taiwan; Borneo, Indonesia (Sumatra), Philippines (Luzon), Sri Lanka.

***burgeoni* De Muizon, 1955**

burgeoni De Muizon, 1955b: 873 (*Jonthocerus*).

Type depository. MNHN (HT!).

Distribution. Democratic Republic of the Congo (ex Zaire), Ghana, Sierra Leone.

Notes. The quotation for the People's Republic of Congo (Sforzi, 1998b: 274) is incorrect and should be referred to as the Democratic Republic of the Congo.

***carinensis* Senna, 1892**

carinensis Senna, 1892e: 450 (*Cerobates canaliculatus* var.).

caesius Kleine, 1944: 151 (*Jonthocerus*); syn. Damoiseau, 1987: 82.

caesus: Kabakov, 2001: 209 (*Jonthocerus*) [lapsus].

Type depositories. *C. canaliculatus* var. *carinensis*: MSNG (LT!, PLT!), IRSNB (PLT!), MNHN (PLT!), MZUF (PLT!); *J. caesius*: MNHN (HT!).

Distribution. S China; India (Darejeeling), Laos, Malaysia, Myanmar [= Burma], Vietnam.

Notes. Lectotype of *J. canaliculatus* var. *carinensis* (MSNG) designated on a specimen with the following labels: Carin, Asciuii Ghecù, 1400-1500 m, III.IV.1988, L. Fea/ typus/ *Cerobates canaliculatus* Mot. var. *carinensis* Senna. Paralectotypes: 3 specimens (MSNG), Carin, Asciuii Ghecù, 1400-1500 m, III.IV.1988, L. Fea; 1 specimen (MNHN), H.te Birmanie, Mines des Rubis, 1200-2300 m, Doherty 1890; 1 specimen (MZUF), Carin, Asciuii Ghecù, 1400-1500 m, III.IV.1988, L. Fea/ *J. carinensis* (Senna) cotypi! ♀ ♀/ *C. canaliculatus* v. *carinensis* n. Senna/ Ann. Mus. Genova, 1892, A. Soc. Ent. Belg., 1894, cotypi ♀ ♀/ ex-coll. Senna/ "La Specola" Firenze, 6926; 1 specimen (MZUF), Carin, Asciuii Ghecù, 1400-1500 m, III.IV.1988, L. Fea/ ex-coll. Senna/ "La Specola" Firenze, 6927; 1 specimen (MZUF), Miniere di rubini, 1200-2300 m, Alta Birmania/ ex-coll. Senna/ "La Specola" Firenze, 6995/ 1 female specimen (MZUF), Miniere di rubini, 1200-2300 m, Alta Bir-

mania/ ♀/ *Cerobates (Jonthocerus) carinensis* Senna, R. Damoiseau det./ ex-coll. Senna/ "La Specola" Firenze, 6995; 1 female specimen (IRSNB), Carin, Asciuii Ghecù, 1400-1500 m, III.IV.1988, L. Fea/ *Cer. canaliculatus* var. *carinensis* Senna n./ *Jonthocerus carinensis* Senna Allotype ♀ R. Damoiseau det. 1961/ Allotype; 1 female specimen (IRSNB), Carin, Asciuii Ghecù, 1400-1500 m, III.IV.1988, L. Fea/ *Jonthocerus. carinensis* Senna ♀/ *Jonthocerus. carinensis* Senna, Paratype ♀, R. Damoiseau det. 1961/ Paratype.

***conradti* Senna, 1898**

conradti Senna, 1898e: 374 (*Jonthocerus*).

Type depositories. MZUF (LT!), DEI (PLT!), SMTD (PLT).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea (including Bioko Is. [= Fernando Poo]), Gabon, Ghana, Ivory Coast, Nigeria, Republic of the Congo, Senegal, Sierra Leone, Sudan, Tanzania, Togo, Uganda.

Notes. Lectotype designated by Bartolozzi *et al.* (1985: 95) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Damoiseau (1967a: 178) did not mention Tanzania (Schönfeldt, 1909: 16) and Togo (Kleine, 1920f: 39) when he gave the species distribution.

***crematus* Lacordaire, 1866**

crematus Lacordaire, 1866: 416 (*Jonthocerus*).

concisus Kleine, 1925b: 130 (*Cerobates*); syn. Damoiseau, 1971b: 270.

Type depositories. *I. crematus*: IRSNB (HT!); *C. concisus*: NHM (LT!, PLT!).

Distribution. China (Hainan); India (including Andaman Is.), Sri Lanka.

Notes. Lectotype of *C. concisus* (NHM) designated on a specimen with the following labels: Syntype/ Typus/ Ceylon/ 59-106/ *Cerobates concisus* Kleine, R. Kleine det. 1922. Paralectotype: 1 specimen (NHM), Syntype/ Ceylon/ 59-106/ *Cerobates ihrasco* Dohrn.

***curtus* Damoiseau, 1967**

curtus Damoiseau, 1967b: 6 (*Jonthocerus*).

Type depository. HNHM (HT).

Distribution. Republic of the Congo.

dalmoni Mantilleri, 2002

dalmoni Mantilleri, 2002: 335.

Type depository. MNHN (HT).

Distribution. Cameroon.

decorsei De Muizon, 1955

decorsei De Muizon, 1955b: 872 (*Jonthocerus*).

Type depository. MNHN (HT!).

Distribution. Angola, Cameroon, Central African Republic, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea.

elegans Damoiseau, 1963

elegans Damoiseau, 1963a: 122 (*Jonthocerus*).

Type depositories. IRSNB (HT!, PT!), MRAC (AT!, PT!), SMTD (PT).

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Ghana, Republic of the Congo, Republic of Guinea, Uganda.

Notes. Damoiseau in his original description writes that the holotype is in MRAC, but actually it is in IRSNB. He also mentions paratypes in MSNG but we did not find them.

foveolatus Senna, 1893

foveolatus Senna, 1893e: 300 (*Jonthocerus*).

Type depositories. MZUF (LT!), RMNH (PLT), SMTD (PLT).

Distribution. Indonesia (Sumatra), Europe (I).

Notes. Lectotype designated by Bartolozzi *et al.* (1985: 96) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

***laevithorax* Damoiseau, 1987**

laevithorax Damoiseau, 1987: 86 (*Jonthocerus*).

Type depository. BPBM (HT).

Distribution. Philippines (Luzon).

***laticostatis* Kleine, 1920**

laticostatis Kleine, 1920f: 38 (*Jonthocerus*).

laticostatus: Kobakov, 2001 (*Jonthocerus*) [lapsus].

laticostalis: Li-zhong, 2002: 296 [lapsus].

Type depositories. DEI (HT!).

Distribution. Taiwan.

Notes. Li-zhong (2002) also quotes the species for Japan, Philippines, Indonesia, and Borneo.

***maai* Damoiseau, 1987**

maai Damoiseau, 1987: 85 (*Jonthocerus*).

Type depository. BPBM (HT).

Distribution. Thailand.

***mentaweicus* Senna, 1898**

mentaweicus Senna, 1898a: 228 (*Jonthocerus*).

Type depository. MSNG (HT!).

Distribution. Indonesia (Mentawai).

***mimus* Senna, 1898**

mimus Senna, 1898a: 229 (*Jonthocerus*).

sumatranus Senna, 1898f: 191 (*Jonthocerus mimus* var.); syn. Damoiseau, 1987: 86.

Type depositories. *J. mimus*: MSNG (HT!); *J. mimus* var. *sumatranus*: MSNG (HT!).

Distribution. Borneo, Indonesia (Mentawai).

Notes. Senna redescribed the variety *sumatranus* in 1903 (p. 164).

***modiglianii* Senna, 1898**

modiglianii Senna, 1898a: 228 (*Jonthocerus*).

Type depository. MSNG (ST!).

Distribution. Borneo, India (Andaman Is.), Indonesia (Java, Mentawai, Sumatra), Philippines (Mindanao).

Notes. Senna (1898a: 228) mentions a second male specimen in the De-sbrochers collection.

***nigripes* Lewis, 1883**

nigripes Lewis, 1883: 298 (*Jonthocerus*).

Type depository. NHM (LT!, PLT!).

Distribution. Japan, Taiwan.

Notes. Damoiseau (1987: 87) reported that the specimens held in NHM are holotype, allotype and paratypes, but in his original description Lewis did not designate any of the 20 syntypes as holotype; however, as there is more than one syntype in NHM, and as Damoiseau did not specifically select one as holotype, we designate the lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Lectotype of *J. nigripes* (NHM) designated on a male specimen with the following labels: Syntype/ Type H. T./ sp. figured/ Japan, G. Lewis 1910-320/ Ichiuchi, 30.IV-2.V.81/ *Jonthocerus nigripes* Lewis. Paralectotypes: 19 specimens (NHM) with the same label data as the lectotype.

***nigrothorax* Damoiseau, 1987**

nigrothorax Damoiseau, 1987: 88 (*Jonthocerus*).

Type depositories. AMNH (HT, AT, PT), IRSNB (PT!).

Distribution. New Guinea.

***ophthalmicus* Pascoe, 1872**

ophthalmicus Pascoe, 1872: 320 (*Jonthocerus*).

australasiae Fairmaire, 1882: 463 (*Cerobates*); syn. Damoiseau, 1987: 81.

Type depositories. *I. ophthalmicus*: NHM (HT!); *C. australasiae*: SAMA (?) (HT).

Distribution. Japan, Taiwan; Indonesia (Flores, Moluccas, Sumatra); Australia.

Notes. According to Kleine (1928a: 75) the type for *C. australasiae* was in SAMA; according to Damoiseau (1987: 82) it could be in Oberthür's collection (MNHN), but we did not find it there.

Morimoto (1976a: 277) did not mention this species among the Brentidae of Japan.

papuensis Macleay, 1887

papuensis Macleay, 1887: 194 (*Jonthocerus*).

Type depository. AMS (HT).

Distribution. New Guinea.

Notes. Kleine (1938d: 50) mentioned New South Wales as type locality, but actually the species was described from the "Fly River, New Guinea". Hahn (1962) and Britton & Stanbury (1981) did not mention this species, neither did Damoiseau (1987). Remarks on this species are in Zimmerman (1994: 164).

pasteuri Senna, 1898

pasteuri Senna, 1898d: 55 (*Jonthocerus*).

Type depository. MZUF (HT!).

Distribution. Indonesia (Java, Sumatra), Philippines (Luzon).

Notes. Damoiseau (1987: 88) wrote that paratypes are also held in MZUF, but species description seems to be based on a single male specimen.

sondaicus Senna, 1893

sondaicus Senna, 1893e: 302 (*Jonthocerus*).

angulaticeps Senna, 1898d: 53 (*Jonthocerus*); syn. Damoiseau, 1987: 79.

borneensis Senna, 1898f: 190 (*Jonthocerus angulaticeps* var.); syn. Damoiseau, 1987: 79.
sandaicus: Edwards, 1988: 489 (*Cerobates*) [lapsus]

Type depositories. *J. sondaicus*: MZUF (LT!), IRSNB (PLT!), MSNG (PLT!), RMNH (PLT!); *J. angulaticeps*: MZUF (LT!, PLT!), RMNH (PLT!); *J. angulaticeps* var. *borneensis*: unknown.

Distribution. Taiwan; Borneo, Indonesia (Java, Sumatra), Malaysia; Europe (I).

Notes. Damoiseau (1987: 80) reported that the specimens of *J. sondaicus* held in MZUF are holotype and allotype and that some paratypes are held in RMNH and IRSNB, but Senna in his original description did not designate any of the five specimens as holotype; however, as there is only one syntype in MZUF, this must be considered as lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999). The lectotype has the following labels: Sumatra, import. in Europa col tabacco secco, leg. E. Grouvelle/ *Jonthocerus sondaicus* Senna, ♂, cotypus! Bull. Soc. Ent. Ital., XXV, 1893, cotypus! ♂/ *Cerobates (Jonthocerus) sondaicus* Senna, R. Damoiseau det., 1966/ ex-coll. Senna/ "La Specola" Firenze, 6931.

Lectotype of *J. angulaticeps* (MZUF) designated on a male specimen with the following labels: Giava occid., leg. I. D. Pasteur, ♂/ *Jonthocerus angulaticeps* ♂ ♀, cotypi! Senna/ Notes Leyd. Mus. XX, 1898, cotypi! ♂ ♂ ♀ ♀/ ex-coll. Senna/ "La Specola" Firenze, 6925. Paralectotypes: 1 female specimen (MZUF), Giava occid., leg. dr. Pasteur/ *Jonthocerus angulaticeps* ♂ ♀, cotypi, Senna dupl./ *Cerobates (Jonthocerus) sondaicus* Senna, R. Damoiseau det./ ex-coll. Senna/ "La Specola" Firenze, 6925; 1 male specimen (MZUF), Giava occ., leg. Pasteur, ♂/ *Cerobates (Jonthocerus) sondaicus* Senna, R. Damoiseau det./ Prép. Micr. N° 1967/ 08.03.02/ ex-coll. Senna/ "La Specola" Firenze, 6925; 1 female specimen (MZUF), Giava occid., leg. I. D. Pasteur, ♀/ *Cerobates (Jonthocerus) sondaicus* Senna, R. Damoiseau det./ ex-coll. Senna/ "La Specola" Firenze, 6925; 1 male and 2 female specimens (MZUF), Giava occid., leg. I. D. Pasteur/ ex-coll. Senna/ "La Specola" Firenze, 6925.

usambaricus Senna, 1896

usambaricus Senna, 1896a: 216.

Type depository. Unknown.

Distribution. Democratic Republic of the Congo (ex Zaire), Nigeria, Tanzania, Zambia.

Notes. In his Monograph on the African Brentidae Damoiseau (1967a: 474) did not redescribe the species but only listed it with a question mark in the genus *Ionthocerus*.

zanzibaricus Senna, 1898

zanzibaricus Senna, 1898f: 193 (*Ionthocerus*).

Type depository. MSNG (LT!, PLT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Ivory Coast, Kenya, Tanzania.

Hyperephanus Senna, 1892

Hyperephanus Senna, 1892d: 153.

Homoeolizus Sharp, 1895a: 18; syn. *Kleine*, 1927a: 16.

Type species: *Hyperephanus hirsutus* Senna, 1892 = *Ephebocerus hirtellus* Erichson, 1847.

Distribution. Costa Rica, Mexico, Panama; Bolivia, Brazil, Chile, Peru.

hirtellus (Erichson, 1847)

hirtellus Erichson, 1847: 126 (*Ephebocerus*).

pilosus Kirsch, 1875a: 281 (*Stereodermus*); syn. *Calabresi*, 1920a: 39.

hirsutus Senna, 1892d: 155 (*Hyperephanus*); syn. *Calabresi*, 1920a: 39.

salvini Sharp, 1895a: 18 (*Homoeolizus*); syn. *Calabresi*, 1920a: 39.

Type depositories. *E. hirtellus*: MNHUB (?) (HT); *S. pilosus*: SMTD (?) (HT); *H. hirsutus*: MZUF (HT!); *H. salvini*: NHM (LT!, PLT!).

Distribution. Costa Rica, Mexico, Panama; Bolivia, Brazil, Chile, Peru.

Notes. Lectotype of *H. salvini* (NHM) designated on a male specimen with the following labels: *Homoeolizus salvini* ♂ Type, D. S., Bugaba, Champion/ Syntype/ type/ B.C.A., Col. IV, 6 *Stereodermus pedator* Sharp/ Sharp Coll. 1905-313. Paralectotypes: 30 specimens (NHM) with the same label data as the lectotype.

According to Kleine (1928a: 74) the type of *E. hirtellus* was in MNHUB, but we did not find it there. He also reported (1928a: 74) that the type for *S. pilosus* was in SMTD; according to O. Jaeger (SMTD) (2002, personal communication) it is not in the SMTD collections.

***proseni* Soares & Scivittaro, 1977**

proseni Soares & Scivittaro, 1977: 91.

Type depository. APC (HT).

Distribution. Bolivia.

***Metatrachelizus* Kleine, 1922**

Metatrachelizus Kleine, 1922f: 207.

Type species: *Metatrachelizus abjectus* Kleine, 1922.

Distribution. Republic of the Congo; Borneo, India (including Assam), Indonesia (Misool; Moluccas: Buru; Salawati; Sumatra), Malaysia, Philippines (Siargao), Vietnam; New Guinea.

***abjectus* Kleine, 1922**

abjectus Kleine, 1922f: 208.

Type depository. SMTD (HT).

Distribution. Borneo, Indonesia (Sumatra), Malaysia.

***africanus* Damoiseau, 1967**

africanus Damoiseau, 1967b: 9.

Type depositories. HNHN (HT, PT), IRSNB (PT!), MNHN (PT!).

Distribution. Republic of the Congo.

artificialis Kleine, 1923

artificialis Kleine, 1923d: 134.

artificialis: Kleine, 1925f: 19 (*Metatrachelizus*) [lapsus].

Type depository. NHM (LT!).

Distribution. India.

Notes. Lectotype of *M. artificialis* (NHM) designated on a specimen with the following labels: Syntype/ Typus!/ Nilgiri Hills/ Andrewes Bequest. B. M. 1922-221/ *Metatrachelizus artificialis* Kln. det. 1922/ Prépar. génit. No BM 15, R. Damoiseau.

castigatus Kleine, 1926

castigatus Kleine, 1926a: 20.

Type depositories. NHM (HT!), Mjöberg coll. (localization unknown) (PT).

Distribution. Borneo, Indonesia (Salawati, Sumatra), Malaysia; New Guinea.

confragosus Kleine, 1926

confragosus Kleine, 1926a: 20.

Type depository. NHM (ST!).

Distribution. Borneo, Indonesia (Misool); New Guinea.

congruens Kleine, 1925

congruens Kleine, 1925b: 132.

Type depository. NHM (LT!, PLT!).

Distribution. Borneo, India (including Assam), Vietnam.

Notes. Lectotype of *M. congruens* (NHM) designated on a specimen with the following labels: Syntype/ Typus!/ Assam Sudiya/ Fry Coll. 1905-100/ Doherty/ *Metatrachelizus congruens* Kln. R. Kleine det. 1922. Paralectotype: 1 specimen (NHM), Syntype/ Assam, Sudiya/ Fry Coll. 1905-100/ Doherty.

constans Kleine, 1926

constans Kleine, 1926a: 20.

Type depository. NHM (LT!).

Distribution. Indonesia (Misool), Philippines (Siargao).

Notes. Lectotype of *M. constans* (NHM) designated on a specimen with the following labels: *Stereodermus* Mysol/ Syntype/ Type/ Sharp Coll. 1905-313/ Prépar. génit. BM 16, R. Damoiseau/ *Metatrachelizus constans* Kln. Typus!, R. Kleine det. 1922/ R. Damoiseau det. 1971 *Metatrachelizus confragosus* Kleine, new synonymy [*in litteris*].

ferox Kleine, 1944

ferox Kleine, 1944: 152.

Type depositories. MNHN (LT!, PLT!), DEI (PLT!).

Distribution. Vietnam.

Notes. Döbler & Gaedike (1995: 445) reported that the specimens held in MNHN and in Kleine's collection are holotype and paratypes, but in his original description Kleine did not designate any of the syntypes as holotype; however, as there are two syntypes in MNHN, and as Döbler & Gaedike did not specifically select one as holotype, we designate the lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Lectotype of *M. ferox* (MNHN) designated on a male specimen with the following labels: Museum Paris, Tonkin, Reg. de Hoa-Binh, A. de Cooman 1927/ ♂/ type/ *Metatrachelizus ferox* Kln., R. Kleine det 1943. Paralectotypes: 1 specimen (MNHN), Museum Paris, Tonkin, Reg. de Hoa-Binh, A. de Cooman 1928 [both specimens glued on the same card]; 1 specimen (DEI), Museum Paris, Tonkin N., env. d'Ha-Giang, Leut. coll. Bonifacy, 1913/ paratypus/ *Metatrachelizus ferox* Kln., R. Kleine det., 1943.

fucosus Kleine, 1925

fucosus Kleine, 1925f: 18.

Type depository. ZMAN (HT!).

Distribution. Indonesia (Moluccas: Buru), Vietnam.

Pseudanchisteus Kleine, 1922

Pseudanchisteus Kleine, 1922h: 136.

Type species: *Pseudanchisteus neglectus* Kleine, 1922.

Distribution. Democratic Republic of the Congo (ex Zaire), Equatorial Guinea, Ghana.

neglectus Kleine, 1922

neglectus Kleine, 1922h: 137.

Type depository. MNHUB (LT!, PLT!).

Distribution. Democratic Republic of the Congo (ex Zaire), Equatorial Guinea, Ghana.

Notes. Damoiseau (1967a: 221) reported that the specimen held in MNHUB is the holotype, but in his original description Kleine did not designate either of the syntypes as holotype; however, as there are two syntypes in MNHUB, and as Damoiseau did not specifically select one as holotype, we designate the lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Lectotype of *P. neglectus* (MNHUB) designated on a male specimen with the following labels: Span. Guinea, Nkolentangan, XI.07-V.08, G. Tessmann S. G./ Type/ Zool. Mus. Berlin. Paralectotype: 1 male specimen (MNHUB), Span. Guinea, Nkolentangan, XI.07-V.08, G. Tessmann S. G./ Type/ *Pseudanchisteus neglectus* Kleine, det. Autor, 1919/ Zool. Mus. Berlin.

Stereobates Sharp, 1895

Stereobates Sharp, 1895a: 16.

Type species: *Stereobates pedator* Sharp, 1895.

Distribution. Mexico, Panama.

pedator Sharp, 1895

pedator Sharp, 1895a: 17.

gracilis Sharp, 1895a: 17 (*Stereobates*); **syn. n.**

chiriquensis Sharp, 1895a: 18 (*Stereobates*); **syn. n.**

Type depositories. *S. pedator*: NHM (LT!, PLT!); *S. gracilis*: NHM (HT!); *S. chiriquensis*: NHM (HT!).

Distribution. Mexico, Panama.

Notes. Lectotype of *S. pedator* (NHM) designated on a specimen with the following labels: *Stereobates pedator* Type D. S., Toxpam, Sallé/ Syntype/ type/ prép. génit. No Bm 21, R. Damoiseau/ Toxpam/ Mexico, Salle Coll./ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Stereodermus pedator* Sharp. Paralectotype: 1 specimen (NHM) with the same label data as the lectotype.

After studying the type material, we consider *S. gracilis* and *S. chiriquensis* to be synonyms for *S. pedator*.

Stereobatinus Kleine, 1927

Stereobatinus Kleine, 1927b: 424.

Type species: *Stereobatinus efferus* Kleine, 1927.

Distribution. Colombia.

Notes. Alonso-Zarazaga & Lyal (1999: 52) include Brazil in the genus distribution but we could not find any quotation for this country.

efferus Kleine, 1927

efferus Kleine, 1927b: 425.

Type depository. NHM (HT!).

Distribution. Colombia.

Stereoderminus Kleine, 1927

Stereoderminus Kleine, 1927b: 422.

Type species: *Stereoderminus effrenatus* Kleine, 1927.

Distribution. Brazil.

effrenatus Kleine, 1927

effrenatus Kleine, 1927b: 423.

Type depository. NHM (HT!).

Distribution. Brazil.

Stereodermus Lacordaire, 1866

Stereodermus Lacordaire, 1866: 419.

Sereodermus: Kleine, 1921n: 8 [lapsus].

Type species: *Arrhenodes pygmaeus* Gyllenhal, 1833.

Distribution. U.S.A. (Florida); Costa Rica, Cuba, Guatemala, Mexico, Nicaragua, Panama, Virgin Is.; Argentina, Bolivia, Brazil, Colombia, Ecuador, Peru, Suriname, Venezuela; Indonesia (Batu, Engano, Nias, Sipura, Sulawesi, Sumatra), Philippines (Luzon, Mindanao).

barbirostris Sharp, 1895

barbirostris Sharp, 1895a: 14.

Type depository. NHM (LT!, PLT!).

Distribution. Costa Rica, Panama.

Notes. Lectotype of *S. barbirostris* (NHM) designated on a specimen with the following labels: Syntype/ V. de Chiriqui, 2-3000 ft, Champion/ Sharp, Coll. 1905-313/ B.C.A., Col. IV, 6. *Stereodermus barbirostris* Sharp. Paralecotype: 1 specimen (NHM), Syntype/ Bugaba, Panama, Champion/ Prépar. génit. No. BM 38, R. Damoiseau/ B.C.A., Col. IV, 6. *Stereodermus barbirostris* Sharp.

breviceps Sharp, 1895

breviceps Sharp, 1895a: 11.

Type depository. NHM (LT!, PLT!).

Distribution. Guatemala.

Notes. Lectotype of *S. breviceps* (NHM) designated on a male specimen with the following labels: Syntype/ Cerro Zunil, 4000 ft, Champion/ Prépar. génit. No. BM 31, R. Damoiseau/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Stereodermus breviceps* Sharp. Paralectotypes: 3 specimens (NHM) with the same label data as the lectotype; 1 specimen (NHM), Syntype/ Pancina, Vera Paz, Champion/ B.C.A., Col. IV, 6. *Stereodermus breviceps* Sharp.

calvus Sharp, 1895

calvus Sharp, 1895a: 15.

Type depository. NHM (LT!, PLT!).

Distribution. Costa Rica, Panama.

Notes. Lectotype of *S. calvus* (NHM) designated on a specimen with the following labels: Syntype/ Prépar. génit. No. BM 39, R. Damoiseau/ V. de Chiriqui, 2-3000 ft, Champion/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Stereodermus calvus* Sharp. Paralectotypes: 4 specimens (NHM) with the same label data as the lectotype.

carinatus Sharp, 1895

carinatus Sharp, 1895a: 10.

tenuis Sharp, 1895a: 10 (*Stereodermus carinatus* var.); **syn. n.**

Type depositories. *S. carinatus*: NHM (LT!, PLT!); *S. carinatus* var. *tenuis*: NHM (HT!).

Distribution. Costa Rica, Guatemala, Mexico, Panama; Brazil, Suriname, Venezuela.

Notes. Lectotype of *S. carinatus* (NHM) designated on a specimen with the following labels: *S. carinatus* Types D. S., Zapote, Champion/ Syntype/ type/ Zapote, Guatemala, G. C. Champion / Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Stereodermus carinatus* Sharp. Paralectotypes: 11 specimens (NHM), with the same label data as the lectotype; 4 specimens (NHM), *Stereodermus carinatus* D. S., Zapote/ Syntype/ type/ Zapote, Guatemala, G. C. Champion / Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Stereodermus carinatus* Sharp; 1 specimen (NHM), Syntype/ Zapote, Guatemala, G. C. Champion / B.C.A., Col. IV, 6. *Stereodermus carinatus* Sharp.

***chontalensis* Sharp, 1895**

chontalensis Sharp, 1895a: 10.

Type depository. NHM (LT!, PLT!).

Distribution. Costa Rica, Mexico, Nicaragua, Panama; Colombia, Ecuador.

Notes. Lectotype of *S. chontalensis* (NHM) designated on a male specimen with the following labels: *S. chontalensis* Type D. S./ Syntype/ type/ Chontales, Janson/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Stereodermus Chontalensis* Sharp. Paralectotypes: 1 specimen (NHM), *S. chontalensis* D. S./ Syntype/ Chontales, Nicaragua, T. Belt/ B.C.A., Col. IV, 6. *Stereodermus Chontalensis* Sharp; 1 specimen (NHM), *S. chontalensis* Type ex parte D. S. Chontales/ Syntype/ Chontales, Nicaragua, T. Belt/ Sharp Coll., 1905-313/ B.C.A., Col. IV, 6. *Stereodermus chontalensis* Sharp; 1 specimen (NHM), *S. chontalensis* D. S./ Syntype/ Prépar. génit. No. BM 30 R. Damoiseau/ Chontales, Nicaragua, T. Belt/ B.C.A., Col. IV, 6. *Stereodermus chontalensis* Sharp/ Damoiseau det., allotype ♂; 1 specimen (NHM), Syntype/ V. de Chiriqui, below 4,000 ft., Champion/ B.C.A., Col. IV, 6. *Stereodermus chontalensis* Sharp.

***dentipennis* Sharp, 1895**

dentipennis Sharp, 1895a: 11.

Type depository. NHM (LT!, PLT!).

Distribution. Costa Rica, Mexico; Colombia, Peru.

Notes. Lectotype of *S. dentipennis* (NHM) designated on a male specimen with the following labels: Syntype/ Cerro de Plumas, Mexico, Hoege/ Prépar. génit. No. BM 32, R. Damoiseau/ B.C.A., Col. IV, 6. *Stereodermus dentipennis* Sharp. Paralectotype: 1 female specimen (NHM), Syntype/ Jalapa, Mexico, M. Trujillo/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Stereodermus dentipennis* Sharp.

***dentipes* Sharp, 1895**

dentipes Sharp, 1895a: 14.

Type depository. NHM (HT!).

Distribution. Costa Rica, Panama.

elytralis Senna, 1903

elytralis Senna, 1903: 166.

Type depository. MSNG (HT!).

Distribution. Indonesia (Sumatra).

exilis Suffrian, 1870

exilis Suffrian, 1870: 220.

Type depository. MNHC (ST).

Distribution. U.S.A. (Florida); Cuba, Virgin Is.

Notes. Warner (1960: 29), Anderson (1992: 212), Poole & Gentili (1996: 62), and Thomas (1996: 3) report the presence of *Stereodermus exilis* from Florida.

fessus Kleine, 1927

fessus Kleine, 1927b: 425.

Type depository. NHM (LT!, PLT!).

Distribution. Bolivia, Brazil.

Notes. Lectotype of *S. fessus* (NHM) designated on a specimen with the following labels: Syntype/ Type/ Gounelle/ Parà/ Fry coll. 1905-100/ *Stereodermus fessus* Kln Typus! 1923, R. Kleine det. Paralectotypes: 3 specimens (NHM), with the same label data as the lectotype.

filum Sharp, 1895

filum Sharp, 1895a: 16.

Type depository. NHM (LT!, PLT!).

Distribution. Panama.

Notes. Lectotype of *S. filum* (NHM) designated on a male specimen with the following labels: Lectotype [*in litteris*]/ Syntype/ Prépar. génit. No. BM 25, R. Damoiseau/ V. de Chiriqui, 2-3000 ft, Champion/ B.C.A., Col. IV, 6. *Stereodermus filum* Sharp/ R. Damoiseau *filum* Sharp, Lectotype ♂. Paralectotypes: 4 specimens (NHM), with the same label data as the lectotype.

flavotibialis Kleine, 1922

flavotibialis Kleine, 1922b: 28.

Type depository. SMTD (ST).

Distribution. Philippines (Luzon).

gestroi Senna, 1893

gestroi Senna, 1893b: 261.

Type depositories. MSNG (LT!, PLT!), MNHN (PLT!), MZUF (PLT!), DEI (PLT!), SMTD (PLT).

Distribution. Indonesia (Batu, Engano, Nias, Sumatra); Europe (I).

Notes. Lectotype of *S. gestroi* (MSNG) designated on a specimen with the following labels: Engano, Bua Bua, V.VI.1891, Modigliani/ *Stereodermus Gestroi* Senna, typus!. Paralectotypes: 6 specimens (MSNG), Engano, Bua Bua, V.VI.1891, Modigliani; 3 specimens (MNHN), Engano, Bua Bua, V-VI.1891, Modigliani; 1 specimen (DEI), Engano, Bua Bua, V-VI.1891, Modigliani/ coll. Vogel/ v. Schönfeldt det./ *Stereodermus gestroi* Senna/ Dtsch. Entomol. Institut Berlin/ Syntypus; 1 specimen (MZUF), Engano, Bua Bua, V.VI.1891, Modigliani/ *Stereodermus gestroi* Senna, cotypus!/ Ann. Mus. Genova (2), XIX, 1893, cotypi!/ ex-coll. Senna/ "La Specola" Firenze, 6934; 2 specimens (MZUF), Engano, Bua Bua, V.VI.1891, Modigliani/ ex-coll. Senna/ "La Specola" Firenze, 6934.

godmani Sharp, 1895

godmani Sharp, 1895a: 13.

Type depository. NHM (HT!).

Distribution. Costa Rica, Guatemala.

infidus Senna, 1898

infidus Senna, 1898a: 226.

Type depository. MSNG (LT!, PLT!).

Distribution. Indonesia (Batu, Nias, Sipura, Sumatra), Vietnam.

Notes. Lectotype of *S. infidus* (MSNG) designated on a male specimen with the following labels: Mentawai, Sipora Sereinu, V-VI.94, Modigliani/ cotypus/ *Stereodermus infidus* cotipi!. Paralectotypes: 3 male and 2 female specimens (MSNG), Mentawai, Sipora Sereinu, V-VI.94, Modigliani/ cotypus/ *Stereodermus infidus* cotipi!; 1 specimen (MSNG), Mentawai, Si Oban, IV-VIII.94, Modigliani.

There are seven specimens labelled as cotypes in MSNG, although in his original description Senna (1898a: 7) only mentions six.

latirostris Sharp, 1895

latirostris Sharp, 1895a: 9.

Type depositories. NHM (LT!, PLT!), IRSNB (PLT!), SMTD (PLT).

Distribution. Costa Rica, Guatemala, Nicaragua, Panama; Ecuador.

Notes. Lectotype of *S. latirostris* (NHM) designated on the second of the three specimens glued to the same label, with the following labels: *Stereodermus latirostris* Type D. S., V. de Chiriqui, Champion/ Syntype/ type/ V. de Chiriqui, 25-4.000 ft., Champion/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Stereodermus latirostris* Sharp. Paralectotypes: 19 specimens (NHM) with the same label data as the lectotype; 3 specimens (NHM), *S. latirostris* ♀/ Teleman, Vera Paz, Champion/ Syntype/ B.C.A., Col. IV, 6. *Stereodermus latirostris* Sharp/ Sharp Coll. 1905-313; 1 specimen (NHM), Syntype/ Chontales, Janson/ B.C.A., Col. IV, 6. *Stereodermus latirostris* Sharp; 1 male specimen (IRSNB), V. de Chiriqui, 2-3000 ft., Champion/ *Stereodermus latirostris* Sh./ Paratype/ Biol. C. Amer., Don Godman et Salvin/ Prépar. génit. No. 80 H 24 R. Damoiseau/ *Stereodermus latirostris* Sharp, R. Damoiseau det.

longiceps Sharp, 1895

longiceps Sharp, 1895a: 9.

Type depository. NHM (LT!, PLT!).

Distribution. Costa Rica, Guatemala, Panama.

Notes. Lectotype of *S. longiceps* (NHM) designated on a female specimen with the following labels: *Stereodermus longiceps* Type D. S., V. de Chiriqui, Champion/ ♀/ Syntype/ type/ Prépar. génit. No. BM 26. R. Damoiseau/ V. de Chiriqui, 25-4.000 ft., Champion / Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Stereodermus longiceps* Sharp. Paralectotypes: 2 female specimens (NHM), *Stereodermus longiceps* D. S., V. de Chiriqui, Champion/ ♀/ Syntype/ V. de Chiriqui, 25-4.000 ft., Champion/ B.C.A., Col. IV, 6. *Stereodermus longiceps* Sharp; 1 male specimen (NHM), syntype/ Prépar. génit. No. BM 27. R. Damoiseau/ V. de Chiriqui, 25-4.000 ft., Champion/ B.C.A., Col. IV, 6. *Stereodermus longiceps* Sharp/ Sharp Coll. 1905-313/ R. Damoiseau det., Allo-type ♂.

mitratus Sharp, 1895

mitratus Sharp, 1895a: 12.

Type depository. NHM (HT!).

Distribution. Panama; Indonesia (Sumatra?).

nemoralis Senna, 1898

nemoralis Senna, 1898b: 379.

Type depository. MSNG (HT!).

Distribution. Indonesia (Nias, Sumatra), Philippines (Mindanao).

Notes. Senna (1898b: 379) first named the species from a single specimen and wrote that he would give a detailed description later. However he did give some distinctive characters for the new taxon. In 1903 (p. 166) Senna published the full description of the species and wrote that the type series consisted of six cotypes, which can not be considered as syntypes.

nigriceps Sharp, 1895

nigriceps Sharp, 1895a: 15.

Type depository. NHM (HT!).

Distribution. Guatemala.

puncticollis Sharp, 1895

puncticollis Sharp, 1895a: 13.

Type depository. NHM (LT!, PLT!).

Distribution. Costa Rica, Panama.

Notes. Lectotype of *S. puncticollis* (NHM) designated on a specimen with the following labels: Syntype/ V. de Chiriqui, 2-3000 ft, Champion/ Sharp Coll. 1905-313/ Prépar. génit. No. BM 37, R. Damoiseau/ B.C.A., Col. IV, 6. *Stereodermus puncticollis* Sharp. Paralectotypes: 3 specimens (NHM), with the same label data as the lectotype.

pygmaeus (Gyllenhal, 1833)

pygmaeus Gyllenhal (in Schoenherr, 1833: 333) (*Arrhenodes*).

brevirostris Senna, 1889: 35 (*Stereodermus*); syn. Schönfeldt, 1908: 53.

Type depositories. *A. pygmaeus*: IRSNB (LT!), NHRS (PLT); *S. brevisrostris*: MZUF (HT!).

Distribution. Costa Rica, Guatemala, Mexico, Nicaragua, Panama; Argentina, Bolivia, Brazil, Ecuador, Peru.

Notes. Lectotype of *A. pygmaeus* (IRSNB) designated on a male specimen [incorrectly labelled as holotype] with the following labels: Mexique/ Coll. Dejean, Coll. Roelofs/ Holotype/ *Pygmaeus* Schoenherr/ *Trachelizus Pygmaeus* Schoenherr/ Prépar. génit. N. 71/6.7/1 R. Damoiseau/ *Stereodermus pygmaeus* (Gyllenhal), R. Damoiseau det. 1969.

There is also a female specimen in IRSNB that Damoiseau labelled 'Allotype', but it can not belong to the type series, as it has a label dating "Deyr. 1861" and the species was described in 1833.

***raapi* Senna, 1897**

raapi Senna, 1897c: 543.

Type depository. MSNG (HT!).

Distribution. Indonesia (Batu, Sulawesi).

***siporanus* Senna, 1898**

siporanus Senna, 1898a: 225.

Type depository. MSNG (HT!).

Distribution. Indonesia (Mentawai).

***zunilensis* Sharp, 1895**

zunilensis Sharp, 1895a: 13.

Type depository. NHM (LT!, PLT!).

Distribution. Guatemala; Bolivia, Peru.

Notes. Lectotype of *S. zunilensis* (NHM) designated on a male specimen with the following labels: Syntype/ Cerro Zunil 4-5000 ft, Champion/ Sharp Coll. 1905-313/ Prépar. génit. No. BM 36, R. Damoiseau/ B.C.A., Col. IV, 6. *Stereodermus zunilensis* Sharp. Paralectotypes: 2 specimens (NHM), with the same label data as the lectotype.

PHOLIDOCHLAMYDINAE Damoiseau, 1962

Pholidochlamydiae Damoiseau, 1962b: 18.

Pholidochlamydiae: Damoiseau, 1962b: 26 [lapsus].

***Pholidochlamys* Lacordaire, 1866**

Pholidochlamys Lacordaire, 1866: 473.

Type species: *Pholidochlamys madagascariensis* Lacordaire, 1866.

Distribution. Madagascar.

madagascariensis Lacordaire, 1866

madagascariensis Lacordaire, 1866: 474.

Type depository. MCZ (HT).

Distribution. Madagascar.

TAPHRODERINAE Lacordaire, 1866

Taphrodérides Lacordaire, 1866: 406.

Ischnomérides Lacordaire, 1866: 414.

Taphroderinae Pascoe, 1872: 317.

Taphroderini Kolbe, 1883b: 234.

Taphroderina Sharp, 1895a: 2.

Taphroderi Senna, 1896a: 210.

Taphroderidae Schönfeldt, 1908: 3.

Ischnomeridae Schönfeldt, 1908: 15.

Ischnomerini Schönfeldt, 1910a: 9.

Tapheoderini: Kleine, 1921a: 78 [lapsus].

Taphoderidae: Ienistea, 1986: 32 [lapsus].

Aulacoderes Chevrolat, 1839

Aulacoderes Chevrolat, 1839: 182.

Ischnomerus Schoenherr, 1840: 571 (non Labram & Imhoff, 1838); syn. Kleine, 1938d: 161.

Ischnocerus: Schoenherr, 1840: 572 [lapsus].

Aulacoderus: Agassiz, 1846a: 18 [lapsus].

Aulacodores: Lacordaire, 1866: 414 [lapsus].

Type species: *Aulacoderes immotus* Chevrolat, 1839.

Distribution. Madagascar.

immotus Chevrolat, 1839

immotus Chevrolat, 1839: 182.

linearis Boheman (in Schoenherr, 1840: 572) (*Ischnomerus*); syn. Gemminger & Harold, 1872: 2704.

Type depositories. *A. immotus*: MNHN (LT!, PLT!); *I. linearis*: unknown.

Distribution. Madagascar.

Notes. Lectotype of *A. immotus* (MNHN) designated on a male specimen with the following labels: Madagascar/ Chevrolat 12/ type/ ex Musaeo G. Power/ Museum Paris 1952, coll. R. Oberthür. Paralectotype: 1 male specimen (MNHN), Madagascar/ Chevrolat 12/ type/ ♂/ ex Musaeo G. Power/ Museum Paris 1952, coll. R. Oberthür.

Bolbocranius Kolbe, 1897

Bolbocranius Kolbe, 1897: 284.

Anisognathus Lacordaire, 1866: 411 (non Reichenbach, 1850); syn. Alonso-Zarazaga & Lyal, 1999: 53.

Isognathus Kolbe, 1888: 305 (non Felder, 1862); syn. Kleine, 1927a: 69.

Bolbocephalus Kolbe, 1916: 64 (non Whitfield, 1890); syn. Kleine, 1938d: 160.

Bolbocranius Kolbe, 1916: 64 (non Kolbe, 1897); syn. Alonso-Zarazaga & Lyal, 1999: 53.

Haplocephalus Quentin, 1966: 1664; syn. Damoiseau, 1967d: 131.

Aulonocanthus Quentin, 1966: 1665; syn. Damoiseau, 1967d: 131.

Colpocranius Quentin, 1966: 1666; syn. Damoiseau, 1967d: 131.

Colpometopus Quentin, 1966: 1666 (non Abeille, 1900); syn. Damoiseau, 1967d: 131.

Type species: *Bolbocranius mandibularis* Kolbe, 1897.

Distribution. Tropical Africa.

Notes. Damoiseau (1967a) did not mention the genera *Haplocephalus*, *Aulonocanthus*, *Colpocranius*, *Colpometopus* in his Monograph on African Brentidae.

anaticeps (Kolbe, 1883)

anaticeps Kolbe, 1883b: 235 (*Anisognathus*).

agnaticeps: K. E. Schedl, 1961a: 83 (*Bolbocephalus*) [lapsus].

Type depository. MNHUB (HT!).

Distribution. Angola, Democratic Republic of the Congo (ex Zaire), Gabon, Ivory Coast, Republic of the Congo, Republic of Guinea, Sierra Leone, Tanzania, Zambia.

Notes. Damoiseau (1967a: 460) incorrectly reported that the holotype was in MNHN.

andreorum (Quentin, 1966)

andreorum Quentin, 1966: 1668 (*Colpocranius*).

Type depository. MNHN (HT!).

Distribution. Republic of the Congo.

Notes. Damoiseau (1967a) did not mention this species in his Monograph on African Brentidae.

bicolor Senna, 1898

bicolor Senna, 1898c: 371.

parvulus De Muizon, 1955b: 912 (*Bolbocranius*); syn. Damoiseau, 1967a: 451.

Type depositories. *B. bicolor*: MZUF (LT!, PLT!), DEI (PLT!), MRAC (PLT!), SMTD (PLT); *B. parvulus*: MNHN (LT!, PLT!).

Distribution. Tropical Africa.

Notes. Lectotype of *B. bicolor* (MZUF) designated on a male specimen with the following labels: Station J. Albrechts Höhe, leg. Conradt N. Camerun/ *Bolbocranius bicolor* Senna, cotypi ♂ ♀/ Deutsch. Ent. Zeitsch., 1898, cotypi ♂ ♂ ♀ ♀/ ex-coll. Senna/ "La Specola" Firenze, 7070. Paralectotypes: 3 male and 4 female specimens (MZUF), N. Camerun, leg. Conradt/ ex-coll. Senna/ "La Specola" Firenze, 7070; 6 specimens (DEI), N Kamerun, Johann-Albrechtshöhe, L. Conradt, 1896/ Syntypus/ coll. Kraatz/ Senna det./ v. Schönfeldt det.; 8 specimens (DEI), N Kamerun, Johann-Albrechtshöhe, L. Conradt, 1896/ coll. Kraatz/ v. Schönfeldt det./ Syntypus/ *Anisognathus bicolor* (Senna), Scivittaro det., 1974; 1 specimen (MRAC), Paratypus/ Musée du Congo, N Kamerun, J. Albr. Hohe (Conradt 96) D. Ent. Nat. Mus. ex Coll. Kraatz/ N Kamerun, Johann Albrechtshöhe L. Conradt, 1896/ Coll. Kraatz/ R. det. T 1330/ *Bolbocranius bicolor* Senna, cotype!/ *Anisognathus bicolor* (Senna), R. Damoiseau det., 1966.

Lectotype of *B. parvulus* (MNHN) designated on a male specimen with the following labels: Oyem, Gabon, J. De Muizon/ cotype. Paralectotypes: 2 specimens (MNHN), with the same label data as the lectotype.

csikii (Bolkay, 1910)

csikii Bolkay, 1910a: 182 (*Anisognathus*).

distortus Auct. (non Westwood, 1849) (*Anisognathus*).

bolkay Kleine, 1914b: 170 (*Anisognathus*); syn. Damoiseau, 1967a: 447.

csiki: Kleine, 1914b: 166 (*Anisognathus*) [lapsus].

czikii: Kleine, 1938d: 160 (*Anisognathus*) [lapsus].

Type depositories. *A. csikii*: HNHM (HT); *A. bolkay*: HNHM (ST).

Distribution. Tropical Africa.

Notes. Arrow (1940: 319) first recognized two different species among the specimens identified as “*A. distortus*” by many authors (e. g. Kleine, 1938d: 160): *A. distortus* is limited to South Africa whilst *A. csikii* is distributed throughout the rest of Tropical Africa.

Bolkay (1910a: 182) described and illustrated the species with two different formae; the “forma *asymmetrica*” corresponds to *B. csikii* while the “forma *symmetrica*” corresponds to a different species similar to *B. rostralis* or *B. delhaisei*.

delhaisei (Damoiseau, 1963)

delhaisei Damoiseau, 1963h: 7 (*Bolbocephalus*).

Type depository. IRSNB (HT!).

Distribution. Democratic Republic of the Congo (ex Zaire), Ghana, Sudan.

distortus (Westwood, 1849)

distortus Westwood, 1849: 206 (*Taphroderes*).

augurius Kolbe, 1885: 187 (*Anisognathus*); syn. Kleine, 1927a: 69.

Type depositories. *T. distortus*: NHM (?) (ST), OXUM (ST!); *A. augurius*: MNHUB (HT!).

Distribution. South Africa.

Notes. Damoiseau (1967a: 446) reported that a specimen of *T. distortus* held in NHM is the holotype, but in his original description Westwood did not

designate any of the syntypes as holotype. We did not find any type in NHM but we did find a syntype in OXUM.

Damoiseau did not mention *A. augurius* in his Monograph on the African Brentidae (1967a).

***mandibularis* Kolbe, 1897**

mandibularis Kolbe, 1897: 284.

Type depository. MNHUB (LT!, PLT!).

Distribution. Kenya, Tanzania.

Notes. Lectotype of *B. mandibularis* (MNHUB) designated on a male specimen with the following labels: Usambara, Derema, 350 m, 25.X-21.XI.91, Conradt S./ *Bolbocranius mandibularis* n. sp. Kolbe/ Type. Paralectotype: 1 male specimen (MNHUB), Usambara, Derema, 850 m, 16.IX-7.X.91, Conradt S./ 90614/ Type.

***mechowi* (Kolbe, 1883)**

mechowi Kolbe, 1883b: 234 (*Anisognathus*).

peregrinus Kolbe, 1916: 66 (*Bolbocephalus mechowi* var.); **syn. n.**

Type depositories. *A. mechowi*: MNHUB (HT!); *A. mechowi* var. *peregrinus*: unknown.

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Gabon, Ghana, Ivory Coast, Republic of the Congo, Rwanda, Tanzania.

Notes. Damoiseau (1967a: 454) reported that the holotype of *A. mechowi* was in MNHN; Quentin (1970: 220) reported that the same was in the Museum of Stettin, but we found it in MNHUB.

Neither Kleine (1938d), De Muizon (1960) nor Damoiseau (1967a) mentioned *B. mechowi* var. *peregrinus* Kolbe.

modicus (Kolbe, 1916)

modicus Kolbe, 1916: 65 (*Bolbocephalus*).

Type depositories. MNHUB (LT!, PLT!), IRSNB (?) (PLT).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea, Gabon, Ivory Coast.

Notes. Quentin (1970: 220) reported that the types were in the Museum of Stettin, but we found them in MNHUB. Damoiseau (1967a: 458) designated the lectotype; he also reported that a paralectotype was in IRSNB, but we did not find it there.

nimbaensis Damoiseau, 1967

nimbaensis Damoiseau, 1967a: 447.

Type depository. MNHN (?) (HT).

Distribution. Republic of Guinea.

Notes. According to Damoiseau (1967a: 448) the holotype of *A. nimbaensis* is in MNHN, but we did not find it there.

opacus Kolbe, 1916

opacus Kolbe, 1916: 66.

ephippium Kleine, 1916c: 81 (*Bolbocranius*); syn. De Muizon, 1955b: 911.

recens Kleine, 1934a: 306 (*Bolbocephalus*); syn. Damoiseau, 1967a: 458.

Type depositories. *B. opacus*: MNHUB (LT!); *B. ephippium*: DEI (HT!); *B. recens*: DEI (HT!, PT!).

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Ghana, Ivory Coast, Republic of the Congo, Republic of Guinea, Tanzania, Togo.

Notes. Lectotype of *B. opacus* (MNHUB) designated on a male specimen with the following labels: G. Bissau, Giulberni/ 82160/ coll. H. C. Vogel/ Lectotype ♂/ *opacus* n. sp. Kleine. We were unable to definitely identify other syntypes.

rostralis (Damoiseau, 1961)

rostralis Damoiseau, 1961a: 289 (*Bolbocephalus*).

Type depository. MRAC (HT!).

Distribution. Democratic Republic of the Congo (ex Zaire).

Notes. In his original description, Damoiseau mentions the type in MRAC and does not give any paratype, but he furnishes more than one measurement for the species. There is one specimen he labelled as paratype in IRSNB.

truncatus (Damoiseau, 1961)

truncatus Damoiseau, 1961a: 288 (*Bolbocephalus*).

Type depository. MRAC (HT!).

Distribution. Angola, Democratic Republic of the Congo (ex Zaire).

unguicularis Kolbe, 1916

unguicularis Kolbe, 1916: 67.

Type depository. MNHUB (?) (HT).

Distribution. Tanzania.

Notes. According to Kleine (1928a: 93) the type was in MNHUB, but we did not find it there.

unicolor De Muizon, 1960

unicolor De Muizon, 1960: 238.

Type depositories. MNHN (LT!, PLT!), NHM (PLT!).

Distribution. Cameroon, Kenya, Tanzania.

Notes. Lectotype of *B. unicolor* (MNHN) designated on a male specimen with the following labels: Van Someren, Meru, 7.43/ Kenya/ cotype/ *Bolbocranius unicolor* De Muizon. Paralectotypes: 1 female specimen (MNHN), Van Someren, Meru, 7.43/ Kenya/ cotype/ *Bolbocranius unicolor* De Muizon; 3 male and 1 female specimens (NHM), Syntype/ Van Someren, Meru, 7.43/ Pres. by. Com. Inst. Ent. B. M. 1950-15/ *Bolbocranius unicolor* n. sp., J. De Muizon det.

***Monrosiaia* Haedo Rossi, 1960**

Monrosiaia Haedo Rossi, 1960: 100.

Type species: *Monrosiaia franciscoi* Haedo Rossi, 1960.

Distribution. Costa Rica, Panama, Trinidad and Tobago; Argentina, Bolivia, Brazil, Venezuela.

***franciscoi* Haedo Rossi, 1960**

franciscoi Haedo Rossi, 1960: 101.

Type depositories. JHRC (HT, AT), AMC (PT).

Distribution. Bolivia.

Notes. Scivittaro (1975: 544) reports that a cotype is held in AMNH.

***inpai* Scivittaro, 1976**

inpai Scivittaro, 1976: 79.

Type depository. INPA (HT).

Distribution. Brazil.

***kleinei* Scivittaro, 1975**

kleinei Scivittaro, 1975: 550.

Type depositories. MZSP (HT, PT), MAPA (PT), NHRS (PT), MEFP (PT), IOC (PT).

Distribution. Brazil.

lineicollis (Haedo Rossi, 1954)

lineicollis Haedo Rossi, 1954a: 2 (*Taphroderes*).

lineocollis: Scivittaro, 1975: 544 [lapsus].

Type depository. IMLA (HT).

Distribution. Argentina, Brazil.

monicae Scivittaro, 1975

monicae Scivittaro, 1975: 547.

Type depositories. AMNH (HT, PT), MZSP (PT), NMNH (PT), MCCV (PT).

Distribution. Costa Rica, Panama, Trinidad and Tobago.

sulcata Scivittaro, 1978

sulcata Scivittaro, 1978: 161.

Type depository. MBUC (HT, PT).

Distribution. Venezuela.

walkyriae Scivittaro, 1975

walkyriae Scivittaro, 1975: 549.

Type depositories. AMNH (HT), MZSP (PT).

Distribution. Brazil.

Plesiobolbus Kolbe, 1916

Plesiobolbus Kolbe, 1916: 64.

Type species: *Plesiobolbus sagax* Kolbe, 1916.

Distribution. Cameroon, Tanzania.

cribriceps (Kolbe, 1897)

cribriceps Kolbe, 1897: 285 (*Bolbocephalus*).

Type depository. MNHUB (HT!).

Distribution. Tanzania.

martini Bartolozzi, 1991

martini Bartolozzi, 1991: 25.

Type depository. ZMUC (HT!).

Distribution. Tanzania.

sagax Kolbe, 1916

sagax Kolbe, 1916: 64.

Type depository. MNHUB (HT!).

Distribution. Cameroon.

Taphroderes Schoenherr, 1823

Taphroderes Schoenherr, 1823: 1137.

Abactrus Sharp, 1895a: 6; syn. Scivittaro, 1975: 440.

Type species: *Brentus foveatus* Lund, 1800.

Distribution. Costa Rica, Mexico, Panama; Bolivia, Brazil, Colombia, French Guyana, Paraguay, Peru, Venezuela.

bolivianus Scivittaro, 1975

bolivianus Scivittaro, 1975: 460.

Type depository. DEI (HT!, PT!).

Distribution. Bolivia.

championi (Sharp, 1895)

championi Sharp, 1895a: 6 (*Abactus*).

Type depository. NHM (HT!).

Distribution. Panama.

dimidiatus Scivittaro, 1975

dimidiatus Scivittaro, 1975: 450.

Type depository. DEI (HT!).

Distribution. Peru.

foveatus (Lund, 1800)

foveatus Lund, 1800: 69 (*Brentus*).

foveolatus: Gyllenhal (in Schoenherr, 1833: 367) (*Taphroderes*) [lapsus].

Type depository. ZMUC (ST).

Distribution. Costa Rica; Brazil, Colombia, French Guyana, Peru.

kissingeri (Sleeper, 1960)

kissingeri Sleeper, 1960: 83 (*Abactus*).

Type depository. CAS (HT).

Distribution. Mexico.

monnei Scivittaro, 1975

monnei Scivittaro, 1975: 452.

Type depository. URMU (HT).

Distribution. Venezuela.

***nevermanni* Kleine, 1927**

nevermanni Kleine, 1927e: 293.

nevermonni: Scivittaro, 1975: 442 (*Taphroderes*) [lapsus].

Type depositories. DEI (LT!), SMTD (PLT), ZMUH (PLT), NMNH (Nevermann coll.) (PLT).

Distribution. Costa Rica; Venezuela.

Notes. Lectotype of *T. nevermanni* (DEI) designated on a specimen with the following labels: Costa Rica, E. Nevermann, 30.IV.1925/ Hamburgfarm, Revendazon, Ebene Limon/ *Taphroderes nevermanni* Kln., R. Kleine det., 1927/ non *Abactrus championi* Sharp/ cotype No. 54645, U.S.N.M./ Syntypus.

***sahlbergi* (Sharp, 1895)**

sahlbergi Sharp, 1895a: 6 (*Abactrus*).

Type depository. NHM (HT!).

Distribution. Brazil, Paraguay.

***venezuelanus* Scivittaro, 1975**

venezuelanus Scivittaro, 1975: 462.

Type depositories. MCCV (HT, PT), SMFD (PT).

Distribution. Venezuela.

***Taphroderoides* Scivittaro, 1975**

Taphroderoides Scivittaro, 1975: 470.

Type species: *Taphroderes doctus* Kleine, 1927.

Distribution. Brazil.

***doctus* (Kleine, 1927)**

doctus Kleine, 1927b: 455 (*Taphroderes*).

Type depository. NHM (HT!).

Distribution. Brazil.

Taphroderomimus Kleine, 1927

Taphroderomimus Kleine, 1927b: 515.

Taphroderominus: Kleine, 1927b: 453 [lapsus].

Type species: *Taphroderomimus disjunctus* Kleine, 1927.

Distribution. Brazil.

disjunctus Kleine, 1927

disjunctus Kleine, 1927b: 455.

Type depositories. DEI (LT!), IPEACS (PLT), SMTD (PLT).

Distribution. Brazil.

Notes. Scivittaro (1975: 468) reported that the specimens held in IPEACS and in DEI are the holotype and a paratype, but in his original description Kleine did not designate any of the syntypes as holotype; however, as there is only one syntype in DEI, this must be considered as lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999). The lectotype has the following labels: Est. Esp. Santo, Guandù, 30.XI.1920, F. Hoffmann/ Syntypus/ *Taphroderominus disjunctus* Kleine, Scivittaro det., 1973.

Taphroderopsis Scivittaro, 1975

Taphroderopsis Scivittaro, 1975: 474.

Taphroderes Schoenherr, 1833: 366 (non Schoenherr, 1823); syn. Alonso-Zarazaga & Lyal, 1999: 53.

Type species: *Taphroderes brevipes* Gyllenhal, 1833.

Distribution. Costa Rica, Guatemala, Mexico, Nicaragua, Panama, Saint Vincent and the Grenadines (Saint Vincent); Bolivia, Brazil, Colombia, French Guyana, Peru, Suriname, Trinidad and Tobago, Venezuela.

abdominalis Scivittaro, 1975

abdominalis Scivittaro, 1975: 512.

Type depository. NMNH (HT).

Distribution. Bolivia.

annulata (Kleine, 1921)

annulatus Kleine, 1921m: 60 (*Taphroderes*).

Type depository. RMNH (HT).

Distribution. Colombia.

apicalis (Sharp, 1895)

apicalis Sharp, 1895a: 4 (*Taphroderes*).

Type depository. NHM (LT!, PLT!).

Distribution. Costa Rica, Nicaragua, Panama; Brazil.

Notes. Lectotype of *T. apicalis* (NHM) designated on a specimen with the following labels: *Taphroderes apicalis* Types D. S./ syntype/ Bugaba, 800-1500 ft., Champion/ B.C.A., Col. IV, 6. *Taphroderes apicalis* Sharp/ Sharp Coll. 1905-313/ Prépar. genit. No BM 77, R. Damoiseau. Paralectotypes: 2 specimens (NHM), *Taphroderes apicalis* D. S./ Syntype/ Bugaba, 800-1500 ft., Champion/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Taphroderes apicalis* Sharp/ Scivittaro det., 1973, *Taphroderopsis apicalis* (Sharp); 1 specimen (NHM), Syntype/ V. de Chiriqui, 25-4000 ft., Champion/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Taphroderes apicalis* Sharp/ Scivittaro det., 1973, *Taphroderopsis apicalis* (Sharp); 1 specimen (NHM), Syntype/ Nicaragua, T. Belt/ B.C.A., Col. IV, 6. *Taphroderes apicalis* Sharp/ R. Damoiseau det., *Taphroderes apicalis* (Sharp); 1 specimen (NHM), Syntype/ V. de Chiriqui, 25-4000 ft., Champion/ B.C.A., Col. IV, 6. *Taphroderes apicalis* Sharp.

bicolor Scivittaro, 1975

bicolor Scivittaro, 1975: 528.

Type depository. ZMAN (HT!).

Distribution. French Guyana.

brevipes (Gyllenhal, 1833)

brevipes Gyllenhal (in Schoenherr, 1833: 366) (*Taphroderes*).

breviceps: Kleine, 1928a: 93 (*Taphroderes*) [lapsus].

Type depository. NHRS (HT).

Distribution. Brazil, Colombia, Suriname.

claviger Scivittaro, 1975

claviger Scivittaro, 1975: 514.

Type depository. DEI (HT!).

Distribution. Bolivia.

curvidens (Kleine, 1920)

curvidens Kleine, 1920h: 2 (*Taphroderes*).

Type depository. NHRS (?) (ST).

Distribution. Bolivia, Brazil, French Guyana, Peru.

Notes. Kleine (1920h: 2) reported that the type for *T. curvidens* was in NHRS but the species is not listed in the NHRS web site.

filiformis (Pascoe, 1872)

filiformis Pascoe, 1872: 319 (*Taphroderes*).

piliformis: Senna, 1893a: 52 (*Taphroderes*) [lapsus].

Type depository. NHM (HT!).

Distribution. Brazil, Venezuela.

goiana Scivittaro, 1975

goianus Scivittaro, 1975: 532.

Type depository. MNHUB (HT!).

Distribution. Brazil.

mammillata Scivittaro, 1975

mammillatus Scivittaro, 1975: 508.

Type depositories. CMNH (HT, PT), MZSP (PT).

Distribution. Brazil.

marisae Scivittaro, 1975

marisae Scivittaro, 1975: 526.

Type depository. NHM (HT!).

Distribution. Brazil.

mendax Scivittaro, 1975

mendax Scivittaro, 1975: 520.

Type depositories. MZSP (HT), MNRJ (Campos Seabra coll.) (PT).

Distribution. Brazil, Venezuela.

mexicana (Sharp, 1895)

mexicanus Sharp, 1895a: 3 (*Taphroderes*).

Type depository. NHM (LT!, PLT!).

Distribution. Guatemala, Mexico.

Notes. Lectotype of *T. mexicanus* (NHM) designated on a specimen with the following labels: Syntype/ Toxпам/ B.C.A., Col. IV, 6. *Taphroderes mexicanus* Sharp/ Sharp Coll. 1905-313. Paralectotypes: 1 specimen (NHM), Syn-

type/ 2500/ Toxpam, Mexico Sall Coll./ B.C.A., Col. IV, 6. *Taphroderes mexicanus* Sharp/ *Taphroderes* sp., Salle; 2 specimens (NHM), *T. mexicanus* D. S./ Syntype/ Mexico, Sall Coll./ Toxpam/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Taphroderes mexicanus* Sharp/ Scivittaro det., 1973, *Taphroderopsis oscillator* (Sharp); 1 specimen (NHM), Syntype/ Mexico, Sall Coll./ Toxpam/ B.C.A., Col. IV, 6. *Taphroderes mexicanus* Sharp/ R. Damoiseau det., *Taphroderes mexicanus* Sharp/ Prépar. génit. No. 77 A 22 R. Damoiseau; 1 specimen (NHM), Syntype/ Tamahu, Vera Paz, Champion/ B.C.A., Col. IV, 6. *Taphroderes mexicanus* var. Sharp/ Scivittaro det., 1973, *Taphroderopsis oscillator* (Sharp); 1 specimen (NHM), Syntype/ Jalapa, Vera Paz, Höge/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Taphroderes mexicanus* var. Sharp/ Scivittaro det., 1973, *Taphroderopsis oscillator* (Sharp).

obtusa (Pascoe, 1872)

obtus Pascoe, 1872: 320 (*Taphroderes*).

Type depository. NHM (HT!).

Distribution. Costa Rica, Trinidad and Tobago; Bolivia, Brazil.

oscillator (Sharp, 1895)

oscillator Sharp, 1895a: 5 (*Taphroderes*).

Type depository. NHM (LT!, PLT!).

Distribution. Panama, Costa Rica; Brazil, French Guyana.

Notes. Lectotype of *T. oscillator* (NHM) designated on a male specimen with the following labels: Syntype/ V. de Chiriqui, 3-4000 ft, Champion/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Taphroderes oscillator* Sharp. Paralectotypes: 2 female specimens (NHM), Syntype/ V. de Chiriqui, 3-4000 ft, Champion/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Taphroderes oscillator* Sharp, Scivittaro det., 1973, *Taphroderopsis oscillator* (Sharp).

patriciae Scivittaro, 1975

patriciae Scivittaro, 1975: 496.

Type depository. NHM (HT!).

Distribution. Brazil.

***quadrisignata* (Erichson, 1847)**

quadrisignatus Erichson, 1847: 127 (*Taphroderes*).

Type depository. MNHUB (?) (HT).

Distribution. Guatemala; Bolivia, Brazil, Ecuador, Peru, Venezuela.

Notes. According to Kleine (1928a: 93) the type was in MNHUB, but we did not find it there.

***recta* (Sharp, 1895)**

rectus Sharp, 1895a: 3 (*Taphroderes*).

Type depository. NHM (LT!, PLT!).

Distribution. Costa Rica, Guatemala, Panama; Brazil.

Notes. Lectotype of *T. rectus* (NHM) designated on a specimen with the following labels: *Taphroderes rectus* Types D. S./ V. de Chiriqui, 25-4000 ft., Champion/ Syntype/ B.C.A., Col. IV, 6. *Taphroderes rectus* Sharp/ Sharp Coll. 1905-313/ Prépar. genit. No BM 76 R. Damoiseau. Paralectotypes: 9 specimens (NHM), *Taphroderes rectus* Types D. S./ V. de Chiriqui, 25-4000 ft., Champion/ Syntype/ B.C.A., Col. IV, 6. *Taphroderes rectus* Sharp/ Sharp Coll. 1905-313; 1 specimen (NHM), syntype/ Bugaba, 800-1,500 ft., Champion/ B.C.A., Col. IV, 6. *Taphroderes rectus* Sharp/ Sharp Coll. 1905-313.

***sexmaculata* (Boheman, 1840)**

sexmaculatus Boheman (in Schoenherr, 1840: 573) (*Taphroderes*).

beltianus Sharp, 1895a: 4 (*Taphroderes*); syn. Kleine, 1927a: 70.

Type depositories. *T. sexmaculatus*: OXUM (HT!); *T. beltianus*: NHM (LT!, PLT!).

Distribution. Costa Rica, Nicaragua, Panama, Saint Vincent and the Grenadines (Saint Vincent); Brazil.

Notes. Lectotype of *T. beltianus* (NHM) designated on a specimen with the following labels: Syntype/ V. de Chiriqui, 25-4000 ft Champion/ B.C.A., Col. IV, 6. *Taphroderes beltianus* Sharp/ R. Damoiseau det. *Taphroderes bel-*

tianus Sharp. Paralectotypes: 1 specimen (NHM), with the same label data as the lectotype; 1 specimen (NHM), Syntype/ Chontales, Nicaragua, T. Belt/ B.C.A., Col. IV, 6. *Taphroderes beltianus* Sharp/ R. Damoiseau det. *Taphroderes beltianus* Sharp; 1 specimen (NHM), Syntype/ Nicaragua/ Salle Coll./ B.C.A., Col. IV, 6. *Taphroderes beltianus* Sharp/ R. Damoiseau det., *Taphroderes beltianus* Sharp.

similis Scivittaro, 1975

similis Scivittaro, 1975: 481.

Type depositories. MNRJ (Campos Seabra coll.) (HT, PT), DEI (PT!), SMFD (PT).

Distribution. Bolivia, Brazil.

soaresi Scivittaro, 1975

soaresi Scivittaro, 1975: 498.

Type depositories. AMNH (HT), NHM (PT!), MZSP (PT).

Distribution. Brazil.

striolata (Labram & Imhoff, 1841)

striolatus Labram & Imhoff, 1841: fasc. 8 [page not numbered] (*Taphroderes*).

Type depository. MLUH (ST).

Distribution. Costa Rica; Brazil.

tosta (Sharp, 1895)

tostus Sharp, 1895a: 4 (*Taphroderes*).

Type depository. NHM (HT!).

Distribution. Guatemala.

***ubirajarai* Scivittaro, 1975**

ubirajarai Scivittaro, 1975: 510.

Type depositories. CMNH (HT), MZSP (PT).

Distribution. Brazil.

***ventralis* (Sharp, 1895)**

ventralis Sharp, 1895a: 5 (*Taphroderes*).

Type depository. NHM (LT!, PLT!).

Distribution. Panama.

Notes. Lectotype of *T. ventralis* (NHM) designated on a specimen with the following labels: Syntype/ V. de Chiriqui, 25-4000 ft, Champion/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Taphroderes ventralis* Sharp. Paralectotypes: 1 specimen (NHM), with the same label data as the lectotype; 1 specimen (NHM), Syntype/ Bugaba, 800-1,500 ft., Champion/ B.C.A., Col. IV, 6. *Taphroderes ventralis* Sharp/ Scivittaro det., 1973, *Taphroderopsis ventralis* (Sharp).

TRACHELIZINAE Lacordaire, 1866

Trachelizinae Lacordaire, 1866: 417.

TRACHELIZINI Lacordaire, 1866

Trachélizides Lacordaire, 1866: 417.

Héphébocérides Lacordaire, 1866: 415.

Trachelizinae Pascoe, 1872: 317.

Ephebocerinae: Pascoe, 1872: 317 [lapsus].

Trachelizi Kolbe, 1883c: 74.

Hephebocerini Kolbe, 1883e: 185.

Trachelizina Sharp, 1895a: 19.

Epheboceridae Schönfeldt, 1908: 15.

Trachelizidae Schönfeldt, 1908: 18.

Ephebocerini Schönfeldt, 1910a: 9.

Trachelizini Schönfeldt, 1910a: 10.

Anchistenini K. E. Schedl, 1961c: 198.

Hepheboceridae Damoiseau, 1963b: 203.

Allodapinus Hedicke, 1923

Allodapinus Hedicke, 1923: 72 [replacement name for *Allodapus*].

Allodapus Kleine, 1922h: 145 (non Fieber, 1860).

Allodapsa Strand, 1928: 74 [unjustified replacement name].

Type species: *Allodapus hospiton* Kleine, 1922.

Distribution. Indonesia (Sulawesi), Malaysia; New Guinea.

flavosignatus (Kleine, 1935)

flavosignata Kleine, 1935e: 306 (*Hypomiolispa*).

Type depository. NHM (HT!).

Distribution. New Guinea.

hospiton (Kleine, 1922)

hospiton Kleine, 1922h: 147 (*Allodapus*).

Type depository. MNHUB (HT!).

Distribution. Indonesia (Sulawesi); New Guinea.

kleinei Damoiseau, 1989

kleinei Damoiseau, 1989b: 74 [replacement name for *opulentus*].

opulentus Kleine, 1936c: 138 (*Allodapinus*).

Type depository. *A. kleinei* (= *A. opulentus*): NHM (HT!).

Distribution. Malaysia.

Notes. Damoiseau (1989: 74) gave the new name *kleinei* to *Allodapinus opulentus* Kleine, 1936 due to the existence of *A. opulentus* (Kleine, 1935), described as *Hypomiolispa*.

longithorax Damoiseau, 1989*longithorax* Damoiseau, 1989b: 75.**Type depository.** MONZ (HT!).**Distribution.** New Guinea.***opulentus*** (Kleine, 1935)*opulenta* Kleine, 1935e: 306 (*Hypomiolispa*).**Type depository.** NHM (HT!).**Distribution.** New Guinea.***papuanus*** Damoiseau, 1989*papuanus* Damoiseau, 1989b: 72.**Type depository.** NHM (HT!).**Distribution.** New Guinea.***Anchisteus*** Kolbe, 1883*Anchisteus* Kolbe, 1883e: 185.**Type species:** *Anchisteus peregrinus* Kolbe, 1883.**Distribution.** Madagascar.***niger*** Damoiseau, 1963*niger* Damoiseau, 1963h: 1.**Type depository.** IRSNB (HT!, PT!).**Distribution.** Madagascar.

peregrinus Kolbe, 1883*peregrinus* Kolbe, 1883e: 185.**Type depository.** MNHUB (?) (HT).**Distribution.** Madagascar.**Notes.** According to Kleine (1928a: 75) the type was in MNHUB, but we did not find it there.*Anocamara* Kleine, 1920*Anocamara* Kleine, 1920l: 120.**Type species:** *Anocamara proportionalis* Kleine, 1920.**Distribution.** Borneo, Indonesia (Sumatra), Malaysia.*borneensis* Damoiseau, 1989*borneensis* Damoiseau, 1989b: 76.**Type depository.** BPBM (HT).**Distribution.** Borneo.*catenata* Kleine, 1925*catenata* Kleine, 1925b: 132.**Type depository.** NHM (HT!).**Distribution.** Malaysia.*proportionalis* Kleine, 1920*proportionalis* Kleine, 1920l: 122.**Type depositories.** DEI (HT!), SMTD (PT), MZPW (PT).**Distribution.** Borneo, Indonesia (Sumatra), Malaysia.

undulata Damoiseau, 1989

undulata Damoiseau, 1989b: 77.

Type depository. MHNG (HT, PT).

Distribution. Malaysia.

Bothriorhinus Fairmaire, 1881

Bothriorhinus Fairmaire, 1881c: 421.

Bothyrorhynus: Deyrolle, 1881: 522 [lapsus].

Bothriorrhinus W. F. Kirby, 1882: 89 [unjustified emendation].

Botriorrhinus: Kleine, 1920a: 32 [lapsus].

Type species: *Bothriorhinus costulipennis* Fairmaire, 1881.

Distribution. Papua New Guinea (Bismarck Archipelago: Duke of York, New Britain).

Notes. Fairmaire redescribed the genus in 1883 (p. 42).

costulipennis Fairmaire, 1881

costulipennis Fairmaire, 1881c: 421.

costulippennis: Deyrolle, 1881: 522 (*Bothyrorhynus*) (sic!) [lapsus].

Type depository. Unknown.

Distribution. Papua New Guinea (Bismarck Archipelago: Duke of York, New Britain).

Notes. Fairmaire redescribed the species in 1883 (p. 42).

Euschizus Kleine, 1922

Euschizus Kleine, 1922g: 225.

Type species: *Euschizus alatus* Kleine, 1922 = *Ceocephalus internatus* Pascoe, 1872.

Distribution. Australia (including Norfolk Is.).

Notes. According to Damoiseau (1989b: 79) the genus includes three species which he listed together with their synonymies; Zimmerman (1994: 201) considered them differently. Here we follow Damoiseau's opinion.

dictatorius Kleine, 1926

dictatorius Kleine, 1926f: 370.

Type depository. NHM (HT!).

Distribution. Australia.

internatus (Pascoe, 1872)

internatus Pascoe, 1872: 324 (*Ceocephalus*).

alatus Kleine, 1922g: 226 (*Euschizus*); syn. Damoiseau, 1989b: 79.

Type depositories. *C. internatus*: NHM (LT!, PLT!); *E. alatus*: DEI (LT!, PLT!).

Distribution. Australia (including Norfolk Is.).

Notes. Lectotype of *C. internatus* (NHM) designated on a specimen with the following labels: Syntype/ Type/ Queensland/ *Ceocephalus internatus* Pascoe Type/ Pascoe Coll. 93-60/ *Ceocephalus internatus* Pasc. Paralectotype: 1 specimen (NHM), Syntype/ Co-type/ Queensland/ Pascoe Coll. 93-60/ *Ceocephalus internatus* Pasc., co-type.

Lectotype of *E. alatus* (DEI) designated on a male specimen with the following labels: Blackall, Range/ Syntype/ *Euschizus alatus* Kleine/ Kleine det. Paralectotype: 1 female specimen (DEI), Wide Bay/ Syntype/ Kleine det.

tenuitarsis (Pascoe, 1872)

tenuitarsis Pascoe, 1872: 324 (*Ceocephalus*).

alarius Kleine, 1922g: 224 (*Trachelizus*); syn. Damoiseau, 1989b: 79.

Type depositories. *C. tenuitarsis*: NHM (HT!); *T. alarius*: DEI (LT!, PLT!).

Distribution. Australia.

Notes. Lectotype of *T. alarius* (DEI) designated on a male specimen (on the right side) with the following labels: Syntype/ Kleine det./ Coll Hacker/ Austral./ *Trachelizus alarius* Kln., det. 1921. Paralectotype: 1 female specimen (on the left side) (DEI), with the same label data as the lectotype [both specimens are glued on the same card].

***Hemisamblus* Kleine, 1925**

Hemisamblus Kleine, 1925b: 132.

Type species: *Hemisamblus contemptus* Kleine, 1925.

Distribution. Laos.

***contemptus* Kleine, 1925**

contemptus Kleine, 1925b: 133.

Type depository. NHM (HT!).

Distribution. Laos.

***Hephebocerus* Schoenherr, 1840**

Hephebocerus Schoenherr, 1840: 501.

Ephebocerus Agassiz, 1846b: 178 [unjustified emendation].

Ephebocerus Gemminger & Harold, 1872: 2705 (non Agassiz, 1846) [unjustified emendation].

Pseudoparatrachelizus Haedo Rossi, 1954a: 1; syn. Scivittaro, 1982: 334.

Type species: *Brenthus nanus* Boheman, 1833.

Distribution. Costa Rica, Cuba, Dominican Republic, Guadeloupe Is., Mexico, Nicaragua, Panama, Trinidad and Tobago; Argentina, Bolivia, Brazil, Colombia, Ecuador, Paraguay, Peru, Venezuela.

***mexicanus* Sharp, 1895**

mexicanus Sharp, 1895a: 19.

oscillator Sharp, 1895a: 20 (*Trachelizus*); syn. Scivittaro, 1982: 338.

Type depositories. *H. mexicanus*: NHM (LT!, PLT!); *T. oscillator*: NHM (LT!).

Distribution. Costa Rica, Dominican Republic, Mexico, Nicaragua, Panama, Trinidad and Tobago; Argentina, Bolivia, Peru, Venezuela.

Notes. Lectotype of *H. mexicanus* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ Motzorongo, Vera Cruz, Flohr/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Hephebocerus mexicanus* Sharp/ *Hephebocerus mexicanus* Sharp Scivittaro ex 1979. Paralectotypes: 1 male specimen (NHM), Syntype/ V. de Chiriqui, 25-4000 ft., Champion/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Hephebocerus mexicanus* Sharp; 1 male specimen (NHM), Syntype/ Chontales, Nicaragua, T. Belt/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Hephebocerus mexicanus* Sharp.

Lectotype of *T. oscillator* (NHM) designated on a female specimen with the following labels: Syntype/ Type/ Cordova, Mexico, Sallé/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus oscillator* Sharp/ Prépar. génit. No BM 74 R. Damoiseau/ *Hephebocerus mexicanus* Sharp, Scivittaro det. 1981; no other syntypes found in NHM.

nanus (Boheman, 1833)

nanus Boheman (in Schoenherr, 1833: 355) (*Brenthus*).

boops Boheman (in Schoenherr, 1840: 503) (*Hephebocerus*); syn. Damoiseau, 1963h: 7.

notatus Boheman (in Schoenherr, 1840: 500) (*Trachelizus*); syn. Damoiseau, 1963h: 7.

dufaui Denier, 1922: 24 (*Ephebocerus*); syn. Scivittaro, 1982: 335.

lilloi Haedo Rossi, 1954a: 2 (*Hephebocerus*); syn. Scivittaro, 1982: 335.

oscillator Haedo Rossi, 1954: 71 (non Sharp, 1895) (*Pseudoparatrachelizus*); syn. Scivittaro, 1982: 335.

Type depositories. *B. nanus*: NHRS (HT); *H. boops*: NHRS (HT); *T. notatus*: NHRS (ST); *E. dufau*: MNHN (?) (ST), Denier coll. (MLPA) (?) (ST); *H. lilloi*: IMLA (HT).

Distribution. Cuba, Guadeloupe Is., Mexico; Argentina, Brazil, Colombia, Ecuador, Paraguay, Peru.

Notes. We found three specimens of *H. boops* in MNHN with the following labels: Brasil/ Chevrolat 14/ ex Musaeo G. Power/ type. We are not sure if we should consider them as types.

Denier (1922: 26) reported that the four syntypes of *E. dufau* were in MNHN and in his collection, but we did not find any syntype in MNHN.

rotundatus Damoiseau, 1963

rotundatus Damoiseau, 1963h: 5.

Type depository. IRSNB (HT!).

Distribution. Brazil.

soaresi Scivittaro, 1981

soaresi Scivittaro, 1981: 35.

Type depository. MZSP (HT, PT).

Distribution. Brazil.

Homophylus Kleine, 1920

Homophylus Kleine, 1920j: 244.

Type species: *Homophylus castaneus* Kleine, 1920.

Distribution. Borneo, Indonesia (Java), Philippines (Mindanao), Vietnam.

castaneus Kleine, 1920

castaneus Kleine, 1920j: 246.

Type depository. RMNH (LT!, PLT!).

Distribution. Indonesia (Java).

Notes. Lectotype of *H. castaneus* (RMNH) designated on a male specimen with the following labels: P. F. Sijthoff, Preanger, Java/ Lectotype *H. castaneus*, R. Damoiseau det. [*in litteris*]. Paralectotype: 1 female specimen (RMNH), with the same label data as the lectotype.

***durus* Kleine, 1920**

durus Kleine, 1920j: 246.

Type depository. RMNH (HT!).

Distribution. Indonesia (Java).

***indosinensis* Damoiseau, 1966**

indosinensis Damoiseau, 1966b: 449.

Type depositories. BPBM (HT, AT, PT), IRSNB (PT!).

Distribution. Vietnam.

***lewisi* Damoiseau, 1989**

lewisi Damoiseau, 1989b: 80.

Type depository. NHM (HT!).

Distribution. Borneo.

***mindanensis* Kleine, 1925**

mindanensis Kleine, 1925c: 595.

Type depository. SMTD (HT).

Distribution. Philippines (Mindanao).

***Hypomiolispa* Kleine, 1918**

Hypomiolispa Kleine, 1918f: 163.

Hypomialispa: Kleine, 1920a: 31 [lapsus].

Hypomyolispa: Zherikhin & Gratshev, 1995: 637 [lapsus].

Hipomiolispa: Kabakov, 2001: 207 [lapsus].

Type species: *Miolispa ceylonica* Desbrochers des Loges, 1890.

Distribution. Japan, Taiwan; Borneo, India (including Andaman Is., Assam, Darjeeling), Indonesia (Batu; Engano; Java; Mentawai; Moluccas: Bacan [= Batjan], Kay, Seram [= Ceram]; Simeulue [= Simalur]; Sulawesi; Suma-

tra), Malaysia, Myanmar [= Burma], Philippines (Basilan, Luzon, Mindanao, Palawan, Samar, Tawi Tawi), Sri Lanka, Thailand, Vietnam; New Guinea (including Bismarck Archipelago).

***amoena* (Perroud, 1853) n. comb.**

amoenus Perroud, 1853: 423 (*Ceocephalus*).

Type depository. MNHN (HT!).

Distribution. Indonesia (Java).

***bickhardti* Kleine, 1918**

bickhardti Kleine, 1918h: 323.

Type depository. MZPW (ST).

Distribution. Borneo, Indonesia (Sumatra).

***ceylonica* (Desbrochers des Loges, 1890)**

ceylonica Desbrochers des Loges, 1890: 222 (*Miolispa*).

Type depositories. MNHN (?) (ST), NZSI (ST).

Distribution. India, Sri Lanka.

Notes. According to Kleine (1928a: 78) types were in Oberthür's collection (now in MNHN and IRSNB), but we did not find them there.

***chapaensis* Kleine, 1933**

chapaensis Kleine, 1933d: 303.

Type depository. SMTD (ST).

Distribution. Vietnam.

clavata Kleine, 1918

clavata Kleine, 1918h: 327.

Type depository. MNHUB (HT!).

Distribution. Borneo, Indonesia (Sumatra), Malaysia, Philippines (Luzon).

colorata Kleine, 1925

colorata Kleine, 1925b: 136.

Type depository. NMH (LT!, PLT!).

Distribution. Sri Lanka.

Notes. Lectotype of *H. colorata* (NHM) designated on a male specimen with the following labels: Syntype/ Typus! ♂ / Bogawantalawa, 4,900-5,200 ft, 28.II-12.III.1882/ Ceylon, G. Lewis 1910-320/ *Hypomiolisa colorata* Kln., R. Kleine det. 1922/ Prépar. génit. No. BM 59, R. Damoiseau. Paralectotype: 1 female specimen (NHM), Syntype/ Typus! ♀ / Dikoya, 3,800-4,200 ft, 6.XII.81-16.I.82/ Ceylon, G. Lewis 1910-320/ *Hypomiolisa colorata* Kln., R. Kleine det. 1922.

compressa Kleine, 1918

compressa Kleine, 1918h: 341.

Type depositories. MNHUB (HT!, PT!), MZPW (AT, PT), NHRS (PT), SMTD (PT).

Distribution. Borneo, India, Indonesia (Java; Moluccas: Bacan [= Batjan], Kay, Seram [= Ceram]; Sumatra), Malaysia, Vietnam.

conformis (Senna, 1892)

conformis Senna, 1892a: 175 (*Miolisa*).

Type depository. RMNH (HT!).

Distribution. Indonesia (Java).

conjugalis Kleine, 1925

conjugalis Kleine, 1925b: 134.

Type depository. NHM (LT!, PLT!).

Distribution. India (Andaman Is.).

Notes. Lectotype of *H. conjugalis* (NHM) designated on a male specimen with the following labels: Syntype/ Typus!/ Andaman Islands/ Fry Coll. 1905-100/ Roepstorff/ *Hypomiolispa conjugalis* Kln. R. Kleine det. 1922/ Prépar. génit. No. BM 56 R. Damoiseau. Paralectotype: 1 female specimen (NHM), Syntype/ Andamans (Roepstorff)/ *Hypomiolispa conjugalis* Kln. ♀ Type, R. Kleine det. 1922.

conjuncta Kleine, 1925

conjuncta Kleine, 1925b: 136.

Type depository. NHM (HT!).

Distribution. Myanmar [= Burma].

consociata Kleine, 1923

consociata Kleine, 1923j: 408.

Type depository. Buitenzorg Museum (?) (HT).

Distribution. Indonesia (Sumatra), Philippines (Tawi Tawi).

conspecta Kleine, 1926

conspecta Kleine, 1926a: 28.

Type depository. NHM (HT!).

Distribution. Indonesia (Sumatra), Malaysia, "Philippines".

conspicua Kleine, 1926

conspicua Kleine, 1926a: 28.

Type depository. NHM (HT!).

Distribution. Borneo.

cruda Kleine, 1925

cruda Kleine, 1925b: 135.

Type depository. NHM (HT!).

Distribution. India (Assam).

demissa Kleine, 1923

demissa Kleine, 1923i: 271.

Type depository. NHM (HT!).

Distribution. Malaysia.

dentigena Kleine, 1918

dentigena Kleine, 1918h: 331.

Type depository. MZPW (ST).

Distribution. Borneo, Indonesia (Sumatra), Myanmar [= Burma], Thailand.

dissimilis Kleine, 1943

dissimilis Kleine, 1943b: 139.

Type depository. HNHM (HT).

Distribution. New Guinea.

elegans (Senna, 1895)

elegans Senna, 1895a: 192 (*Miolispa*).

Type depository. RMNH (HT!).

Distribution. Indonesia (Sumatra).

enganica (Senna, 1893)

enganica Senna, 1893b: 267 (*Miolispa*).

Type depositories. MSNG (LT!, PLT!), MZUF (PLT!).

Distribution. Indonesia (Engano).

Notes. In his original description Senna listed three syntypes, but the typical series seems to consist of four specimens (MSNG and MZUF), all with the same collecting data.

Lectotype (MSNG) designated on a specimen with the following labels: Engano, Bua Bua, V-VI.1891, Modigliani/ *Miolispa enganica* Senna Typus!. Paralectotypes: 1 specimen (MSNG), Engano, Bua Bua, V-VI.1891, Modigliani; 1 specimen (MZUF), Engano, Bua Bua, V-VI.1891, Modigliani/ *M. enganica* Senna, cotypus!/ Ann. Mus. Gen. (2), XIII, 1893 cotypus! ♂/ Prépar. génit. N° 76 J 23, R. Damoiseau/ ex-coll. Senna/ "La Specola" Firenze, 7001.

exarata (Desbrochers des Loges, 1890)

exarata Desbrochers des Loges, 1890: 223 (*Miolispa*).

humeralis Senna, 1903: 171 (*Miolispa exarata* var.); syn. Kleine, 1918e: 347.

sumatrana Senna, 1903: 170 (*Miolispa exarata* var.); syn. Kleine, 1918e: 347.

Type depositories. *M. exarata*: MNHN (Desbrochers coll.) (?) (HT); *M. exarata* var. *humeralis*: MSNG (LT!, PLT!); *M. exarata* var. *sumatrana*: MSNG (LT!, PLT!).

Distribution. Borneo, Indonesia (Batu, Java, Mentawai, Sumatra), Malaysia, Philippines (Basilan, Mindanao, Samar); New Guinea.

Notes. Desbrochers des Loges (1891: 223) reported that the type of *M. exarata* was in his collection, which should now be in MNHN, but we did not find it there.

Lectotype of *M. exarata* var. *humeralis* (MSNG) designated on a specimen with the following labels: Sumatra, Pangherang, Pisang, X.90 and III.91, E. Modigliani/ *Miolispa exarata* Desbr. var. *humeralis* Senna. Paralectotypes: 3

specimens (MSNG), Sumatra, Pangherang, Pisang, X.90 and III.91, E. Modigliani; 2 specimens (MSNG), Sumatra, Sirambè, XII.90-III.91, E. Modigliani.

Lectotype of *M. exarata* var. *sumatrana* (MSNG) designated on a specimen with the following labels: Sumatra, Sirambè, XII.90-III.91, E. Modigliani/ *Miolispa exarata* Desbr. var. *sumatrana* Senna. Paralectotypes: 4 specimens (MSNG), Sumatra, Pangherang, Pisang, X.90 and III.91, E. Modigliani.

exigua Kleine, 1918

exigua Kleine, 1918h: 334.

Type depository. MNHUB (LT!, PLT!).

Distribution. Borneo, Indonesia (Java).

Notes. Lectotype of *H. exigua* (MNHUB) designated on a specimen with the following labels: Borneo/ 111820/ *Hypomiolispa exigua* Kleine. Paralectotype: 1 specimen (MNHUB), with the same label data as the lectotype.

exquisita Kleine, 1939

exquisita Kleine, 1939g: 365.

Type depository. SMKM (HT).

Distribution. Malaysia.

Notes. Kleine described this species as new twice (1943a: 2).

fasciata Kleine, 1920

fasciata Kleine, 1920j: 242.

Type depository. RMNH (HT!).

Distribution. Indonesia (Sumatra).

fausti (Senna, 1895)

fausti Senna, 1895a: 188 (*Miolispa*).

Type depository. RMNH (LT!, PLT!).

Distribution. Borneo, Indonesia (Batu, Mentawai, Sumatra), Malaysia, Philippines (Luzon), Thailand.

Notes. According to Senna's original description, one male and one female of the typical series were in RMNH, another female was in the Faust collection.

Lectotype of *M. fausti* (RMNH) designated on a male specimen with the following labels: Sumatra, Deli, Schagen v. Leeuwen/ *Hypomiolispa fausti* Lectotype, R. Damoiseau det. [*in litteris*]. Paralectotype: 1 female specimen (RMNH), J. L. Weyers, Thambang Salida, Sumatra/ *Hypomiolispa fausti* Allotype, Damoiseau det. [*in litteris*].

ferox Kleine, 1926

ferox Kleine, 1926a: 29.

Type depository. ZMAN (LT!, PLT!).

Distribution. Indonesia (Sumatra).

Notes. Lectotype of *H. ferox* (ZMAN) designated on a male specimen with the following labels: Boekit Gabah, Z.W.K. Sumatra, XII.1918, leg. H. Lucht, coll. F. C. Drescher. Paralectotype: 1 female specimen (ZMAN) with the same label data as the lectotype.

gardneri Kleine, 1931

gardneri Kleine, 1931a: 94.

Type depository. NHM (HT!).

Distribution. India (Darjeeling).

glabra Kleine, 1944

glabra Kleine, 1944: 152.

Type depositories. MNHN (LT!), DEI (PLT!), unknown (PLT).

Distribution. Indonesia (Java).

Notes. Döbler & Gaedike (1995: 446) reported that the specimen held in MNHN is the holotype, and that two paratypes are in DEI and in the Kleine's collection, but in his original description Kleine did not designate any of the three syntypes as holotype; however, as there is only one syntype in MNHN, this must be considered as lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999). The lectotype has the following labels: Museum Paris, Java, J. D. Pasteur 268-94/ type/ prépar. génit. N. 78 D 20 R. Damoiseau/ *Hypomiolispa glabra* Kln, R. Kleine det. 1943.

grandis Damoiseau, 1989

grandis Damoiseau, 1989b: 80.

Type depository. NHMB (HT!).

Distribution. Indonesia (Sumatra).

helleri Kleine, 1918

helleri Kleine, 1918h: 329.

Type depository. SMTD (ST).

Distribution. Philippines (Basilan, Mindanao).

incerta Kleine, 1937

incerta Kleine, 1937i: 204.

Type depositories. DEI (LT!), IFRI (PLT).

Distribution. India (Darjeeling).

Notes. Lectotype of *H. incerta* (DEI) designated on a specimen with the following labels: Kurseong, Bengal, R. O. Coll. 16.V.1936./ R. R. D. 967, B. C. R. 69, Cage 838, ex *Anthocephalus cadamba* 469/ Syntypus/ *Hypomiolispa incerta* Kleine, R. Kleine det., 1936.

invisitata Kleine, 1935*invisitata* Kleine, 1935e: 305.**Type depositories.** NHM (LT!), DEI (PLT!).**Distribution.** Papua New Guinea (Bismarck Archipelago).

Notes. Döbler & Gaedike (1995: 448) reported that the specimen held in NHM is the holotype and that two paratypes are in DEI, but in his original description Kleine did not designate any of the three syntypes as holotype; however, as there is only one syntype in NHM, this must be considered as lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

malaisei Kleine, 1939*malaisei* Kleine, 1939d: 3.**Type depository.** NHRS (HT).**Distribution.** Myanmar [= Burma].*mikagei* Morimoto, 1976*mikagei* Morimoto, 1976a: 271.**Type depository.** ELKU (HT, PT).**Distribution.** Japan, Taiwan.*nitida* Kleine, 1918*nitida* Kleine, 1918h: 311.**Type depositories.** MNHUB (HT!), ZSM (ST).**Distribution.** India (Assam), Indonesia (Sumatra), Vietnam.*nupta* (Senna, 1892)*nupta* Senna, 1892a: 171 (*Miolispa*).**Type depositories.** MZUF (LT!, PLT!), RMNH (PLT!).

Distribution. Borneo, India (Assam), Indonesia (Batu, Java, Mentawai, Sumatra), Malaysia, Philippines (Luzon, Mindanao), Thailand.

Notes. Lectotype of *M. nupta* (MZUF) designated on a specimen with the following labels: J. D. Pasteur, Java Occid./ *M. nupta* Senna, cotypi!/ Not. Leyd. Mus., XIV, 1892, cotypi! ♂ ♂ ♀ ♀ / ex-coll. Senna/ "La Specola" Firenze, 7085. Paralectotypes: 14 specimens (MZUF), Java Occid., J. D. Pasteur, 1895/ ex-coll. Senna; 4 specimens (MZUF), J. D. Pasteur, 1895, Java Occid./ ex-coll. Senna; 1 specimen (RMNH), J. D. Pasteur, Java Occid./ Lectotype, R. Damoiseau det. [*in litteris*].

ocularis Kleine, 1928

ocularis Kleine, 1928d: 57.

Type depository. BPBM (HT).

Distribution. Laos, Philippines (Luzon).

opposita Kleine, 1922

opposita Kleine, 1922q: 149.

Type depository. SMTD (HT).

Distribution. Indonesia (Sumatra).

pasteuri (Senna, 1898)

pasteuri Senna, 1898e: 56 (*Miolispa*).

Type depositories. MZUF (LT!), RMNH (PLT).

Distribution. Borneo, Indonesia (Java, Sumatra).

Notes. Lectotype of *M. pasteuri* (MZUF) designated on a specimen with the following labels: Java Occ., J. D. Pasteur, ♂ / *Miolispa pasteuri* Senna, typus! ♂ ♀ / Not. Leyd. Mus., XX, 1898, typi! ♂ ♀ / Prépar. génit. N° 76 J 26, R. Damoiseau/ ex-coll. Senna/ "La Specola" Firenze, 6974.

reticulata Kleine, 1918

reticulata Kleine, 1918h: 333.

Type depositories. MNHUB (HT!), MZPW (PT).

Distribution. Borneo, Indonesia (Sumatra).

rufovittata (Perroud, 1853) n. comb.

rufovittatus Perroud, 1853: 419 (*Ceocephalus*).

Type depository. MNHN (HT!).

Distribution. Indonesia (Java).

rugosa Kleine, 1918

rugosa Kleine, 1918h: 336.

Type depository. MZPW (ST).

Distribution. Indonesia (Sumatra).

sponsa Kleine, 1918

sponsa Kleine, 1918h: 324.

Type depositories. MNHUB (LT!, PLT!), DEI (PLT!), MZPW (PLT), NHRS (PLT).

Distribution. Borneo, Indonesia (Java, Mentawai, Sumatra), Malaysia, Philippines (Mindanao, Samar).

Notes. Lectotype of *H. sponsa* (MNHUB) designated on a specimen with the following labels: Sumatra/ 111849/ Sumatra, Deli, Sibolangit, Jachan V./ cotypus/ *Hypomiolisa sponsa* Kleine. Paralectotypes: 2 specimens (MNHUB), N Borneo, Kina-Balu-Geb., Waterstradt S./ 11828/ cotypus/ *sponsa* Kleine; 1 specimen (MNHUB), N Borneo, Kina-Balu-Geb., Waterstradt S./ 11829/ cotypus/ *sponsa* Kleine; 1 specimen (MNHUB), Sunda Jne., ex coll. v. Studt G./ 111845/ cotypus/ *sponsa* Kleine; 1 specimen (MNHUB), Sumatra,

Deli, Sibolangit, Jachan V./ 111852/ *cotypus sponsa* Kleine; 1 specimen (DEI), N. O. Sumatra, Tebing-tinggi, Dr. Schultheiss/ Syntypus/ *Hypomiolispa sponsa* Kleine.

***sumatrana* (Senna, 1895)**

sumatrana Senna, 1895a: 190 (*Miolispa*).

Type depository. RMNH (HT!).

Distribution. Borneo, Indonesia (Java, Sumatra).

***tomentosa* Kleine, 1922**

tomentosa Kleine, 1922r: 156.

Type depository. SMTD (HT).

Distribution. Philippines (Mindanao).

***tonkinensis* Kleine, 1933**

tonkinensis Kleine, 1933d: 301.

Type depositories. SMTD (ST), unknown (ST).

Distribution. Vietnam.

***trachelizoides* (Senna, 1895)**

trachelizoides Senna, 1895a: 193 (*Miolispa*).

trachelizoides: Kleine, 1922r: 163 (*Hypomiolispa*) [lapsus].

trachelizoides: Kleine, 1930e: 26 (*Miolispa*) [lapsus].

Type depositories. MZUF (LT!), ZMAN (PLT!).

Distribution. Borneo, Indonesia (Batu, Java, Simeulue [= Simalur]; Sulawesi, Sumatra), Malaysia, Philippines (Mindanao, Palawan, Samar), Thailand.

Notes. Lectotype of *M. trachelizoides* (MZUF) designated on a male specimen with the following labels: Sumatra, ex dr. Faust/ *Miolispa trachelizoides* ♂ Senna, cotypus! / Not. Leyd. Mus., XVI, 1894, cotypus! ♂ / ex-coll. Senna/ "La Specola" Firenze, 6975. Paralectotype: 1 male specimen (ZMAN), Sumatra, Schagen v. Leeuwen, Deli.

***Hypotrachelizus* Kleine, 1933**

Hypotrachelizus Kleine, 1933e: 323.

Type species: *Hypotrachelizus mjobergi* Kleine, 1933.

Distribution. Borneo.

***mjobergi* Kleine, 1933**

mjobergi Kleine, 1933e: 324.

Type depository. NHRS (ST).

Distribution. Borneo

***Leptomiolispa* Kleine, 1933**

Leptomiolispa Kleine, 1933d: 300.

Type species: *Leptomiolispa aureliae* Kleine, 1933.

Distribution. Vietnam.

***aureliae* Kleine, 1933**

aureliae Kleine, 1933d: 301.

Type depository. SMTD (HT).

Distribution. Vietnam.

Miolispa Pascoe, 1862

Miolispa Pascoe, 1862: 393.

Cacotrachelus Sharp, 1900: 386; syn. Kleine, 1926b: 225.

Molispa: Kano, 1930: 152 [lapsus].

Miolipsia: Damoiseau, 1966: 425 [lapsus].

Miolipsa: Döbler & Gaedike, 1995: 439 [lapsus]

Type species: *Miolispa suturalis* Pascoe, 1862.

Distribution. Japan, Taiwan; Borneo, India (including Andaman Is.), Indonesia (Bali; Engano; Flores; Moluccas: Ambon, Aru, Bacan [= Batjan], Buru, Halmahera [= Gilolo], Kisar [= Kisser], Seram [= Ceram], Sula Is., Ternate; Java; Misool; Morotai; Sulawesi; Sumatra), Philippines (Basilan, Catanduanes, Leyte, Luzon, Mindanao, Mindoro, Negros, Panaon, Polillo, Samar, Sibuyan, Siargao, Tawi Tawi), Thailand; Australia, Fiji Is., New Caledonia, New Guinea (including Bismarck Archipelago: Duke of York, New Britain; Woodlark), Solomon Is., Vanuatu [= New Hebrides].

adversaria Kleine, 1922

adversaria Kleine, 1922f: 210.

Type depository. MZPW (ST).

Distribution. Borneo, India (including Andaman Is.).

aeneicollis Kleine, 1919

aeneicollis Kleine, 1919f: 294.

Type depositories. SMTD (HT, PT), DEI (PT!), MNHUB (PT!).

Distribution. Indonesia (Moluccas: Aru); New Guinea.

affinis Kleine, 1919

affinis Kleine, 1919f: 278.

Type depositories. SMTD (HT), ZMAN (AT!).

Distribution. Philippines (Mindanao); New Guinea, Solomon Is.

annae Kleine, 1919

annae Kleine, 1919f: 263.

Type depository. SMTD (HT).

Distribution. New Guinea.

apicipilosa Damoiseau, 1972

apicipilosa Damoiseau, 1972b: 272.

Type depository. HNHM (HT).

Distribution. New Guinea.

aruensis Kleine, 1919

aruensis Kleine, 1919f: 276.

Type depository. MNHUB (HT!).

Distribution. Indonesia (Moluccas: Aru; Misool); New Guinea, Solomon Is.

australiana Senna, 1897

australiana Senna, 1897a: 228.

Type depository. MZUF (HT!).

Distribution. Australia, New Guinea (?).

bicolor Kleine, 1919

bicolor Kleine, 1919f: 316.

Type depositories. SMTD (ST), MZPW (ST).

Distribution. Philippines (Luzon, Mindanao, Mindoro, Samar).

borneensis Senna, 1898

borneensis Senna, 1898e: 70.

Type depositories. MZUF (HT!), SMTD (PT).

Distribution. Borneo, Indonesia (Java), Philippines (Mindanao).

clavicornis Kleine, 1922

clavicornis Kleine, 1922b: 30.

Type depository. SMTD (ST).

Distribution. Philippines (Luzon, Mindanao, Sibuyan, Tawi Tawi).

coerulans Kleine, 1919

coerulans Kleine, 1919f: 273.

Type depository. MNHUB (HT!).

Distribution. New Guinea.

compressicornis Damoiseau, 1989

compressicornis Damoiseau, 1989b: 82.

Type depository. ZMAN (HT!).

Distribution. Solomon Is.

Notes. According to Damoiseau (1989b: 182) the type was in RMNH, but actually this specimen is held in ZMAN (B. Brugge, 2002, personal communication).

conjuncta Kleine, 1919

conjuncta Kleine, 1919f: 302.

Type depository. ZMUH (ST).

Distribution. New Guinea.

cordiformis* Macleay, 1887cordiformis* Macleay, 1887: 193.**Type depository.** AMS (HT!).**Distribution.** New Guinea.***crassifemoralis* Kleine, 1917***crassifemoralis* Kleine, 1917i: 292.**Type depository.** RMNH (HT!).**Distribution.** Taiwan; Borneo; New Guinea.***credula* Kleine, 1926***credula* Kleine, 1926f: 364.**Type depository.** NHM (HT!).**Distribution.** New Guinea.***cruciata* Senna, 1898***cruciata* Senna, 1898e: 69.**Type depository.** MZUF (HT!).**Distribution.** Japan, Taiwan; Borneo, Indonesia (Java, Sumatra), Philippines (Mindanao).***discors* Senna, 1895***discors* Senna, 1895f: 358.**Type depository.** MZUF (HT!).**Distribution.** Taiwan; Borneo, Indonesia (Sulawesi), Malaysia, Philippines (Mindanao).

***dissoluta* Kleine, 1926**

dissoluta Kleine, 1926a: 24.

Type depository. NHM (HT!).

Distribution. Borneo.

***dubia* Senna, 1893**

dubia Senna, 1893b: 273.

Type depository. MSNG (HT!).

Distribution. Indonesia (Engano, Sumatra).

***ebenina* Macleay, 1887**

ebenina Macleay, 1887: 194.

ebenina: Kleine, 1938d: 64 (*Miolispa*) [lapsus].

Type depository. AMS (HT).

Distribution. New Guinea.

***elongata* Kleine, 1919**

elongata Kleine, 1919f: 244.

Type depository. SMTD (ST).

Distribution. Borneo, Philippines (Basilan, Luzon, Mindanao, Negros, Panaon, Tawi Tawi).

***ephippium* Kleine, 1919**

ephippium Kleine, 1919f: 247.

Type depository. SMTD (HT).

Distribution. Philippines (Luzon).

fijiana Kleine, 1928

fijiana Kleine, 1928d: 55.

Type depository. BPBM (HT).

Distribution. Fiji Is.

flavolineata Kleine, 1919

flavolineata Kleine, 1919f: 282.

flavolineata: Kleine, 1921a: 17 [lapsus].

Type depository. SMTD (ST).

Distribution. Philippines (Basilan, Luzon, Mindanao).

flexilis Kleine, 1923

flexilis Kleine, 1923e: 161.

Type depository. SMTD (HT, PT).

Distribution. Philippines (Mindanao, Samar).

formosa Kleine, 1923

formosa Kleine, 1923e: 160.

Type depository. SMTD (HT).

Distribution. Philippines (Mindanao).

fornicata Kleine, 1923

fornicata Kleine, 1923e: 161.

Type depository. SMTD (HT).

Distribution. Indonesia (Sula Is.), Philippines (Leyte, Luzon, Mindanao).

fraudatrix Kleine, 1919

fraudatrix Kleine, 1919f: 249.

Type depository. SMTD (HT).

Distribution. Philippines (Luzon); New Guinea.

frivola Kleine, 1924

frivola Kleine, 1924b: 33.

Type depository. MNHUB (HT!).

Distribution. New Guinea.

fulgida Kleine, 1925

fulgida Kleine, 1925f: 23

Type depository. ZMAN (LT!, PLT!).

Distribution. Indonesia (Buru).

Notes. Lectotype of *M. fulgida* (ZMAN) designated on a male specimen with the following labels: L. J. Toxopeus, Buru Station 9, May 1921. Paralectotype: 1 female specimen (ZMAN), L. J. Toxopeus, Buru Station 9, 20.VI-10.VII.21.

fundata Kleine, 1925

fundata Kleine, 1925f: 20.

Type depository. ZMAN (LT!, PLT!).

Distribution. Indonesia (Buru).

Notes. Lectotype of *M. fundata* (ZMAN) designated on a male specimen with the following labels: L. J. Toxopeus, Buru Station 1, Oct.-Nov. 1921. Paralectotype: 1 female specimen (ZMAN), with the same label data as the lectotype.

***funnebris* Kleine, 1925**

funnebris Kleine, 1925f: 21.

Type depository. ZMAN (LT!, PLT!).

Distribution. Indonesia (Buru).

Notes. Lectotype of *M. funnebris* (ZMAN) designated on a male specimen with the following labels: L. J. Toxopeus, Buru Station 9, May 1921/ typus. Paralectotype: 1 female specimen (ZMAN), L. J. Toxopeus, Buru Station 9, 20.VI-10.VII.21/ paratypus.

***granulata* Kleine, 1919**

granulata Kleine, 1919f: 303.

Type depositories. ZSM (HT), MZPW (AT).

Distribution. Borneo, Indonesia (Java, Sumatra), Malaysia, Thailand.

***honesta* Kleine, 1926**

honesta Kleine, 1926f: 364.

Type depository. NHM (HT!).

Distribution. Borneo.

***honorata* Kleine, 1926**

honorata Kleine, 1926c: 163.

Type depository. NHM (HT!).

Distribution. Malaysia.

impunctata Kleine, 1919

impunctata Kleine, 1919f: 251.

Type depository. MNHUB (HT!).

Distribution. Malaysia.

intermedia Senna, 1897

intermedia Senna, 1897b: 239.

Type depository. MZUF (HT!).

Distribution. Borneo, Indonesia (Java; Moluccas: Ambon; Sulawesi), Philippines (Mindanao).

javanica Senna, 1892

javanica Senna, 1892a: 167.

javanus Sharp, 1900: 387 (*Cacotrachelus*); syn. Kleine, 1938d: 65.

Type depositories. *M. javanica*: MZUF (LT!, PLT!), RMNH (PLT); *C. javanus*: NHM (HT!).

Distribution. Indonesia (Java).

Notes. Lectotype of *M. javanica* (MZUF) designated on a male specimen with the following labels: Java Occ., J. D. Pasteur, 95 / *Miolispa javanica* Senna, cotypi! / Not. Leyd. Mus., XIV, 1892, cotypi! ♂ ♀ / ex -coll. Senna / "La Specola" Firenze, 6990. Paralectotypes: 17 specimens (MZUF), Java Occ., W. Pasteur, 95 / ex-coll. Senna / "La Specola" Firenze, 6990; 7 specimens (MZUF), Java Occ., W. Pasteur / ex-coll. Senna / "La Specola" Firenze, 6990.

jordani Senna, 1895

jordani Senna, 1895f: 360.

Type depository. MZUF (HT!).

Distribution. Malaysia.

lineata Senna, 1898

lineata Senna, 1898d: 57.

Type depositories. MZUF (LT!, PLT!), RMNH (PLT).

Distribution. Indonesia (Java), Malaysia, Philippines (Mindanao).

Notes. Lectotype of *M. lineata* (MZUF) designated on a male specimen with the following labels: Java Occ., W. Pasteur, 95 / *Miolispa lineata* Senna, typus! / Not. Leyd. Mus., XX, 1898, typus! ♂ / Prépar. génit. N° 76 J 15, R. Damoiseau/ ex-coll. Senna/ "La Specola" Firenze, 6985. Paralectotypes: 3 specimens (MZUF), Java Occ., W. Pasteur, 95/ ex-coll. Senna/ "La Specola" Firenze, 6985.

mariae Senna, 1891

mariae Senna, 1891: 165.

Type depository. MZUF (HT!).

Distribution. Indonesia (Sumatra), Malaysia, Thailand.

metallica Senna, 1892

metallica Senna, 1892a: 169.

integrivitta Senna, 1895f: 361 (*Miolispa metallica* var.); syn. Kleine, 1919f: 323.

impressa Senna, 1903: 172 (*Miolispa metallica* var.); syn. Kleine, 1919f: 323.

Type depositories. *M. metallica*: MZUF (LT!, PLT!), RMNH (PLT!), MSNG (?) (PLT), SMTD (PLT); *M. metallica* var. *integrivitta*: IRSNB (HT!); *M. metallica* var. *impressa*: MSNG (LT!, PLT!).

Distribution. Borneo, Indonesia (Java, Sumatra), Malaysia, Thailand.

Notes. Lectotype of *M. metallica* (MZUF) designated on a specimen with the following labels: Java Occ., W. Pasteur, 95 / *Miolispa metallica* Senna, cotypi! / Not. Leyden Mus., XIV, 1892, cotypi! ♂ ♀ / ex-coll. Senna/ "La Specola" Firenze, 6994. Paralectotypes: 25 specimens (MZUF), Java Occ., W. Pasteur, 95/ ex-coll. Senna/ "La Specola" Firenze, 6994; 1 specimen (RMNH), W. Pasteur, Java Occid.

According to Senna (1892a: 171) some syntypes of *M. metallica* from Borneo were in MSNG, but we did not find them there.

Lectotype of *M. metallica* var. *impressa* (MSNG) designated on a specimen with the following labels: Sumatra, Sirambè, XII.90-III.91, E.

Modigliani/ *metallica* Senna var. *impressa* Senna. Paralectotypes: 2 specimens (MSNG), Sumatra, Pangherang, Pisang, X.90 and III.91 E. Modigliani; 1 specimen (MSNG), Sumatra, Ajer Mantior, Agosto 1878, O. Beccari; 1 specimen (RMNH), J. D. Pasteur, Java Occid.

***nigricollis* Kleine, 1919**

nigricollis Kleine, 1919f: 224.

Type depositories. MZPW (ST), SMTD (ST).

Distribution. India (Andaman Is.), Indonesia (Mentawai, Sumatra).

Notes. Senna (1898a: 234) quoted this species as *M. nigricollis* Senna but gave no description. Kleine (1919f: 224) described it under the same name given by Senna [*in litteris*].

***nonfriedi* Senna, 1895**

nonfriedi Senna, 1895f: 363.

Type depository. MZUF (LT!, PLT!).

Distribution. New Guinea.

Notes. Lectotype designated by Bartolozzi *et al.* (1985: 99) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

***novaguineensis novaguineensis* (Guérin-Méneville, 1831)**

novaguineensis Guérin-Méneville, 1831: 109 (*Brenthus*).

puncticollis Boisduval, 1835b: 312 (*Brenthus*); syn. Gemminger & Harold, 1872: 2710.

antennata Senna, 1897a: 229 (*Miolispa novaguineensis* var.); syn. Schönfeldt, 1908: 25.

novaguineensis: Schönfeldt, 1908: 25 (*Miolispa*) [lapsus].

Type depositories. *B. novaguineensis*: MNHN (?) (HT); *B. puncticollis*: unknown; *M. novaguineensis* var. *antennata*: MZUF (HT!).

Distribution. Indonesia (Moluccas: Aru, Kisar [= Kisser]; Sulawesi), Malaysia; Australia, New Caledonia, New Guinea.

***novaguineensis pumilla* (Montrouzier, 1855)**

pumillus Montrouzier, 1855: 37 (*Cephalobarus*).

pumilus: Gemminger & Harrold, 1872: 2706 (*Trachelizus*) [lapsus].

pumilia: Kleine, 1930c: 285 (*Miolispa*) [lapsus].

Type depository. MNHN (LT!, PLT!).

Distribution. New Guinea (including Bismarck Archipelago: Duke of York, New Britain; Woodlark), Solomon Is.

Notes. Lectotype of *C. pumillus* (MNHN) designated on a specimen with the following labels: Museum Paris, Woodlark/ *pumilus* Mont. Woodlark/ type/ *Miolispa novaeguineensis pumila* (Montrouzier), R. Damoiseau det. 1978/ Prépar. Génit. N° 78 E 20, R. Damoiseau. Paralectotype: 1 specimen (MNHN), Museum Paris, Woodlark/ *Miolispa novaeguineensis pumila* (Montrouzier), R. Damoiseau det. 1978.

***occulta* Kleine, 1926**

occulta Kleine, 1926c: 162.

Type depository. NHM (HT!).

Distribution. Malaysia.

***papuana* Kleine, 1919**

papuana Kleine, 1919f: 274.

Type depositories. SMTD (HT), ZMAN (PT!).

Distribution. Indonesia (Bacan [= Batjan]); New Guinea (including Bismarck Archipelago), Solomon Is.

***pascoei* Kleine, 1919**

pascoei Kleine, 1919f: 226.

Type depository. SMTD (ST).

Distribution. Philippines (Luzon, Mindanao).

***paucicostata* Kleine, 1919**

paucicostata Kleine, 1919f: 312.

Type depository. SMTD (HT).

Distribution. Philippines (Luzon).

***persimilis* Kleine, 1922**

persimilis Kleine, 1922r: 154.

Type depository. SMTD (HT).

Distribution. Philippines (Mindanao).

***pirithorax* Damoiseau, 1966**

pirithorax Damoiseau, 1966e: 425.

Type depositories. ZMUC (HT, AT), IRSNB (PT!), RMNH (PT!).

Distribution. Borneo, Philippines (Tawi Tawi).

***polita* Shaw, 1953**

polita Shaw, 1953: 329.

Type depository. NHMB (HT!).

Distribution. Indonesia (Flores).

***poweri* Senna, 1895**

poweri Senna, 1895f: 364.

Type depository. IRSNB (HT!).

Distribution. Borneo, Indonesia (Java, Sumatra), Malaysia.

pulchella Kleine, 1922

pulchella Kleine, 1922b: 29.

pulchellea: Kleine, 1922r: 162 (*Miolispa*) [lapsus].

Type depository. SMTD (HT).

Distribution. Philippines (Luzon).

pulla Kleine, 1935

pulla Kleine, 1935b: 174.

Type depositories. SMTD (HT, PT), DEI (PT!).

Distribution. Solomon Is.

pygmaea Senna, 1895

pygmaea Senna, 1895f: 360.

pygmoea: Bartolozzi *et al.*, 1985: 99 (*Miolispa*) [lapsus].

Type depositories. MZUF (LT!, PLT!), IRSNB (PLT!), NHN (?) (PLT).

Distribution. Indonesia (Moluccas: Ambon, Halmahera [= Gilolo], Seram [= Ceram], Ternate), Malaysia; New Guinea (including Bismarck Archipelago: New Britain).

Notes. In his original description, Senna reported that the five specimens of the typical series were in IRSNB, MZUF and in the Tring Museum (now in NHM). We did not find the last two syntypes in NHM.

Lectotype of *M. pygmaea* (MZUF) designated on a male specimen with the following labels: Is. Penang, da Boucard, ♂ / *Miolispa pygmaea* Senna, cotypi ♂ ♀ / Ann. Soc. Ent. Belg., XXXIX, 1895, cotypi! ♂ ♀ / ex-coll. Senna/ "La Specola" Firenze, 6992. Paralectotypes: 1 female specimen (MZUF), Is. Penang, da Boucard, ♀ / ex-coll. Senna/ "La Specola" Firenze, 6992; 1 male specimen (IRSNB), Ceram/ Coll. Castelnau, Coll. Roelofs/ *Miolispa pygmaea* Senna cotypus ♂ R. Damoiseau rev. 1971/ paratype/ Prép. génit. No. 79 K 12 R. Damoiseau.

***robusta* Kleine, 1919**

robusta Kleine, 1919f: 230.

Type depository. SMTD (ST).

Distribution. Borneo, Indonesia (Moluccas: Ambon), Philippines (Basilan, Catanduanes, Luzon, Mindanao, Mindoro, Polillo, Samar, Siargao).

***rubrofemoralis* Kleine, 1941**

rubrofemoralis Kleine, 1941a: 130.

Type depository. MNHN (HT!).

Distribution. Vanuatu [= New Hebrides].

***salomonensis* Senna, 1894**

salomonensis Senna, 1894f: 11.

sculptipennis Sharp, 1900: 387 (*Cacotrachelus*); syn. Kleine, 1927a: 25.

Type depositories. *M. salomonensis*: MNHN (LT!, PLT!), SMTD (PLT); *C. sculptipennis*: CUMZ (ST).

Distribution. New Guinea (including Bismarck Archipelago: New Britain), Solomon Is.

Notes. Lectotype of *M. salomonensis* (MNHN) designated on a specimen with the following labels: Salomon/ ex Musaeo H. W. Bates 1892/ *Miolispa salomonensis* typus! Dr. Senna vidit 1893. Paralectotype: 1 specimen (MNHN), Salomon/ ex Musaeo H. W. Bates 1892/ Dr. Senna vidit 1893.

***simulans* Senna, 1895**

simulans Senna, 1895f: 362.

Type depository. MZUF (LT!, PLT!).

Distribution. Indonesia (Moluccas: Kisar [= Kisser], Morotai); New Guinea (including Bismarck Archipelago).

Notes. Lectotype designated by Bartolozzi *et al.* (1985: 101) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

***siporana* Senna, 1898**

siporana Senna, 1898a: 233.

sipora: Kleine, 1925f: 23 (*Miolispa*) [lapsus]

siporaba: Kleine, 1931h: 410 (*Miolispa*) [lapsus]

Type depository. MSNG (LT!, PLT!).

Distribution. Borneo, Indonesia (Java; Mentawai; Moluccas: Ambon; Sumatra), Malaysia, Philippines (Luzon, Mindanao).

Notes. Lectotype of *M. siporana* (MSNG) designated on a specimen with the following labels: Mentawai, Si Oban, IV-VIII.94, Modigliani/ *Miolispa siporana* n. cotypi! Senna. Paralectotypes: 7 specimens (MSNG), Mentawai, Si Oban, IV-VIII.94, Modigliani.

***sororia* Kleine, 1919**

sororia Kleine, 1919f: 257.

Type depositories. SMTD (HT), MNHUB (PT).

Distribution. Indonesia (Moluccas: Kisar [= Kisser]); New Guinea (including Bismarck Archipelago).

***splendida* Kleine, 1917**

splendida Kleine, 1917i: 289.

Type depository. RMNH (HT!).

Distribution. New Guinea (including Bismarck Archipelago).

***strand* Kleine, 1919**

strand Kleine, 1919f: 297.

Type depositories. DEI (LT!), MNHUB (PLT!), SMTD (PLT).

Distribution. Philippines (Luzon); Papua New Guinea (Bismarck Archipelago: New Britain), Solomon Is.

Notes. Lectotype designated by Döbler & Gaedike (1995: 454) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

sulcicollis Kleine, 1919

sulcicollis Kleine, 1919f: 290.

Type depository. SMTD (HT).

Distribution. New Guinea.

suturalis Pascoe, 1862

suturalis Pascoe, 1862: 393.

striata Senna, 1895f: 359 (*Miolispa suturalis* ssp.); syn. Kleine, 1919f: 324.

Type depositories. *M. suturalis*: NHM (HT!); *M. suturalis* ssp. *striata*: IRSNB (HT!).

Distribution. India (Andamans Is.), Indonesia (Bali; Java; Moluccas: Ambon, Bacan [= Batjan], Halmahera; Sumatra), Malaysia.

sycophanta Senna, 1893

sycophanta Senna, 1893b: 270.

Type depositories. MZUF (LT!), MSNG (PLT!).

Distribution. Indonesia (Engano, Sumatra).

Notes. Lectotype designated by Bartolozzi *et al.* (1985: 101) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

***testacea* Kleine, 1919**

testacea Kleine, 1919f: 261.

Type depository. SMTD (HT).

Distribution. Indonesia (Java; Moluccas: Ambon; Sulawesi); New Guinea.

***tuberculatipennis* Senna, 1897**

tuberculatipennis Senna, 1897b: 237.

Type depositories. MZUF (LT!, PLT!), IRSNB (PLT!), MNHN (PLT!), SMTD (PLT).

Distribution. Indonesia (Sulawesi).

Notes. Lectotype designated by Bartolozzi *et al.* (1985: 101) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

***unicolor* Kleine, 1919**

unicolor Kleine, 1919f: 314.

Type depository. SMTD (HT).

Distribution. Philippines (Luzon).

***unilineata* Kleine, 1944**

unilineata Kleine, 1944: 151.

Type depository. MZPW (HT, PT).

Distribution. New Guinea.

***variabilis* Damoiseau, 1989**

variabilis Damoiseau, 1989b: 83.

Type depositories. NMNH (HT, AT), IRSNB (PT!), SAMA (?) (PT).

Distribution. New Guinea.

Notes. Damoiseau (1972b: 274) quoted this species *in litteris* and stated that one paratype was in HNHM, but in his subsequent description (Damoiseau, 1989b: 84) he did not mention this paratype.

Miolispoides Senna, 1894

Miolispoides Senna, 1894b: 368.

Type species: *Miolispoides birmanicus* Senna, 1894.

Distribution. S India, Laos, Myanmar [= Burma], Vietnam.

birmanicus Senna, 1894

birmanicus Senna, 1894b: 369.

Type depositories. MZUF (LT!), MNHN (PLT).

Distribution. Myanmar [= Burma].

Notes. Lectotype designated by Bartolozzi *et al.* (1985: 94) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

According to Senna (1894b: 369) some syntypes were in Oberthür's collection (now in MNHN and IRSNB), but we did not find them there.

indicus Damoiseau, 1989

indicus Damoiseau, 1989b: 85.

Type depositories. NHM (HT, AT, PT!), IRSNB (?) (PT).

Distribution. S India.

Notes. Damoiseau (1989b: 86) reported that holotype and the allotype were in NHM, and a paratype in IRSNB, but we did not find them there.

sculpturatus Calabresi, 1921*sculpturatus* Calabresi, 1921a: 5.

Type depositories. MSNG (LT!, PLT!), IRSNB (PLT!), Saigon Museum (?) (PLT).

Distribution. Laos, Vietnam.

Notes. Lectotype of *M. sculpturatus* (MSNG) designated on a specimen with the following labels: Tongla, 30.IV.918 (Laos)/ *Miolispoides sculpturatus* ♂ typus! Calabr. Paralectotypes: 3 specimens (MSNG), Tongla, 30.IV.918 (Laos); 1 male specimen (IRSNB), Tong-La, 30.IV.918/ Vitalis de Salvaza/ R. I. Sc. N. B., I. G. 12.457/ Type/ *Miolispoides sculpturatus* n. sp. typus! ♂.

Paratrachelizus Kleine, 1921*Paratrachelizus* Kleine, 1921h: 32.

Type species: *Brentus ferrugineus* Lund, 1800.

Distribution. U.S.A. (Florida); Belize, Costa Rica, Cuba, El Salvador, Guatemala, Mexico, Nicaragua, Panama, Puerto Rico; Argentina, Bolivia, Brazil, Colombia, Ecuador, Peru, Uruguay.

adustus (Boheman, 1840)*adustus* Boheman (in Schoenherr, 1840: 497) (*Trachelizus*).

Type depositories. NHRS (ST), MLUH (ST).

Distribution. Brazil.

advena (Sharp, 1895)*advena* Sharp, 1895a: 20 (*Trachelizus*).

Type depository. NHM (LT!, PLT!).

Distribution. Panama; Brazil.

Notes. Lectotype of *T. advena* (NHM) designated on a specimen with the following labels: Syntype/ Type/ Caldera, 1200 ft., Champion/ Sharp Coll.

1905-313/ B.C.A., Col. IV, 6. *Trachelizus advena* Sharp. Paralectotypes: 4 specimens (NHM), with the same label data as the lectotype; 3 specimens (NHM), Syntype/ V. de Chiriqui, 25-4000 ft., Champion/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus advena* Sharp/ *Paratrachelizus advena* (Sharp), Scivittaro ex. 1979.

***afflictus* Kleine, 1922**

afflictus Kleine, 1922f: 209.

Type depository. MZPW (ST).

Distribution. Brazil.

***agnatus* Kleine, 1922**

agnatus Kleine, 1922g: 223.

Type depository. DEI (HT!).

Distribution. Costa Rica; Colombia, Peru.

Notes. Döbler & Gaedike (1995: 454) reported that there is one syntype in DEI, but the species seems to have been described on a single female specimen.

***arduus* (Sharp, 1895)**

arduus Sharp, 1895a: 29 (*Trachelizus*).

Type depository. NHM (LT!, PLT!).

Distribution. Costa Rica, Guatemala.

Notes. Lectotype of *T. arduus* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ Capetillo, Guatemala, Champion/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus arduus* Sharp/ *Paratrachelizus arduus* (Sharp) Scivittaro det. 1981. Paralectotypes: 1 female specimen (NHM), with the same label data as the lectotype; 1 female specimen (NHM), Syntype/ S. Geronimo, Guatemala, Champion/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus arduus* Sharp; 1 female specimen (NHM), Syntype/ Senahu, Vera Paz, Champion/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus arduus* Sharp.

***aureopilosus* (Senna, 1890)**

aureopilosus Senna, 1890: 105 (*Trachelizus*).

hirtellus Sharp, 1895a: 23 (*Trachelizus*); **syn. n.**

Type depositories. *T. aureopilosus*: MZUF (HT!); *T. hirtellus*: NHM (LT!, PLT!).

Distribution. Costa Rica, Nicaragua, Panama; Bolivia, Brazil, Colombia.

Notes. Lectotype of *T. hirtellus* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ Chontales, Nicaragua, T. Belt/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus hirtellus* Sharp/ Prépar. génit. No. BM 65 R. Damoiseau. Paralectotypes: 2 male and 3 female specimens (NHM), Syntype/ Chontales, Nicaragua, T. Belt/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus hirtellus* Sharp (1 male and 1 female identified as *Paratrachelizus aureopilosus* Senna by Scivittaro, 1979); 2 male and 3 female specimens (NHM), Syntype/ V. de Chiriqui, 25-4000 ft., Champion/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus hirtellus* Sharp (1 male identified as *Paratrachelizus aureopilosus* Senna by Scivittaro, 1979); 1 male specimen (NHM), Syntype/ Nicaragua/ 2509/ Salle Coll./ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus hirtellus* Sharp.

After studying the type material, we consider *T. hirtellus* to be a synonym for *T. aureopilosus*.

***clavicornis* (Boheman, 1840)**

clavicornis Boheman (in Schoenherr, 1840: 492) (*Trachelizus*).

Type depository. NHRS (HT).

Distribution. Brazil.

Notes. Damoiseau (1989b: 89) reported that the holotype and the allotype are in NHRS but the species seems to have been described on a single specimen.

***cognatus* (Sharp, 1895)**

cognatus Sharp, 1895a: 22 (*Trachelizus*).

Type depository. NHM (LT!, PLT!).

Distribution. Panama; Peru.

Notes. Lectotype of *T. cognatus* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ V. de Chiriqui/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus cognatus* Sharp. Paralectotypes: 1 female specimen (NHM), with the same label data as the lectotype; 16 specimens (NHM), Syntype/ V. de Chiriqui, 25-4000 ft., Champion/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus cognatus* Sharp (one specimen labelled as *Paratrachelizus dispar* by Scivittaro, 1979).

dispar (Sharp, 1895)

dispar Sharp, 1895a: 21 (*Trachelizus*).

Type depository. NHM (LT!, PLT!).

Distribution. Costa Rica, Panama; Brazil.

Notes. Lectotype of *T. dispar* (NHM) designated on a male specimen with the following labels: Syntype/ V. de Chiriqui/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus dispar* Sharp/ Prépar. génit. No BM 63 R. Damoiseau. Paralectotypes: 1 female specimen (NHM), with the same label data as the lectotype; 28 specimens (NHM), Syntype/ V. de Chiriqui, 3-4000 ft., Champion/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus dispar* Sharp.

dorsalis (Boheman, 1840)

dorsalis Boheman (in Schoenherr, 1840: 499) (*Trachelizus*).

Type depository. NHRS (HT).

Distribution. El Salvador, Mexico; Argentina, Brazil, Colombia.

ducalis (Sharp, 1895)

ducalis Sharp, 1895a: 31 (*Trachelizus*).

Type depository. NHM (HT!).

Distribution. Guatemala; Argentina, Bolivia, Uruguay.

elevatus (Sharp, 1895)

elevatus Sharp, 1895a: 22 (*Trachelizus*).

Type depository. NHM (LT!, PLT!).

Distribution. Guatemala, Panama.

Notes. Lectotype of *T. elevatus* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ V. de Chiriqui/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus elevatus* Sharp. Paralectotypes: 1 female specimen (NHM), with the same label data as the lectotype; 10 specimens (NHM), Syntype/ V. de Chiriqui, 25-4000 ft., Champion/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus elevatus* Sharp; 1 specimen (NHM), Syntype/ 2511/ Izabal/ Sharp Coll. 1905-313/ Salle Coll./ B.C.A., Col. IV, 6. *Trachelizus elevatus* Sharp.

ferrugineus Lund, 1800

ferrugineus Lund, 1800: 68 (*Brentus*).

Type depository. ZMUC (HT).

Distribution. Brazil.

filiformis (Sharp, 1895)

filiformis Sharp, 1895a: 29 (*Trachelizus*).

Type depository. NHM (LT!, PLT!).

Distribution. El Salvador, Mexico.

Notes. Lectotype of *T. filiformis* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ Cerro de Plumas, Mexico, Hoege/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus filiformis* Sharp. Paralectotypes: 15 specimens (NHM), with the same label data as the lectotype [3 specimens labelled as *Paratrachelizus occlusus* (Sharp) by Scivittaro, 1981]; 1 female specimen (NHM), Syntype/ Bobo, Mexico, Hoege/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus filiformis* Sharp.

fracticornis (Sharp, 1895)

fracticornis Sharp, 1895a: 28 (*Trachelizus*).

Type depository. NHM (LT!, PLT!).

Distribution. Belize, Guatemala, Mexico, Nicaragua, Panama; Colombia, Ecuador.

Notes. Lectotype of *P. fracticornis* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ Las Mercedes, 3000 ft., Champion/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus fracticornis* Sharp. Paralectotypes: 6 specimens (NHM), with the same label data as the lectotype; 2 specimens (NHM), Syntype/ Cordova/ Mexico, Salle Coll./ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus fracticornis* Sharp; 2 specimens (NHM), Syntype/ Belize, Blancaneaux/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus fracticornis* Sharp; 3 specimens (NHM), Syntype/ Chontales, Janson/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus fracticornis* Sharp; 1 specimen (NHM), Syntype/ Chontales, Nicaragua, T. Belt/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus fracticornis* Sharp; 1 specimen (NHM), Syntype/ V. de Chiriqui, Champion/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus fracticornis* Sharp; 1 specimen (NHM), Syntype/ Cerro Zunil, 4-5000 ft., Champion/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus fracticornis* Sharp.

frontalis (Sharp, 1895)

frontalis Sharp, 1895a: 25 (*Trachelizus*).

Type depository. NHM (LT!, PLT!).

Distribution. Nicaragua, Panama.

Notes. Lectotype of *T. frontalis* (NHM) designated on a female specimen with the following labels: Syntype/ Type/ Chontales, Nicaragua, Janson/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus frontalis* Sharp. Paralectotype: 1 female specimen (NHM), Syntype/ V. de Chiriqui, 25-4000 ft., Champion/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus frontalis* Sharp/ *Paratrachelizus frontalis* (Sharp) Scivittaro det. 1979.

***geminatus* (Sharp, 1895)**

geminatus Sharp, 1895a: 30 (*Trachelizus*).

Type depository. NHM (HT!).

Distribution. Mexico.

***laticollis* (Sharp, 1895)**

laticollis Sharp, 1895a: 31 (*Trachelizus*).

Type depository. NHM (HT!).

Distribution. Costa Rica, Guatemala.

***linearis* (Suffrian, 1870)**

linearis Suffrian, 1870: 218 (*Trachelizus*).

Type depository. MNHC (ST).

Distribution. Cuba, Panama, Puerto Rico.

***lineatus* (Sharp, 1895)**

lineatus Sharp, 1895a: 22 (*Trachelizus*).

Type depository. NHM (LT!, PLT!).

Distribution. Panama.

Notes. Lectotype of *T. lineatus* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ V. de Chiriqui/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus lineatus* Sharp/ Prépar. génit. No BM 64 R. Damoiseau. Paralectotypes: 1 female specimen (NHM), with the same label data as the lectotype; 14 specimens (NHM), Syntype/ V. de Chiriqui, 25-4000 ft., Champion/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus lineatus* Sharp.

***nigricornis* (Sharp, 1895)**

nigricornis Sharp, 1895a: 26 (*Trachelizus*).

Type depository. NHM (LT!, PLT!).

Distribution. Costa Rica, Guatemala.

Notes. Lectotype of *T. nigricornis* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ Zapote, Guatemala, Champion/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus nigricornis* Sharp/ Prépar. génit. No. BM 67, R. Damoiseau/ *Paratrachelizus nigricornis* (Sharp) Scivittaro det. 1981. Paralectotypes: 1 female specimen (NHM), Syntype/ Cerro Zunil, 4000 ft., Champion/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus nigricornis* Sharp; 1 female specimen (NHM), Syntype/ V. de Atitlan, 25-3500 ft., Champion/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus nigricornis* Sharp.

***occlusus* (Sharp, 1895)**

occlusus Sharp, 1895a: 27 (*Trachelizus*).

Type depository. NHM (LT!, PLT!).

Distribution. Mexico.

Notes. Lectotype of *T. occlusus* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ Catemaco/ Mexico, Salle Coll./ Sharp Coll. 1905-313/ Prépar. génit. No. BM 68, R. Damoiseau/ B.C.A., Col. IV, 6. *Trachelizus occlusus* Sharp/ *Paratrachelizus occlusus* (Sharp) Scivittaro det. 1981. Paralectotypes: 1 male specimen (NHM), Syntype/ Las Vigas, Mexico, Hoegel/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus occlusus* Sharp; 1 female specimen (NHM), Syntype/ Catemaco, Mexico, Salle Coll./ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus occlusus* Sharp.

***optatus* (Sharp, 1895)**

optatus Sharp, 1895a: 30 (*Trachelizus*).

Type depository. NHM (HT!).

Distribution. Costa Rica, Guatemala.

***prolixus* (Sharp, 1895)**

prolixus Sharp, 1895a: 31 (*Trachelizus*).

Type depository. NHM (LT!, PLT!).

Distribution. Belize, Costa Rica, Guatemala, Mexico, Nicaragua.

Notes. Lectotype of *P. prolixus* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ Cerro de Plumas, Hoege/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus prolixus* Sharp. Paralectotypes: 2 female specimens, with the same label data as the lectotype; 1 female specimen (NHM), Syntype/ Cerro Zunil, 4000 ft., Champion/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus prolixus* Sharp; 1 female specimen (NHM), Syntype/ Belize, Blancaneaux/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus prolixus* Sharp; 1 male specimen (NHM), Syntype/ Chontales, Nicaragua, T. Belt/ Sharp Coll. 1905-313; 1 male specimen (NHM), Syntype/ Mexico, Salle Coll./ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus prolixus* Sharp/ *Paratrachelizus adustus* (Boh.) Scivittaro det. 1981.

***punctatus* Soares & Meyer, 1959**

punctatus Soares & Meyer, 1959: 165.

Type depository. MNRJ (Campos Seabra coll.) (HT, AT).

Distribution. Brazil.

***robustus* (Sharp, 1895)**

robustus Sharp, 1895a: 26 (*Trachelizus*).

Type depository. NHM (LT!, PLT!).

Distribution. Belize, Nicaragua, Panama.

Notes. Lectotype of *T. robustus* (NHM) designated on a male specimen with the following labels: Syntype/ David, Panama, Champion/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus robustus* Sharp. Paralectotypes: 1 female specimen (NHM), Syntype/ Chontales, Nicaragua, T. Belt/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus robustus* Sharp/ *Paratrachelizus robustus* (Sharp) Scivittaro det. 1979; 1 female specimen (NHM), Syntype/ Type/ Belize, Blancaneaux/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus robustus* Sharp.

seriatus (Sharp, 1895)

seriatus Sharp, 1895a: 24 (*Trachelizus*).

Type depository. NHM (HT!).

Distribution. Mexico.

serratus (Sharp, 1895)

serratus Sharp, 1895a: 24 (*Trachelizus*).

Type depository. NHM (LT!, PLT!).

Distribution. Mexico.

Notes. Lectotype of *T. serratus* (NHM) designated on a female specimen with the following labels: Syntype/ Type/ Cerro de Plumas, Mexico, Hoege/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus serratus* Sharp/ *Paratrachelizus seriatus* (Sharp) Scivittaro det. 1981. Paralectotypes: 2 female specimens (NHM), with the same label data as the lectotype.

simplex (Suffrian, 1870)

simplex Suffrian, 1870: 219 (*Trachelizus*).

Type depository. MNHC (ST).

Distribution. Costa Rica, Cuba, Guatemala.

sternalis (Sharp, 1895)

sternalis Sharp, 1895a: 25 (*Trachelizus*).

sternalis: Haedo Rossi, 1961: 39 (*Paratrachelizus*) [lapsus].

Type depository. NHM (LT!, PLT!).

Distribution. Guatemala, Mexico, Nicaragua, Panama; Argentina, Brazil, Paraguay.

Notes. Lectotype of *T. sternalis* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ Zapote, Guatemala, Champion/

Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus sternalis* Sharp. Paralectotypes: 7 specimens (NHM), with the same label data as the lectotype; 4 specimens (NHM), Syntype/ Chontales, Janson/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus sternalis* Sharp; 2 specimens (NHM), Syntype/ Cerro de Plumas, Mexico, Hoege/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus sternalis* Sharp; 1 specimen (NHM), Syntype/ Caldera, 1200 ft., Champion/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus sternalis* Sharp; 1 specimen (NHM), Syntype/ Atoyac, Vera Cruz, April, H. H. S./ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus sternalis* Sharp; 1 specimen (NHM), Syntype/ S. Geronimo, Guatemala, Champion/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus sternalis* Sharp; 1 specimen (NHM), Syntype/ V. de Chiriqui, 25-4000 ft., Champion/ Sharp Coll. 1905-313/ B.C.A., Col. IV, 6. *Trachelizus sternalis* Sharp.

***sulcirostris* (Boheman, 1833)**

sulcirostris Boheman (in Schoenherr, 1833: 354) (*Brenthus*).

Type depository. NHRS (HT).

Distribution. Brazil.

***tenuis* (Suffrian, 1870)**

tenuis Suffrian, 1870: 216 (*Trachelizus*).

Type depository. MNHC (ST).

Distribution. Cuba.

***turgidirostris* (Boheman, 1840)**

turgidirostris Boheman (in Schoenherr, 1840: 494) (*Trachelizus*).

Type depositories. MNHN (ST!), NHRS (ST).

Distribution. Guatemala, Mexico, Nicaragua, Panama; Argentina, Peru.

uncimanus (Boheman, 1840)

uncimanus Boheman (in Schoenherr, 1840: 496) (*Trachelizus*).

miamana: Beyer, 1904: 168 (*Trachelizus*) [lapsus].

uncimannus: Poole & Gentili, 1996: 62 (*Trachelizus*) [lapsus].

Type depositories. MNHN (ST!), NHRS (ST).

Distribution. USA (Florida: Keys Is.); Cuba.

ventralis Haedo Rossi, 1954

ventralis Haedo Rossi, 1954a: 2.

Type depositories. JHRC (HT), APC (AT).

Distribution. Argentina, Uruguay.

Peritrachelizus Kleine, 1922

Peritrachelizus Kleine, 1922f: 205.

Type species: *Peritrachelizus columbiensis* Kleine, 1922.

Distribution. Colombia.

columbiensis Kleine, 1922

columbiensis Kleine, 1922f: 206.

Type depository. SMTD (ST).

Distribution. Colombia.

Perroudia Damoiseau, 1962

Perroudia Damoiseau, 1962c: 286.

Type species: *Trachelizus victoris* Perroud & Montrouzier, 1865.

Distribution. New Caledonia.

victoris (Perroud & Montrouzier, 1865)

victoris Perroud & Montrouzier, 1865: 137 (*Trachelizus*).

Type depository. IRSNB (LT!, PLT!).

Distribution. New Caledonia.

Notes. Lectotype designated by Damoiseau (1966a: 25).

Alonso-Zarazaga & Lyal (1999: 53 and 248) only give Perroud as author of the species, but we believe that Perroud and Montrouzier should be considered as co-authors.

Schizephebocerus Kleine, 1923

Schizephebocerus Kleine, 1923c: 127.

Type species: *Schizephebocerus ambitiosus* Kleine, 1923.

Distribution. Borneo, Malaysia.

Notes. Kleine (1922m: 162) mentioned this genus in a list of genera, but he did not describe it until one year later (1923c: 127).

ambitiosus Kleine, 1923

ambitiosus Kleine, 1923c: 128.

Type depository. NHM (HT!).

Distribution. Borneo, Malaysia.

Stenobrentus Damoiseau, 1966

Stenobrentus Damoiseau, 1966c: 147.

Type species: *Stenobrentus lineatus* Damoiseau, 1966.

Distribution. Australia.

lineatus Damoiseau, 1966

lineatus Damoiseau, 1966c: 148.

Type depository. SAMA (HT).

Distribution. Australia.

Trachelizus Dejean, 1834

Trachelizus Dejean, 1834: 243.

Trachelisus: Kabakov, 2001: 207 [lapsus].

Type species: *Brentus bisulcatus* Lund, 1800.

Distribution. S China, Japan, Taiwan; Borneo, India (including Assam, Darjeeling), Indonesia (Bali; Engano, Flores; Java; Moluccas: Amboin, Aru, Bacan [= Batjan], Buru, Halmahera [= Gilolo], Larat; Mentawai; Sulawesi; Sumatra; Sumba), Kampuchea [= Cambodia], Laos, Malaysia, Myanmar [= Burma], Philippines (Basilan, Leyte, Luzon, Mindanao, Samar), Sri Lanka, Thailand, Vietnam; Australia, New Guinea (including Bismarck Archipelago), Solomon Is.

Notes. Senna (1898a: 230) quoted the species *T. rostralis* Senna claiming that it was published in the *Bullettino della Società Entomologica Italiana*, Vol. XXX; but actually this species has never been described.

bisulcatus (Lund, 1800)

bisulcatus Lund, 1800: 67 (*Brentus*).

bicanaliculata Schaufuss, 1885: 206 (*Miolispa*); syn. Senna, 1899: 300.

semivelata Schaufuss, 1885: 206 (*Miolispa*); syn. Senna, 1899: 300.

striata Schaufuss, 1885: 206 (*Miolispa*); syn. Zimmerman, 1994: 209.

exophthalmus Lea, 1899: 636 (*Ceocephalus*); syn. Kleine, 1927a: 22.

bisulcatus: Swezey, 1928: 62 (*Trachelizus*) [lapsus].

Type depositories. *B. bisulcatus*: ZMUC (ST); *M. bicanaliculata*: NMPC (Salvator coll.) (?) (HT); *M. semivelata*: MNHUB (LT!, PLT!), NMPC (Salvator coll.) (PLT); *M. striata*: NMPC (Salvator coll.) (?) (ST); *C. exophthalmus*: SAMA (Lea coll.) (ST), MMUS (ST).

Distribution. S China (Hainan), Japan, Taiwan; India (including Assam), Indonesia (Bali; Flores; Java; Moluccas: Amboin, Aru, Bacan [= Batjan], Bu-

ru, Halmahera [= Gilolo], Larat, Mentawai; Sulawesi; Sumatra; Sumba), Laos, Malaysia, Myanmar [= Burma], Philippines (Basilan, Leyte, Luzon, Mindanao, Samar, Tagoloan), Sri Lanka, Thailand, Vietnam; Australia, New Guinea (including Bismarck Archipelago), Solomon Is.

Notes. According to Dr J. Jelinek (NMPC) (2002, personal communication) the types for *M. bicanaliculata* and *M. striata* are not in NMPC; we were unable to find these types in MNHUB either, where other Schaufuss' types are held.

Lectotype of *M. semivelata* (MNHUB) designated on a specimen with the following labels: Type/ *Miolispa semivelata* Schauf., Macassar. Paralectotypes: 2 specimens (MNHUB), Coll. L. W. Schaufuss/ 111900/ Typus.

confinis Kleine, 1926

confinis Kleine, 1926a: 21.

Type depository. NHM (HT!).

Distribution. Borneo.

despiratus Kleine, 1926

despiratus Kleine, 1926a: 22.

Type depository. NHM (HT!).

Distribution. Indonesia ("Moluccas").

dividuus Kleine, 1925

dividuus Kleine, 1925b: 131.

Type depository. NHM (HT!).

Distribution. India (including Assam, Darjeeling), Vietnam.

dohertyi Senna, 1894

dohertyi Senna, 1894b: 366.

Type depository. MNHN (?) (ST).

Distribution. Myanmar [= Burma].

Notes. According to Senna (1894b: 367) the syntypes were in Oberthür's collection (now in MNHN and IRSNB), but we did not find them there.

insularis Senna, 1892

insularis Senna, 1892a: 163.

Type depositories. MZUF (LT!), RMNH (PLT!), SMTD (PLT).

Distribution. Indonesia (Java, Sumatra), Malaysia.

Notes. Lectotype designated by Bartolozzi *et al.* (1985: 97) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

japonicus Morimoto, 1976

japonicus Morimoto, 1976a: 272.

Type depository. ELKU (HT, PT).

Distribution. Japan.

laevigatus Senna, 1898

laevigatus Senna, 1898d: 58.

Type depository. MZUF (LT!).

Distribution. Borneo, Indonesia (Java, Sumatra), Malaysia, Myanmar [= Burma], Vietnam.

Notes. Lectotype designated by Bartolozzi *et al.* (1985: 98) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

***makiharai* Morimoto, 1976**

makiharai Morimoto, 1976a: 274.

Type depository. ELKU (HT).

Distribution. Japan.

Notes. Morimoto (1976a) described the species on seven specimens, but he did not specify where the six paratypes are held. According to H. Kojima (ELKU) (2002, personal communication) they are not in ELKU.

***modestus* Senna, 1892**

modestus Senna, 1892a: 165.

Type depositories. RMNH (LT!), SMTD (PLT).

Distribution. Indonesia (Batu, Engano, Java, Nias, Sumatra).

Notes. Lectotype of *T. modestus* (RMNH) designated on a specimen with the following labels: Muller, Java/ *Hypomiolispa insularis* R. Damoiseau det.

***moestus* Senna, 1892**

moestus Senna, 1892a: 165.

Type depository. RMNH (HT).

Distribution. Indonesia (Java, Sumatra), Malaysia.

***peregrinus* Senna, 1898**

peregrinus Senna, 1898a: 231.

Type depository. MSNG (HT!).

Distribution. Sumatra (Mentawai).

***politus* Senna, 1892**

politus Senna, 1892e: 462.

Type depository. MSNG (HT!).

Distribution. Borneo, Kampuchea [= Cambodia], India, Malaysia, Myanmar [= Burma], Vietnam.

quaesitus Kleine, 1935

quaesitus Kleine, 1935e: 304.

Type depositories. NHM (ST!), DEI (ST!), unknown (ST).

Distribution. New Guinea.

Notes. Döbler & Gaedike (1995: 452) reported that species description was based on six specimens, the holotype and five paratypes, but in his original description Kleine did not designate any of the six syntypes as holotype.

rufomaculatus Senna, 1892

rufomaculatus Senna, 1892e: 460.

Type depository. MSNG (HT!).

Distribution. Myanmar [= Burma].

Tracheloschizus Damoiseau, 1966

Tracheloschizus Damoiseau, 1966d: 21.

Type species: *Schizotrachelus dichrous* Lacordaire, 1866.

Distribution. India (Andaman Is.), Indonesia (Flores; Java; Moluccas: Ambon, Aru, Buru, Kay, Halmahera [= Gilolo], Larat; Sulawesi, Sumatra, Timor), Malaysia, Myanmar [= Burma], Philippines (Basilan, Camiguin, Catanduanes, Leyte, Luzon, Mindanao, Negros, Panaon, Polillo, Samar, San Miguel, Siargao, Sibuyan, Tawi Tawi), Thailand, Vietnam; Australia, Fiji Is., New Caledonia, New Guinea (including Bismarck Archipelago: New Britain; Woodlark), Solomon Is., Vanuatu [= New Hebrides].

Notes. Damoiseau described this genus twice in the same year (1966d: 21; 1966a: 19); the first paper was published on 1st July, the second on 26th October.

The systematics of the species included in this genus needs accurate revi-

sion, based on the study of the genital structures. Our classification must be considered as provisional.

***alienus* (Kleine, 1922) n. comb.**

alienus Kleine, 1922g: 226 (*Schizotrachelus*).

Type depository. DEI (LT!, PLT!).

Distribution. Indonesia (Moluccas: Aru, Kay).

Notes. Lectotype of *S. alienus* (DEI) designated on a male specimen with the following labels: Aru-Insel, Wamina Dobbo, C. Ribbe, 1883/ Syntypus/ Kleine det./ *Schizotrachelus alienus* Kleine, det. Aut., 1921. Paralectotype: 1 female specimen (DEI) with the same label data as the lectotype.

We include the species in this genus on the basis of the external morphology of the type material.

***atilis* (Kleine, 1922)**

atilis Kleine, 1922g: 227 (*Schizotrachelus*).

Type depository. DEI (LT!, PLT!).

Distribution. Australia, New Guinea, Solomon Is.

Notes. Lectotype of *S. atilis* (DEI) designated on a female specimen with the following labels: Neuguinea/ Syntype/ coll. Kraatz/ *Schizotrachelus atilis* det. aut. 1921. Paralectotype: 1 female specimen (DEI), Coen District, Cape York, Queensland, H. Hacker/ Coll. Hacker/ Kleine det./ Syntype.

***anguliceps* (Senna, 1899)**

anguliceps Senna, 1899: 308 (*Schizotrachelus*).

Type depository. NHMB (LT!, PLT!).

Distribution. Borneo, Indonesia (Sulawesi), Malaysia, Philippines (Basilan, Leyte, Luzon, Negros, San Miguel); Australia.

Notes. Lectotype designated by Damoiseau (1989b: 90).

***bakeri* (Kleine, 1922) n. comb.**

bakeri Kleine, 1922b: 33 (*Schizotrachelus*).

concolor Kleine, 1922b: 34 (*Schizotrachelus bakeri* f.); **syn. n.**

Type depositories. *S. bakeri*: SMTD (ST); *S. bakeri* f. *concolor*: SMTD (ST).

Distribution. Philippines (Basilan, Catanduanes, Luzon, Mindanao, Negros, Polillo, Samar, Siargao, Sibuyan).

Notes. We include the species in this genus going by the characters given in the original description.

***birmanicus* (Senna, 1892) n. comb.**

birmanicus Senna, 1892e: 485 (*Schizotrachelus*).

Type depository. MSNG (LT!, PLT!).

Distribution. Myanmar [= Burma], Vietnam.

Notes. Lectotype of *S. birmanicus* (MSNG) designated on a male specimen with the following label: Bhamò, Birmania, VIII.1885, Fea. Paralectotype: 1 female specimen (MSNG) with the same label data as the lectotype.

We include the species in this genus on the basis of the external morphology of the type material.

***brunneus* (Kleine, 1922) n. comb.**

brunneus Kleine, 1922b: 36 (*Schizotrachelus*).

Type depository. SMTD (ST).

Distribution. Philippines (Luzon).

Notes. We include the species in this genus going by the characters given in the original description.

***cameratus* (Lacordaire, 1866) n. comb.**

cameratus Lacordaire, 1866: 456 (*Schizotrachelus*).

tameratus: Kleine, 1926a: 60 (*Schizotrachelus*) [lapsus].

Type depository. IRSNB (LT!, PLT!).

Distribution. Borneo, India (Andaman Is.), Indonesia (Sumatra), Malaysia, Thailand, Vietnam; New Guinea.

Notes. Lectotype of *S. cameratus* (IRSNB) designated on a male specimen [incorrectly labelled as holotype] with the following labels: Malacca/ Castelnau/ type /Coll. Lacordaire/ *Schizotrachelus cameratus*/ Prépar. génit. n° 76 H 16, R. Damoiseau/ *Tracheloschizus cameratus* (Lacordaire), HT, R. Damoiseau det. Paralectotypes: 1 male specimen (IRSNB), Malacca/ Coll. Castelnau, Coll. Roelofs/ Prépar. génit. n° 76 H 23, R. Damoiseau/ *Tracheloschizus cameratus* (Lacordaire), R. Damoiseau det.

We include the species in this genus on the basis of the external morphology of the type material.

***castaneicolor* (Kleine, 1933) n. comb.**

castaneicolor Kleine, 1933e: 326 (*Schizotrachelus*).

Type depository. NHM (HT!).

Distribution. Fiji Is.

Notes. We include the species in this genus on the basis of the external morphology of the type material.

***consideratus* (Kleine, 1926) n. comb.**

consideratus Kleine, 1926a: 61 (*Schizotrachelus*).

Type depositories. NHM (LT!, PLT!), SMTD (PLT).

Distribution. Borneo, "Philippines".

Notes. Lectotype of *S. consideratus* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ Typus ♂/ Quop, W Sarawak, G. E. Bryant, 2.IV.14/ G. Bryant Coll. 1919-147/ *Schizotrachelus consideratus* Kln, R. Kleine det. 1922/ *Tracheloschizus consideratus* (Kleine) R. Damoiseau det. Paralectotype: 1 female specimen (NHM), Syntype/ Puak, W Sarawak, G. E. Bryant, 5.V.14/ G. Bryant, Coll. 1919-147/ *Tracheloschizus consideratus* (Kleine), R. Damoiseau det.

We include the species in this genus on the basis of the external morphology of the type material.

***consimilis* (Kleine, 1923) n. comb.**

consimilis Kleine, 1923e: 166 (*Schizotrachelus*).

Type depository. NHM (HT!).

Distribution. Indonesia (Moluccas: Ambon), Philippines (Luzon).

Notes. We include the species in this genus on the basis of the external morphology of the type material.

***corpulentus* (Kleine, 1922) n. comb.**

corpulentus Kleine, 1922b: 32 (*Schizotrachelus*).

Type depository. SMTD (ST).

Distribution. Philippines (Luzon, Mindanao).

Notes. We include the species in this genus going by the characters given in the original description.

***dichrous* (Lacordaire, 1866)**

dichrous Lacordaire, 1866: 456 (*Schizotrachelus*).

Type depository. IRSNB (LT!, PLT!).

Distribution. Indonesia (Moluccas: Larat); Australia.

Notes. Damoiseau (1989b: 92) reported that the specimens held in IRSNB are holotype and allotype, but in his original description Lacordaire did not designate any of the syntypes as holotype; however, as there is more than one syntype in IRSNB, and as Damoiseau did not specifically select one as holotype, we designate the lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Lectotype of *S. dichrous* (IRSNB) designated on a male specimen with the following labels: Moret. B. [Moreton Bay]/ Coll. Roelofs/ nov. sp./ *Schizotra-*

chelus dichrous/ Holotype/ *Tracheloschizus dichrous* Lacord., R. Damoiseau det. Paralectotypes: 1 male specimen (IRSNB), Dr. A. Breyer/ Coll. Roelofs/ *Schizotrachelus dichrous* ♂/ Prépar. génit. n° 76 H 14, R. Damoiseau/ *Tracheloschizus dichrous* (Lac.), R. Damoiseau det.; 1 male specimen (IRSNB), M. R. Belg./ *Schizotrachelus dichrous* Lacord. ♂/ *Tracheloschizus dichrous* (Lac.), R. Damoiseau det.

***fortis* (Kleine, 1935) n. comb.**

fortis Kleine, 1935e: 309 (*Schizotrachelus*).

Type depository. NHM (ST!).

Distribution. New Guinea.

Notes. We include the species in this genus after studying the external morphology of the type material.

***gemellus* (Kleine, 1925) n. comb.**

gemellus Kleine, 1925f: 29 (*Schizotrachelus*).

Type depository. ZMAN (LT!, PLT!).

Distribution. Indonesia (Moluccas: Buru).

Notes. Lectotype of *S. gemellus* (ZMAN) designated on a male specimen with the following labels: L. J. Toxopeus, Buru Station 1, Apr.-Sept. 21. Paralectotype: 1 female specimen (ZMAN), L. J. Toxopeus, Buru Station 1, 10.II-16.III.21.

We include the species in this genus on the basis of the external morphology of the type material.

***generalis* (Kleine, 1925) n. comb.**

generalis Kleine, 1925f: 30 (*Schizotrachelus*).

Type depository. ZMAN (HT!).

Distribution. Indonesia (Moluccas: Buru).

Notes. We include the species in this genus on the basis of the external morphology of the type material.

***gloriosus* Damoiseau, 1989**

gloriosus Damoiseau, 1989b: 93.

Type depositories. MZUF (HT!, PT!), IRSNB (PT!).

Distribution. Vietnam.

***imbricellus* (Kleine, 1925) n. comb.**

imbricellus Kleine, 1925c: 604 (*Schizotrachelus*).

Type depositories. SMTD (ST), MPMP (?) (ST).

Distribution. Philippines (Luzon).

Notes. We include the species in this genus going by the characters given in the original description.

***imitator* (Kleine, 1925) n. comb.**

imitator Kleine, 1925c: 603 (*Schizotrachelus*).

Type depository. SMTD (HT, AT, PT).

Distribution. Philippines (Camiguin, Catanduanes, Leithe, Luzon, Panaon, Polillo).

Notes. We include the species in this genus going by the characters given in the original description.

***implicatus* Damoiseau, 1966**

implicatus Damoiseau, 1966e: 435.

Type depositories. ZMUC (HT), IRSNB (PT!).

Distribution. Philippines (Mindanao, Tawi Tawi).

***inconstans* (Kleine, 1922) n. comb.**

inconstans Kleine, 1922b: 31 (*Schizotrachelus*).

Type depository. SMTD (ST).

Distribution. Philippines (Luzon, Mindanao).

Notes. We include the species in this genus going by the characters given in the original description.

***interruptus* (Senna, 1898) n. comb.**

interruptus Senna, 1898e: 72 (*Schizotrachelus*).

Type depository. MZUF (HT!).

Distribution. Borneo, Malaysia.

Notes. We include the species in this genus on the basis of the external morphology of the type material.

***intrusus* (Senna, 1898) n. comb.**

intrusus Senna, 1898e: 73 (*Schizotrachelus*).

Type depository. MNHN (HT!).

Distribution. Borneo.

Notes. We include the species in this genus on the basis of the external morphology of the type material.

***maindroni* Damoiseau, 1989**

maindroni Damoiseau, 1989b: 92.

Type depositories. MNHN (HT!, AT!, PT!), IRSNB (PT!).

Distribution. Indonesia (Moluccas: Halmahera [= Gilolo]).

***marginatus* (Senna, 1898) n. comb.**

marginatus Senna, 1898e: 75 (*Schizotrachelus*).

Type depository. MZUF (LT!, PLT!).

Distribution. Borneo, Malaysia.

Notes. Lectotype designated by Bartolozzi *et al.* (1985: 98) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

We include the species in this genus on the basis of the external morphology of the type material.

***metallicus* (Senna, 1897) n. comb.**

metallicus Senna, 1897b: 240 (*Schizotrachelus*).

Type depository. MZUF (LT!, PLT!).

Distribution. Indonesia (Sulawesi), Philippines (Luzon, Mindanao).

Notes. Lectotype of *S. metallicus* (MZUF) designated on a male specimen with the following labels: S-Celebes, Lompa-Battau, 3000', marz 1896, H. Fruhstorfer/ *Schizotrachelus metallicus* Senna, typus! ♂ ♀ / Prép. génit. N° 76 J 04, R. Damoiseau/ *Tracheloschizus metallicus* (Senna), R. Damoiseau/ ex-coll. Senna/ "La Specola" Firenze, 7062. Paralectotype: 1 female specimen (MZUF), S-Celebes, Lompa-Battau, 3000', marz 1896, H. Fruhstorfer/ *Tracheloschizus metallicus* (Senna), R. Damoiseau/ ex-coll. Senna/ "La Specola" Firenze, 7062.

We include the species in this genus on the basis of the external morphology of the type material.

***multipunctatus* Damoiseau, 1989**

multipunctatus Damoiseau, 1989b: 92.

Type depository. GDFPNG (HT).

Distribution. New Guinea.

***nanus* (Senna, 1894) n. comb.**

nanus Senna, 1894f: 12 (*Schizotrachelus*).

Type depository. MNHN (LT!, PLT!).

Distribution. Solomon Is.

Notes. Lectotype of *S. nanus* (MNHN) designated on a male specimen with the following labels: Salomon Is./ ex Musaeo H. W. Bates 1892/ *Schizotrachelus nanus* Senna n. typus ♂ ♀/ Dr. Senna vidit 1893/ Museum Paris 1952, coll. R. Oberthür. Paralectotypes: 2 specimens (MNHN), Salomon Is./ ex Musaeo H. W. Bates 1892/ Dr. Senna vidit 1893/ Museum Paris 1952, coll. R. Oberthür.

We include the species in this genus on the basis of the external morphology of the type material.

***neohibernicus* Damoiseau, 1966**

neohibernicus Damoiseau, 1966e: 440.

Type depository. ZMUC (HT).

Distribution. Papua New Guinea (Bismarck Archipelago: New Ireland).

***puncticollis* (Boheman, 1840) n. comb.**

puncticollis Boheman (in Schoenherr, 1840: 494) (*Trachelizus*).

kuncticollis: Kleine, 1926h: [page not numbered, addenda] (*Schizotrachelus*) [lapsus].

Type depository. NHRS (HT).

Distribution. Borneo, Indonesia (Java), Malaysia.

Notes. We include the species in this genus going by the characters given in the original description.

***quaesitus* (Kleine, 1936) n. comb.**

quaesitus Kleine, 1936d: 94 (*Schizotrachelus*).

Type depository. NHM (HT!).

Distribution. Vanuatu [= New Hebrides].

Notes. We include the species in this genus on the basis of the external morphology of the type material.

***salomonensis* (Kleine, 1933) n. comb.**

salomonensis Kleine, 1933e: 326 (*Schizotrachelus*).

Type depository. NHM (HT!).

Distribution. Solomon Is.

Notes. We include the species in this genus on the basis of the external morphology of the type material.

***scrobicollis* (Gyllenhal, 1833) n. comb.**

scrobicollis Gyllenhal (in Schoenherr, 1833: 331) (*Arrhenodes*).

Type depository. NHRS (HT).

Distribution. Borneo, Indonesia (Java, Sumatra), Philippines (Mindanao).

Notes. We include the species in this genus going by the characters given in the original description.

***sculptiventris* (Senna, 1892) n. comb.**

sculptiventris Senna, 1892e: 488 (*Schizotrachelus*).

Type depository. MSNG (HT!).

Distribution. Myanmar [= Burma].

Notes. We include the species in this genus on the basis of the external morphology of the type material.

***suspensus* (Kleine, 1943) n. comb.**

suspensus Kleine, 1943b: 143 (*Schizotrachelus*).

Type depositories. DEI (LT!), HNHM (PLT).

Distribution. New Guinea.

Notes. Lectotype of *S. suspensus* (DEI) designated on a female specimen with the following labels: N. Guinea, Biró 97/ Erima, Astrolabe B./ Syntypus/ *Schizotrachelus suspensus* Kln., R. Kleine det., 1942.

We include the species in this genus going by the characters given in the original description.

***timoriensis* (Senna, 1898) n. comb.**

timoriensis Senna, 1898e: 74 (*Schizotrachelus*).

Type depository. MZUF (HT!).

Distribution. Indonesia (Timor).

Notes. We include the species in this genus going by the characters in the original description.

***unicolor* (Montrouzier, 1855)**

unicolor Montrouzier, 1855: 39 (*Arrhenodes*).

dentatus Kleine, 1944: 155 (*Schizotrachelus*); syn. Damoiseau, 1966a: 27.

Type depositories. *A. unicolor*: IRSNB (LT!); *S. dentatus*: MNHN (HT!).

Distribution. New Caledonia, New Guinea (including Bismarck Archipelago: New Britain; Woodlark).

Notes. Damoiseau (1966a: 21) reported that the specimen of *T. unicolor* held in IRSNB is the holotype, but in his original description Montrouzier did not designate any of the syntypes as holotype; however, as there is only one syntype in IRSNB, this must be considered as lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999). The lectotype [incorrectly labelled as holotype] has the following labels: *Arrhenodes* (Schoenherr) *unicolor* (Woodlark) rec. Montrouzier, 1847-1852/ *Schizotrachelus unicolor* Montr., Type/ Prépar. génit. n° 76 H 13, R. Damoiseau/ *Tracheloschizus unicolor* (Montrouzier), Type, R. Damoiseau det./ Holotype.

***viridescens* (Shaw, 1953) n. comb.**

viridescens Shaw, 1953: 330 (*Schizotrachelus*).

Type depository. NHMB (HT!).

Distribution. Indonesia (Flores); New Guinea.

Notes. We include the species in this genus going by the characters given in the original description.

***vitalisi* (Calabresi, 1921) n. comb.**

vitalisi Calabresi, 1921a: 12 (*Schizotrachelus*).

Type depository. MSNG (HT!).

Distribution. Vietnam.

Notes. We include the species in this genus going by the characters given in the original description.

***Tulotus* Senna, 1894**

Tulotus Senna, 1894b: 370.

Type species: *Tulotus maculipennis* Senna, 1894.

Distribution. Bhutan, India (including Assam), Myanmar [= Burma], Vietnam.

***maculipennis* Senna, 1894**

maculipennis Senna, 1894b: 371.

Type depository. MNHN (LT!, PLT!).

Distribution. Bhutan, India (including Assam), Myanmar [= Burma], Vietnam.

Notes. Lectotype of *T. maculipennis* (MNHN) designated on a male specimen with the following labels: H.te Birmanie, Mines des Rubis, 1200-2300 m, Doherty 1880/ *Tulotus* n. g. *maculipennis* n. *typus*!! Senna vidit 1895. Paralectotypes: 5 male specimens (MNHN), H.te Birmanie, Mines des Rubis, 1200-2300 m, Doherty 1880/ Senna vidit 1895.

***Vasseletia* Sharp, 1895**

Vasseletia Sharp, 1895a: 32.

Vaseletia: Beyer, 1904: 168 [lapsus].

Vassetia: Scivittaro, 1990: 338 [lapsus].

Type species: *Trachelizus vasseleti* Boheman, 1840.

Distribution. U.S.A. (California); Mexico.

***vasseleti* (Boheman, 1840)**

vasseleti Boheman (in Schoenherr, 1840: 498) (*Trachelizus*).

vaseleta: Beyer, 1904: 168 (*Vaseletia*) [lapsus].

Type depositories. MNHN (LT!, PLT!), NHM (PLT!).

Distribution. U.S.A. (California); Mexico.

Notes. Lectotype of *T. vasseleti* (MNHN) designated on a male specimen with the following labels: Type/ Mexique/ Chevrolat 38/ ex Musaeo G. Power/ Museum Paris 1952, coll. R. Oberthür. Paralectotypes: 1 female specimen (MNHN), with the same label data as the lectotype; 1 male specimen (NHM), Syntype/ Type/ Mexico, Sallé Coll./ *Trachelizus vasseleti* Boh. apud Sallé type/ B.C.A., Col. IV, 6. *Vasseletia vasseleti* Sharp; 1 female specimen (NHM), Syntype/ Type/ Mexico, Sallé Coll./ *Trachelizus vasseleti* Boh. apud Sallé type/ *Vasseletia vasseleti* (Boh.) Scivittaro det. 1979.

ACRATINI Alonso-Zarazaga, Lyal, Bartolozzi & Sforzi, 1999

Acratini Alonso-Zarazaga, Lyal, Bartolozzi & Sforzi, (in Alonso-Zarazaga & Lyal, 1999: 53).

Némocéphalides Lacordaire, 1866: 459.

Nemocephali Kolbe, 1883c: 75.

Nemocephalinae Senna, 1892c: 61.

Nemocephalina Sharp, 1895a: 71.

Nematocephalidae Schönfeldt, 1908: 65.

Nematocephalini Schönfeldt, 1910a: 40.

Nemocephalini Kleine, 1922d: 142.

Nemocephalidae Kleine, 1922d: 147.

Acratus Lacordaire, 1866

Acratus Lacordaire, 1866: 463.

Type species: *Brentus suturalis* Lund, 1800.

Distribution. Costa Rica, Guadaloupe Is., Nicaragua; Argentina, Bolivia, Brazil, Colombia, French Guyana, Peru, Venezuela.

apicalis Sharp, 1895

apicalis Sharp, 1895a: 77.

Type depository. NHM (LT!, PLT!).

Distribution. Costa Rica, Nicaragua.

Notes. Lectotype of *A. apicalis* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ Chontales, Nicaragua, T. Belt/ Sharp Coll. 1905-313/ B. C. A., Col. IV, 6. *Acratus apicalis* Sharp. Paralectotypes: 2 male specimens (NHM), with the same label data as the lectotype; 2 male specimens (NHM), Syntype/ Chontales, Janson/ Sharp Coll. 1905-313/ B. C. A., Col. IV, 6. *Acratus apicalis* Sharp; 1 female specimen (NHM), Syntype/ Nicaragua/ Sharp Coll. 1905-313/ B. C. A., Col. IV, 6. *Acratus apicalis* Sharp.

armatus Senna, 1897

armatus Senna, 1897a: 234.

Type depositories. IRSNB (LT!, PLT!), MZUF (PLT!).

Distribution. Brazil, Colombia.

Notes. Lectotype of *A. armatus* (IRSNB) designated on a male specimen with the following labels: Coll. Lacordaire/ M. R. Belg./ *Acratus armatus* (Chev.) ♂/ Colombie/ *Acratus armatus* Senna ♂ ♀ cotypi! Prépar. génit. No. 900 R. Damoiseau/ Paratype. Paralectotypes: 1 male specimen (IRSNB), Coll. Roelofs/ M. R. Belg./ *Teramocerus* Sh. *Interruptolineatus* Ch. *armatus* Dej.; 1 male specimen (IRSNB), n. Gr./ Coll. Roelofs/ *Nemocephalus armatus* Reiche Boga; 1 male specimen (IRSNB), Coll. Roelofs/ M. R. Belg./ nov. sp. *armatus* Reiche; 1 female specimen (IRSNB), Coll. Roelofs/ ♀/ M. R. Belg.; 1 female specimen (IRSNB), Coll. Lacordaire/ ♀/ M. R. Belg.; 1 female specimen (IRSNB), Colombie/ Coll. Roelofs/ M. R. Belg./ *Acratus armatus* (Reiche) (Chev.); 1 male specimen (MZUF), Colombie/ "La Specola", Firenze 12353; 1 female specimen (MZUF), Colombie/ "La Specola", Firenze 12354.

***bechynei* Soares, 1970**

bechynei Soares, 1970b: 200.

Type depository. MPEG (HT).

Distribution. Brazil.

***bellus* Soares, 1970**

bellus Soares, 1970c: 203.

Type depository. MNRJ (Campos Seabra coll.) (HT, AT, PT).

Distribution. Brazil.

***carioca* Soares, 1970**

carioca Soares, 1970c: 205.

Type depository. MNRJ (Campos Seabra coll.) (HT, AT, PT).

Distribution. Brazil.

***diringshofeni* Soares, 1970**

diringshofeni Soares, 1970c: 206.

Type depository. MZSP (Diringshofen coll.) (HT).

Distribution. Brazil.

disjunctus Kleine, 1927

disjunctus Kleine, 1927b: 466.

Type depositories. NHM (LT!), DEI (?) (PLT).

Distribution. Brazil.

Notes. Lectotype of *A. disjunctus* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ 82109/ Brasilia, Rio San/ Fry Coll. 1905-100/ *Acratus disjunctus* Kln, Typus ♂ R. Kleine det. 1923.

According to Kleine (1928a: 94) syntypes were also held in DEI, but we did not find them there.

durabilis Kleine, 1927

durabilis Kleine, 1927b: 468.

Type depository. NHM (HT!).

Distribution. Brazil.

errabundus Kleine, 1927

errabundus Kleine, 1927b: 472.

Type depository. NHM (HT!).

Distribution. Brazil.

expectatus Kleine, 1927

expectatus Kleine, 1927b: 469.

expectatus: Kleine, 1927b: 469 (*Acratus*) [lapsus].

espectatus: Kleine, 1927b: 470 (*Acratus*) [lapsus].

Type depository. NHM (HT!).

Distribution. Brazil, Peru.

expressus Kleine, 1927

expressus Kleine, 1927b: 465.

Type depository. NHM (LT!, PLT!).

Distribution. Brazil.

Notes. Lectotype of *A. expressus* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ 2247/ Fry Rio San/ Fry Coll. 1905-100/ *Acratus expressus* Kln, Type ♂ R. Kleine det. 1923. Paralectotypes: 2 male specimens (NHM), with the same label data as the lectotype.

exquisitus Kleine, 1927

exquisitus Kleine, 1927b: 465.

Type depository. NHM (LT!, PLT!).

Distribution. Brazil, Peru.

Notes. Soares (1970c: 216) reported that a specimen held in NHM is the holotype, but in his original description Kleine did not designate either of the syntypes as holotype; however, as there are two syntypes in NHM, and as Soares did not specifically select one as holotype, we designate the lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Lectotype of *A. exquisitus* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ 2247/ Para/ 9/ Pascoe Coll. 93-60/ *Acratus exquisitus* Kln, Typus ♂, R. Kleine det. 1923. Paralectotype: 1 male specimen (NHM), Syntype/ 15988/ Amazon, Bates/ Fry Coll. 1905-100/ *Acratus exquisitus* Kln.

extraordinarius Kleine, 1927

extraordinarius Kleine, 1927b: 472.

Type depository. NHM (HT!).

Distribution. Costa Rica; Brazil.

extrarius Kleine, 1927

extrarius Kleine, 1927b: 471.

Type depository. NHM (LT!, PLT!).

Distribution. Bolivia, Brazil, French Guyana, Peru.

Notes. Lectotype of *A. extrarius* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ Amazon, Bates/ Fry Coll. 1905-100/ ex Mus. W. W. S./ *Acratus extrarius* Kln, Type ♂ R. Kleine det. 1923. Paralectotype: 1 female specimen (NHM), with the same label data as the lectotype.

fallax Kleine, 1927

fallax Kleine, 1927b: 463.

Type depository. NHM (HT!).

Distribution. Brazil.

Notes. According to Soares (1970c: 250) this species is probably synonymous of *Brentus rufiventris* Boheman, 1840.

fidelis Kleine, 1927

fidelis Kleine, 1927b: 469.

Type depository. NHM (LT!, PLT!).

Distribution. Brazil.

Notes. Lectotype of *A. fidelis* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ 2245/ Fry Rio San/ Fry Coll. 1905-100/ *Acratus fidelis* Kln Type ♂ R. Kleine det. 1923. Paralectotypes: 3 male and 1 female specimens (NHM), with the same label data as the lectotype.

***gracilipes* Soares, 1970**

gracilipes Soares, 1970c: 219.

Type depository. MNRJ (Campos Seabra coll.) (HT).

Distribution. Brazil.

***interruptolineatus* (Gyllenhal, 1833)**

interruptolineatus Gyllenhal (in Schoenherr, 1833: 352) (*Brenthus*).

Type depository. MNHN (HT!).

Distribution. Brazil.

***laevigatus* (Boheman, 1840)**

laevigatus Boheman (in Schoenherr, 1840: 562) (*Teramocerus*).

laerigatus: Kleine, 1927b: 472 (*Acratus*) [lapsus].

Type depository. NHRS (ST).

Distribution. Brazil, Venezuela.

***mendax* Soares, 1970**

mendax Soares, 1970c: 225.

Type depository. MNRJ (Campos Seabra coll.) (HT).

Distribution. Brazil.

***nitidissimus* Soares, 1970**

nitidissimus Soares, 1970c: 226.

Type depository. MNRJ (Campos Seabra coll.) (HT).

Distribution. Brazil.

plumirostris (Boheman, 1840)

plumirostris Boheman (in Schoenherr, 1840: 568) (*Teramocerus*).

Type depository. MLUH (HT).

Distribution. Brazil, Venezuela.

pohli Soares, 1970

pohli Soares, 1970c: 229.

Type depository. MZSP (Diringshofen coll.) (HT).

Distribution. Brazil.

propinquus Senna, 1890

propinquus Senna, 1890: 108.

Type depository. MZUF (LT!, PLT!).

Distribution. Brazil.

Notes. Lectotype designated by Bartolozzi *et al.* (1985: 99) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

pseudoarticulatus Soares, 1970

pseudoarticulatus Soares, 1970c: 232.

Type depository. MNRJ (HT).

Distribution. Brazil.

ruber (Erichson, 1847)

ruber Erichson, 1847: 127 (*Brenthus*).

Type depository. MNHUB (?) (HT).

Distribution. Brazil, Peru.

Notes. According to Kleine (1928a: 94) the type was in MNHUB, but we did not find it there.

***seabrai* Soares & Meyer, 1959**

seabrai Soares & Meyer, 1959: 167.

Type depository. MNRJ (Campos Seabra coll.) (HT, AT).

Distribution. Brazil.

***seorsus* Soares, 1970**

seorsus Soares, 1970c: 238.

Type depository. APC (HT).

Distribution. Brazil.

***subfasciatus* (Boheman, 1840)**

subfasciatus Boheman (in Schoenherr, 1840: 569) (*Teramocerus*).

Type depository. NHRS (ST).

Distribution. Guadeloupe Is.

***suratus* (Boheman, 1840)**

suratus Boheman (in Schoenherr, 1840: 539) (*Brenthus*).

Type depository. NHRS (HT).

Distribution. Brazil.

***suturalis* (Lund, 1800)**

suturalis Lund, 1800: 63 (*Brenthus*).

helmenreichii Redtenbacher, 1868: 172 (*Trachelizus*); syn. Kleine, 1927a: 74.

helmenreichi: Schönfeldt, 1908: 22 (*Trachelizus*) [lapsus].

Type depositories. *B. suturalis*: ZMUC (HT); *T. helmenreichii*: NMW (HT).

Distribution. Bolivia, Brazil.

Notes. Olivier (1807: 454) also mentioned French Guyana and Suriname but the old quotations are sometimes unreliable.

***tarsatus* (Gyllenhal, 1833)**

tarsatus Gyllenhal (in Schoenherr, 1833: 351) (*Brenthus*).

kleinei Haedo Rossi, 1954a: 2 (*Acratus*); syn. Soares, 1970c: 245.

Type depositories. *B. tarsatus*: NHRS (HT); *A. kleinei*: IMLA (HT, AT, PT), JHRC (PT), MLPA (PT).

Distribution. Argentina, Brazil.

***telesi* Soares & Meyer, 1959**

telesi Soares & Meyer, 1959: 169.

Type depository. MNRJ (Campos Seabra coll.) (HT, AT).

Distribution. Brazil, Peru.

***villens* Soares, 1970**

villens Soares, 1970c: 248.

Type depository. MNRJ (Campos Seabra coll.) (HT, AT).

Distribution. Brazil.

***Fonteboanius* Senna, 1893**

Fonteboanius Senna, 1893d: 125.

Fontebuanus: Bartolozzi *et al.*, 1985: 98 [lapsus].

Type species: *Fonteboanius lamellipes* Senna, 1893.

Distribution. Brazil.

***lamellipes* Senna, 1893**

lamellipes Senna, 1893c: 125.

Type depositories. MZUF (LT!), SMFD (von Schönfeldt coll.) (PLT).

Distribution. Brazil.

Notes. Lectotype designated by Bartolozzi *et al.* (1985: 98) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

***Leptocymatium* Kleine, 1922**

Leptocymatium Kleine, 1922d: 147.

Type species: *Leptocymatium perlucidum* Kleine, 1922.

Distribution. Guadeloupe Is., Virgin Is.; Brazil.

***observans* Kleine, 1941**

observans Kleine, 1941c: 192.

Type depository. MZPW (ST).

Distribution. Brazil.

***perlucidum* Kleine, 1922**

perlucidum Kleine, 1922d: 148.

Type depositories. SMTD (HT), DEI (PT!).

Distribution. Guadeloupe Is., Virgin Is.

***Neacratus* Alonso-Zarazaga, Lyal,
Bartolozzi & Sforzi, 1999**

Neacratus Alonso-Zarazaga, Lyal, Bartolozzi & Sforzi, 1999: 54 .

Nemocephalus: Lacordaire, 1866: 462 (non Guérin-Méneville, 1827) [misidentification; Alonso-Zarazaga & Lyal, 1999: 54]

Nematocephalus: Gemminger & Harold, 1872: 2718 (non Agassiz, 1846b) [misidentification; Alonso-Zarazaga & Lyal, 1999: 54]

Type species: *Brentus glabratus* Lund, 1800.

Distribution. Belize, Costa Rica, Dominican Republic, El Salvador, Guatemala, Mexico, Nicaragua, Panama; Argentina, Bolivia, Brazil, Colombia, Ecuador, French Guyana, Paraguay, Peru, Venezuela.

***argentinus* (Haedo Rossi, 1954) n. comb.**

argentinus Haedo Rossi, 1954a: 2 (*Nemocephalus*).

Type depositories. MLPA (HT, AT, PT), APC (PT), JHRC (PT).

Distribution. Argentina, Bolivia, Brazil.

***brevicostatus* (Kleine, 1922) n. comb.**

brevicostatus Kleine, 1922d: 144 (*Nemocephalus*).

Type depository. SMTD (HT).

Distribution. Brazil, French Guyana.

***clavipes* (Boheman, 1840) n. comb.**

clavipes Boheman (in Schoenherr, 1840: 547) (*Brenthus*).

Type depositories. MNHN (LT!, PLT!), NHRS (PLT).

Distribution. Costa Rica; Brazil, Peru.

Notes. Lectotype of *B. clavipes* (MNHN) designated on a male specimen with the following labels: Bresil/ Chevrolat 284/ type/ ex Museo G. Power/ Museum Paris 1952, coll. R. Oberthür. Paralectotype (MNHN): 1 female specimen with the same label data as the lectotype.

***crinitus* (Kleine, 1922) n. comb.**

crinitus Kleine, 1922d: 143 (*Nemocephalus*).

Type depository. SMTD (HT).

Distribution. Brazil, Paraguay.

***deplanatus* (Sharp, 1895) n. comb.**

deplanatus Sharp, 1895a: 75 (*Nemocephalus*).

Type depository. NHM (HT!).

Distribution. Nicaragua.

***dolosus* (Kleine, 1927) n. comb.**

dolosus Kleine, 1927b: 456 (*Nemocephalus*).

Type depository. NHM (LT!, PLT!).

Distribution. Dominican Republic.

Notes. Lectotype of *N. dolosus* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ Dominica, Laiou, G. A. Ramage, 97-67/ *Nemocephalus dolosus* Kln, Type ♂ R. Kleine det. 1923. Paralectotypes: 3 male specimens (NHM), with the same label data as the lectotype; 2 male and 1 female specimens (NHM), Syntype/ Dominica, 95-132.

***erectus* (Kleine, 1927) n. comb.**

erectus Kleine, 1927b: 457 (*Nemocephalus*).

Type depository. NHM (HT!).

Distribution. Ecuador.

***famulus* (Boheman, 1840) n. comb.**

famulus Boheman (in Schoenherr, 1840: 546) (*Brenthus*).

Type depository. NHRS (HT).

Distribution. Brazil.

***fasciatus* (Kleine, 1922) n. comb.**

fasciatus Kleine, 1922d: 145 (*Nemocephalus*).

proseni Haedo Rossi, 1954a: 2 (*Nemocephalus*); syn. Soares & Scivittaro, 1977: 92.

Type depositories. *N. fasciatus*: SMTD (HT); *N. proseni*: IMLA (HT, AT, PT), APC (PT), JHRC (PT), MLPA (PT).

Distribution. Argentina, Brazil, Colombia, Paraguay, Venezuela.

***femoratus* (Sharp, 1895) n. comb.**

femoratus Sharp, 1895a: 74 (*Nemocephalus*).

Type depository. NHM (LT!, PLT!).

Distribution. Belize, El Salvador, Guatemala, Nicaragua.

Notes. Lectotype of *N. femoratus* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ El Reposo, 800 ft, Champion/ B. C. A., Col. IV, 6. *Nemocephalus femoratus* Sharp/ Sharp Coll. 1905-313. Paralectotypes: 9 specimens (NHM), with the same label data as the lectotype; 1 male specimen (NHM), Syntype/ Paraiso, 300 ft, Champion/ B. C. A., Col. IV, 6. *Nemocephalus femoratus* Sharp / Sharp Coll. 1905-313; 1 male specimen (NHM), Syntype/ R. Sarstoon, B. Honduras, Blancaneau/ B. C. A., Col. IV, 6. *Nemocephalus femoratus* Sharp / Sharp Coll. 1905-313.

***fulgidus* (Kleine, 1928) n. comb.**

fulgidus Kleine, 1928f: 225 (*Nemocephalus*).

Type depository. ZMUC (ST).

Distribution. Brazil.

***glabratus* (Lund, 1800)**

glabratus Lund, 1800: 65 (*Brentus*).

Type depository. ZMUC (ST).

Distribution. Brazil, Colombia, French Guyana, Paraguay.

Notes. In NHM there are two specimens, a male and a female, labelled as «types» but it is uncertain if they really are types.

***guatemalensis* (Senna, 1893) n. comb.**

guatemalensis Senna, 1893d: 128 (*Nematocephalus*).

Type depositories. MZUF (LT!, PLT!), SMTD (PLT), SMFD (von Schönfeldt coll.) (PLT).

Distribution. Costa Rica, Guatemala, Mexico, Nicaragua, Panama.

Notes. Lectotype of *N. guatemalensis* (MZUF) designated on a specimen with the following labels: Guatemala, A. Boucard/ *Nemocephalus guatemalensis* typus! Senna/ Bull. Soc. Ent. It., XXV, 1893, typus!/ ex-coll. Senna/ "La Specola" Firenze, 7069. Paralectotype (MZUF): Guatemala, A. Boucard/ *Nemocephalus guatemalensis* typus! Senna/ ex-coll. Senna/ "La Specola" Firenze, 7069.

***longiceps* (Perroud, 1853) n. comb.**

longiceps Perroud, 1853: 438 (*Nemocephalus*).

Type depository. MNHN (HT!).

Distribution. Brazil.

***muticus* (Kleine, 1922) n. comb.**

muticus Kleine, 1922d: 145 (*Nemocephalus*).

Type depository. SMTD (HT).

Distribution. Brazil.

***nodosus* (Kleine, 1937) n. comb.**

nodosus Kleine, 1937c: 141 (*Nemocephalus*).

Type depository. NMW (ST).

Distribution. Argentina.

***obtusus* (Lund, 1800) n. comb.**

obtusus Lund, 1800: 64 (*Brentus*).

Type depository. ZMUC (ST).

Distribution. Brazil.

***piceus* (Perroud, 1853) n. comb.**

piceus Perroud, 1853: 442 (*Nemocephalus*).

Type depository. MNHN (LT!, PLT!).

Distribution. Brazil.

Notes. Lectotype of *N. piceus* (MNHN) designated on a male specimen with the following label: Brésil. Paralectotype: 1 female specimen (MNHN), with no label.

***puncticeps* (Sharp, 1895) n. comb.**

puncticeps Sharp, 1895a: 76 (*Nemocephalus*).

punctipes: Kleine, 1933c: 288 (*Nemocephalus*) [lapsus].

Type depository. NHM (LT!, PLT!).

Distribution. Belize, Costa Rica, Guatemala, Mexico, Nicaragua, Panama.

Notes. Lectotype of *N. puncticeps* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ Temax, N. Yucatan, Gaumer/ B. C. A., Col. IV, 6. *Nemocephalus puncticeps* Sharp/ Sharp Coll. 1905-313. Paralectotypes: 3 male specimens (NHM), with the same label data as the lectotype; 1 male specimen (NHM), Syntype/ Cerro de Plumas, Mexico, Hoege/ B. C. A., Col. IV, 6. *Nemocephalus puncticeps* Sharp/ Sharp Coll. 1905-313; 1 male specimen (NHM), Syntype/ R. Sarstoon, B. Honduras, Blancaneau/ B.

C. A., Col. IV, 6. *Nemocephalus puncticeps* Sharp; 1 male specimen (NHM), Syntype/ Chontales, Nicaragua, T. Belt/ B. C. A., Col. IV, 6 *Nemocephalus puncticeps* Sharp; 1 male specimen (NHM), Syntype/ V. de Chiriqui, 25-4000 ft, Champion/ B. C. A., Col. IV, 6. *Nemocephalus puncticeps* Sharp; 1 female specimen (NHM), Syntype/ Teleman, Vera Paz, Champion/ B. C. A., Col. IV, 6. *Nemocephalus puncticeps* Sharp; 2 female specimens (NHM), Syntype/ Bugaba, 800-1500 ft, Champion/ B. C. A., Col. IV, 6. *Nemocephalus puncticeps* Sharp; 1 male and 1 female specimen (NHM), Syntype/ Mexico, Salle Coll./ B. C. A., Col. IV, 6. *Nemocephalus puncticeps* Sharp; 1 male and 1 female specimen (NHM), Syntype/ Chontales, Janson/ B. C. A., Col. IV, 6. *Nemocephalus puncticeps* Sharp/ Sharp Coll. 1905-313.

***punctulatus* (Boheman, 1840) n. comb.**

punctulatus Boheman (in Schoenherr, 1840: 544) (*Brenthus*).

Type depository. MNHN (HT!).

Distribution. Costa Rica, Mexico; Paraguay.

***scotti* (Kleine, 1923) n. comb.**

scotti Kleine, 1923c: 131 (*Nemocephalus*).

armatus Haedo Rossi, 1954a: 2 (*Nemocephalus*); syn. Soares & Scivittaro, 1977: 93.

Type depositories. *N. scotti*: NHM (LT!, PLT!); *N. armatus*: IMLA (HT, AT, PT), APC (PT), JHRC (PT).

Distribution. "Antilles"; Argentina, Bolivia, Brazil.

Notes. Lectotype of *N. scotti* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ Typus/ Amazonas, Goodman, 1879/ ex Mus. Cambr. 1922-127/ *Nemocephalus scotti* det. Aut. 1922. Paralectotype: 1 female specimen (NHM), with the same label data as the lectotype.

***tonsus* (Kleine, 1928) n. comb.**

tonsus Kleine, 1928f: 223 (*Nemocephalus*).

Type depository. ZMUC (ST).

Distribution. Brazil, Peru.

Nemobrenthus Sharp, 1895

Nemobrenthus Sharp, 1895a: 71.

Neobrenthus: Kleine, 1937e: 102 [lapsus].

Type species: *Nemobrenthus aeneipennis* Sharp, 1895.

Distribution. Costa Rica, Panama; Brazil, Venezuela.

aeneipennis Sharp, 1895

aeneipennis Sharp, 1895a: 72.

aeneippennis: Kleine, 1927b: 462 (*Nemobrenthus*) [lapsus].

Type depository. NHM (LT!, PLT!).

Distribution. Costa Rica, Panama; Brazil, Ecuador, Venezuela.

Notes. Lectotype of *N. aeneipennis* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ David, Panama, Champion/ B. C. A., Col. IV, 6. *Nemocephalus aeneipennis* Sharp/ Sharp Coll. 1905-313. Paralectotype: 1 male specimen (NHM), Syntype/ Type/ V. de Chiriqui, 25-4000 ft, Champion/ B. C. A., Col. IV, 6. *Nemocephalus aeneipennis* Sharp / Sharp Coll. 1905-313.

bechynei Soares & Scivittaro, 1972

bechynei Soares & Scivittaro, 1972b: 281.

Type depositories. MPEG (HT, PT), MNRJ (Campos Seabra coll.) (HT), MZSP (PT).

Distribution. Brazil.

Notes. Soares & Scivittaro redescribed the species in 1979 (1979b: 207).

expletus Kleine, 1927

expletus Kleine, 1927b: 461.

Type depository. NHM (LT!, PLT!).

Distribution. Brazil.

Notes. Lectotype of *N. expletus* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ Jatahy, Prov. Goyas. Brésil, Déc. 97- Janv. 98/ Fry Coll. 1905-100/ *Nemocephalus expletus* Typus ♂, det. R. Kleine, 1927. Paralectotype: 1 female specimen (NHM), with the same label data as the lectotype.

***melzeri* Kleine, 1926**

melzeri Kleine, 1926f: 371.

Type depository. IPEACS (HT).

Distribution. Brazil.

***pereirai* Soares & Scivittaro, 1972**

pereirai Soares & Scivittaro, 1972b: 281.

Type depository. MZSP (HT).

Distribution. Brazil.

Notes. Soares & Scivittaro redescribed the species in 1979 (1979b: 210).

***sublaevis* (Boheman, 1840)**

sublaevis Boheman (in Schoenherr, 1840: 541) (*Brenthus*).

Type depository. NHRS (ST).

Distribution. Brazil, Ecuador.

***Nemocephalinus* Kleine, 1927**

Nemocephalinus Kleine, 1927b: 459.

Type species: *Nemocephalinus dubitabilis* Kleine, 1927.

Distribution. Brazil, French Guyana.

dubitabilis Kleine, 1927

dubitabilis Kleine, 1927b: 459.

dubtitabilis: Kleine, 1927a: 73 (*Nemocephalinus*) [lapsus].

Type depository. NHM (LT!, PLT!).

Distribution. Brazil, French Guyana.

Notes. Lectotype of *N. dubitabilis* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ Cayen ♂/ Bowring 63-47/ *Nemocephalinus dubitabilis* Kln, Typus! R. Kleine det. 1923. Paralectotype: 1 male specimen (NHM), Syntype/ Fry Rio San/ Fry Coll. 1905-100/ *Nemocephalinus dubitabilis* Kln, R. Kleine det. 1923.

Nemocephalus Guérin-Ménéville, 1827

Nemocephalus Guérin-Ménéville, 1827: 506.

Nematocephalus Agassiz, 1846b: 247 [unjustified emendation].

Exopleura Kleine, 1922d: 149; syn. Alonso-Zarazaga & Lyal, 1999: 54.

Type species: *Brentus monilis* Fabricius, 1787 [by Alonso-Zarazaga, Lyal, Bartolozzi & Sforzi, 1999].

Distribution. Bahamas, Dominican Republic, Guadaloupe Is., Haiti, Mexico, Virgin Is.; Colombia.

avarus (Kleine, 1927) **n. comb.**

avara Kleine, 1927b: 450 (*Exopleura*).

avora: Kleine, 1927b: 452 (*Exopleura*) [lapsus].

Type depositories. SMTD (ST), NHM (ST!).

Distribution. Dominican Republic, Mexico; Colombia.

enodis (Kleine, 1927) **n. comb.**

enodis Kleine, 1927b: 451 (*Exopleura*).

Type depository. NHM (HT!).

Distribution. "Antilles", Bahamas.

***gounellei* Kleine, 1944**

gounellei Kleine, 1944: 157.

Type depository. MNHN (LT!, PLT!).

Distribution. Brazil.

Notes. Lectotype of *N. gounellei* (MNHN) designated on a specimen with the following labels: Salobro prov. De Bahia, Bresil, E G. Gounelle, 6.7.1885/ 27/ *Acratus* sp. 1 ♂ 2 ♀ [*sic!*]/ type/ Museum Paris, coll. E. Gounelle/ *Nemocephalus gounellei* Kln., R. Kleine det 1941. Paralectotype: 1 female specimen (MNHN), Pery Pery, Pernambuco/ cotype/ Museum Paris, coll. E. Gounelle 1915.

***moderatus* (Kleine, 1922) n. comb.**

moderata Kleine, 1922d: 150 (*Exopleura*).

Type depository. SMTD (HT).

Distribution. Haiti.

***monilis* (Fabricius, 1787)**

monilis Fabricius, 1787: 95 (*Brentus*).

Type depository. ZMUC (ST).

Distribution. Guadeloupe Is., Haiti, Virgin Is.

***Nemocoryna* Sharp, 1895**

Nemocoryna Sharp, 1895a: 72.

Type species: *Nemocoryna godmani* Sharp, 1895.

Distribution. Costa Rica, Panama; Bolivia, Brazil.

extranea Kleine, 1927

extranea Kleine, 1927b: 460.

Type depository. NHM (HT!).

Distribution. Brazil.

godmani Sharp, 1895

godmani Sharp, 1895a: 73.

Type depository. NHM (HT!).

Distribution. Costa Rica, Panama; Bolivia, Brazil.

sericata Sharp, 1895

sericata Sharp, 1895a: 73.

Type depository. NHM (HT!).

Distribution. Panama; Brazil.

Proteramocerus Kleine, 1921

Proteramocerus Kleine, 1921g: 129.

Type species: *Brenthus acutipennis* Boheman, 1833.

Distribution. Belize, Costa Rica, Panama, Nicaragua; Bolivia, Brazil, Ecuador, French Guyana, Guiana, Paraguay, Peru.

acutipennis (Boheman, 1833)

acutipennis Boheman (in Schoenherr, 1833: 350) (*Brenthus*).

Type depository. NHRS (HT).

Distribution. Brazil.

***aeneus* Soares & Dias, 1971**

aeneus Soares & Dias, 1971: 47.

Type depository. MZSP (Diringshofen coll.) (HT, PT).

Distribution. Brazil.

***affinis* Soares & Dias, 1971**

affinis Soares & Dias, 1971: 49.

Type depository. MNRJ (Campos Seabra coll.) (HT).

Distribution. Brazil.

***appendiculatus* Soares & Scivittaro, 1972**

appendiculatus Soares & Scivittaro, 1972a: 1.

Type depository. MNRJ (Campos Seabra coll.) (HT, AT, PT).

Distribution. Brazil.

***badius* (Boheman, 1840)**

badius Boheman (in Schoenherr, 1840: 563) (*Teramocerus*).

Type depository. MLUH (HT).

Distribution. Brazil.

***chalcites* (Perty, 1832)**

chalcites Perty, 1832: 69 (*Arrhenodes*).

sesquistriatus Boheman (in Schoenherr, 1833: 349) (*Brenthus*); syn. Schoenherr, 1840: 541.

Type depositories. *A. chalcites*: ZSM (HT); *B. sesquistriatus*: NHRS (ST).

Distribution. Bolivia, Brazil.

***chontalensis* (Sharp, 1895)**

chontalensis Sharp, 1895a: 76 (*Acratus*).

Type depository. NHM (LT!, PLT!).

Distribution. Costa Rica, Nicaragua; Bolivia, Brazil, Peru.

Notes. Lectotype of *A. chontalensis* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ Chontales, Nicaragua, T. Belt/ Sharp Coll. 1905-313/ B. C. A., Col. IV, 6. *Acratus chontalensis* Sharp. Paralectotype: 1 female specimen (NHM), Syntype/ Type/ Chontales, Janson/ Sharp Coll. 1905-313/ B. C. A., Col. IV, 6. *Acratus chontalensis* Sharp.

***diringshofeni* Soares & Dias, 1971**

diringshofeni Soares & Dias, 1971: 54.

Type depository. MZSP (Diringshofen coll.) (HT, AT, PT), MNRJ (PT).

Distribution. Brazil.

***disparilis* Soares & Dias, 1971**

disparilis Soares & Dias, 1971: 56.

Type depositories. MNRJ (Campos Seabra coll.) (HT, AT, PT), IBSP (PT), IOC (PT), MNRJ (PT), MZSP (Diringshofen coll.) (PT).

Distribution. Brazil.

***emendatus* Kleine, 1927**

emendatus Kleine, 1927b: 448.

Type depository. NHM (HT!).

Distribution. Brazil.

eminens Kleine, 1927

eminens Kleine, 1927b: 447.

Type depository. NHM (HT!).

Distribution. Brazil.

enervatus Kleine, 1927

enervatus Kleine, 1927b: 449.

Type depository. NHM (LT!, PLT!).

Distribution. Brazil.

Notes. Lectotype of *P. enervatus* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ Selected ♂ Para, Bates, killed trees/ Bowring 63-47/ *Proteramocerus enervatus* Kln ♂ type, R. Kleine det. 1923. Paralectotypes: 1 female specimen (NHM), Syntype/ Co-type/ Para/ Bowring 63-47/ *Proteramocerus enervatus* Kln ♀ type, R. Kleine det. 1923/ *Teramocerus amoenus* Jek. n. sp./ *Amoenus* Jek. Mss.; 1 male specimen (NHM), Syntype/ Para/ *Proteramocerus enervatus* Kln, R. Kleine det. 1923.

fidus Kleine, 1927

fidus Kleine, 1927b: 447.

Type depository. NHM (LT!).

Distribution. Brazil.

Notes. Lectotype of *P. fidus* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ St Paulo/ *Proteramocerus fidus* Kln ♂ type, R. Kleine det. 1923.

filum (Sharp, 1895)

filum Sharp, 1895a: 78 (*Acratus*).

Type depository. NHM (LT!, PLT!).

Distribution. Belize, Nicaragua.

Notes. Lectotype of *A. filum* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ Chontales, Nicaragua, T. Belt/ Sharp Coll. 1905-313/ B. C. A., Col. IV, 6. *Acratus filum* Sharp. Paralectotype: 1 male specimen (NHM), Syntype/ Type/ Rio Hondo, B. Honduras, Blancaneau/ Sharp Coll. 1905-313/ B. C. A., Col. IV, 6. *Acratus filum* Sharp.

forficula Soares & Scivittaro, 1972

forficula Soares & Scivittaro, 1972a: 3.

Type depository. MNRJ (Campos Seabra coll.) (HT, AT), MZSP (Diringshofen coll.) (PT).

Distribution. Brazil.

goianus Soares & Dias, 1971

goianus Soares & Dias, 1971: 61.

Type depository. MZSP (Diringshofen coll.) (HT).

Distribution. Brazil.

laevis (Germar, 1824)

laevis Germar, 1824: 190 (*Brentus*).

Type depository. MLUH (HT).

Distribution. Brazil, Paraguay.

luteolineatus Meyer, 1959

luteolineatus Meyer, 1959a: 128.

Type depository. MNRJ (Campos Seabra coll.) (HT).

Distribution. Brazil.

micans Kleine, 1921

micans Kleine, 1921g: 130.

Type depository. SMTD (HT).

Distribution. Costa Rica; Brazil, French Guyana, Guiana.

nitidus Kleine, 1921

nitidus Kleine, 1921g: 131.

Type depository. SMTD (ST).

Distribution. Costa Rica, Panama; Brazil.

obtusipennis Kleine, 1944

obtusipennis Kleine, 1944: 154.

Type depository. MNHN (HT!).

Distribution. Brazil.

obuncus Kleine, 1938

obuncus Kleine, 1938f: 290.

obscurus: Kleine, 1938d: 127 (*Proteramocerus*) [lapsus].

Type depository. DEI (HT!).

Distribution. Ecuador.

oliveirai Soares & Dias, 1971

oliveirai Soares & Dias, 1971: 65.

Type depositories. MNRJ (Campos Seabra coll.) (HT, AT, PT), MZSP (Diringshofen coll.) (PT).

Distribution. Brazil.

opacus (Perty, 1832)

opacus Perty, 1832: 69 (*Arrhenodes*).

Type depository. ZSM (HT).

Distribution. Brazil.

pulcher Soares & Dias, 1971

pulcher Soares & Dias, 1971: 67.

Type depositories. IPEACS (HT), MNRJ (Campos Seabra coll.) (PT), MZSP (Diringshofen coll.) (PT).

Distribution. Brazil.

villens Soares & Dias, 1971

villens Soares & Dias, 1971: 69.

Type depositories. MZSP (HT, PT), IPEACS (AT, PT), MNRJ (Campos Seabra coll.) (PT), MZSP (?) (Amante coll.) (PT).

Distribution. Brazil.

zellibori Soares & Dias, 1971

zellibori Soares & Dias, 1971: 71.

Type depository. MNRJ (Campos Seabra coll.) (HT, AT, PT).

Distribution. Brazil.

Sclerotrachelus Kleine, 1921

Sclerotrachelus Kleine, 1921g: 128.

Type species: *Sclerotrachelus brasiliensis* Kleine, 1921.

Distribution. Brazil, Paraguay.

brasiliensis Kleine, 1921

brasiliensis Kleine, 1921g: 129.

Type depository. SMTD (HT).

Distribution. Brazil, Paraguay.

Teramocerus Schoenherr, 1840

Teramocerus Schoenherr, 1840: 556.

Teramozerus: Kleine, 1920e: 13 [lapsus].

Type species: *Teramocerus janthinus* Boheman, 1840.

Distribution. Costa Rica, Nicaragua; Brazil, French Guyana.

belti Sharp, 1895

belti Sharp, 1895a: 78.

Type depository. NHM (LT!, PLT!).

Distribution. Costa Rica, Nicaragua.

Notes. Lectotype of *T. beltii* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ Chontales, Nicaragua, T. Belt/ Sharp Coll. 1905-313/ B. C. A., Col. IV, 6. *Teramocerus beltii* Sharp. Paralectotypes: 1 male and 1 female specimen (NHM), with the same label data as the lectotype; 1 male specimen (NHM), Syntype/ Type/ Chontales, Nicaragua, Janson/ Sharp Coll. 1905-313/ B. C. A., Col. IV, 6. *Teramocerus beltii* Sharp; 1 male specimen (NHM), Syntype/ 2548/ Nicaragua/ Salle Coll./ B. C. A., Col. IV, 6. *Teramocerus beltii* Sharp/ *Teramocerus* sp. apud Sallé; 1 male specimen (NHM), Syntype / Bates/ *Croesus* Lac. MS, Cayen./ Bowring 63-47/ *Teramocerus beltii* Sharp, Syntype.

According to Senna (1895, manuscript note to reprint of Sharp's paper) the female syntype does not belong to this species.

eletus Kleine, 1927

eletus Kleine, 1927b: 446.

elutus: Kleine, 1927b: 508 (*Teramocerus*) [lapsus].

Type depository. NHM (HT!).

Distribution. Brazil.

exilis Boheman, 1840

exilis Boheman (in Schoenherr, 1840: 561).

Type depository. NHRS (HT).

Distribution. Brazil.

gracilis (Boheman, 1833)

gracilis Boheman (in Schoenherr, 1833: 341) (*Belorhynchus*).

Type depository. NHRS (HT).

Distribution. Brazil.

janthinus Boheman, 1840

janthinus Boheman (in Schoenherr, 1840: 558).

Type depository. NHRS (HT).

Distribution. Brazil.

mamillatus Meyer, 1959

mamillatus Meyer, 1959b: 300.

Type depository. MNRJ (Campos Seabra coll.) (HT).

Distribution. Brazil.

***mannerheimi* Boheman, 1840**

mannerheimi Boheman (in Schoenherr, 1840: 559).

Type depository. NHRS (ST).

Distribution. Brazil.

***obscurus* Perroud, 1853**

obscurus Perroud, 1853: 446.

Type depository. MNHN (HT!).

Distribution. Brazil.

***pulchellus* Perroud, 1853**

pulchellus Perroud, 1853: 450.

Type depository. MNHN (HT!).

Distribution. Brazil, French Guyana.

***punctirostris* Boheman, 1840**

punctirostris Boheman (in Schoenherr, 1840: 566).

Type depository. NHRS (ST).

Distribution. Brazil.

***Thaumastopsis* Kleine, 1921**

Thaumastopsis Kleine, 1921g: 126.

Thaumasthopsis: Kleine, 1922m: 163 [lapsus].

Type species: *Thaumastopsis gemmaria* Kleine, 1921.

Distribution. Brazil, Colombia.

exitialis Kleine, 1927*exitialis* Kleine, 1927b: 462.**Type depository.** NHM (HT!).**Distribution.** Brazil.*gemmaria* Kleine, 1921*gemmarius* Kleine, 1921g: 127.**Type depository.** SMTD (HT).**Distribution.** Brazil, Colombia.*separata* Kleine, 1941*separatus* Kleine, 1941c: 193.**Type depository.** MZPW (ST).**Distribution.** Brazil.**ITHYSTENINI** Lacordaire, 1866

Ithysténides Lacordaire, 1866: 464.

Ithysteninae Pascoe, 1872: 317.

Ischnomeri Kolbe, 1883c: 74.

Ithysteni Kolbe, 1883c: 80.

Ithystenina Sharp, 1895a: 78.

Leptorrhynchidae Schönfeldt, 1908: 69.

Ithystenini Schönfeldt, 1910a: 42.

Jthystenini: Kleine, 1932c: 188 [lapsus].

Hystenini: Kleine, 1944: 154 [lapsus].

Ettystenini: Scivittaro, 1975: 436 [lapsus].

Ithystenidae Ienistea, 1986: 32.

Achrionota Pascoe, 1872

Achrionota Pascoe, 1872: 325.

Achrionata: Kleine, 1921a: 55 [lapsus].

Type species: *Achrionota bilineata* Pascoe, 1872.

Distribution. Borneo, Indonesia (Sulawesi, Sumatra), Malaysia, Philippines (Bohol, Leyte, Mindanao, Panaon, Polillo, Siargao).

bilineata Pascoe, 1872

bilineata Pascoe, 1872: 325.

Type depository. NHM (LT!, PLT!).

Distribution. Borneo, Indonesia (Sumatra), Malaysia, Philippines (Bohol, Leyte, Mindanao, Panaon, Polillo, Siargao).

Notes. Lectotype of *A. bilineata* (NHM) designated on a male specimen with the following labels: Syntype/ Borneo/ Pascoe Coll. 93-60/ *Achrionota bilineata* Pasc. Type. Paralectotype: 1 male specimen (NHM), with the same label data as the lectotype [missing thorax, head glued to the label].

celebica (Senna, 1897) n. comb.

celebicus Senna, 1897b: 242 (*Heteroplites*).

Type depositories. MZUF (LT!, PLT!), MSNG (PLT!), Fruhstorfer coll. (localization unknown) (PLT).

Distribution. Borneo, Indonesia (Sulawesi).

Notes. Senna [*in litteris*] included this species, described in the genus *Heteroplites* [= *Ischnomerus*], in the genus *Achrionota* because the elytra only have one sutural stria and lack the second.

Lectotype of *H. celebica* (MZUF) designated on a male specimen with the following labels: S-Celebes, Lompa-Battau, 3000', marz 1896, H. Fruhstorfer/ *Heteroplites celebicus*, cotypi! Senna, nunc: *Achrionota*/ ex-coll. Senna/ "La Specola" Firenze, 7042. Paralectotypes: 4 male specimens (MZUF), S-Celebes, Lompa-Battau, 3000', marz 1896, H. Fruhstorfer/ ex-coll. Senna/ "La Specola" Firenze, 7042; 2 male specimens (MZUF), S. Celebes, Bua-

Kraeng, 5000', Febr. 1896, H. Fruhstorfer/ ex-coll. Senna/ "La Specola" Firenze, 7043; 1 male specimen (MSNG), S-Celebes Lompa-Battau 3000', marz 1896, H. Fruhstorfer.

***concolor* Kleine, 1919**

concolor Kleine, 1919e: 130.

Type depositories. SMTD (ST), MNHUB (ST!).

Distribution. Indonesia (Sulawesi).

***spinifer* (Kleine, 1922)**

spinifer Kleine, 1922b: 36 (*Heteroplites*).

Type depository. SMTD (ST).

Distribution. Philippines (Bohol, Leyte, Mindanao, Panaon, Polillo, Siargao).

***Belorhynchus* Berthold, 1827 new placement**

Belorhynchus Berthold, 1827: 384.

Tinoteramocerus Kleine, 1927b: 444; **syn. n.**

Type species: *Curculio nasutus* Fabricius, 1781.

Distribution. Cuba, Dominican Republic, Jamaica.

***nasutus* (Fabricius, 1781)**

nasutus Fabricius, 1781: 182 (*Curculio*).

enormis Kleine, 1927b: 445 (*Tinoteramocerus*); **syn. n.**

Type depositories. *C. nasutus*: NHM (Banks coll.) (HT!); *T. enormis*: NHM (HT!).

Distribution. Jamaica.

Notes. According to Schoenherr (1833: 335) *Curculio nasutus* Fabricius belongs to the genus *Belopherus*. Schoenherr (1833: 336 and 341) also states that the illustrations of *Brentus nasutus* by Olivier (1808, pl. 2, fig. 7a and 7b) represent two different species: fig. 7a is *Belopherus nasutus*; fig. 7b is *Brentus myrmecophaga* Herbst. Schönfeldt (1908: 45) included *nasutus* Fabricius in *Belopherus* and placed *nasutus* Olivier (non Fabricius) among the synonyms of *Tychaeus curvidens* Lund (= *myrmecophaga* Herbst); Kleine (1938d: 121 and 124) likewise.

According to Damoiseau (1962b: 28 and 31) *C. nasutus* Fabricius should be placed among the “*incertae sedis*” species and not in the genus *Belopherus*, but he was unable to study the type specimen. During our stay at NHM, in Banks collection we found the Fabricius type for *C. nasutus* which is the type species of the genus *Belorhynchus* (Latreille, 1828: 593).

After studying the type material, we also consider *Tinoteramocerus enormis* Kleine to be a synonym for *C. nasutus*.

***stratiorrhinoides* (Kleine, 1942) n. comb.**

stratiorrhinoides Kleine, 1942: 131 (*Tinoteramocerus*).

Type depository. MZSF (HT).

Distribution. Cuba.

***Bulbogaster* Lacordaire, 1866**

Bulbogaster Lacordaire, 1866: 467.

Bolbogaster Gemminger & Harold, 1872: 2719 [unjustified emendation].

Type species: *Bulbogaster ctenostomoides* Lacordaire, 1866.

Distribution. Fiji Is., Vanuatu [= New Hebrides].

***ctenostomoides* Lacordaire, 1866**

ctenostomoides Lacordaire, 1866: 467.

Type depository. IRSNB (LT!, PLT!).

Distribution. Fiji Is.

Notes. Lectotype designated by Damoiseau (1989b: 65).

hebridarum (Fairmaire, 1878)

hebridarum Fairmaire, 1878: 282 (*Bolbogaster*).

Type depository. MNHN (HT!).

Distribution. Vanuatu [= New Hebrides].

juncea Damoiseau, 1989

juncea Damoiseau, 1989b: 65.

Type depositories. DSIRW (HT, AT, PT), IRSNB (PT!).

Distribution. Fiji Is.

Ceocephalus Guérin-Méneville, 1833

Ceocephalus Guérin-Méneville, 1833: 139.

Diurus Dejean, 1834: 244; syn. Alonso-Zarazaga & Lyal, 1999: 54.

Diuris: Westwood, 1847: 31 [lapsus].

Fornicaria Gistel, 1848: XI; syn. Alonso-Zarazaga & Lyal, 1999: 54.

Diurus Pascoe, 1862: 392 (non Dejean, 1834); syn. Sforzi & Bartolozzi, 1997: 174.

Type species: *Ceocephalus furcillatus* Guérin-Méneville, 1833.

Distribution. Taiwan; Borneo, India (including Andaman Is., Assam, Nicobar Is.), Indonesia (Batu; Java; Moluccas: Aru; Sumatra), Laos, Malaysia, Myanmar [= Burma], Philippines (Basilan, Luzon, Mindanao, Mindoro, Samar, Tawi Tawi), Thailand, Vietnam; Caroline Is., New Guinea, Solomon Is.

albopilosus (Kleine, 1926)

albopilosus Kleine, 1926c: 174 (*Diurus*).

Type depository. NHM (HT!).

Distribution. Malaysia.

ambiguus (Senna, 1911)

ambiguus Senna, 1911: 37 (*Diurus*).

Type depository. MZUF (LT!, PLT!).

Distribution. Borneo.

Notes. Lectotype designated by Bartolozzi *et al.* (1985: 94) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

antennatus (Ritsema, 1882)

antennatus Ritsema, 1882b: 214 (*Diurus*).

sphacelatus Pascoe, 1887: 19 (*Diurus*); syn. Sforzi & Bartolozzi, 1997: 177.

attenuatus: Kleine, 1921c: 49 (*Diurus*) [lapsus].

sphacellatus: Kleine, 1926c: 173 (*Diurus*) [lapsus].

Type depositories. *D. antennatus*: RMNH (HT!); *D. sphacelatus*: NHM (LT!, PLT!).

Distribution. Taiwan; Borneo, India (including Andaman Is., Nicobar Is.), Indonesia (Java, Sumatra), Laos, Malaysia, Myanmar [= Burma], Thailand, Vietnam.

Notes. Lectotype of *D. sphacelatus* designated by Sforzi & Bartolozzi (1997: 177).

articulatus (Senna, 1911)

articulatus Senna, 1911: 38 (*Diurus*).

Type depository. MZUF (HT!, AT!).

Distribution. Borneo, Malaysia, Vietnam.

celsus (Kleine, 1923)

celsus Kleine, 1923i: 274 (*Diurus*).

eelsus: Kleine, 1938c: 144 (*Diurus*) [lapsus].

Type depository. NHM (LT!, PLT!).

Distribution. Malaysia.

Notes. Lectotype of *D. celsus* (NHM) designated on a male specimen with the following labels: Syntype/ Typus! ♂/ Penang, Lamb. 93.60/ *Diurus celsus* Kln., R. Kleine det., 1922. Paralectotypes: 1 male and 1 female specimen (NHM), Syntype/ Gunong Angsi, Negri Sembilan, 2000-2790', April 1918/ *Diurus celsus* Kln; 1 female specimen (NHM), Syntype/ Type H. T./ Typus!/ Malay Pen., H.C. Robinson, 1907-48/ Gunong Tahan, Pahang, 2500-3500 ft., May-July 1905/ *Diurus celsus* n. sp., R. Kleine det.

***completus* (Kleine, 1923)**

completus Kleine, 1923i: 276 (*Diurus*).

Type depositories. NHM (LT!, PLT!), ZRC (PLT).

Distribution. Borneo, Indonesia (Java), Malaysia.

Notes. Lectotype of *D. completus* (NHM) designated on a male specimen with the following labels: Syntype/ Typus! / Penang/ Penang, Lamb. 93.60/ *Diurus completus* Kln., R. Kleine det., 1922. Paralectotypes: 1 female specimen (NHM), Syntype/ Type H. P./ Typus!/ Penang/ Penang Lamb 93-60/ *Diurus completus* Kln, R. Kleine det. 1922/ not ♀ of *completus* but a large ♀ of *forcipatus* Westw. D. J. Atkinson det. 1951 from key: descr.; 1 female specimen (NHM), Syntype/ Malay Pen., H. C. Robinson, 1907-48/ Gunong Tahan, Pahang, 2500-3500 ft., May-July 1905/ *Diurus completus* Kln R. Kleine det. 1922/ *forcipatus* Westw. D. J. Atkinson det. 1951; 1 female specimen (NHM), Syntype/ Sing./ Baly/ Bowring 63-47/ *Diurus completus* Kln, R. Kleine det. 1922/ *forcipatus* Westw. not *completus* D. J. Atkinson det. 1951.

***compressicauda* (Fairmaire, 1881)**

compressicauda Fairmaire, 1881a: 349 (*Diurus*).

Type depository. Unknown.

Distribution. Caroline Is.

***deruptus* (Kleine, 1923)**

deruptus Kleine, 1923i: 274 (*Diurus*).

Type depository. NHM (LT!, PLT!).

Distribution. Borneo, Malaysia.

Notes. Lectotype of *D. deruptus* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ Sarawak, June, 1892/ *Diurus deruptus* Kln, Typus! det. auctore/ Brit. Mus. 1923-88. Paralectotype: 1 female specimen (NHM), Syntype/ Co-type/ Sarawak, June 1892/ Brit. Mus. 1923-88.

***enricae* Sforzi & Bartolozzi, 1997**

enricae Sforzi & Bartolozzi, 1997: 181.

Type depository. NHM (HT!).

Distribution. Borneo.

***erythropus* (Ritsema, 1882)**

erythropus Ritsema, 1882b: 210 (*Diurus*).

Type depository. RMNH (HT!).

Distribution. Indonesia (Batu, Sumatra).

***filicauda* (Senna, 1911)**

filicauda Senna, 1911: 47 (*Diurus*).

Type depository. MZUF (LT!, PLT!).

Distribution. Borneo, India (Andaman Is.) Indonesia (Java, Sumatra), Malaysia, Philippines (Luzon)(?).

Notes. Lectotype designated by Bartolozzi *et al.* (1985: 96) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

***forcipatus* (Westwood, 1848)**

forcipatus Westwood, 1848: 31 (*Diurus*).

furcillatus Pascoe, 1862: 393 (non Guérin-Ménéville, 1833) (*Diurus*); syn. Senna, 1892c: 63.

Type depository. OXUM (HT).

Distribution. Borneo, India (Assam), Indonesia (Java, Sumatra), Malaysia.

forficuloides (K. E. Schedl, 1959)

forficuloides K. E. Schedl, 1959: 65 (*Diurus*).

Type depository. MVMA (HT).

Distribution. Solomon Is.

Notes. According to K. E. Schedl (1959: 65) a paratype was in his private collection, but W. Schedl (2001, personal communication) did not find it in his father's collection.

furcillatus Guérin-Ménéville, 1833

furcillatus Guérin-Ménéville, 1833: pl. 36, fig. 8.

dispar Pascoe, 1862: 393 (*Diurus*); syn. *Senna*, 1892c: 63.

turcillatus: Gyllenhal (in Schoenherr, 1833: 359) (*Ceocephalus*) [lapsus].

furcellatus: Wallace, 1874: 37 (*Diurus*) [lapsus].

Type depositories. *C. furcillatus*: MNHN (HT!); *D. dispar*: NHM (ST!).

Distribution. Borneo, Indonesia (Java, Sumatra), Malaysia, Philippines (Luzon, Mindanao, Mindoro, Samar, Tawi Tawi), Thailand.

griseus (Kleine, 1922)

griseus Kleine, 1922k: 173 (*Diurus*).

Type depository. SMTD (HT!).

Distribution. Indonesia (Moluccas: Aru); New Guinea.

indianus (Kleine, 1944)

indianus Kleine, 1944: 155 (*Diurus*).

Type depository. MNHN (HT!).

Distribution. S India.

intermedius (Senna, 1911)

intermedius Senna, 1911: 41 (*Diurus*).

Type depository. MZUF (HT!).

Distribution. Borneo.

nigripes (Kleine, 1926)

nigripes Kleine, 1926a: 57 (*Diurus*).

Type depositories. Unknown (HT), NHM (AT!).

Distribution. Indonesia (Sumatra), Malaysia.

Notes. According to Kleine (1926a), the holotype for this species was located in the Stettin Museum. After World War II, most of the Museum's material was transferred to MZPW. However, the collection manager T. Huflejt (MZPW) (2002, personal communication) could not find the *D. nigripes* holotype there.

ominosus (Senna, 1892)

ominosus Senna, 1892e: 492 (*Diurus*).

Type depository. MSNG (HT!).

Distribution. Myanmar [= Burma].

philippinicus (Senna, 1911)

philippinicus Senna, 1911: 45 (*Diurus*).

grootaerti Damoiseau, 1989b: 66 (*Diurus*); **syn. n.**

Type depositories. *D. philippinicus*: MZUF (LT!, PLT!); *D. grootaerti*: IRSNB (HT!).

Distribution. Philippines (Luzon); New Guinea.

Notes. Lectotype of *D. philippinicus* (MZUF) designated on a male specimen with the following labels: N. Luzon, Is. Filippine, Whitehead '94/ *Diurus philippinicus*, cotypi ♂ ♀, Senna/ Bull. Soc. Ent. It., XLI, 1909, cotypi ♂ ♂ ♀ ♀, ex-coll. Senna/ "La Specola" Firenze, 7054. Paralectotypes: 1 female (MZUF), N. Luzon, Is. Filippine, Whitehead '94/ *Diurus philippinicus* ♀/ *Diurus philippinicus* Senna, R. Damoiseau det.; 2 male and 6 female specimens (MZUF), N. Luzon, Whitehead, '94/ ex-coll. Senna/ "La Specola" Firenze, 7054.

After examination of the type material, we state that *C. grootaerti* and *C. philippinicus* are synonyms.

poultoni (Senna, 1911)

poultoni Senna, 1911: 44 (*Diurus*).

Type depository. MZUF (LT!, PLT!).

Distribution. Borneo

Notes. Lectotype of *D. poultoni* (MZUF) designated on a male specimen with the following labels: N. W. Borneo/ Kuching, Capt. July 13.99 by Dyak coll. Pres. 1900 by R. Shelford/ *Diurus poultoni* Senna, cotypi! ♂ ♂ ♀/ Bull. Soc. Ent. It. XLI, 1909, cotypi! ♂ ♂ ♀/ ex-coll. Senna/ "La Specola" Firenze, 7053. Paralectotypes: 1 male specimen (MZUF), N. W. Borneo/ Kuching, Capt. Jan. 25.00 by Dyak coll. Pres. 1900 by R. Shelford/ ex-coll. Senna/ "La Specola" Firenze, 7052; 1 male specimen (MZUF), N. W. Borneo/ Kuching, Capt. July 13.99 by Dyak coll. Pres. 1900 by R. Shelford/ *Diurus poultoni* Senna, R. Damoiseau det./ ex-coll. Senna/ "La Specola" Firenze, 7053; 1 female specimen (MZUF), N. W. Borneo/ Kuching, Capt. July 13.99 by Dyak coll. Pres. 1900 by R. Shelford/ *Diurus poultoni* Senna, ♀ typus!, R. Damoiseau det./ ex-coll. Senna/ "La Specola" Firenze, 7052.

samarensis (Kleine, 1926)

samarensis Kleine, 1926f: 370 (*Diurus*).

Type depository. SMTD (HT!).

Distribution. Philippines (Samar).

sennai Sforzi & Bartolozzi, 1997

sennai Sforzi & Bartolozzi, 1997: 185.

Type depository. MNHN (HT!).

Distribution. Borneo.

shelfordi (Senna, 1902)

shelfordi Senna, 1902: 279 (*Diurus*).

Type depositories. OXUM (LT!), MZUF (PLT!).

Distribution. Borneo, Philippines (Basilan, Luzon, Mindanao).

Notes. Lectotype designated by Damoiseau (1989b: 68); he did not list the paralectotypes held in MZUF.

silvanus (Senna, 1902)

silvanus Senna, 1902: 280 (*Diurus*).

silvanus: Senna, 1902: 246 (*Diurus*) [lapsus].

Type depository. MZUF (HT!).

Distribution. Borneo, Indonesia (Sumatra), Malaysia.

Notes. Kleine (1928a: 89) incorrectly reported that the types are in MSNG and OXUM.

spoetteli (Kleine, 1921)

spoetteli Kleine, 1921: 65 (*Diurus*).

Type depository. SMFD (HT!).

Distribution. Indonesia (Java), Malaysia.

tarsatus (Ritsema, 1882)

tarsatus Ritsema, 1882b: 212 (*Diurus*).

Type depository. RMNH (HT!).

Distribution. Borneo, India (Andaman Is.?).

Eterodiurus Senna, 1911

Eterodiurus Senna, 1911: 51.

Heterodiurus: Kleine, 1927a: 58 [lapsus].

Type species: *Eterodiurus singularis* Senna, 1911.

Distribution. S China; India, Laos, Malaysia, Vietnam.

singularis Senna, 1911

singularis Senna, 1911: 52.

compendarius Kleine, 1925b: 162 (*Diurus*); syn. Sforzi & Bartolozzi, 1993: 296.

Type depositories. *E. singularis*: MZUF (HT!); *D. compendarius*: NHM (LT!).

Distribution. S China; India, Laos, Malaysia, Vietnam.

Notes. Lectotype of *Diurus compendarius* designated by Sforzi & Bartolozzi (1993: 296) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Hovasius Senna, 1895

Hovasius Senna, 1895e: 291.

Type species: *Hovasius alluaudi* Senna, 1895.

Distribution. Madagascar.

alluaudi Senna, 1895

alluaudi Senna, 1895e: 292.

Type depository. MNHN (HT!).

Distribution. Madagascar.

Ischnomerus Labram & Imhoff, 1838

Ischnomerus Labram & Imhoff, 1838: fasc. 2 [page not numbered].

Heteroplites Lacordaire, 1866: 471; syn. Kleine, 1938d: 135.

Cediocera Pascoe, 1887: 20; syn. Kleine, 1938d: 135.

Gediocera: Kleine, 1925b: 47 [lapsus].

Cedeocera: Sharp, 1936: 297 [lapsus].

Heteroclites: Bartolozzi *et al.*, 1985: 95 [lapsus].

Type species: *Ischnomerus erythroderes* Labram & Imhoff, 1838.

Distribution. S China; Borneo, India (including Andaman Is.), Indonesia (Engano, Java, Mentawai, Sumatra), Malaysia, Philippines (Basilan, Luzon, Mindanao, Negros, Palawan, Tawi Tawi), Vietnam; New Guinea.

erythroderes Labram & Imhoff, 1838

erythroderes Labram & Imhoff, 1838: fasc. 2 [page not numbered].

erythroderes Boheman (in Schoenherr, 1840: 564) (non Labram & Imhoff, 1838) (*Teramocerus*); syn. Kleine, 1938d: 135.

Type depositories. *I. erythroderes*: unknown; *T. erythroderes*: NHRS (ST).

Distribution. Philippines (Luzon, Mindanao, Palawan).

longicornis (Pascoe, 1887)

longicornis Pascoe, 1887: 20 (*Cediocera*).

Type depository. NHM (LT!, PLT!).

Distribution. S China; India (including Andaman Is.), Indonesia (Mentawai, Sumatra), Malaysia, Vietnam.

Notes. Lectotype of *C. longicornis* (NHM) designated on a male specimen with the following labels: Syntype/ Type H. T./ Andaman/ Pascoe Coll. 93-60/ *Cediocera longicornis* Pasc. Typus. Paralectotypes: 1 male and 2 female specimens (NHM), with the same label data as the lectotype.

tristis (Senna, 1892)

tristis Senna, 1892a: 181 (*Cediocera*).

Type depositories. MZUF (LT!, PLT!), MSNG (PLT!), SMTD (PLT), RMNH (PLT).

Distribution. S China; Borneo, India (Andaman Is.), Indonesia (Engano, Java, Sumatra), Malaysia, Philippines (Basilan, Negros, Tawi Tawi); New Guinea.

Notes. Lectotype of *C. tristis* (MZUF) designated on a male specimen with the following labels: Giava occ., J. D. Pasteur/ *Cediocera tristis* Senna, 1892, cotypi!/ Notes Leyden Mus. XIV, 1892, cotypi ♂♂ ♀♀/ ex-coll. Senna/ "La Specola" Firenze, 7039. Paralectotypes: 15 male and 13 female specimens (MZUF), Giava occ., J. D. Pasteur/ ex-coll. Senna/ "La Specola" Firenze, 7039; 3 male and 3 female specimens (MZUF), Engano, Bua Bua, V-VII.1891, Modigliani/ ex-coll. Senna/ "La Specola" Firenze, 7040; 4 male and 1 female specimens (MZUF), Perak, d. Staudinger/ ex-coll. Senna/ "La Specola" Firenze, 7041.

Ithystenomorphus Kleine, 1919

Ithystenomorphus Kleine, 1919e: 121.

Type species: *Ithystenomorphus femoralis* Kleine, 1919.

Distribution. New Guinea.

femoralis Kleine, 1919

femoralis Kleine, 1919e: 123.

Type depositories. SMTD (ST), ZMAN (?) (ST).

Distribution. New Guinea.

Notes. According to Kleine (1919e: 124) syntypes were in SMTD and ZMAN, but we did not find any syntype in ZMAN.

Ithystenus Pascoe, 1862

Ithystenus Pascoe, 1862: 390.

Leptorhynchus Boisduval, 1835b: 318 (non Clift, 1828); syn. Alonso-Zarazaga & Lyal, 1999: 54.

Leptorynchus: Boisduval, 1835b: 318 [lapsus].

Leptorhynchus Guérin-Méneville, 1838: 110 (non Clift, 1828, nec Du Bus, 1835, nec Boisduval, 1835, nec Ménetriés, 1835); syn. Schönfeldt, 1908: 74.

Leptorrhynchus: Gemminger & Harold, 1872: 2719 [lapsus].

Ithistenus: Senna, 1892b: 33 [lapsus].

Coptorhynchus: Desbrochers des Loges, 1892: 109 (non Guérin-Méneville, 1841) [lapsus].

Ithsytenus: K. E. Schedl, 1959: 63 [lapsus].

Itystenus: Döbler & Gaedike, 1995: 449 [lapsus].

Isthystenus: Döbler & Gaedike, 1995: 451 [lapsus].

Type species: *Brenthus angustatus* Guérin-Méneville, 1831.

Distribution. Indonesia (Moluccas: Ambon, Aru, Bacan [= Batjan], Buru, Halmahera [= Gilolo], Kay, Larat, Obi, Seram [= Ceram]; Misool); Australia, New Caledonia, New Guinea (including Bismarck Archipelago: New Britain; Woodlark), Polynesia, Solomon Is. (Shortlands), Vanuatu [= New Hebrides].

Notes. We were able to examine the labels handwritten by Desbrochers des Loges (MNHN and IRSNB) and we noted that this author had included his new species *francoisi* in the genus *Leptorhynchus*, thus the name *Coptorhynchus* that appears in the species description (1892: 109) would seem to be a lapsus, as Alonso-Zarazaga & Lyal (1999: 54) suggested.

adoptivus Kleine, 1919

adoptivus Kleine, 1919e: 121.

Type depositories. MNHUB (LT!), SMTD (PLT), MZPW (PLT).

Distribution. New Guinea.

Notes. Lectotype of *I. adoptivus* (MNHUB) designated on a male specimen with the following labels: Sattelberg/ typus!! 117471/ *Ithystenus adoptivus* Kl.

alatus Kleine, 1919

alatus Kleine, 1919e: 102.

Type depository. DEI (LT!, PLT!).

Distribution. New Guinea.

Notes. Lectotype of *I. alatus* (DEI) designated on a male specimen with the following labels: Typus!/ Syntypus/ D. N. Guinea, Kani- Geb./ Coll. Bennigsen/ *Ithystenus alatus* Kleine/ Kleine det./ Dtsch. Entomol. Institut Berlin. Paralectotype: 1 male specimen (DEI), D. N. Guinea, Kani- Geb./ Syntypus/ Coll. Bennigsen/ Kleine det./ Dtsch. Entomol. Institut Berlin.

angustatus Guérin-Méneville, 1831

angustatus Guérin-Méneville, 1831: 111 (*Brenthus*).

Type depository. MNHN (?) (HT).

Distribution. Indonesia (Moluccas: Aru, Bacan [= Batjan], Buru; Kay, Misool); New Guinea.

Notes. According to Kleine (1928a: 88) the type was in MNHN, but we did not find it there.

appendiculatus Kleine, 1919

appendiculatus Kleine, 1919e: 104.

appendicalatus: Kleine, 1926b: 249 (*Ithystenus*) [lapsus].

Type depositories. MNHUB (LT!, PLT!), SMTD (PLT).

Distribution. New Guinea.

Notes. Lectotype of *I. appendiculatus* (MNHUB) designated on a male specimen with the following labels: D. N. Guinea 212, Flusslager 18, 160 m, 25-27.XI.12, Kais. Augustafl. Exp. Bürgers S. G./ 116999/ 18/ typus!/ *Ithystenus appendiculatus* Kln. Paralectotypes: 1 female specimen (MNHUB), D. N. Guinea 128, Quellag, 13-16.VIII.12, Kais. Augustafl. Exp. Bürgers S. G./ 116998/ 17/ typus; 1 female specimen (MNHUB), D. N. Guinea 209, Etappenberg, 16-18.IX.12, Kais. Augustafl. Exp. Bürgers S. G./ 117001/ 20/ typus; 9 male and 7 female specimens (MNHUB), D. N. Guinea 1913, Hauptlg. b. Malu I-II., Kais. Augustafl. Exp. Bürgers S. G.; 2 male specimens (MNHUB), D. N. Guinea 308, Hauptlg. b. Malu, 29.III.13, Kais. Augustafl. Exp. Bürgers S. G.; 1 male specimen (MNHUB), D. N. Guinea 308, Hauptlg. b. Malu, 31.III.13, Kais. Augustafl. Exp. Bürgers S. G.; 1 female specimen (MNHUB), D. N. Guinea 243, Hauptlg. b. Malu, 20.I.13, Kais. Augustafl. Exp. Bürgers S. G.; 1 male and 5 female specimens (MNHUB), D. N. Guinea 253, Hauptlg. b. Malu, 2.II.13, Kais. Augustafl. Exp. Bürgers S. G.; 1 male

specimen (MNHUB), D. N. Guinea 254, Hauptlg. b. Malu, 3.II.13, Kais. Augustafl. Exp. Bürgers S. G.; 3 male and 3 female specimens (MNHUB), D. N. Guinea 256, Hauptlg. b. Malu, 6.II.13, Kais. Augustafl. Exp. Bürgers S. G.

***barbirostris* Kleine, 1920**

barbirostris Kleine, 1920d: 69.

Type depository. SMTD (?) (HT).

Distribution. Australia.

Notes. The type should be in SMTD, but according to O. Jaeger (SMTD) (2002, personal communication) it is not in the SMTD collections.

***bicolor* Sforzi & Bartolozzi n. sp.**

bicolor Guérin-Méneville *sensu* Kleine, 1919e: 91 (non *Brenthus bicolor* Guérin-Méneville, 1831).

Type depository. MZUF (HT!, AT!, PT!).

Distribution. Indonesia (Bacan [= Batjan]); New Guinea (including Bismarck Archipelago: New Britain).

Notes. Kleine (1938d: 130) listed the species *bicolor* Guérin-Méneville in the genus *Ithystenus*; he also listed (1938d: 128) a *Phocylides bicolor* Guérin-Méneville. According to Kleine (1919e: 133) the species illustrated and described by Guérin-Méneville was pars *Phocylides* and pars *Ithystenus*. We believe he was wrong; in our opinion both the drawings (Guérin-Méneville, 1831: pl. VI, figs. 11 and 11a) and the description (Guérin-Méneville, 1838: 109) clearly refer to *Phocylides*. We validate the name *bicolor* (*sensu* Kleine) for this species of *Ithystenus* and designate the following type specimens: Holotype ♂ (MZUF), Andai, N. G., 1892, W. Doherty, ex-coll. Senna; Allotype ♀ and Paratype ♀ (MZUF), with the same label data as the holotype. For the description of the new species check Kleine (1919e: 91).

***bistriatus* Kleine, 1919**

bistriatus Kleine, 1919e: 107.

Type depository. MNHUB (HT!).

Distribution. New Guinea.

caudatus Kleine, 1919

caudatus Kleine, 1919e: 100.

Type depository. MNHUB (LT!, PLT!).

Distribution. Papua New Guinea (Bismarck Archipelago).

Notes. Lectotype of *I. caudatus* (MNHUB) designated on a male specimen with the following labels: Gazelle Hlbins. Massaiva, 29.XII.07, Prof. Preuss/ 117003/ *Ithystenus caudatus* Kl. Paralectotypes: 3 male specimens (MNHUB), with the same label data as the lectotype.

chevrolatii (Boisduval, 1835)

chevrolatii Boisduval, 1835b: 313 (*Brenthus*).

chevrolati: Kleine, 1928a: 96 (*Ithystenus*) [lapsus].

Type depository. Unknown.

Distribution. Papua New Guinea (Woodlark), Solomon Is.

confluens Kleine, 1919

confluens Kleine, 1919e: 119.

Type depositories. MNHUB (ST), SMTD (ST).

Distribution. New Guinea (including Bismarck Archipelago: New Britain).

cultellatus Kleine, 1919

cultellatus Kleine, 1919e: 96.

Type depositories. MNHUB (LT!, PLT!), SMTD (PLT).

Distribution. New Guinea, Solomon Is.

Notes. Lectotype of *I. cultellatus* (MNHUB) designated on a male specimen with the following labels: D. N. Guinea 300, Hunsteinspitze, 12.III.13,

Kais. Augustaf. Exp. Bürgers S. G./ 117019/ *Ithystenus cultellatus* Kl. Paralectotypes: 5 male and 4 female specimens (MNHUB), D. N. Guinea, Hunsteinspitze, several dates from 22.II.13 to 12.III.13, Kais. Augustaf. Exp. Bürgers S. G.; 1 female specimen (MNHUB), D. N. Guinea 131, Hunsteinspitze, 1350 m, 22.VIII.12, Kais. Augustaf. Exp. Bürgers S. G./ 117019.

***cupreus* Kleine, 1943**

cupreus Kleine, 1943b: 140.

Type depositories. DEI (LT!), HNHM (PLT), MZPW (PLT).

Distribution. New Guinea.

Notes. Lectotype of *I. cupreus* (DEI) designated on a male specimen with the following labels: D. N. Guinea, Biró, 1899/ Sattelberg Huon Golf./ Syntypus/ *Ithystenus cupreus* Kln., R. Kleine det., 1941/ Dtsch. Entomol. Institut, Berlin.

***curvidens* (Montrouzier, 1855)**

curvidens Montrouzier, 1855: 38 (*Leptorynchus*).

Type depository. IRSNB (HT!).

Distribution. Indonesia (Moluccas: Aru, Bacan [= Batjan], Seram [= Ceram], Halmahera [= Gilolo]); New Guinea (including Bismarck Archipelago: New Britain; Woodlark).

***cyaneiventris* Kleine, 1926**

cyaneiventris Kleine, 1926b: 250.

Type depository. SMTD (HT).

Distribution. New Guinea.

***debilis* Sharp, 1900**

debilis Sharp, 1900: 387.

Type depository. CUMZ (HT).

Distribution. Papua New Guinea (Bismarck Archipelago: New Britain).

decorus Kleine, 1919

decorus Kleine, 1919e: 117.

Type depositories. MNHUB (LT!), SMTD (PLT), MZPW (PLT).

Distribution. New Guinea (including Bismarck Archipelago: New Britain).

Notes. Lectotype of *I. decorus* (MNHUB) designated on a male specimen with the following labels: D. N. Guinea 247, Hauptlg. b. Malu, 27.I.13, Kais. Augustaf. Exp. Bürgers S. G./ 117002/ 21/ typus!/ *Ithystenus decorus* Kl.

densepunctatus Kleine, 1919

densepunctatus Kleine, 1919e: 89.

Type depositories. DEI (LT!, PLT!), MNHUB (PLT!), SMTD (PLT), MZPW (PLT).

Distribution. New Guinea, Solomon Is.

Notes. According to Döbler & Gaedike (1995: 450) some syntypes are in DEI, in coll. Staudinger and in coll. Bang-Haas but they are not given in the original description.

Lectotype of *I. densepunctatus* (DEI) designated on a male specimen with the following labels: Typus!/ Syntypus/ N. Guinea, Sattelberg, v. Bennigsen/ Syntypus/ Kleine det./ *Ithystenus densepunctatus* Kl./ Dtsch. Entomol. Institut Berlin. Paralectotypes: 2 female specimens (DEI), with the same label data as the lectotype; 1 male specimen (MNHUB), N. Guinea, Sattelberg, v. Wiedenfeld s. g./ 117016/ 42/ *Ithystenus densepunctatus* Kl.; 1 male specimen (MNHUB), D. Neu Guinea, Sattelberg 08, Prof. Neuhauss S./ 117016.

diversicolor Kleine, 1930

diversicolor Kleine, 1930b: 409.

Type depository. DEI (HT!, PT!).

Distribution. New Guinea.

forficulatus Kleine, 1931

forficulatus Kleine, 1931a: 97.

Type depository. SMTD (HT).

Distribution. New Guinea.

francoisi (Desbrochers des Loges, 1892)

francoisi Desbrochers des Loges, 1892: 109 (*Coptorhynchus*).

franzoisi: Schönfeldt, 1908: 83 (*Leptorrhynchus*) [lapsus].

Type depositories. MNHN (LT!, PLT!), IRSNB (PLT!).

Distribution. New Caledonia, New Guinea (including Bismarck Archipelago: New Britain), Polynesia, Solomon Is. (Shortlands), Vanuatu [= New Hebrides].

Notes. Lectotype of *C. francoisi* (MNHN) designated on a male specimen with the following labels: Museum Paris, Nouvelles Hebrides, Espiritu Santo, François 767-92/ *Leptorhynchus francoisi* Desbrochers, Espiritu Santo (N.lle Hebr.)/ *Ithystenus francoisi* Desbrochers, R. Damoiseau rev./ prépar. génit. N° 78 D 19, R. Damoiseau det. Paralectotypes: 1 specimen (MNHN), Museum Paris, Nouvelles Hebrides, Espiritu Santo, François 767-92/ *Ithystenus francoisi* Desbr., Kleine det. 1942; 1 female specimen (MNHN), Nuova Guinea/ Cotype; 1 male specimen (IRSNB), *Leptorhynchus francoisi* m. ♂, N. Hébr./ Prépar. génit. No. 79 C 12 R. Damoiseau/ *Ithystenus francoisi* Desbr. R. Damoiseau det. 1979/ Lectotype [*in litteris*]; 1 female specimen, *Leptorhynchus francoisi* m. ♀ Nouv. Hébr./ *Ithystenus francoisi* Desbr. R. Damoiseau det. 1979/ Allotype [*in litteris*].

franklini Kleine, 1925

franklini Kleine, 1925e: 53.

Type depositories. DEI (LT!, PLT!), SMTD (PLT).

Distribution. New Guinea (including Bismarck Archipelago).

Notes. Lectotype of *I. franklini* (DEI) designated on a male specimen with the following labels: D. Neu-Guinea, Wahnes, Franklin Müller/ Syntypus/

Kleine det./ Dtsch. Entomol. Institut Berlin/ *Ithystenus franklini* Typus, Kln., 1924. Paralectotypes: 2 male and 2 female specimens (DEI), with the same label data as the lectotype.

franklinmuelleri Kleine, 1925

franklinmuelleri Kleine, 1925e: 54.

Type depositories. MNHN (LT!, PLT!), DEI (PLT!), IRSNB (PLT!), MNHUB (PLT!), MZUF (PLT!), NHM (PLT!), OXUM (PLT!), CUIC (PLT), MZPW (PLT), SMTD (PLT).

Distribution. New Guinea.

Notes. Lectotype of *I. franklinmuelleri* (MNHN) designated on a male specimen with the following labels: D. Neu-Guinea/ Wahnes Franklin Müller/ Paratype/ *Ithystenus franklin-muelleri* Kleine. Paralectotypes: 10 specimens (MNHN), with the same label data as the lectotype; 81 specimens (DEI), D. Neu-Guinea, Wahnes, Franklin Müller/ Syntype; 3 male and 3 female specimens (MNHUB), D. Neu-Guinea/ Wahnes, Franklin Müller/ Paratypus; 2 male specimens (MNHUB), D. Neu-Guinea/ Wahnes Franklin Müller/ *Ithystenus franklin-muelleri* Kl., cotypus; 2 male and 1 female specimens (OXUM), D. Neu-Guinea, Wahnes, Franklin Müller/ Paratypus/ Rec. in exch fr. W. Horn 1925; 8 specimens (NHM), D. Neu-Guinea, Wahnes, Franklin Müller/ Paratypus/ Rec. in exch fr. W. Horn 1925; 100 specimens (IRSNB), D. Neu-Guinea/ Wahnes Franklin Müller/ Paratype/ *Ithystenus franklinmuelleri* Kl.; 1 male specimen (MZUF), D. Neu-Guinea/ Wahnes Franklin Müller/ "La Specola" Firenze, 12351; 1 female (MZUF), D. Neu-Guinea/ Wahnes Franklin Müller/ "La Specola" Firenze, 12352.

frontalis Pascoe, 1862

frontalis Pascoe, 1862: 391.

Type depository. NHM (HT!).

Distribution. Indonesia (Moluccas: Aru, Bacan [= Batjan], Kay, Seram [= Ceram]); Australia, New Guinea (including Bismarck Archipelago: New Britain), Polynesia.

fumosus Pascoe, 1862

fumosus Pascoe, 1862: 391.

Type depository. NHM (LT!, PLT!).

Distribution. Indonesia (Moluccas: Bacan [= Batjan]).

Notes. Lectotype of *I. fumosus* (NHM) designated on a male specimen with the following labels: Syntype/ Type H. T./ Batchian/ Pascoe Coll. 93-60/ *fumosus* Typus/ *Ithystenus fumosus* Pasc. Paralectotypes: 3 male and 1 female specimens (NHM), with the same label data as the lectotype.

furvus Kleine, 1925

furvus Kleine, 1925f: 26.

Type depository. ZMAN (HT!).

Distribution. Indonesia (Moluccas: Buru).

hebridarum Senna, 1897

hebridarum Senna, 1897a: 235.

Type depository. IRSNB (HT!).

Distribution. Solomon Is., Vanuatu [= New Hebrides].

hollandiae (Boisduval, 1835)

hollandiae Boisduval, 1835b: 315 (*Brenthus*).

Type depository. Unknown.

Distribution. Australia, New Guinea, Solomon Is.

Notes. The original description gives the type depository as coll. M. Dupont; Kleine (1928a: 88) indicates IRSNB, but we did not find this type there.

impar Kleine, 1943

impar Kleine, 1943b: 141.

Type depositories. DEI (LT!), NHM (PLT!), HNHM (PLT), MZPW (PLT).

Distribution. New Guinea.

Notes. Lectotype of *I. impar* (DEI) designated on a male specimen with the following labels: Nuova Guinea, Biró 1900/ Sattelberg, Huon-Golf./ Syn-typus/ *Ithystenus impar* Kln., R. Kleine det., 1942.

leveri Kleine, 1939

leveri Kleine, 1939e: 587.

Type depository. NHM (LT!).

Distribution. Solomon Is.

Notes. Lectotype of *I. leveri* (NHM) designated on a male specimen with the following labels: British Solomons, 1934, K. J. A. W. Lewer/ Pres. by Imp. Inst. Ent. B.M. 1937-559/ *Ithystenus leveri* Kln, R. Kleine det. 1937.

This species was described on a male and a female specimen, we were not able to find the female specimen in NHM.

linearis Pascoe, 1862

linearis Pascoe, 1862: 391.

Type depository. NHM (HT!).

Distribution. Indonesia (Moluccas: Aru, Bacan [= Batjan], Buru, Halmahera [= Gilolo], Obi, Seram [= Ceram]); New Guinea.

mocsaryi (Bolkay, 1910)

mocsaryi Bolkay, 1910b: 185 (*Leptorhynchus*).

Type depository. HNHM (ST).

Distribution. Indonesia (Moluccas: Halmahera [= Gilolo]).

muelleri Kleine, 1925

muelleri Kleine, 1925e: 53.

Type depository. DEI (HT!).

Distribution. Fiji Is., New Guinea, Vanuatu [= New Hebrides].

nigrosulcatus Fairmaire, 1881

nigrosulcatus Fairmaire, 1881c: 421.

Type depository. MNHN (HT!).

Distribution. Fiji Is., Vanuatu [= New Hebrides].

noonadani Damoiseau, 1966

noonadani Damoiseau, 1966e: 451.

Type depository. ZMUC (HT).

Distribution. Papua New Guinea (Bismarck Archipelago).

ophiopsis Pascoe, 1862

ophiopsis Pascoe, 1862: 391.

Type depository. NHM (HT!).

Distribution. Indonesia (Moluccas: Aru, Bacan [= Batjan], Seram [= Ceram]); New Guinea.

perlongus Kleine, 1919

perlongus Kleine, 1919e: 105.

Type depositories. DEI (LT!, PLT!), MNHUB (PLT!), SMTD (PLT).

Distribution. New Guinea.

Notes. Lectotype of *I. perlongus* (DEI) designated on a male specimen with the following labels: Mäanderberg 403, 21-30.8.13, Bürgers S. G./ ♂/ Typus/ Syntypus/ Dtsch. Entomol. Institut Berlin/ *Ithystenus perlongus* Kleine, ex typis ♂ ♀. Paralectotypes: 1 female specimen (DEI), D. N. Guinea, Mäanderberg 404, 21-30.VIII.13, Kais. Augustafl. Exp., Bürgers S. G./ ♀/ Syntypus; 17 male and 24 female specimens (MNHUB), D. N. Guinea, Mäanderberg 403, 21-30.VIII.13, Bürgers S. G.; 8 male and 23 female specimens (MNHUB), D. N. Guinea, Mäanderberg 404, 21-30.VIII.13, Kais. Augustafl. Exp., Bürgers S. G.

***pumilus* (Boisduval, 1835)**

pumilus Boisduval, 1835b: 314 (*Brenthus*).

Type depository. Unknown.

Distribution. Papua New Guinea (Bismarck Archipelago: New Britain), Solomon Is.

Notes. Kleine did not mention this species in his *Genera Insectorum* (1938d).

***punctifrons* Kleine, 1919**

punctifrons Kleine, 1919e: 113.

Type depository. SMTD (HT).

Distribution. Indonesia (Moluccas: Buru); New Guinea.

***rugosipunctatus* Kleine, 1935**

rugosipunctatus Kleine, 1935b: 175.

Type depository. SMTD (ST).

Distribution. Solomon Is.

sabulosus Kleine, 1919

sabulosus Kleine, 1919e: 92.

Type depositories. DEI (LT!, PLT!), MNHUB (PLT!), NHMB (PLT!), MZPW (PLT), SMTD (PLT).

Distribution. Indonesia (Moluccas: Aru, Bacan [= Batjan], Seram [= Ceram]); Australia, New Guinea, Vanuatu [= New Hebrides].

Notes. According to Döbler & Gaedike (1995: 453) syntypes are in the Bang-Haas collection, but Kleine (1919e) did not give this information.

Lectotype of *I. sabulosus* (DEI) designated on a male specimen with the following labels: Coen District, Cape York, Queensland, H. Hacker/ Kleine det./ Dtsch. Entomol. Institut Berlin/ Syntypus/ *Leptorrhynchus hollandiae*, Baiad, N. Queensland/ *sabulosus*. Paralectotypes: 1 male specimen (DEI), Coen District, Cape York, Queensland, H. Hacker/ coll. Hacker / Kleine det./ Syntypus; 1 male specimen (DEI), Ceram/ Syntypus/ Kleine det.; 1 male specimen (MNHUB), Aru Inseln, Ureiuning, C. Ribbe, 1884/ cotypus!/ 117463/ *Ithystenus sabulosus* Kl.; 3 male specimens (MNHUB), with the same locality as the lectotype; 3 male specimens (MNHUB), Ceram, Jlo, C. Ribbe, 1884; 1 male specimen (MNHUB), N. Guinea, Andai, XII, Coll. Bruijn, 1875; 1 specimen (NHMB), Arfak, N. Guinea/ cotypus/ 84/ *Ithystenus sabulosus* Kln!; 2 specimens (NHMB), Dtsch. N-Guinea/ cotypus; 1 specimen (NHMB), Aru-Insel, Ureiuning, C. Ribbe 1884/ cotypus!/ *Ithystenus sabulosus* Kln.

similis Kleine, 1919

similis Kleine, 1919e: 98.

Type depository. MNHUB (LT!, PLT!).

Distribution. New Guinea.

Notes. Lectotype of *I. similis* (MNHUB) designated on a male specimen with the following labels: D. N. Guinea 224, Lordberg, 9.XII.12, Kais. Augustafl.[uss] Exp. Bürgers S. G./ 1170009/ 18/ *Ithystenus similis* Kl. ♂ Type. Paralectotypes: 1 male specimen (MNHUB), D. N. Guinea 228, Standlagera, Aprilfl.[uss], 21.XIII.12, Bürgers S. G./ 116996/ 15 *Ithystenus similis* Kl.; 2 male and 2 female specimens (MNHUB), D. N. Guinea 228, Standlagera, Aprilfl.[uss], 21.XIII.12, Bürgers S. G.; 2 female specimens (MNHUB), D. N. Guinea 225, Lordberg, 10.XII.12, Kais. Augustafl.[uss] Exp. Bürgers S. G.; 1 male specimen (MNHUB), D. N. Guinea 223, Lordberg, 8.XII.12, Kais. Augustafl.[uss] Exp. Bürgers S. G.

***spinosus* Kleine, 1919**

spinosus Kleine, 1919e: 69.

Type depositories. MNHUB (LT!), SMTD (PLT), MZPW (PLT).

Distribution. Indonesia (Moluccas: Aru, Halmahera [= Gilolo], Kay, Larat; Misool); New Guinea (including Bismarck Archipelago: New Britain, New Ireland), Solomon Is.

Notes. Lectotype of *I. spinosus* (MNHUB) designated on a male specimen with the following labels: Neu-Meckfenburg, Lamasong, Dr. Krämer S. G./ 117005/ 31/ typus!! *Ithystenus spinosus* Kl.

***unicolor* Kleine, 1919**

unicolor Kleine, 1919e: 74.

Type depository. SMTD (ST).

Distribution. Indonesia (Moluccas: Larat); New Guinea.

***wallacei* Pascoe, 1862**

wallacei Pascoe, 1862: 390.

Type depository. NHM (HT!).

Distribution. Indonesia (Moluccas: Ambon, Aru, Buru, Seram [= Ceram]; Misool); New Guinea.

***Kolbrentus* Alonso-Zarazaga, Lyal,
Bartolozzi & Sforzi, 1999**

Kolbrentus Alonso-Zarazaga, Lyal, Bartolozzi & Sforzi (in: Alonso-Zarazaga & Lyal, 1999: 54)
[replacement name for *Homales*].

Homales Kolbe, 1883c: 80 (non Kolbe, 1883a: 382).

Type species: *Homales glaber* Kolbe, 1883.

Distribution. Madagascar.

Notes. In 1883 Kolbe published a paper where he created the subgenus *Homales* for the genus *Centrophorus* (1883a: 382). In the same year Kolbe created the genus *Homales* in the tribe Ithysteni [sic!] (1883c: 80). In this paper he added a short note (1883c: 79) in which he explained that the spelling *Homales* in the previous paper (1883a: 382) was a misspelling of *Synhomales*. Alonso-Zarazaga *et al.* (1999: 54) gave the new name *Kolbrentus* to the *Homales* of the second publication.

***glaber* (Kolbe, 1883)**

glaber Kolbe, 1883c: 81 (*Homales*).

Type depository. MNHUB (LT!, PLT!).

Distribution. Madagascar.

Notes. Lectotype of *H. glaber* (MNHUB) designated on a male specimen with the following labels: 107093/ n. g. *Arrhenodinorum*/ Type. Paralectotypes: 1 male specimen (MNHUB), C. Madag., Hildbr./ *Homales glaber* Kolbe, Typus; 1 female specimen (MNHUB), Madagascar/ 107093/ Type.

***metallicus* (Kolbe, 1883) n. comb.**

metallicus Kolbe, 1883c: 82 (*Homales*).

Type depository. MNHUB (HT!).

Distribution. Madagascar.

***Lasiorrhynchus* Lacordaire, 1866**

Lasiorrhynchus Lacordaire, 1866: 469.

Lasiorrhynchus: Gemminger & Harold, 1872: 2720 [lapsus].

Type species: *Curculio barbicornis* Fabricius, 1775.

Distribution. New Zealand.

barbicornis (Fabricius, 1775)

barbicornis Fabricius, 1775: 134 (*Curculio*).

assimilis Fabricius, 1775: 134 (*Curculio*); syn. Gemminger & Harold, 1872: 2720.

Type depositories. *C. barbicornis*: NHM (LT!, PLT!), ZMUK (PLT); *C. assimilis*: NHM (LT!), ZMUK (PLT).

Distribution. New Zealand.

Notes. Lectotypes of *C. barbicornis* and *C. assimilis* designated by Kuschel (1970: 194).

Mesetia Blackburn, 1896

Mesetia Blackburn, 1896: 37.

Type species: *Mesetia amoena* Blackburn, 1896.

Distribution. Australia.

amoena Blackburn, 1896

amoena Blackburn, 1896: 37.

Type depository. NHM (HT!).

Distribution. Australia.

Ozodecerus Chevrolat, 1839

Ozodecerus Chevrolat, 1839: 175.

Ozodeocerus Agassiz, 1846b: 269 [unjustified emendation].

Ozodeocerus Gemminger & Harold, 1872: 2719 (non Agassiz, 1846) [unjustified emendation].

Type species: *Ozodecerus forficulatus* Chevrolat, 1839.

Distribution. Madagascar.

***forficulatus* Chevrolat, 1839**

forficulatus Chevrolat, 1839: 176.

forficatus: Kleine, 1927a: 56 (*Ozodecerus*) [lapsus].

Type depository. MNHN (LT!, PLT!).

Distribution. Madagascar.

Notes. Lectotype of *O. forficulatus* (MNHN) designated on a male specimen with the following labels: Madagascar/ Chevrolat T. 322/ type/ ex Musaeo G. Power/ Museum Paris 1952, coll. R. Oberthür/ *Ozodecerus forficulatus* Chevrolat, det R. Damoiseau/ Prép. Génit. 78 D 16. Paralectotypes: 1 male specimen (MNHN), Madagascar/ Chevrolat T. 322 bis/ ex Musaeo G. Power/ Museum Paris 1952 coll. R. Oberthür; 1 female specimen (MNHN), Madagascar/ Chevrolat 327/ ex Musaeo G. Power/ Museum Paris 1952, coll. R. Oberthür.

***pygmaeus* Senna, 1893**

pygmaeus Senna, 1893d: 132.

Type depository. SMFD (von Schönfeldt coll.) (HT).

Distribution. Madagascar.

***rugicollis* Chevrolat, 1839**

rugicollis Chevrolat, 1839: 176.

Type depository. MNHN (LT!, PLT!).

Distribution. Madagascar.

Notes. Lectotype of *O. rugicollis* (MNHN) designated on a male specimen with the following labels: Madagascar/ Chevrolat 324/ type/ ex Musaeo G. Power/ Museum Paris 1952, coll. R. Oberthür/ *Ozodecerus rugicollis* Chevrolat det. R. Damoiseau. Paralectotype: 1 female specimen (MNHN), Madagascar/ Chevrolat 324/ ex Musaeo G. Power/ Museum Paris 1952, coll. R. Oberthür/ Dessin.

***tricuspidatus* Chevrolat, 1839**

tricuspidatus Chevrolat, 1839: 176.

Type depository. MNHN (HT!).

Distribution. Madagascar.

***Plesiophocylides* Kleine, 1925**

Plesiophocylides Kleine, 1925b: 160.

Type species: *Plesiophocylides conditus* Kleine, 1925.

Distribution. India (Assam).

Notes. Kleine (1922m: 163) mentioned this taxon in a list of genera, but he did not describe it until three years later (1925b: 160).

***conditus* Kleine, 1925**

conditus Kleine, 1925b: 161.

conditus: Kleine, 1925b: 161 (*Plesiophocylides*) [lapsus].

Type depository. NHM (HT!).

Distribution. India (Assam).

***Pseudophocylides* Kleine, 1920**

Pseudophocylides Kleine, 1920n: 47.

Type species: *Pseudophocylides insularis* Kleine, 1920.

Distribution. New Guinea.

***clarus* Kleine, 1923**

clarus Kleine, 1923j: 406.

Type depository. DEI (LT!).

Distribution. New Guinea.

Notes. Lectotype of *P. clarus* (DEI) designated on a male specimen with the following labels: Neuguinea/ Syntypus/ Kleine det./ *Pseudophocylides clarus* Kleine.

***insularis* Kleine, 1920**

insularis Kleine, 1920n: 48.

Type depository. ZMAN (LT!, PLT!).

Distribution. New Guinea.

Notes. Lectotype of *P. insularis* (ZMAN) designated on a male specimen with the following labels: Z Nieuw Guinea, Versteeg, 1912.13, 22.XII.12. Paralectotype: 1 female specimen (ZMAN), Z Nieuw Guinea, Versteeg, 1912.13, Pesechen.2.

***Syngenithystenus* Kleine, 1919**

Syngenithystenus Kleine, 1919e: 124.

Syngenithystenus: Kleine, 1927a: 58 [lapsus].

Type species: *Ithystenus nigronitens* Kleine, 1919 = *Leptorhynchus guerinii* Montrouzier, 1855.

Distribution. New Guinea (including Woodlark).

***guerinii* (Montrouzier, 1855)**

guerinii Montrouzier, 1855: 39 (*Leptorynchus*).

guerini: Senna, 1894e: 564 (*Ithystenus*) [lapsus].

nigronitens Kleine, 1919e: 125 (*Syngenithystenus*); syn. Damoiseau, 1966a: 17.

Type depositories. *L. guerinii*: IRSNB (HT!); *S. nigronitens*: MNHUB (LT!, PLT!), SMTD (PLT!).

Distribution. New Guinea (including Woodlark).

Notes. Lectotype of *S. nigronitens* (MNHUB) designated on a male specimen with the following labels: K. Wilhelm Land, Bongu/ typus!/ 117468/ *Syngenithystenus nigronitens* Kleine. Paralectotype: 1 female specimen (MNHUB), Kais. Wilhelmsland, Paup, Dr. Schlaginbaufen/ 1910, 1/ typus/ 117468.

MICROTRACHELIZINI Zimmerman, 1994

Microtrachelizina Zimmerman, 1994: 182.

Microtrachelizini Morrone, 1998: 132.

Aneorhachis Kleine, 1923

Aneorhachis Kleine, 1923d: 135.

Ancorhachis: Kleine, 1931a: 110 [lapsus].

Type species: *Aneorhachis astricta* Kleine, 1923.

Distribution. Bhutan, India.

Notes. Kleine (1922m: 162) mentioned this taxon in a list of genera, but he did not describe it until one year later (1923d: 135).

astricta Kleine, 1923

astricta Kleine, 1923d: 136.

Type depository. NHM (HT!).

Distribution. India.

monticola Damoiseau, 1987

monticola Damoiseau, 1987: 61.

Type depository. MNHN (HT!).

Distribution. Bhutan.

Araiorrhinus Senna, 1893

Araiorrhinus Senna, 1893e: 325.

Araeorrhinus Senna, 1895a: 187 [unjustified emendament].

Araiorrhynchus: Kleine, 1939f: 131 [lapsus].

Araiorrhinus: Neave, 1939: 272 [lapsus].

Arraiorrhinus: Damoiseau, 1963b: 195 [lapsus].

Araiorrhynus: Kabakov, 2001: 208 [lapsus].

Type species: *Araiorrhinus longirostris* Senna, 1893.

Distribution. India, Indonesia (Sumatra, Timor), Malaysia, Myanmar [= Burma], Philippines, Thailand, Vietnam; Australia, New Guinea, Solomon Is.; Europe (I).

armatus Damoiseau, 1987

armatus Damoiseau, 1987: 68.

Type depository. MZUF (HT!).

Distribution. New Guinea.

beesoni Kleine, 1925

beesoni Kleine, 1925b: 138.

becsoni: Kleine, 1923j: 406 (*Araiorrhinus*) [lapsus, nomen nudum].

Type depository. NHM (HT!).

Distribution. India (including Assam), Myanmar [= Burma], Thailand, Vietnam.

Notes. According to Damoiseau (1987: 67) the type was lost, but we found it in NHM.

exportatus Senna, 1893

exportatus Senna, 1893e: 327.

Type depositories. MZUF (LT!), RMNH (PLT!).

Distribution. Indonesia (Sumatra), Malaysia, Philippines; New Guinea; Europe (I).

Notes. Lectotype designated by Bartolozzi *et al.* (1985: 96) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

In his original description Senna only referred to two specimens, one in his collection and one in RMNH. It is hard to understand how Damoiseau (1987: 65) could speak about a third paralectotype in IRSNB.

howittii (Pascoe, 1872)

howittii Pascoe, 1872: 320 (*Trachelizus*).

australicus Senna, 1893e: 327 (*Araiorrhinus*); syn. Damoiseau, 1987: 66.

howetti: Froggatt, 1907: 179 (*Trachelizus*) [lapsus].

howitti: Schönfeldt, 1908: 23 (*Microtrachelizus*) [lapsus].

Type depositories. *T. howittii*: NHM (HT!); *A. australicus*: MZUF (LT!), SMTD (PLT).

Distribution. Australia.

Notes. Senna (1893e: 327) only furnished a short diagnosis of *A. australicus* in a key of the genus *Araiorrhinus*.

Lectotype of *A. australicus* designated by Bartolozzi *et al.* (1985: 94) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

According to Zimmerman (1994: 184) Kleine's quotations (1927a: 27; 1938d: 71) from Malaysia and Sumatra for *A. australicus* are doubtful.

levisulcatus Damoiseau, 1987

levisulcatus Damoiseau, 1987: 64.

Type depository. NHM (HT!).

Distribution. Malaysia.

lieftincki Kleine, 1939

lieftincki Kleine, 1939f: 131.

Type depository. Buitenzorg Museum (?) (HT).

Distribution. Indonesia (Sumatra).

longirostris Senna, 1893

longirostris Senna, 1893e: 326.

Type depository. RMNH (HT!).

Distribution. India (Assam), Indonesia (Sumatra), Malaysia, Philippines; Solomon Is.

Notes. Senna redescribed this species in 1895 (1895a: 187).

recurvicosta Damoiseau, 1966

recurvicosta Damoiseau, 1966e: 423.

Type depository. ZMUC (HT).

Distribution. Philippines (Tawi Tawi).

sondaicus Senna, 1893

sondaicus Senna, 1893e: 329.

Type depositories. MZUF (LT!), RMNH (PLT!), SMTD (PLT).

Distribution. India (Assam), Indonesia (Sumatra), Malaysia, Vietnam; Europe (I).

Notes. Lectotype designated by Bartolozzi *et al.* (1985: 101) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

timoriensis Damoiseau, 1987

timoriensis Damoiseau, 1987: 63.

Type depository. MNHN (HT!).

Distribution. Indonesia (Timor).

Entomopisthius De Muizon, 1959

Entomopisthius De Muizon, 1959: 77.

Type species: *Entomopisthius leleupi* De Muizon, 1959.

Distribution. Democratic Republic of the Congo (ex Zaire).

emarginatus (K. E. Schedl, 1961)

emarginatus K. E. Schedl, 1961c: 199 (*Carcinopisthius*).

perstriatus Damoiseau, 1961a: 284 (*Entomopisthius*); syn. Damoiseau, 1963a: 128.

Type depositories. *C. emarginatus*: MRAC (HT!, PT!); *E. perstriatus*: MRAC (HT!, PT!), IRSNB (PT!).

Distribution. Democratic Republic of the Congo (ex Zaire).

Notes. K. E. Schedl (1961c: 199) reported that paratypes of *C. emarginatus* are in his collection, but W. Schedl (2001, personal communication) did not find them in his father's collection.

leleupi De Muizon, 1959

leleupi De Muizon, 1959: 77.

Type depository. MRAC (HT!).

Distribution. Democratic Republic of the Congo (ex Zaire).

suturalis Damoiseau, 1961

suturalis Damoiseau, 1961a: 285.

Type depository. MRAC (HT!).

Distribution. Democratic Republic of the Congo (ex Zaire).

Notes. In his original description, Damoiseau quotes the holotype in MRAC and does not mention any paratype, but he furnishes more than one measurement for the species. There are two specimens he labelled as paratypes in IRSNB.

Higonius Lewis, 1883

Higonius Lewis, 1883: 299.

Type species: *Higonius poweri* Lewis, 1883.

Distribution. Japan, Taiwan; Bhutan, Borneo, India (including Assam, Darjeeling), Indonesia (Engano; Java; Mentawai; Moluccas: Seram [= Ceram]; Sumatra), Myanmar [= Burma], Philippines (Luzon, Masbate, Mindanao), Sri Lanka, Thailand, Vietnam; New Guinea, Vanuatu [= New Hebrides]; Europe (I).

angustirostris Damoiseau, 1987

angustirostris Damoiseau, 1987: 51.

Type depository. BPBM (HT!).

Distribution. Borneo.

bilobicollis Senna, 1898

bilobicollis Senna, 1898a: 235.

Type depository. MSNG (HT!).

Distribution. Taiwan; Indonesia (Mentawai).

cilo Lewis, 1883

cilo Lewis, 1883: 300.

Type depository. NHM (LT!, PLT!).

Distribution. Japan, Taiwan; Bhutan, India (including Assam, Darjeeling), Myanmar [= Burma], Philippines (Luzon, Masbate), Vietnam.

Notes. Damoiseau designated (1987: 50) the lectotypes in NHM, but the six specimens that we found there have no lectotype labels, only for syntypes.

***crux* Olliff, 1883**

crux Olliff, 1883: 300.

Type depository. NHM (HT!).

Distribution. Borneo, India (Andaman Is., Nicobar Is.), Indonesia (Java; Mentawai; Moluccas: Seram [= Ceram]; Sumatra), Malaysia, Myanmar [= Burma], Vietnam; Europe (I).

***grouvellei* Senna, 1893**

grouvellei Senna, 1893e: 311.

Type depository. MZUF (LT!).

Distribution. Indonesia (Java, Sumatra); Europe (I).

Notes. Lectotype designated by Bartolozzi *et al.* (1985: 97) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

***hirsutus* Senna, 1893**

hirsutus Senna, 1893d: 117.

Type depository. SMFD (von Schönfeldt coll.) (HT).

Distribution. Malaysia.

***niassicus* Senna, 1893**

niassicus Senna, 1893e: 310.

Type depositories. MZUF (LT!), SMTD (PLT).

Distribution. Indonesia (Nias).

Notes. Senna (1893e: 310) furnished a short diagnosis of this species in a key of the genus *Higonius*; later (Senna, 1898b: 380) he wrote that a male

specimen was held in his collection and a female specimen in the Faust collection.

Lectotype designated by Bartolozzi *et al.* (1985: 99) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

***nudus* Kleine, 1920**

nudus Kleine, 1920j: 244.

Type depository. RMNH (LT!, PLT!).

Distribution. Indonesia (Sumatra), Vietnam.

Notes. Damoiseau (1987: 53) reported that the specimens held in RMNH are holotype and paratype, but in his original description Kleine did not designate either of the syntypes as holotype; however, as there are two syntypes in RMNH, and as Damoiseau did not specifically select one as holotype, we designate the lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Lectotype of *H. nudus* (RMNH) designated on a male specimen with the following labels: Sumatra, Manna, M. Knappert. Paralectotype: 1 male specimen (RMNH), with the same label data.

***perpusillus* Kleine, 1944**

perpusillus Kleine, 1944: 152.

Type depositories. MNHN (HT!, PT!), DEI (PT!).

Distribution. Vanuatu [= New Hebrides].

***poweri* Lewis, 1883**

poweri Lewis, 1883: 299.

Type depository. NHM (HT!).

Distribution. Borneo, Indonesia (Batu, Engano, Java, Nias, Sumatra), Malaysia, Myanmar [= Burma], Philippines (Mindanao), Sri Lanka, Thailand, Vietnam; Europe (I).

reconditus Kleine, 1933

reconditus Kleine, 1933e: 322.

Type depository. MZB (?) (coll. Drescher) (HT), NHM (PT).

Distribution. Indonesia (Java).

Notes. According to Damoiseau (1987: 52) the holotype is not in ZMAN but he found a paratype in NHM which is actually a specimen of *Higonius grouvellei*, so Kleine's quotation for Malaysia (1933e: 323) is incorrect. We were unable to locate this specimen in NHM.

trisulcatus Damoiseau, 1987

trisulcatus Damoiseau, 1987: 46.

Type depository. SAMA (HT).

Distribution. New Guinea.

Higonodes Zimmerman, 1994

Higonodes Zimmerman, 1994: 184.

Type species: *Higonius novenarius* Damoiseau, 1987.

Distribution. Indonesia (Moluccas: Aru); Australia.

novenarius (Damoiseau, 1987)

novenarius Damoiseau, 1987: 48 (*Higonius*).

Type depositories. MMA (HT), MNHN (PT).

Distribution. Indonesia (Moluccas: Aru); Australia.

Microtrachelizus Senna, 1893

Microtrachelizus Senna, 1893e: 315.

Ceunonus Kleine, 1922h: 138; syn. Damoiseau, 1963a: 127.

Microtrachelisus: Kabakov, 2001: 210 [lapsus].

Type species: *Trachelizus lyratus* Perroud & Montrouzier, 1865.

Distribution. S China; Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea (including Bioko Is. [= Fernando Poo]), Ethiopia, Ghana, Ivory Coast, Kenya, Madagascar, Nigeria, Principe, Republic of the Congo, Tanzania, Togo; Borneo, India (including Assam, Darjeeling), Indonesia (Engano, Java, Mentawai, Sumatra), Malaysia, Myanmar [= Burma], Philippines (Mindanao, Negros, Samar), Thailand, Vietnam; Australia, New Caledonia, New Guinea; Europe (I).

accomodatus Kleine, 1922

accomodatus Kleine, 1922f: 208.

Type depository. SMTD (HT).

Distribution. Borneo, India (including Assam), Malaysia, Myanmar [= Burma].

aethiopicus Calabresi, 1920

aethiopicus Calabresi, 1920d: 27.

minutus Kleine, 1922h: 139 (*Ceunonus*); syn. Damoiseau, 1963a: 127.

sordidus Kleine, 1922h: 144 (*Microtrachelizus*); syn. Damoiseau, 1967a: 224.

copulatus Kleine, 1924a: 103 (*Microtrachelizus*); syn. Damoiseau, 1967a: 224.

captiosus Kleine, 1924a: 104 (*Microtrachelizus*); syn. Damoiseau, 1967a: 224.

Type depositories. *M. aethiopicus*: MSNG (LT!, PLT!); *C. minutus*: DEI (LT!, PLT!), MNHUB (PLT!); *M. sordidus*: MNHUB (?) (HT); *M. copulatus*: MRAC (ST!); *M. captiosus*: MRAC (ST!), SMTD (ST).

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea (including Bioko Is. [= Fernando Poo]), Ethiopia, Ivory Coast, Kenya, Ghana, Madagascar, Nigeria, Principe, Republic of the Congo, Tanzania, Togo.

Notes. Damoiseau (1967a: 225) reported that the specimens of *M. aethiopicus* held in MSNG are holotype and allotype, but in her original description Calabresi did not designate any of the syntypes as holotype; however, as there is more than one syntype in MSNG, and as Damoiseau did not specifically select one as holotype, we designate the lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Lectotype of *M. aethiopicus* (MSNG) designated on a specimen with the following labels: Is. Principe, Roca Inf., D. Henrique, 100-300 m, IV.1901, L. Fea/ typus/ *Microtrachelizus aethiopicus* Calabr. Typus! Paralectotypes: 2 specimens (MSNG), Is. Principe, Roca Inf., D. Henrique, 100-300 m, IV.1901, L. Fea.

Lectotype of *C. minutus* (DEI) designated on a specimen with the following labels: N Kamerun, L. Conradt, 96/ Syntypus/ Dtsch. Entomol. Institut Berlin/ coll. Kraatz/ Dtsch. Entomol. Institut Berlin Kleine det. Paralectotypes: 8 specimens (DEI) with the same label data as the lectotype; 2 specimens (MNHUB), Togo, Bismarckburg, L. Conradt S./ typus!/ *Microtrachelizus aethiopicus* Calabr., R. Damoiseau det., 1961; 1 specimen (MNHUB), Kamerun, L. Conradt S./ typus!/ *Microtrachelizus aethiopicus* Calabr., R. Damoiseau det., 1961.

According to Kleine (1928a: 79) the type of *M. sordidus* was in MNHUB, but we did not find it there.

Damoiseau (1967a: 225) did not mention Bioko Is. [= Fernando Poo] and Togo (Kleine, 1922h: 140) when he gave the species distribution.

apertus Kleine, 1925

apertus Kleine, 1925b: 140.

Type depositories. NHM (LT, PLT!), SMTD (PLT), IFRI (PLT).

Distribution. S China; Borneo, India (including Assam, Darjeeling), Indonesia (Java, Sumatra), Malaysia, Thailand, Vietnam.

Notes. The species was redescribed by Kleine a year later (1926a: 33).

Lectotype designated by Damoiseau (1987: 57). We did not find the lectotype labelled by Damoiseau in NHM, only a paralectotype with the following labels: syntype/ type/ typus!/ ex *Dalbergia* Assam/ U. Dihing Res., Lakhimpur, Assam, 5.VI.1921, C. F. C. Beeson/ *Microtrachelizus apertus* Kleine, R. Kleine det.

***attritus* Kleine, 1923**

attritus Kleine, 1923d: 136.

Type depository. NHM (HT!).

Distribution. India.

***beneficus* Kleine, 1925**

beneficus Kleine, 1925b: 139.

Type depositories. NHM (LT!), SMTD (PLT), IFRI (PLT) (?).

Distribution. Borneo, India (including Assam), Vietnam.

Notes. Species description was based on ten specimens (IFRI). We found one of these in NHM, and so we designate this specimen as lectotype.

Lectotype of *M. beneficus* (NHM) designated on a specimen with the following labels: Syntype/ Typus!! ex *Shorea assamica*/ Margherata, Lakhimpur Div: Assam, 24.XI.1921, Dy. R. Coll./ India B. M. 1931-82/ R. R. D. 934, B. C. R. 75, Cage 143/ *Microtrachelizus beneficus* Kln., Typus!! *Microtrachelizus beneficus* Kleine.

***bhamoensis* (Senna, 1892)**

bhamoensis Senna, 1892e: 456 (*Trachelizus*).

bhamoënsis: Senna, 1894e: 557 (*Microtrachelizus*) [lapsus].

occultus Kleine, 1935e: 307 (*Microtrachelizus*); syn. Damoiseau, 1987: 56.

Type depositories. *T. bhamoensis*: MSNG (HT!); *M. occultus*: NHM (ST!), DEI (ST!).

Distribution. Indonesia (Sumatra), Myanmar [= Burma], Vietnam (?); New Guinea; Europe (I).

Notes. According to Döbler & Gaedike (1995: 450) two paratypes of *M. occultus* are in DEI, but Kleine did not designate any of the syntypes as holotype and paratypes. The original series consist of five specimens, but we found five specimens labelled as "syntypes" in NHM and two others in DEI.

borneensis Damoiseau, 1987

borneensis Damoiseau, 1987: 56.

Type depository. NHM (HT!).

Distribution. Borneo.

Notes. Damoiseau (1987: 55) did not describe this species in full, but its main characters were included in a key, so the species is valid.

brevisulcatus Senna, 1894

brevisulcatus Senna, 1894e: 558.

Type depositories. MZUF (LT!), MSNG (PLT!).

Distribution. New Guinea.

Notes. Lectotype designated by Bartolozzi *et al.* (1985: 95) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

brevitibia (Senna, 1892)

brevitibia Senna, 1892e: 454 (*Trachelizus*).

Type depository. MSNG (HT!).

Distribution. Myanmar [= Burma]; New Guinea.

contiguus (Senna, 1893)

contiguus Senna, 1893b: 266 (*Trachelizus*).

Type depositories. MZUF (LT!), MSNG (PLT!).

Distribution. India, Indonesia (Engano, Java), Malaysia, Thailand.

Notes. Lectotype designated by Bartolozzi *et al.* (1985: 95) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

***coomani* Damoiseau, 1987**

coomani Damoiseau, 1987: 56.

Type depository. MNHN (HT!, AT).

Distribution. Vietnam.

Notes. Damoiseau (1987: 56) did not describe this species in full, but its main characters were included in a key, so the species is valid. We did not find the allotype in MNHN.

***costatus* Damoiseau, 1987**

costatus Damoiseau, 1987: 56.

Type depository. MZPW (HT).

Distribution. Indonesia (Sumatra).

Notes. Damoiseau (1987: 55) did not describe this species in full, but its main characters were included in a key, so the species is valid.

***cylindricornis* (Power, 1880)**

cylindricornis Power, 1880: 187 (*Trachelizus*).

cylindriformis: Donckier de Donceel, 1884: 3 (*Trachelizus*) [lapsus].

reductus Kleine, 1941b: 228 (*Microtrachelizus*); syn. Damoiseau, 1987: 58.

Type depositories. *T. cylindricornis*: RMNH (HT!); *M. reductus*: MNHN (LT!, PLT!).

Distribution. S China; Indonesia (Mentawai, Sumatra), Myanmar [= Burma], Vietnam.

Notes. Damoiseau (1987: 58) reported that the specimens of *M. reductus* held in MNHN are holotype and allotype, but in the original description Kleine did not designate either of the syntypes as holotype; however, as there are two syntypes in MNHN, and as Damoiseau did not specifically select one as holotype, we designate the lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Lectotype of *M. reductus* (MNHN) designated on a specimen with the following labels: Tonkin, Région de Hoa-Binh/ Museum Paris 1934, A. de Cooman/ type/ *Microtracheliz reductus* Kln., R. Kleine det 1941. Paralectotype: 1 specimen (MNHN), Tonkin, Région de Hoa-Binh/ Museum Paris, 1934, A. de Cooman/ cotype.

dubius Kleine, 1935

dubius Kleine, 1935e: 307.

Type depository. NHM (HT!).

Distribution. New Guinea.

fluxus Kleine, 1923

fluxus Kleine, 1923e: 162.

Type depository. SMTD (HT).

Distribution. Philippines (Negros).

fractus Kleine, 1924

fractus Kleine, 1924a: 104.

Type depository. MRAC (HT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire).

Notes. The quotation for the People's Republic of Congo (Sforzi, 1998b: 274) is incorrect and should be referred to as the Democratic Republic of the Congo.

ghecuanus (Senna, 1892)

ghecuanus Senna, 1892e: 457 (*Trachelizus*).

ghekuanus: Schönfeldt, 1908: 23 (*Microtrachelizus*) [lapsus].

Type depository. MSNG (HT!).

Distribution. Myanmar [= Burma].

imbecillus Kleine, 1926

imbecillus Kleine, 1926f: 365.

Type depository. MRAC (HT!).

Distribution. Democratic Republic of the Congo (ex Zaire).

incertus Kleine, 1935

incertus Kleine, 1935e: 307.

Type depository. NHM (HT!).

Distribution. New Guinea.

incisus Kleine, 1935

incisus Kleine, 1935e: 307.

Type depository. NHM (HT!).

Distribution. New Guinea.

laevis Damoiseau, 1987

laevis Damoiseau, 1987: 56.

Type depository. SAMA (Lea coll.) (HT).

Distribution. Australia.

lyratus (Perroud & Montrouzier, 1865)

lyratus Perroud & Montrouzier, 1865: 139 (*Trachelizus*).

Type depositories. IRSNB (LT!, PLT!), MNHN (PLT!).

Distribution. New Caledonia.

Notes. Alonso-Zarazaga & Lyal (1999: 55 and 248) give Perroud as author of the species, but we believe that Perroud and Montrouzier must be considered as co-authors.

Lectotype designated by Damoiseau (1966a: 26).

mentaweicus Senna, 1898

mentaweicus Senna, 1898a: 236.

Type depository. MSNG (HT!).

Distribution. Indonesia (Mentawai).

monilicornis Damoiseau, 1987

monilicornis Damoiseau, 1987: 57.

Type depository. MNHN (HT!).

Distribution. Indonesia (Moluccas).

montrouzieri Senna, 1903

montrouzieri Senna, 1903: 167.

Type depository. MSNG (HT!).

Distribution. Indonesia (Java, Sumatra).

plenicostatus Damoiseau, 1987

plenicostatus Damoiseau, 1987: 56.

Type depositories. NHM (HT!), BPBM (?) (AT).

Distribution. Borneo.

Notes. This species was not fully described by Damoiseau (1987: 56), but its main characters were included in a key, so the species is valid. The holotype is labelled as follows: F.G.B. 670, 21.VIII.1959 Sarawak, Kuching.

According to A. Samuelson (BPBM) (2001, personal communication) the allotype of *M. plenicostatus* is not in BPBM now because it was never returned after the untimely death of Damoiseau.

pubescens Senna, 1893

pubescens Senna, 1893e: 320.

Type depositories. MZUF (LT!), RMNH (PLT!).

Distribution. Indonesia (Sumatra), Malaysia, Philippines (Negros); Europe (I).

Notes. Lectotype designated by Bartolozzi *et al.* (1985: 99) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

There is a specimen Damoiseau labelled as paratype in IRSNB, but the species was described on only two specimens which are in MZUF and RMNH.

queenslandicus Damoiseau, 1987

queenslandicus Damoiseau, 1987: 56.

Type depository. UQDZ (HT).

Distribution. Australia.

Notes. Damoiseau (1987: 55) did not describe this species in full, but its main characters were included in a key, so the species is valid.

rectestriatus (Fairmaire, 1897) **n. comb.**

rectestriatus Fairmaire, 1897: 195 (*Cerobates*).

Type depository. MNHN (HT!).

Distribution. Comores Is., Madagascar.

rudis Kleine, 1937

rudis Kleine, 1937b: 506.

Type depository. NHM (HT!).

Distribution. Borneo.

semistriatus Damoiseau, 1987

semistriatus Damoiseau, 1987: 56.

Type depository. SAMA (HT).

Distribution. New Guinea.

Notes. Damoiseau (1987: 56) did not describe this species in full, but its main characters were included in a key, so the species is valid.

siamensis Kleine, 1926

siamensis Kleine, 1926c: 165.

Type depository. NHM (HT!).

Distribution. Borneo, Malaysia, Philippines (Samar), Thailand.

Notes. Damoiseau (1987: 60) incorrectly reported that the holotype was in MZUF.

silvicola Senna, 1903

silvicola Senna, 1903: 168.

Type depositories. MSNG (LT!), MZUF (PLT!).

Distribution. Indonesia (Sumatra), Malaysia.

Notes. Lectotype (MSNG) designated on a specimen with the following labels: Sumatra, Sirambè, XII.90-III.91, E. Modigliani/ cotypus/ *Microtrachelizus silvicola* Senna cotypus!. Paralectotype: 1 specimen (MZUF), Malacca, ex coll. Boucard/ *Microtrach. silvicola* Senna cotypus/ Bull. Soc. Ent. It., XXXIV, 1902, cotypus!/ ex-coll. Senna/ "La Specola" Firenze, 6951.

sirambeicus Senna, 1903

sirambeicus Senna, 1903: 169.

Type depository. MSNG (HT!).

Distribution. Indonesia (Sumatra).

sternopilosus Damoiseau, 1987

sternopilosus Damoiseau, 1987: 56.

Type depositories. BPBM (HT), MNHN (PT!).

Distribution. Borneo, Indonesia (Sumatra), Thailand.

Notes. Damoiseau (1987: 56) did not describe this species in full, but its main characters were included in a key, so the species is valid.

According to A. Samuelson (BPBM) (2001, personal communication) the allotype of *M. sternopilosus* is no longer in BPBM because it was never returned after the untimely death of Damoiseau.

tabaci Senna, 1893

tabaci Senna, 1893e: 323.

Type depositories. MZUF (LT!), RMNH (PLT!).

Distribution. Borneo, Indonesia (Sumatra), Malaysia, Myanmar [= Burma], Philippines (Mindanao), Vietnam; New Guinea; Europe (I).

Notes. Lectotype designated by Bartolozzi *et al.* (1985: 101) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

targionii Senna, 1893

targionii Senna, 1893e: 322.

Type depository. RMNH (LT!).

Distribution. Indonesia (Mentawai, Sumatra); New Guinea; Europe (I).

Notes. In his original description Senna listed three syntypes (MZUF, MSNG, RMNH). We did not find any specimen in MSNG with the label data given in the description. The only specimen in MZUF with the correct locality was labelled by Senna as 'cf. c. typo' [compared with type]. Thus we decide to designate the lectotype on the material from RMNH.

Lectotype (RMNH) designated on a specimen with the following labels: Ant. Grouvelle, Mit Tabak von Sumatra/ Paratype/ *Microtrachelizus targionii* R. Damoiseau det.

temporalis Damoiseau, 1987

temporalis Damoiseau, 1987: 56.

Type depository. HNHM (HT).

Distribution. New Guinea.

Notes. Damoiseau (1987: 56) did not describe this species in full, but its main characters were included in a key, so the species is valid.

Parapisthius Kleine, 1935

Parapisthius Kleine, 1935e: 308.

Type species: *Parapisthius intermedius* Kleine, 1935.

Distribution. Malaysia, Vietnam; New Guinea.

gracilis Damoiseau, 1987

gracilis Damoiseau, 1987: 60.

Type depository. BPBM (HT).

Distribution. Vietnam.

intermedius Kleine, 1935

intermedius Kleine, 1935e: 309.

Type depositories. NHM (ST!), DEI (ST!).

Distribution. New Guinea.

Notes. There are two specimens in NHM labelled as type and paratype; Döbler & Gaedike (1995: 448) list another paratype in DEI; Kleine based the description on only two specimens, and did not designate either of them as holotype.

paulus Kleine, 1938*paulus* Kleine, 1938d: 73.**Type depository.** NHM (HT!).**Distribution.** Malaysia.*Pseudohigonius* Damoiseau, 1987*Pseudohigonius* Damoiseau, 1987: 53.**Type species:** *Microtrachelizus rugosisculpturatus* Kleine, 1939.**Distribution.** New Guinea.*rugosisculpturatus* (Kleine, 1939)*rugosisculpturatus* Kleine, 1939c: 107 (*Microtrachelizus*).**Type depository.** NHM (HT!).**Distribution.** New Guinea.*Taphrocomister* Senna, 1895*Taphrocomister* Senna, 1895a: 185.**Type species:** *Taphrocomister singularis* Senna, 1895.**Distribution.** India (Darjeeling); Indonesia (Sumatra).*singularis* Senna, 1895*singularis* Senna, 1895a: 185.*abruptus* Kleine, 1935c: 75 (*Higonius*); **syn. n.****Type depositories.** *T. singularis*: RMNH (HT!); *H. abruptus*: NHM (HT!).**Distribution.** India (Darjeeling); Indonesia (Sumatra).**Notes.** We decided synonymy after examination of the type material.

PSEUDOCEOCEPHALINI Kleine, 1922

- Pseudoceocephalini Kleine, 1922m: 163.
 Céocéphalides Lacordaire, 1866: 444.
 Ceocephalinae Pascoe, 1872: 317.
 Ceocephalini Kolbe, 1883b: 239.
 Ceocephali Kolbe, 1883c: 74.
 Ceocephalidae Schönfeldt, 1908: 53.
 Ceocephalini: Kleine, 1920e: 12 [lapsus].
 Ceocephalini: Kleine, 1921a: 78 [lapsus].
 Pseudoceocephalidae Kleine, 1922g: 225.
 Pseudoceocephalinae Kleine, 1925b: 165.
 Pseudoceocepharini: Miwa & Chûjô, 1939: 7 [lapsus].
 Ceocephalina Zimmerman, 1994: 166.

Agrioblepis Kleine, 1921

Agrioblepis Kleine, 1921m: 62.

Type species: *Agrioblepis hova* Kleine, 1921.

Distribution. Madagascar.

hova Kleine, 1921

hova Kleine, 1921m: 63.

rubripes Kleine, 1944: 157 (*Agrioblepis*); syn. Damoiseau, 1989b: 68.

Type depositories. *A. hova*: RMNH (HT); *A. rubripes*: MNHN (LT!, PLT!), DEI (PLT!).

Distribution. Madagascar.

Notes. Lectotype of *A. rubripes* (MNHN) designated on a male specimen with the following labels: Museum Paris, Madagascar, Region de l'Androy, Ambovombe, Dr. J. Decorse, 1901/ 1 au 15 dec. 00/ type/ *Agrioblepis rubripes* Kln., R. Kleine det. 1941/ comp. au type/ *Agrioblepis hova* ♂ Kleine, R. Damoiseau det. 1964. Paralectotypes: 1 male specimen (MNHN), Museum Paris, Madagascar, Region de l'Androy, Ambovombe, Dr. J. Decorse 1901/ 1 au 15 dec. 00/ *Agrioblepis rubripes* Kln., R. Kleine det. 1943; 1 specimen (MNHN), Museum Paris, Madagascar (S. O.) Plaines de Fiherena, F.

Geay 1905/ type; 1 male specimen (MNHN), Museum Paris, Madagascar, Region de l'Androy, Ambovombe, Dr. J. Decorse 1901/ 1 au 15 dec. 00/ *Agrioblepis rubripes* Kln., R. Kleine det. 1942.

Amerismus Lacordaire, 1866

Amerismus Lacordaire, 1866: 461.

Type species: *Ozodecerus cavicaudatus* Chevrolat, 1839.

Distribution. Madagascar.

admirabilis Kleine, 1922

admirabilis Kleine, 1922f: 219.

Type depository. SMTD (ST).

Distribution. Madagascar.

adulterinus Kleine, 1922

adulterinus Kleine, 1922f: 220.

Type depositories. SMTD (ST), MNHUB (?) (ST).

Distribution. Madagascar.

Notes. According to Kleine (1928a: 93) syntypes were in MNHUB and SMTD, but we did not find any of them in MNHUB.

adultus Kleine, 1922

adultus Kleine, 1922f: 220.

Type depository. SMTD (ST).

Distribution. Madagascar.

affinis Kleine, 1925

affinis Kleine, 1925a: 196.

Type depository. NHM (?) (ST).

Distribution. Madagascar.

Notes. According to Kleine (1925a: 197) the types were in NHM, but we did not find them there.

Kleine (1938d) did not mention *affinis* among the species listed in his Genera Insectorum.

ardens Kleine, 1923

ardens Kleine, 1923h: 53.

Type depositories. NHM (LT!, PLT!), NMPC (?) (PLT).

Distribution. Madagascar.

Notes. Kleine (1923h: 53) based description on a series of four males and three females from the NMPC collection. Two syntypes are now in NHM, but according to J. Jelinek (NMPC) (2002, personal communication) the syntypes in NMPC are missing.

Lectotype of *A. ardens* (NHM) designated on a male specimen with the following labels: Madagascar/ Syntype/ Type/ Sharp Coll. 1905-313/ *Amerismus ardens* Kln. Type ♂ R. Kleine det. 1923. Paralectotype (NHM): 1 female specimen, Syntype/ Madagascar/ Sharp Coll. 1905-313/ *Amerismus ardens* Kln., Type ♀, R. Kleine det. 1923.

cavicaudatus (Chevrolat, 1839)

cavicaudatus Chevrolat, 1839: 176 (*Ozodecerus*).

metallicus Chevrolat, 1839: 177 (*Ozodecerus*); syn. Kolbe, 1883a: 388.

metallicus Boheman (in Schoenherr, 1840: 550) (non Chevrolat, 1839) (*Brenthus*); **syn. n.**

Type depository. *O. cavicaudatus*: MNHN (HT!); *O. metallicus*: MNHN (HT!); *B. metallicus*: MNHN (?) (HT).

Distribution. Madagascar.

Notes. According to Kolbe (1883a: 388) and Senna (1892c: 61) *Ozodecerus metallicus* Chevrolat is a synonym of *O. cavicaudatus* Chevrolat.

Kleine (1938d: 165) did not report this synonymy; he did not mention Chevrolat's species in his *Genera Insectorum* (1938d) nor that of Boheman. We did not find the syntypes of *Brenthus metallicus* Boheman in MNHN, but from the description (1840: 550) we believe that this species is *O. metallicus* Chevrolat (Boheman also reported that this species was "... a *Dom. Chevrolat ad describendum benevole communicatus*").

felix Kleine, 1925

felix Kleine, 1925a: 197.

Type depository. NHM (LT!, PLT!).

Distribution. Madagascar.

Notes. Lectotype of *A. felix* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ Madagascar/ Sharp Coll. 1905-313/ *Amerismus felix* Kln., Type ♂, R. Kleine det. 1923. Paralectotypes: 1 female specimen (NHM), Syntype/ Madagascar/ Pascoe Coll. 93-60/ *Amerismus felix* Kln., Type ♀, R. Kleine det. 1923; 1 male specimen (NHM), Syntype/ Ambohimombo Forest, Cent. Madag., 1200 m, C. J. F. Major, 1905.

hildebrandti Kolbe, 1883

hildebrandti Kolbe, 1883c: 79.

Type depository. MNHUB (?) (ST).

Distribution. Madagascar.

Notes. According to Kleine (1928a: 93) the types are in MNHUB, but we did not find them there.

Anactorus Damoiseau, 1967

Anactorus Damoiseau, 1967a: 314.

Type species: *Anactorus gilvus* Damoiseau, 1967.

Distribution. Tanzania.

gilvus Damoiseau, 1967*gilvus* Damoiseau, 1967a: 314.**Type depository.** MNHUB (HT!).**Distribution.** Tanzania.*Anampyx* Damoiseau, 1963*Anampyx* Damoiseau, 1963a: 133.**Type species:** *Anampyx edentulus* Damoiseau, 1963.**Distribution.** Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Gabon, Ghana, Ivory Coast, Republic of the Congo.*allardi* Damoiseau, 1967*allardi* Damoiseau, 1967a: 310.**Type depositories.** MRAC (HT!, AT!, PT!), IRSNB (PT!).**Distribution.** Democratic Republic of the Congo (ex Zaire).*edentulus* Damoiseau, 1963*edentulus* Damoiseau, 1963a: 135.**Type depository.** MRAC (HT!, PT!).**Distribution.** Cameroon, Democratic Republic of the Congo (ex Zaire).**Notes.** According to Damoiseau (1967a: 310) the paratype for *A. edentulus* actually belongs to *A. occipitalis*. The quotation for People's Republic of Congo (Sforzi, 1998b: 275) is incorrect and should be referred to as the Democratic Republic of the Congo.

***maculatus* (Kleine, 1920)**

maculatus Kleine, 1920c: 40 (*Mygaleicus*).

Type depositories. MNHUB (LT!), DEI (PLT!).

Distribution. Angola, Cameroon, Ghana, Ivory Coast, Republic of the Congo.

Notes. Lectotype designated by Damoiseau (1967a: 313).

***occipitalis* Damoiseau, 1967**

occipitalis Damoiseau, 1967a: 313.

Type depositories. IRSNB (HT!), MRAC (PT!).

Distribution. Democratic Republic of the Congo (ex Zaire), Gabon.

***Aphelampyx* Quentin, 1966**

Aphelampyx Quentin, 1966: 1659.

Type species: *Centrophorus rufescens* J. Thomson, 1858.

Distribution. Gabon, Republic of the Congo.

Notes. Damoiseau (1967a) did not mention this genus in his Monograph on African Brentidae.

***rufescens* (J. Thomson, 1858)**

rufescens J. Thomson, 1858: 120 (*Centrophorus*).

Type depository. MNHN (HT!).

Distribution. Gabon, Republic of the Congo.

***Atenophthalmus* Kleine, 1920**

Atenophthalmus Kleine, 1920h: 4.

Atenophthalmus: Kleine, 1920h: 4 [lapsus].

Holomiolispa Kleine, 1926a: 31; syn. Damoiseau, 1989b: 69.

Type species: *Atenophthalmus tenuicostatus* Kleine, 1920.

Distribution. Indonesia (Moluccas); Caroline Is., Fiji Is., New Guinea (including Woodlark), Solomon Is.

Notes. Alonso-Zarazaga & Lyal (1999: 55) give 1921 as description date for the genus, but in the original reprint in our hands the date is 28th October 1920.

***devotus* (Kleine, 1926)**

devota Kleine, 1926a: 31 (*Holomiolispa*).

Type depository. NHM (HT!).

Distribution. Indonesia (Moluccas).

***fuscojanthinus* (Fairmaire, 1881)**

fuscojanthinus Fairmaire, 1881b: 373 (*Eubactus*).

Type depository. MNHN (LT!).

Distribution. Fiji Is.

Notes. Lectotype of *E. fuscojanthinus* (MNHN) designated on a specimen with the following labels: 160/ ex Musaeo L. Fairmaire 1893/ *Eubactus fuscojanthinus* Fairm. I. Viti/ Prépar. Génit. N° 80 B 18, R. Damiseau/ *Atenophthalmus fuscojanthinus* (Fairmaire), R. Damoiseau det.

***punctatus* (Montrouzier, 1855)**

punctatus Montrouzier, 1855: 40 (*Arrhenodes*).

Type depository. IRSNB (HT!).

Distribution. Papua New Guinea (Woodlark), Solomon Is.

***tenuicostatus* Kleine, 1920**

tenuicostatus Kleine, 1920h: 6.

Type depository. NHRS (HT).

Distribution. Caroline Is., New Guinea, Solomon Is.

Autarcus Senna, 1892

Autarcus Senna, 1892c: 59.

Type species: *Ceocephalus laticollis* Perroud & Montrouzier, 1865.

Distribution. New Caledonia, Polynesia.

laticollis (Perroud & Montrouzier, 1865)

laticollis Perroud & Montrouzier, 1865: 135 (*Ceocephalus*).

Type depository. MNHN (HT!).

Distribution. New Caledonia, Polynesia.

Notes. Alonso-Zarazaga & Lyal (1999: 55 and 248) give Perroud as author of the species, but we believe that Perroud and Montrouzier must be considered as co-authors.

Damoiseau (1966a: 25) designated the neotype on a specimen held in IRSNB, but we found a specimen in MNHN that seems to be the holotype for this species.

Cacoschizus Sharp, 1900

Cacoschizus Sharp, 1900: 386.

Type species: *Schizotrachelus schmeltzii* Fairmaire, 1881.

Distribution. New Guinea (including Bismarck Archipelago: Duke of York, New Britain), Solomon Is.

laevipennis Damoiseau, 1989

laevipennis Damoiseau, 1989b: 77.

Type depository. SAMA (HT), GDFPNG (AT).

Distribution. New Guinea.

Notes. Damoiseau (1989b: 77) writes that the types are in GDFPNG, but according to R. Oberprieler (CSIRO) (2003, personal communication) the holotype is now at SAMA.

***schmeltzii* (Fairmaire, 1881)**

schmeltzii Fairmaire, 1881c: 421 (*Schirotrachelus*) (sic!).

schmeltzi: Schönfeldt, 1908: 63 (*Schizotrachelus*) [lapsus].

Type depository. ZMUH (HT).

Distribution. New Guinea (including Bismarck Archipelago: Duke of York, New Britain), Solomon Is.

Notes. Fairmaire redescribed this species in 1883 (p. 44).

***Calyptulus* Kleine, 1922**

Calyptulus Kleine, 1922p: 135.

Calyptus Kleine, 1921a: 68 (non Haliday, 1835) [no type species].

Type species: *Calyptulus irritans* Kleine, 1922.

Distribution. New Guinea.

Notes. Kleine quoted the genus *Calyptus* in a paper on brentid wings (1920e), and in a paper on elytral patterns (1921a), but he described it under the name *Calyptulus* only in 1922.

***irritans* Kleine, 1922**

irritans Kleine, 1922p: 136.

Type depository. DEI (HT!).

Distribution. New Guinea.

***rostratus* Kleine, 1922**

rostratus Kleine, 1922p: 137.

Type depository. DEI (HT!).

Distribution. New Guinea.

Chalybdicus Kleine, 1922

Chalybdicus Kleine, 1922f: 218.

Type species: *Chalybdicus hahnei* Kleine, 1922.

Distribution. New Guinea, Samoa Is., Solomon Is., Vanuatu [= New Hebrides].

cupronitens Kleine, 1935

cupronitens Kleine, 1935e: 309.

Type depository. NHM (HT!).

Distribution. New Guinea.

hahnei Kleine, 1922

hahnei Kleine, 1922f: 219.

Type depository. MZPW (HT!).

Distribution. Vanuatu [= New Hebrides].

popularis Kleine, 1939

popularis Kleine, 1939e: 588.

Type depository. NHM (HT!).

Distribution. Solomon Is.

reverens Kleine, 1928

reverens Kleine, 1928c: 158.

Type depositories. NHM (LT!, PLT!), SMTD (PLT).

Distribution. Samoa Is.

Notes. Lectotype of *C. reverens* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ Malololelei, Upolu Samoa, 2000 ft.,

22.III.1925, Buxton & Hopkins/ Pres. by Imp. Bur. Ent. Brit. Mus. 1927-294/
Chalybdicus reverens Kln., Type ♂, R. Kleine det. 1926; Paralectotype: 1 fe-
 male specimen (NHM), Type/ Samoan Is., Malololelei, Upolu, 25.IV.1924, P.
 A. Buxton and G. H. Hopkins/ Pres. by Imp. Bur. Ent. Brit. Mus. 1927-294/
Chalybdicus reverens Kln., Type ♀, R. Kleine det. 1926.

According to Zimmerman (1994: 16) this species could be a synonym of
C. hahnei.

Dacetellum Hedicke, 1922

Dacetellum Hedicke, 1922: 427 [replacement name for *Dacetum*].

Dacetum Kleine, 1922f: 220 (non Agassiz, 1846, nec Koch, 1847).

Type species: *Dacetum adjunctum* Kleine, 1922.

Distribution. Madagascar.

adjunctum (Kleine, 1922)

adjunctum Kleine, 1922f: 222 (*Dacetum*).

Type depository. SMTD (ST).

Distribution. Madagascar.

Eubactrus Lacordaire, 1866

Eubactrus Lacordaire, 1866: 456.

Type species: *Eubactrus semiaeneus* Lacordaire, 1866.

Distribution. Indonesia (Moluccas: Halmahera [= Gilolo]); Fiji Is., New
 Guinea (including Bismarck Archipelago: Duke of York), Solomon Is., Va-
 nuatu [= New Hebrides].

cavus Kleine, 1931

cavus Kleine, 1931a: 97.

Type depository. SMTD (HT).

Distribution. New Guinea.

excavus Kleine, 1935

excavus Kleine, 1935b: 176.

Type depository. SMTD (HT).

Distribution. Solomon Is.

metallicollis Fairmaire, 1881

metallicollis Fairmaire, 1881c: 421.

Type depository. Unknown.

Distribution. Fiji Is.

semiaeneus Lacordaire, 1866

semiaeneus Lacordaire, 1866: 457.

Type depository. MCZ (ST).

Distribution. Fiji Is., Vanuatu [= New Hebrides].

spissicornis Fairmaire, 1881

spissicornis Fairmaire, 1881b: 373.

Type depository. ZMUH (HT).

Distribution. New Guinea (including Bismarck Archipelago: Duke of York).

tripartitus Lacordaire, 1866

tripartitus Lacordaire, 1866: 457.

Type depository. MCZ (ST).

Distribution. Indonesia (Moluccas: Halmahera [= Gilolo]).

Eumecopodus Calabresi, 1920

Eumecopodus Calabresi, 1920d: 39.

Pelochroma Kleine, 1922h: 149; syn. De Muizon, 1960: 253.

Eumecopus: Mayné & Donis, 1962: 89 [lapsus].

Type species: *Eumecopodus longicornis* Calabresi, 1920.

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea (Bioko Is. [= Fernando Poo]), Gabon, Ivory Coast, Republic of the Congo, Republic of Guinea, São Thomé, Tanzania.

burgeoni De Muizon, 1959

burgeoni De Muizon, 1959: 79.

Type depositories. MRAC (HT!), NHMB (PT!).

Distribution. Democratic Republic of the Congo (ex Zaire), Gabon, Republic of Guinea.

Notes. The quotation for People's Republic of Congo (Sforzi, 1998b: 275) is incorrect and should be referred to as the Democratic Republic of the Congo.

fuliginosus Calabresi, 1920

fuliginosus Calabresi, 1920d: 41.

Type depository. MSNG (HT!).

Distribution. São Thomé.

guttatus De Muizon, 1955

guttatus De Muizon, 1955b: 904.

Type depository. MNHN (HT!).

Distribution. Gabon, Tanzania.

longicornis Calabresi, 1920

longicornis Calabresi, 1920d: 40.

lutulentum Kleine, 1922h: 151 (*Pelochroma*); syn. De Muizon, 1960: 253.

Type depositories. *E. longicornis*: MSNG (HT!); *P. lutulentum*: MNHUB (?) (ST).

Distribution. Cameroon, Equatorial Guinea (Bioko Is. [= Fernando Poo]), Ivory Coast, Republic of the Congo, Tanzania, Democratic Republic of the Congo (ex Zaire).

Notes. According to Kleine (1928a: 92) the types are in MNHUB, but we did not find them there.

Gynandrorhynchus Lacordaire, 1866

Gynandrorhynchus Lacordaire, 1866: 450.

Gynandrorrhynchus: Gemminger & Harold, 1872: 2715 [lapsus].

Mygaleicus Kleine, 1920c: 34; syn. Damoiseau, 1967a: 286.

Type species: *Gynandrorhynchus bocandei* Lacordaire, 1866.

Distribution. Angola, Burundi, Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea, Ethiopia, Gabon, Ghana, Guinea Bissau, Ivory Coast, Kenya, Malawi, Mozambique, Nigeria, Republic of the Congo, Republic of Guinea, Rwanda, São Thomé and Príncipe, Sierra Leone, Somalia, South Africa, Sudan, Tanzania, Togo, Uganda.

audax (Kleine, 1923)

audax Kleine, 1923d: 137 (*Mygaleicus*).

Type depository. NHM (LT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Ivory Coast, Mozambique, Republic of the Congo.

Notes. Damoiseau (1967a: 298) reported that the specimens held in NHM are holotype and paratype, but in his original description Kleine did not designate any of the syntypes as holotype. However in NHM we found only one syntype, which we consider as lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

***bocandei* Lacordaire, 1866**

bocandei Lacordaire, 1866: 451.

Type depositories. IRSNB (LT!, PLT!), MCZ (PLT).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Ghana, Guinea Bissau, Ivory Coast, Nigeria, Republic of the Congo, Republic of Guinea, Togo.

Notes. Damoiseau (1967a: 290) reported that the specimens held in IRSNB are holotype and paratype, but in his original description Lacordaire did not designate any of the syntypes as holotype; however, as there is only one male syntype in IRSNB, this must be considered as lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999). The lectotype has the following labels: Guinée/ Boucadé [*sic!*]/ Guinée (Bocandé)/ Type/ Coll. Lacordaire/ *Gynandrorhynchus bocandei* Lacordaire, Lectotype, R. Damoiseau det. 1962/ Lectotype [*in litteris*].

***corneolus* (Kleine, 1924)**

corneolus Kleine, 1924a: 107 (*Mygaleicus*).

Type depository. MRAC (HT!).

Distribution. Democratic Republic of the Congo (ex Zaire).

Notes. In Damoiseau's Monograph (1967a: 299) the figures of this species are inverted with those of *G. granulatus* (Bartolozzi, 1991: 14).

***grallator* Damoiseau, 1967**

grallator Damoiseau, 1967a: 304.

Type depository. ZSM (HT).

Distribution. Cameroon.

granulatus (Kleine, 1920)

granulatus Kleine, 1920c: 42 (*Mygaleicus*).

Type depository. MNHUB (HT!).

Distribution. Cameroon.

Notes. In Damoiseau's Monograph (1967a: 299) the figures of this species are inverted with those of *G. corneolus* (Bartolozzi, 1991: 14).

nigrothorax Damoiseau, 1989

nigrothorax Damoiseau, 1989b: 79.

Type depository. MRAC (HT!).

Distribution. Democratic Republic of the Congo (ex Zaire).

politus Damoiseau, 1967

politus Damoiseau, 1967a: 300.

Type depository. MNHUB (HT!).

Distribution. Cameroon.

protractus Damoiseau, 1967

protractus Damoiseau, 1967a: 303.

Type depository. MRAC (HT!).

Distribution. Ivory Coast.

somaliensis Bartolozzi, 1991

somaliensis Bartolozzi, 1991: 11.

Type depository. MRSN (HT!).

Distribution. Somalia.

tarsalis (Kleine, 1920)

tarsalis Kleine, 1920c: 37 (*Mygaleicus*).

Type depositories. MNHUB (LT!, PLT!), NHRS (?) (PLT), ZSM (PLT).

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Ethiopia, Ghana, Ivory Coast, Mozambique, South Africa, Tanzania.

Notes. Damoiseau (1967a: 295) reported that the specimens held in MNHUB are holotype and allotype, and that paratypes are in NHRS and in ZSM, but in his original description Kleine did not designate any of the syntypes as holotype; however, as there are two syntypes in MNHUB, and as Damoiseau did not specifically select one as holotype, we designate the lectotype (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Lectotype of *M. tarsalis* (MNHUB) designated on a specimen with the following labels: Type/ Kamerun, Ebolowa, I-IV.1912, v. Rothkirch S. G./ *Mygaleicus tarsalis* Kleine/ ♂ genitalia enlevés par Kleine, R. Damoiseau rev., 1965/ *Gynandrorhynchus tarsalis* Kleine, R. Damoiseau det., 1965. Paralectotype: 1 specimen (MNHUB), Type/ Kamerun, Ebolowa, I-IV.1912, v. Rothkirch S. G./ *Mygaleicus tarsalis* Kleine/ ♂ genitalia enlevés par Kleine, R. Damoiseau rev., 1965.

Kleine (1920c: 38) reported that the type for *M. tarsalis* was in NHRS but the species is not listed in the NHRS web site.

valentinei Bartolozzi & Sforzi, 1994

valentinei Bartolozzi & Sforzi, 1994: 253.

Type depositories. BDVC (HT!, AT!, PT!), MZUF (PT!), EOC (PT!).

Distribution. Kenya.

Notes. The original description states that the holotype is in NHM, but it seems that Prof. B. D. Valentine never deposited it there.

***vittipennis* (Fähræus, 1871)**

vittipennis Fähræus, 1871: 435 (*Brentus*).

nitida Calabresi, 1920d: 37 (*Mygaleicus vittipennis* var.); **syn. n.**

persimilis Damoiseau, 1963i: 46 (*Mygaleicus*); **syn.** Damoiseau, 1967a: 290.

Type depositories. *B. vittipennis*: NHRS (HT); *M. vittipennis* var. *nitida*: MSNG (LT!, PLT!), IRSNB (PLT!), MZUF (PLT!); *M. persimilis*: MRAC (HT!).

Distribution. Burundi, Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea (including Bioko Is. [= Fernando Poo]), Ethiopia, Gabon, Ghana, Ivory Coast, Kenya, Malawi, Republic of the Congo, Rwanda, São Thomé and Príncipe, Sierra Leone, South Africa, Sudan, Tanzania, Uganda.

Notes. Lectotype of *M. vittipennis* var. *nitida* (MSNG) designated on a specimen with the following labels: Príncipe, Roca Inf., D. Henrique, II.1901, 200-300 m, L. Fea/ *Mygaleicus vittipennis* Fahr. var. *nitida* n. Paralectotypes: 55 specimens (MSNG), 2 specimens (MZUF), Is. Príncipe Bahia do Oeste, V-VI.1901, 100-200 m, L. Fea/ "La Specola" Firenze, 9578; 2 specimens (MZUF), Is. Príncipe Bahia do Oeste, V-VI.1901, 100-200 m, L. Fea/ ex-coll. Senna/ "La Specola" Firenze, 9578; 5 specimens (IRSNB), Is. Príncipe, Bahia do Oeste, V.1901, L. Fea/ *Gynandrorhynchus vittipennis* (Fähræus) *nitida* var. (Calabresi) R. Damoiseau det., 1962/ Paratype.

Damoiseau (1967a: 292) did not mention Fernando Poo and Nord Niassee (Kleine, 1920c: 44) when he gave the species distribution.

***Hetaeroceocephalus* Kleine, 1921**

Hetaeroceocephalus Kleine, 1921j: 113.

Type species: *Hetaeroceocephalus propinquus* Kleine, 1921.

Distribution. Madagascar.

***propinquus* Kleine, 1921**

propinquus Kleine, 1921j: 115.

Type depositories. SMTD (HT), DEI (PT!).

Distribution. Madagascar.

Heterothesis Kleine, 1914

Heterothesis Kleine, 1914b: 178.

Type species: *Heterothesis elegans* Kleine, 1914.

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Gabon, Republic of the Congo.

elegans Kleine, 1914

elegans Kleine, 1914b: 181.

Type depository. MZPW (ST).

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Gabon, Republic of the Congo.

Hormocerus Schoenherr, 1823

Hormocerus Schoenherr, 1823: 1137.

Apterorhinus Senna, 1895d: 59; syn. Zimmerman, 1994: 168.

Apterorhynchus: Kleine, 1937e: 99 [lapsus].

Homaecerus: Fairmaire, 1883: 44 [lapsus].

Type species: *Brentus reticulatus* Lund, 1800.

Distribution. Sri Lanka to Polynesia.

compressitarsus (Senna, 1895)

compressitarsus Senna, 1895d: 61 (*Apterorhinus*).

compressitarsis: Schultze, 1915: 129 (*Apterorhinus*) [lapsus].

albatus Kleine, 1922g: 226 (*Apterorhinus*); syn. Zimmerman, 1994: 175.

Type depositories. *A. compressitarsus*: MZUF (LT!); *A. albatus*: DEI (LT!, PLT!).

Distribution. Borneo, Indonesia (Batu; Java; Moluccas: Buru; Sumatra), Malaysia, Philippines (Leyte, Luzon, Mindanao, Palawan, Siargao); Australia, New Guinea.

Notes. Lectotype of *A. compressitarsus* designated by Bartolozzi *et al.* (1985: 95) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

Lectotype of *A. albatus* (DEI) designated on a specimen with the following labels: Coen District, Cape York, Queensland, H. Hacker/ Syntype/ coll. Hacker/ Kleine det./ coll. DEI Eberswalde/ *Apterorrhinus albatus* Kln., det. Kleine, 1921. Paralectotype: 1 male specimen (DEI), with the same label data as the lectotype.

fossulatus Blackburn, 1896

fossulatus Blackburn, 1896: 36.

Type depository. NHM (LT!).

Distribution. Taiwan (?); Australia.

Notes. Lectotype of *H. fossulatus* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ 6109 T n. 170/ Blackburn Coll. 1910-236/ *Hormocerus fossulatus* Blackb.

reticulatus (Lund, 1800)

reticulatus Lund, 1800: 81 (*Brentus*).

dehaani Gyllenhal (in Schoenherr, 1833: 360) (*Ceocephalus*); syn. Kleine, 1926b: 255.

scrobicollis Boheman, 1845: 373 (*Ceocephalus*); syn. Kleine, 1926b: 255.

annulipes Schaufuss, 1887: 138 (*Hormocerus*); syn. Kleine, 1926b: 255.

scobricollis: Kleine, 1922i: 166 (*Hormocerus*) [lapsus].

Type depositories. *B. reticulatus*: ZMUC (ST); *C. dehanii*: NHRS (HT); *C. scrobicollis*: NHRS (ST); *H. annulipes*: MNHUB (LT!, PLT!), MNHN (ST?), NMPC (Salvator coll.) (PLT).

Distribution. Sri Lanka to Polynesia.

Notes. Lectotype of *H. annulipes* (MNHUB) designated on a male specimen with the following labels: Type/ *Hormocerus annulipes* Schauf., Macassar. Paralectotype: 1 male specimen (MNHUB) with the same label data as the lectotype. We do not designate a specimen of MNHN (Celeb. Mer.) as paralectotype because we are not sure if it really belongs to the type series.

According to Zimmerman (1994: 177) the species has been "widely dispersed through commerce".

Isoceocephalus Kleine, 1920

Isoceocephalus Kleine, 1920c: 46.

Type species: *Centrophorus rufescens* J. Thomson, 1858 *sensu* Kleine, 1920 = *Isoceocephalus rufescens* Quentin, 1966.

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea, Gabon, Ghana, Ivory Coast, Kenya, Namibia, Republic of the Congo, Tanzania, Togo, Uganda.

rufescens Quentin, 1966

rufescens Quentin, 1966: 1656.

rufescens J. Thomson *sensu* Kleine, 1920c: 47 (non *Centrophorus rufescens* J. Thomson, 1858); syn. Quentin, 1966: 1655.

pruinus Kleine, 1922e: 156 (*Pseudoceocephalus*); syn. Damoiseau, 1967a: 269.

Type depositories. *I. rufescens*: MNHN (HT); *P. pruinus*: MNHUB (HT!).

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea, Gabon, Ghana, Ivory Coast, Kenya, Namibia, Republic of the Congo, Tanzania, Togo, Uganda.

Metatrachelus Kleine, 1925

Metatrachelus Kleine, 1925b: 162.

Type species: *Metatrachelus comparativus* Kleine, 1925.

Distribution. Borneo, Indonesia (Sumatra), Malaysia, Philippines, Vietnam; Solomon Is.

Notes. Kleine (1922m: 163) mentioned *Metatrachelus* in a list of genera, but he did not describe it until three years later (1925b: 162).

castaneicolor Kleine, 1941

castaneicolor Kleine, 1941b: 228.

Type depository. MNHN (?) (ST).

Distribution. Vietnam.

Notes. According to Kleine (1941b: 226) the types were in MNHN, but we did not find them there.

***comparativus* Kleine, 1925**

comparativus Kleine, 1925b: 163.

Type depository. NHM (HT!).

Distribution. Malaysia.

***differens* Kleine, 1926**

differens Kleine, 1926a: 63.

Type depository. NHM (HT!).

Distribution. Indonesia (Sumatra), Malaysia, Philippines.

***diffidens* Kleine, 1926**

diffidens Kleine, 1926a: 64.

Type depository. NHM (HT!).

Distribution. Borneo.

***dohrni* Kleine, 1941**

dohrni Kleine, 1941c: 193.

Type depository. MZPW (HT).

Distribution. Indonesia (Sumatra).

firmus Kleine, 1933*firmus* Kleine, 1933e: 325.**Type depository.** SMTD (HT).**Distribution.** Vietnam.*froggatti* Kleine, 1939*froggatti* Kleine, 1939e: 589.**Type depository.** NHM (HT!).**Distribution.** Solomon Is.*hospes* Kleine, 1926*hospes* Kleine, 1926c: 175.**Type depository.** NHM (LT!, PLT!).**Distribution.** Malaysia.

Notes. Lectotype of *M. hospes* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ Rim Malacca, Febr. 1908/ Malay Pen. B.M. 1927-37/ *Metatrachelus hospes* Kleine 1924, Typus/ prépar. génit. No BM 112, R. Damoiseau/ *Tracheloschizus hospes* (Kleine), R. Damoiseau det. Paralectotypes: 2 male specimens (NHM), Syntype/ Malacca s. s. Rim, Febr. 1908/ 468/ ex F.M.S. Museum B. M. 1955-354; 1 female specimen (NHM), Selangor F.M.S., Ayer Itam Kampar/ Malay Pen. B.M. 1927-37/ *Metatrachelus hospes* Kln, Typus, R. Kleine det.; 1 female specimen (NHM), Malay Penin., Selangor F.M.S., Ayer Itam Kampar/ ex F.M.S. Museum B. M. 1955-354.

unisulcatus (Damoiseau, 1966) n. comb.*unisulcatus* Damoiseau, 1966e: 438 (*Tracheloschizus*).**Type depository.** ZMUC (HT).**Distribution.** Philippines (Balabac).

Notes. We include this species in this genus after studying a series of specimens (MZUF) that were identified by Damoiseau.

Neomygaleicus De Muizon, 1960

Neomygaleicus De Muizon, 1960: 206.

Type species: *Neomygaleicus olsenii* De Muizon, 1960.

Distribution. Equatorial Guinea.

olsenii De Muizon, 1960

olsenii De Muizon, 1960: 206.

Type depository. NHMB (HT!).

Distribution. Equatorial Guinea.

Nothogaster Lacordaire, 1866

Nothogaster Lacordaire, 1866: 450.

Type species: *Nothogaster paradoxa* Lacordaire, 1866.

Distribution. Madagascar (?).

Notes. Kleine (1938d) did not mention this genus in his *Genera Insectorum*.

paradoxa Lacordaire, 1866

paradoxa Lacordaire, 1866: 450.

Type depository. MCZ (HT).

Distribution. Madagascar (?).

Orphanobrentus Damoiseau, 1962

Orphanobrentus Damoiseau, 1962a: 65.

Centrophorus Chevrolat, 1839: 173 (non Mueller & Henle, 1837); syn. Alonso-Zarazaga & Lyal, 1999: 55.

Pseudoceocephalus: De Muizon, 1955b: 898 (non Kleine, 1920); syn. Damoiseau, 1962a: 65.

Type species: *Brentus picipes* Olivier, 1807.

Distribution. Tropical Africa (including Comores Is., Madagascar).

curvirostris (Chevrolat, 1839)

curvirostris Chevrolat, 1839: 181 (*Centrophorus*).

fraterculus Kolbe, 1883c: 77 (*Ceocephalus*); syn. Kleine, 1927a: 66.

Type depositories. *C. curvirostris*: MNHN (LT!, PLT!), NHRS (PLT); *C. fraterculus*: MNHUB (LT!, PLT!).

Distribution. Comores Is., Madagascar.

Notes. Lectotype of *C. curvirostris* (MNHN) designated on a specimen with the following labels: Madagascar/ Chevrolat T. 255/ type/ ex Musaeo G. Power/ Museum Paris 1952, coll. R. Oberthür. Paralectotypes: 2 specimens (MNHN), with the same label data as the lectotype.

Lectotype of *P. fraterculus* (MNHUB) designated on a specimen with the following labels: 34560/ Type/ *Ceocephalus fraterculus* n. sp. Kolbe, Type. Paralectotype: 1 specimen (MNHUB), 34560/ *Ceocephalus fraterculus* n. sp./ Madag., Goud.

emarginatus (Chevrolat, 1839) n. comb.

emarginatus Chevrolat, 1839: 180 (*Centrophorus*).

Type depository. MNHN (LT!).

Distribution. Madagascar.

Notes. Lectotype of *C. emarginatus* (MNHN) designated on a male specimen with the following labels: Madagascar/ *emarginatus* T. Chevr./ Chevrolat T. 258/ type/ ex Musaeo G. Power/ Museum Paris 1952, coll. R. Oberthür. As we are unable to identify with certainty other syntypes we do not designate paralectotypes.

Schönfeldt (1908) and Kleine (1938d) did not mention this species in their Catalogues.

This species, formerly included in the genus *Centrophorus*, has provisionally been placed in the genus *Orphanobrentus* due to the synonymy of the two genera, but most probably it belongs to a different genus. Its correct systematic position will be investigated by Sforzi (in preparation).

laevicollis (J. Thomson, 1858)

laevicollis J. Thomson, 1858: 120 (*Centrophorus*).

laevicollis: Kleine, 1920c: 31 (*Pseudoceocephalus*) [lapsus].

Type depository. MNHN (LT!, PLT!).

Distribution. Tropical Africa.

Notes. Damoiseau (1967a: 267) reported that the "type" was probably in MNHN, but Thomson did not designate any of the syntypes as holotype.

Lectotype of *C. laevicollis* (MNHN) designated on a male specimen with the following labels: Gabon/ Thomson ♂ type 74/ type/ ex Musaeo G. Power/ Museum Paris 1952, coll. R. Oberthür. Paralectotypes: 1 female specimen (MNHN), Gabon/ Thomson ♀ type 100/ type/ ex Musaeo G. Power/ Museum Paris 1952, coll. R. Oberthür; 2 specimens (MNHN), Gabon/ Mniszech 82/ ex Musaeo G. Power/ Museum Paris 1952, coll. R. Oberthür.

picipes (Olivier, 1807)

picipes Olivier, 1807: 442 (*Brentus*).

piscipes: J. Thomson, 1858: 120 (*Centrophorus*) [lapsus].

Type depository. Unknown.

Distribution. Tropical Africa (including Madagascar).

punctatissimus (Kleine, 1920)

punctatissimus Kleine, 1920c: 30 (*Pseudoceocephalus*).

Type depository. MNHUB (?) (HT).

Distribution. Angola, Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea (Bioko Is. [= Fernando Poo]), Ethiopia, Gabon, Ivory Coast, Republic of the Congo.

Notes. According to Kleine (1928a: 92) the holotype was in MNHUB, but neither Damoiseau (1967a: 267) nor ourselves found it there.

suturalis Quentin, 1966

suturalis Quentin, 1966: 1656.

Type depository. MNHN (HT!, PT!).

Distribution. Republic of the Congo.

Notes. Damoiseau (1967a) did not mention this species in his Monograph on African Brentidae.

***Oxyscapanus* Damoiseau, 1989**

Oxyscapanus Damoiseau, 1989b: 86.

Type species: *Oxyscapanus malachus* Damoiseau, 1989.

Distribution. Ivory Coast.

***malachus* Damoiseau, 1989**

malachus Damoiseau, 1989b: 87.

Type depository. Unknown.

Distribution. Ivory Coast.

Notes. According to Damoiseau (1989b: 87) the holotype was in K. E. Schedl's collection, but W. Schedl (2001, personal communication) did not find it in his father's collection.

***Palaeoceocephalus* Kleine, 1920**

Palaeoceocephalus Kleine, 1920c: 48.

Palaeoceocephalus: Kleine, 1937e: 100 [lapsus].

Type species: *Palaeocephalus nitidissimus* Kleine, 1920.

Distribution. Madagascar.

***nitidissimus* Kleine, 1920**

nitidissimus Kleine, 1920c: 49.

Type depositories. MNHUB (LT!), IRSNB (PLT!).

Distribution. Madagascar.

Notes. Lectotype and paralectotype designated by Damoiseau (1989b: 88).

***pilosus* Kleine, 1938**

pilosus Kleine, 1938d: 154.

Type depository. SMTD (HT).

Distribution. Madagascar.

***Paraceocephalus* Kleine, 1944**

Paraceocephalus Kleine, 1944: 155.

Type species: *Paraceocephalus pauliani* Kleine, 1944.

Distribution. Madagascar.

***colasi* Damoiseau, 1966**

colasi Damoiseau, 1966c: 145.

Type depository. MNHN (HT!).

Distribution. Madagascar.

***pauliani* Kleine, 1944**

pauliani Kleine, 1944: 157.

Type depositories. MNHN (LT!, PLT!), IRSNB (PLT!), DEI (PLT!).

Distribution. Madagascar.

Notes. Döbler & Gaedike (1995: 450) reported that the specimen held in MNHN was the holotype, but in his original description Kleine did not designate any of the ten syntypes as holotype. Damoiseau (1966c: 145) designated lectotype and paralectotypes on the material of MNHN and IRSNB.

Paryphobrenthus Kolbe, 1897

Paryphobrenthus Kolbe, 1897: 287.

Type species: *Paryphobrenthus beringei* Kolbe, 1897 = *Ceocephalus latirostris* Gerstäcker, 1855.

Distribution. Angola, Democratic Republic of the Congo (ex Zaire), Kenya, Mozambique, South Africa, Sudan, Tanzania.

latirostris (Gerstäcker, 1855)

latirostris Gerstäcker, 1855: 83 (*Ceocephalus*).

beringei Kolbe, 1897: 287 (*Paryphobrenthus*); syn. Kleine, 1920c: 10.

Type depositories. *C. latirostris*: MNHUB (HT!); *P. beringei*: MNHUB (LT!, PLT!).

Distribution. Angola, Democratic Republic of the Congo (ex Zaire), Kenya, Mozambique, South Africa, Sudan, Tanzania.

Notes. Lectotype of *P. beringei* (MNHUB) designated on a male specimen with the following labels: D. O. Afrika, Ugogo, v. Beringer & Jost. S. G./ type/ 67723/ *Paryphobrenthus beringei* n. sp. Kolbe. Paralectotypes: 1 female specimen (MNHUB), D. O. Afrika, Ugogo, v. Beringer & Jost. S. G./ *Paryphobrenthus latirostris* (Gerstaek.), R. Damoiseau det., 1964; 1 male and 2 female specimens (MNHUB), D. O. Afrika, Ugogo, v. Beringer & Jost. S. G.

Peraprophthalmus Kleine, 1923

Peraprophthalmus Kleine, 1923h: 49.

Type species: *Peraprophthalmus applicatus* Kleine, 1923.

Distribution. India, Myanmar [= Burma], Vietnam.

applicatus Kleine, 1923

applicatus Kleine, 1923h: 51.

Type depository. NMPC (HT).

Distribution. India, Myanmar [= Burma], Vietnam.

Periceocephalus Kleine, 1922

Periceocephalus Kleine, 1922f: 216.

Type species: *Periceocephalus adumbratus* Kleine, 1922.

Distribution. Madagascar.

adumbratus Kleine, 1922

adumbratus Kleine, 1922f: 216.

Type depository. MNHUB (?) (HT).

Distribution. Madagascar.

Notes. According to Kleine (1928a: 92) the holotype was in MNHUB, but we did not find it there.

Phacecerus Schoenherr, 1840

Phacecerus Schoenherr, 1840: 554.

Phaoecerus: Vitzthum, 1921b: 7 [lapsus].

Type species: *Phacecerus olivaceus* Boheman, 1840.

Distribution. Madagascar.

coquerelii (Fairmaire, 1871)

coquerelii Fairmaire, 1871: 43 (*Brentus*).

Type depository. MNHN (HT!).

Distribution. Madagascar.

Notes. Kleine (1938d) did not mention this species in his *Genera Insectorum*.

olivaceus Boheman, 1840

olivaceus Boheman (in Schoenherr, 1840: 555).

Type depository. MNHN (?) (HT).

Distribution. Madagascar.

Notes. According to Kleine (1928a: 90) the holotype was in MNHN, but we did not find it there.

planicaudatus (Chevrolat, 1839)

planicaudatus Chevrolat, 1839: 179 (*Brenthus*).

Type depository. MNHN (LT!).

Distribution. Madagascar.

Notes. Lectotype of *P. planicaudatus* (MNHN) designated on a specimen with the following labels: Madagascar/ Chevrolat 279/ type/ Museum Paris 1952, coll. R. Oberthür/ *Phaccerus planicaudatus* Chevrolat, det. R. Damoiseau.

Phocylides Pascoe, 1872

Phocylides Pascoe, 1872: 324.

Phoclidus: K. E. Schedl, 1959: 63 [lapsus].

Type species: *Phocylides collaris* Pascoe, 1872 = *Brenthus bicolor* Guérin-Méneville, 1831.

Distribution. Indonesia (Moluccas: Ambon, Bacan [= Batjan], Buru, Halmahera [= Gilolo], Seram [= Ceram]); New Guinea.

bicolor (Guérin-Méneville, 1831)

bicolor Guérin-Méneville, 1831: pl. VI, fig. 11 (*Brenthus*).

collaris Pascoe, 1872: 325 (*Phocylides*); syn. Damoiseau, 1972b: 276.

ebeninus Pascoe, 1872: 325 (*Phocylides*); syn. Damoiseau, 1972b: 276.

pascoei Macleay, 1887: 193 (*Phocylides*); syn. Damoiseau, 1972b: 276.

ebenius: Kleine, 1926b: 246 (*Phocylides*) [lapsus].

Type depositories. *P. bicolor*: MNHN (?) (HT); *P. collaris*: NHM (LT!, PLT!); *P. ebeninus*: NHM (HT!); *P. pascoei*: AMS (HT).

Distribution. Indonesia (Moluccas: Ambon, Bacan [= Batjan], Buru, Halmahera [= Gilolo], Seram [= Ceram]); New Guinea.

Notes. Kleine (1938d: 128) listed *P. bicolor* Guérin-Ménéville among the four species of *Phocylides*. On page 130 he listed the species *bicolor* Guérin-Ménéville in the genus *Ithystenus*. According to Kleine (1919e: 133) the species illustrated and described by Guérin-Ménéville was pars *Phocylides* and pars *Ithystenus*. We think he was wrong; in our opinion both the drawings (Guérin-Ménéville, [1831]: pl. VI, figs. 11 and 11a) and the description (Guérin-Ménéville, [1838]: 109) clearly refer to *Phocylides*. According to Damoiseau (1972b: 276) all four species listed by Kleine (1938d: 128) in *Phocylides* are *P. bicolor*.

According to Kleine (1928a: 88) the holotype of *P. bicolor* was in MNHN, but we did not find it there.

Lectotype of *P. collaris* (NHM) designated on a male specimen with the following labels: Syntype/ Type H.T./ Batchian/ Pascoe Coll. 93-60/ *Phocylides collaris* Pascoe Type. Paralectotype: 1 female specimen (NHM), with the same data as the lectotype.

Piazocnemis Lacordaire, 1866

Piazocnemis Lacordaire, 1866: 453.

Homales Kolbe, 1883c: 80; **syn. n.**

Synhomales Kolbe, 1883c: 79 [unjustified replacement name].

Piacocnemis: Kleine, 1920a: 57 [lapsus].

Type species: *Brenthus atratus* Klug, 1833.

Distribution. Comores Is., Madagascar.

Notes. In 1883 Kolbe published a paper where he created the subgenus *Homales* for the genus *Centrophorus* (1883a: 382). In the same year Kolbe created the genus *Homales* in the tribe Ithysteni (*sic!*) (1883c: 80). In this paper he added a short note (1883c: 79) explaining that the spelling *Homales* used in his previous paper (1883a: 382) was a misspelling of *Synhomales*. Kleine (1938d: 150) listed all the species, that Kolbe (1883c: 77-79) included in *Centrophorus*, in the genus *Piazocnemis*, except *C. fraterculus*, which is a synonym of *Orphanobrentus curvirostris* (Chevrolat).

***assidua* (Kolbe, 1883)**

assiduus Kolbe, 1883a: 382 (*Centrophorus*).

Type depository. MNHUB (HT!).

Distribution. Madagascar.

***assimilis* Kleine, 1922**

assimilis Kleine, 1922j: 169.

Type depository. MNHUB (?) (HT).

Distribution. Madagascar.

Notes. According to Kleine (1928a: 91) the holotype was in MNHUB, but we did not find it there.

***atrata* (Klug, 1833)**

atratus Klug, 1833: 107 (*Brenthus*).

holosericeofasciatus Boheman (in Schoenherr, 1840: 551) (*Brenthus*); syn. Chevrolat, 1839: 180.

holoseriofasciatus: Kleine, 1938d: 150 (*Piazocnemis*) [lapsus].

Type depositories. *B. atratus*: MNHUB (HT!); *B. holosericeofasciatus*: unknown.

Distribution. Madagascar.

***dives* Lacordaire, 1866**

dives Lacordaire, 1866: 454.

cylindricus Kolbe, 1883a: 388 (*Centrophorus dives* var.); syn. Schönfeldt, 1908: 61.

Type depositories. *P. dives*: IRSNB (?) (HT); *C. dives* var. *cylindricus*: unknown.

Distribution. Madagascar.

Notes. Kleine (1928a: 91) wrote that the type for *P. dives* should be in IRSNB, but we did not find it there.

encausta (Boheman, 1840)

encaustus Boheman (in Schoenherr, 1840: 552) (*Brenthus*).

Type depository. NHRS (ST).

Distribution. Madagascar.

falsa Kleine, 1925

falsus Kleine, 1925a: 194.

Type depositories. NME (ST).

Distribution. Madagascar.

nigrita (Klug, 1833)

nigritus Klug, 1833: 108 (*Brenthus*).

nigritis: Kleine, 1922j: 171 (*Piazocnemis*) [lapsus].

Type depository. MNHUB (HT!).

Distribution. Madagascar.

picicornis (Klug, 1833)

picicornis Klug, 1833: 107 (*Brenthus*).

Type depository. MNHUB (?) (HT).

Distribution. Comores Is., Madagascar.

Notes. According to Kleine (1928a: 91) the holotype was in MNHUB, but we did not find it there.

rectirostris (Kolbe, 1883)

rectirostris Kolbe, 1883a: 385 (*Centrophorus*).

Type depository. MNHUB (LT!, PLT!).

Distribution. Madagascar.

Notes. Lectotype of *C. rectirostris* (MNHUB) designated on a female specimen with the following labels: Madagascar int austr./ Type/ 65894. Paralectotypes: 1 female specimen (MNHUB), Madagascar int austr./ Hildebrandt/ *rectirostris* Kolbe/ type; 1 female specimen (MNHUB), 34558/ Type/ Madag. Goud./ *Centrophorus rectirostris* n. sp.

***striatopunctata* (Kolbe, 1883)**

striatopunctatus Kolbe, 1883a: 388 (*Centrophorus*).

Type depository. MNHUB (HT!).

Distribution. Madagascar.

***striatula* (Olivier, 1807)**

striatulus Olivier, 1807: 441 (*Brentus*).

compressipes Chevrolat, 1839: 181 (*Centrophorus*); syn. Gemminger & Harold, 1872: 2716.

striolatus: Chevrolat, 1839: 181 (*Brenthus*) [lapsus].

compressipes Boheman (in Schoenherr, 1840: 553) (*Brenthus*); syn. Schönfeldt, 1908: 61.

aeneolus Kolbe, 1883a: 387 (*Centrophorus*); syn. Kleine, 1922j: 169.

Type depositories. *B. striatulus*: unknown; *C. compressipes* Chevrolat: MNHN (LT!), NHRS (PLT); *B. compressipes* Boheman: NHRS (?) (ST); *C. aeneolus*: MNHUB (LT!, PLT!).

Distribution. Comores Is., Madagascar.

Notes. Lectotype of *C. compressipes* Chevrolat (MNHN) designated on a specimen with the following labels: Madagascar/ Chevrolat T. 259/ Type/ ex Musaeo G. Power/ Museum Paris 1952, coll. R. Oberthür.

Lectotype of *C. aeneolus* (MNHUB) designated on a specimen with the following labels: 34623/ Type/ Madagasc. Goud./ *aeneolus* Kolbe. Paralectotype: 1 specimen (MNHUB), Madag. int. austr. Hildebrandt/ Type/ 65895/ *aeneolus* Kolbe [head and prothorax are missing].

Gemminger & Harold (1872: 2716), Schönfeldt (1908: 61), and Kleine (1938d: 150) consider *C. compressipes* Chevrolat as a synonym of *B. striatulus* Olivier, but according to Kolbe (1883a: 388) and Senna (1892c: 58) the two species are distinct. We were unable to find the type for *B. striatulus*, so the problem remains unsolved.

***validirostris* (Kolbe, 1883)**

validirostris Kolbe, 1883a: 383 (*Centrophorus*).

Type depository. MNHUB (LT!, PLT!).

Distribution. Madagascar.

Notes. Lectotype of *C. validirostris* (MNHUB) designated on a male specimen with the following labels: Madagaskar int austr./ *validirostris*/ Type/ 65893. Paralectotype: 1 male specimen (MNHUB), Madagaskar int austr./ Hildebrandt/ *validirostris* Kolbe/ Type.

***Pithoderes* Calabresi, 1920**

Pithoderes Calabresi, 1920d: 42.

Type species: *Pithoderes gestri* Calabresi, 1920.

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea, Gabon, Ivory Coast, Republic of the Congo, Republic of Guinea, Sierra Leone.

***gestri* Calabresi, 1920**

gestri Calabresi, 1920d: 43.

corporatus Kleine, 1924a: 108 (*Pithoderes*); syn. De Muizon, 1955b: 906.

gestroi K. E. Schedl, 1961a: 57 (*Pithoderes*) [unjustified emendation].

Type depositories. *P. gestri*: MZUF (LT!), MSNG (PLT!); *P. corporatus*: MRAC (LT!, PLT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea, Gabon, Ivory Coast, Republic of the Congo, Sierra Leone.

Notes. Lectotype of *P. gestri* designated by Damoiseau (1967a: 324) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999). The lectotype has the following labels: Bénito, Congo Franc., determ. Calabresi/ *Pithoderes* n. g. *gestri* n. sp. typus ♂/ *Pithoderes gestroi* emend. typus ♂ Calabresi/ Ann. Mus. Genova, ser. 3 IX, 1920, typus! ♂/ ex coll. Senna/ "La Specola" Firenze, 7067.

Lectotype of *P. corporatus* (MRAC) designated on a specimen [incorrectly labelled as holotype] with the following labels: Holotypus/ Musée du Congo, Vieux-Kassongo, 15.XII.1910, Dr. Bequaert/ Typus!/ R. det. E 670/ *Pithoderes corporatus* Kln. typus!, 1922/ *Pithoderes gestroi* Calabresi, R. Damoiseau det., 1965. Paralectotypes: 3 specimens (MRAC), Paratypus/ Musée du Congo, Vieux-Kassongo, 15.XII.1910, Dr. Bequaert/ R. det. E 670/ *Pithoderes gestroi* Calabresi, R. Damoiseau det., 1966; 1 specimen (MRAC), Musée du Congo, Kasai, Makumbi, 11.X.1921, Dr. H. Schouteden/ *Pithoderes gestroi* Calabresi, R. Damoiseau det., 1966.

The quotation for the People's Republic of Congo (Sforzi, 1998b: 276) is incorrect and should be referred to as the Democratic Republic of the Congo.

guineensis De Muizon, 1960

guineensis De Muizon, 1960: 218.

Type depository. NHMB (HT!).

Distribution. Republic of Guinea.

Prodeutor Pascoe, 1862

Prodeutor Pascoe, 1862: 392.

Type species: *Prodeutor laminatus* Pascoe, 1862.

Distribution. Indonesia (Java; Moluccas: Seram [= Ceram]; Sulawesi).

fruhstorferi Senna, 1897

fruhstorferi Senna, 1897b: 241.

Type depositories. MZUF (LT!, PLT!), Fruhstorfer coll. (localization unknown) (PLT).

Distribution. Indonesia (Java, Sulawesi).

Notes. Lectotype of *P. fruhstorferi* (MZUF) designated on a male specimen with the following labels: S. Celebes, Bua-Kraeng, 5000', Febr. 1896, H. Fruhstorfer/ *Prodeutor fruhstorferi* cotypi! Senna/ Ann. Soc. Ent. Belg., XLI, 1897, cotypi!/ ex-coll. Senna/ "La Specola" Firenze, 7044. Paralectotypes: 3

male and 4 female specimens (MZUF), S. Celebes, Bua-Kraeng, 5000', Febr. 1896, H. Fruhstorfer/ ex-coll. Senna/ "La Specola" Firenze, 7044.

***laminatus* Pascoe, 1862**

laminatus Pascoe, 1862: 392.

setigera Senna, 1890: 102 (*Achrionota*); syn. Senna, 1897b: 237.

Type depositories. *P. laminatus*: NHM (LT!, PLT!); *A. setigera*: MZUF (HT!).

Distribution. Indonesia (Moluccas: Seram [= Ceram]; Sulawesi).

Notes. Lectotype of *P. laminatus* (NHM) designated on a male specimen with the following labels: Syntype/ Type H.T./ Manado Celebes/ Pascoe Coll. 93-60/ *Prodictor laminatus* Pascoe Type; Paralectotypes: 2 male and 1 female specimens (NHM), with the same label data as the lectotype.

***Proephebocer* Calabresi, 1920**

Proephebocer Calabresi, 1920a: 38.

Type species: *Proephebocer nitidissimus* Calabresi, 1920 = *Trachelizus populeus* Boheman, 1840.

Distribution. Madagascar.

***populeus* (Boheman, 1840)**

populeus Boheman (in Schoenherr, 1840: 495) (*Trachelizus*).

nitidissimus Calabresi, 1920a: 39 (*Proephebocer*); syn. Kleine, 1927a: 22.

Type depositories. *T. populeus*: NHRS (HT); *P. nitidissimus*: MZUF (LT!, PLT!).

Distribution. Madagascar.

Notes. Lectotype of *P. nitidissimus* designated by Bartolozzi *et al.* (1985: 99) (Article 74.5 of the International Code of Zoological Nomenclature, Fourth Edition, 1999).

temporalis Damoiseau, 1966

temporalis Damoiseau, 1966c: 140.

Type depositories. MNHN (HT!, AT!, PT!), IRSNB (PT!), PBZT (PT).

Distribution. Madagascar.

Pseudoceocephalus Kleine, 1920

Pseudoceocephalus Kleine, 1920c: 10.

Pseudocephalus: Kleine, 1934a: 302 [lapsus].

Rhomalorrhinus De Muizon, 1955b: 899; syn. Damoiseau, 1962a: 64.

Romalorrhinus: Zimmerman, 1994: 174 [lapsus].

Pseudoceocephalus: Zimmerman, 1994: 174 [lapsus].

Type species: *Brentus depressus* Lund, 1800.

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea (Bioko Is. [= Fernando Poo]), Gabon, Ghana, Ivory Coast, Republic of the Congo, Republic of Guinea, Togo, Uganda.

depressus (Lund, 1800)

depressus Lund, 1800: 83 (*Brentus*).

Type depository. ZMUC (ST).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea (Bioko Is. [= Fernando Poo]), Gabon, Ghana, Ivory Coast, Republic of the Congo, Republic of Guinea, Togo, Uganda.

formosus Kleine, 1934

formosus Kleine, 1934a: 302.

Type depositories. NMW (HT), DEI (PT!), SMTD (PT).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Gabon, Ghana, Republic of the Congo, Republic of Guinea.

Pseudomygaleicus De Muizon, 1960

Pseudomygaleicus De Muizon, 1960: 194.

Type species: *Ceocephalus georgei* Karsch, 1881.

Distribution. Angola, Benin, Central African Republic, Democratic Republic of the Congo (ex Zaire), Gabon, Ivory Coast, Kenya, Rwanda, São Thomé and Príncipe, Sudan, Tanzania, Uganda, Zambia.

alacer Damoiseau, 1967

alacer Damoiseau, 1967a: 286.

Type depositories. MRAC (HT!), IRSNB (PT!).

Distribution. Democratic Republic of the Congo (ex Zaire), Sudan.

bicolor (Kleine, 1920)

bicolor Kleine, 1920c: 45 (*Mygaleicus*).

Type depository. MNHUB (HT!).

Distribution. Democratic Republic of the Congo (ex Zaire), Rwanda.

brunneus De Muizon, 1960

brunneus De Muizon, 1960: 196.

Type depository. MNHN (HT!).

Distribution. Democratic Republic of the Congo (ex Zaire), Gabon, Ivory Coast, Tanzania.

georgei (Karsch, 1881)

georgei Karsch, 1881: 61 (*Ceocephalus*).

Type depository. MNHUB (HT!).

Distribution. São Thomé and Príncipe.

grandis (Damoiseau, 1963)

grandis Damoiseau, 1963a: 130 (*Mygaleicus*).

Type depositories. MRAC (HT!), AT!), IRSNB (PT!).

Distribution. Angola, Central African Republic, Democratic Republic of the Congo (ex Zaire), Equatorial Guinea, Ivory Coast.

Notes. The quotation for the People's Republic of Congo (Sforzi, 1998b: 274) is incorrect and should be referred to as the Democratic Republic of the Congo.

hirtus (Damoiseau, 1963)

hirtus Damoiseau, 1963a: 131 (*Mygaleicus*).

Type depository. MRAC (HT!).

Distribution. Democratic Republic of the Congo (ex Zaire).

humilis Damoiseau, 1963

humilis Damoiseau, 1963i: 44.

Type depository. MRAC (HT!, AT!).

Distribution. Democratic Republic of the Congo (ex Zaire).

nasutus Damoiseau, 1967

nasutus Damoiseau, 1967a: 280.

Type depositories. MRAC (HT!), IRSNB (PT!).

Distribution. Angola, Ivory Coast.

raptor (Kleine, 1934)

raptor Kleine, 1934a: 305 (*Mygaleicus*).

Type depository. DEI (HT!, AT!, PT!).

Distribution. Tanzania, Zambia.

somereni De Muizon, 1960 stat. res.

somereni De Muizon, 1960: 198 (*Mygaleicus*).

Type depositories. NHN (LT!), MNHN (PLT!).

Distribution. Kenya.

Notes. Damoiseau (1967a: 284) placed this species in synonymy with *P. bicolor* (Kleine) but after studying the type material, we consider *P. somereni* to be a valid species

Lectotype of *M. somereni* (NHM) designated on a male specimen with the following labels: Paratype/ Type/ on *Eucalyptus*/ van Someren, Teita Hills [= Taita Hills], Kenya Feb. 1955/ *Mygaleicus somereni* ♂ J. De Muizon det. Paralectotypes: 1 female specimen (NHM), Paratype/ Type/ on *Eucalyptus*/ van Someren, Teita Hills, Kenya Feb. 1955/ *Mygaleicus somereni* ♀ J. De Muizon det.; 1 specimen (MNHN), Kenya, Teita Hills, van Someren, Feb. 1955.

Damoiseau (1967a: 285) quoted Tanzania as type locality, although really the Taita Hills are in southern Kenya.

surdus Damoiseau, 1967

surdus Damoiseau, 1967a: 281.

Type depositories. MRAC (HT!), NHM (PT!).

Distribution. Angola, Benin, Ivory Coast, Uganda.

Pterygostomus Lacordaire, 1866

Pterygostomus Lacordaire, 1866: 448.

Type species: *Ceocephalus opacus* Chevrolat, 1839.

Distribution. Madagascar.

opacus (Chevrolat, 1839)

opacus Chevrolat, 1839: 179 (*Ceocephalus*).

Type depository. MNHN (LT!, PLT!).

Distribution. Madagascar.

Notes. Lectotype of *C. opacus* (MNHN) designated on a male specimen with the following labels: Madagascar/ Chevrolat T. 243/ type/ ex Musaeo G. Power/ Museum Paris 1952, coll. R. Oberthür. Paralectotype: 1 female specimen (MNHN), with the same label data as the lectotype.

Pyresthema Kleine, 1922

Pyresthema Kleine, 1922h: 152.

Type species: *Pyresthema kuntzeni* Kleine, 1922.

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Gabon.

kuntzeni Kleine, 1922

kuntzeni Kleine, 1922h: 153.

Type depository. MNHUB (LT!, PLT!).

Distribution. Cameroon, Democratic Republic of the Congo (ex Zaire), Gabon.

Notes. Damoiseau (1967a: 336) reported that he was unable to find the types in MNHUB, but we found them there.

Lectotype of *P. kuntzeni* (MNHUB) designated on a male specimen with the following labels: S. O. Kamerun, Lolodorf, 9.II-7.VI.95, L. Conradt/ type/ *Pyrestema kuntzeni* Kleine, det. Autor, 1914. Paralectotype: 1 female specimen (MNHUB), with the same label data as the lectotype.

Rhinopteryx Lacordaire, 1866

Rhinopteryx Lacordaire, 1866: 449.

Rhynopteryx: Senna, 1892c: 56 [lapsus].

Rhinopterix: Uhlig & Jaeger, 1995: 231 [lapsus].

Type species: *Ceocephalus foveipennis* J. Thomson, 1858.

Distribution. Tropical Africa; Arabia.

foveipennis (J. Thomson, 1858)

foveipennis J. Thomson, 1858: 119 (*Ceocephalus*).

errans Kolbe, 1883d: 36 (*Rhinopteryx*); syn. De Muizon, 1954: 331.

carinaerostris Kleine, 1914a: 190 (*Rhinopteryx*); syn. De Muizon, 1955b: 909.

carinoerostris: Kleine, 1960: 252 (*Rhinopteryx*) [lapsus].

Type depositories. *C. foveipennis*: MNHN (HT!); *R. errans*: MNHUB (HT!); *R. carinaerostris*: MZPW (HT).

Distribution. Tropical Africa; Arabia.

Notes. In MNHN (Oberthür coll.) there are five males and three females of *C. foveipennis* from Gabon and Guinea. In his original description J. Thomson only described one male specimen from Gabon, so as holotype we identified the male specimen from that locality of the size indicated by J. Thomson (1858: 119). As the lectotype designated by Orbach, Bartolozzi & Sforzi (1995: 22) comes from Guinea, it is not valid.

weneri Orbach, Bartolozzi & Sforzi, 1995

weneri Orbach, Bartolozzi & Sforzi, 1995: 18.

foveipennis Kleine, 1914a: 190 (non J. Thomson, 1858) (*Rhinopteryx*); syn. Orbach, Bartolozzi & Sforzi, 1995: 20.

Type depositories. MZUF (HT!, AT!), CMNH (PT), Orbach coll. (Israel) (PT).

Distribution. Kenya, Republic of the Congo, South Africa, Tanzania.

Schizotrachelus Lacordaire, 1866

Schizotrachelus Lacordaire, 1866: 454.

Schirotrachelus: Fairmaire, 1881c: 421 [lapsus].

Opisthenoplus Kleine, 1922p: 139; syn. Damoiseau, 1966d: 23.

Apisthenoplus: Browne, 1935: 92 [lapsus].

Schizotrachelizus: Kleine, 1939c: 106 [lapsus].

Type species: *Schizotrachelus brevicaudatus* Lacordaire, 1866.

Distribution. Taiwan; Borneo, India (including Andaman Is., Assam, Sikkim), Indonesia (Engano; Java; Mentawai; Moluccas: Banda; Sulawesi; Sumatra), Laos, Malaysia, Myanmar [= Burma], Philippines (Luzon, Mindanao, Negros, Sibuyan), Sri Lanka, Thailand, Vietnam; New Guinea.

Notes. The systematics of the species included in this genus needs accurate revision based on the study of the genital structures (Damoiseau, 1966e: 440).

brevicaudatus Lacordaire, 1866

brevicaudatus Lacordaire, 1866: 455.

Type depository. MCZ (HT).

Distribution. Borneo, Indonesia (Java, Sumatra), Philippines (Luzon, Negros, Sibuyan).

calabresii Kleine, 1922

calabresii Kleine, 1922b: 35.

Type depository. SMTD (HT).

Distribution. Philippines (Luzon).

carinensis Senna, 1892

carinensis Senna, 1892e: 483.

Type depository. MSNG (HT!).

Distribution. Malaysia, Myanmar [= Burma], Vietnam.

carinirostris Senna, 1892

carinirostris Senna, 1892e: 489.

Type depositories. MSNG (LT!, PLT!), MZUF (PLT!).

Distribution. Myanmar [= Burma].

Notes. Lectotype of *S. carinirostris* (MSNG) designated on a male specimen with the following labels: Bhamò, Birmania, VI.1886, Fea. Paralectotypes: 1 female specimen (MSNG), Thagatà, apr. 1887, Fea; 1 male specimen (MZUF), Bhamo, Birmania, Fea, VII, 1886/ *Schizotrachelus carinirostris* Senna, typus! / Ann. Mus. Gen., S. 2, XII, 1892, cotypus ♂ / ex-coll. Senna / "La Specola" Firenze, 7066.

cavus (Walker, 1859)

cavus Walker, 1859: 262 (*Ceocephalus*).

carus: Stebbing, 1914: 390 (*Ceocephalus*) [lapsus].

Type depository. NHM (LT!, PLT!).

Distribution. Borneo, India (including Andaman Is., Assam, Sikkim), Indonesia (Sumatra), Malaysia, Myanmar [= Burma], Philippines (Luzon, Mindanao, Negros), Sri Lanka, Thailand, Vietnam.

Notes. Lectotype of *C. cavus* (NHM) designated on a male specimen with the following labels: Syntype/ Type H.T./ Ceylon/ 50-106/ *Ceocephalus cavus* Walker a. n. H. (Type). Paralectotype: 1 female specimen (NHM), with the same label data as the lectotype.

celebicus Senna, 1899

celebicus Senna, 1899: 306.

Type depositories. MNHN (LT!, PLT!), IRSNB (PLT!).

Distribution. Indonesia (Java, Sulawesi).

Notes. Lectotype (MNHN) of *S. celebicus* designated on a male specimen with the following labels: S Celebes, Lompa-Battau, 3000 ft., Marz 1896, H. Fruhstorfer. Paralectotypes: 2 male specimens (MNHN), S Celebes, Lompa-

Battau, 3000 ft., Marz 1896, H. Fruhstorfer; 2 female specimens (MNHN), S Celebes, Bua-Kraeng, 5000 ft., Febr. 1896, H. Fruhstorfer; 2 male specimens (IRSNB), S Celebes, Lompa-Battau, 3000 ft., Marz 1896, H. Fruhstorfer/ *Schizotrachelus celebicus* Senna, R. Damoiseau det.; 1 male and 1 female specimen (IRSNB), S Celebes, Bua-Kraeng, 5000 ft., Febr. 1896, H. Fruhstorfer/ *Schizotrachelus celebicus* Senna, R. Damoiseau det./ Paralectotype.

cheesmani (Kleine, 1935)

cheesmani Kleine, 1935e: 304 (*Hypomiolista*).

Type depository. NHM (HT!).

Distribution. New Guinea.

cicatricosus (Kleine, 1922)

cicatricosus Kleine, 1922p: 141 (*Opisthenoplus*).

Type depository. DEI (HT!).

Distribution. Borneo, India, Indonesia (Java, Sumatra).

cognatus (Kleine, 1923)

cognatus Kleine, 1923j: 406 (*Opisthenoplus*).

Type depository. DEI (HT!).

Distribution. Indonesia (Java, Sumatra).

commodus (Kleine, 1923)

commodus Kleine, 1923i: 278 (*Opisthenoplus*).

Type depository. NHM (LT!, PLT!).

Distribution. Indonesia (Java), Malaysia.

Notes. Lectotype of *O. commodus* (NHM) designated on a male specimen with the following labels: Syntype/ Type/ Perak, Aug. 1908/ Brit. Mus. 1925-

88/ Maxwell's H. 3500''/ *Opisthenoplus commodus* Kln. ♂ Typus det. Aut. 1922. Paralectotype: 1 female specimen (NHM), Syntype/ Type/ Typus!/ Java/ Bowring 63-47/ *Opisthenoplus commodus* Kln., R. Kleine det. 1922.

***consanguineus* Senna, 1892**

consanguineus Senna, 1892e: 485.

Type depository. MSNG (LT!, PLT!).

Distribution. Myanmar [= Burma], Vietnam.

Notes. Lectotype (MSNG) designated on a male specimen with the following labels: Carin Chebà, 900-1100 m, V-XII.1888, L. Fea/ typus/ *Schizotrachelus consanguineus* Senna nov. subsp./ prépar. génit. N° 76 J 30, R. Damoiseau. Paralectotype: 1 specimen (MSNG), Carin Chebà, 900-1100 m, V-XII.1888, L. Fea.

***consobrinus* Lacordaire, 1866**

consobrinus Lacordaire, 1866: 456.

Type depository. MCZ (HT).

Distribution. Borneo, India (including Andaman Is.), Indonesia (Engano, Java, Sumatra), Myanmar [= Burma], Vietnam.

***fascinatus* (Kleine, 1922)**

fascinatus Kleine, 1922p: 140 (*Opisthenoplus*).

fasciatus: Kleine, 1922r: 162 (*Opisthenoplus*) [lapsus].

Type depositories. DEI (HT!), SMTD (?) (PT).

Distribution. Taiwan; India, Indonesia (Sumatra), Philippines (Luzon), Vietnam.

Notes. Kleine (1922p: 140) reported a paratype for this species in SMTD; according to O. Jaeger (SMTD) (2002, personal communication) two specimens of the species are present in the SMTD collections, but they are not labelled as types.

fecundus (Kleine, 1923)

fecundus Kleine, 1923e: 165 (*Opisthenoplus*).

Type depository. MNHLB (Fuller-Baker coll.) (HT).

Distribution. Philippines (Negros).

intermedius Senna, 1892

intermedius Senna, 1892e: 481.

Type depositories. MSNG (LT!, PLT!), MNHN (PLT!).

Distribution. Borneo, India, Indonesia (Java, Sumatra), Laos, Myanmar [= Burma], Vietnam.

Notes. Lectotype (MSNG) designated on a male specimen with the following labels: Bhamò, Birmania, VI.1885, Fea/ typus/ *Schizotrachelus intermedius* Senna typus!/ prépar. génit. N° 77 J 33, R. Damoiseau. Paralectotypes: 1 female specimen (MSNG), Carin Chebà, 900-1100 m, V-XII.1888, L. Fea; 1 specimen (MNHN), Birmania, Bhamò, Fea, VII.1886/ *Schizotrachelus intermedius* n. sp. Senna; 1 specimen (MNHN), Birmania, Bhamò, Fea, VI-II.1885; 1 female specimen, (MSNG), Bhamò, Birmania, VI.1885, Fea.

javanicus (Perroud, 1853) n. comb.

javanicus Perroud, 1853: 415 (*Ceocephalus*).

Type depository. MNHN (HT!).

Distribution. Indonesia (Java).

madens Lacordaire, 1866

madens Lacordaire, 1866: 455.

Type depositories. IRSNB (ST!), MCZ (ST).

Distribution. Borneo, India, Indonesia (Java; Mentawai; Moluccas: Banda; Sumatra), Laos, Malaysia, Philippines (Sibuyan), Vietnam.

Notes. Kleine (1938d) did not mention this species in his *Genera Insectorum*.

opulentus Kleine, 1943

opulentus Kleine, 1943b: 142.

Type depositories. HNHM (ST), MZPW (ST).

Distribution. New Guinea.

Schizuropterus Kleine, 1925

Schizuropterus Kleine, 1925a: 194.

Type species: *Ceocephalus reichei* Boheman, 1840 = *Brenthus reichei* Chevrolat, 1839.

Distribution. Madagascar.

reichei (Chevrolat, 1839)

reichei Chevrolat, 1839: 178 (*Brenthus*).

reichei Boheman (in Schoenherr, 1840 : 514) (*Ceocephalus*); syn. Schönfeldt, 1908: 60.

Type depository. *B. reichei* Chevrolat: MNHN (?) (HT); *C. reichei* Boheman: MNHN (?) (ST).

Distribution. Madagascar.

Notes. According to Kleine (1928a: 90) the holotype of *B. reichei* Chevrolat was in Oberthür's collection (now in MNHN and IRSNB), but we did not find it there. We did not even find the syntypes of *C. reichei* Boheman in MNHN, where they should be held.

Sennaiella Alonso-Zarazaga, Lyal,
Bartolozzi & Sforzi, 1999

Sennaiella Alonso-Zarazaga, Lyal, Bartolozzi & Sforzi (in: Alonso-Zarazaga & Lyal, 1999: 56)
[replacement name for *Holotrachelus*].

Holotrachelus Kleine, 1922m: 163.

Holotrachelus Kleine, 1923i: 276 (non Holm, 1898).

Type species: *Holotrachelus comparabilis* Kleine, 1923.

Distribution. Borneo, Indonesia (Java), Malaysia, Thailand, Vietnam.

Notes. Kleine (1922m: 163) mentioned *Holotrachelus* in a list of genera, but he did not describe it until one year later (1923i: 276).

comparabilis (Kleine, 1923)

comparabilis Kleine, 1923i: 277 (*Holotrachelus*).

incomparabilis: Kleine, 1923i: 278 (*Holotrachelus*) [lapsus].

Type depository. NHM (LT!, PLT!).

Distribution. Borneo, Malaysia, Thailand, Vietnam.

Notes. Lectotype of *H. comparabilis* (NHM) designated on a male specimen with the following labels: Syntype/ Type H.T./ Gunong Angsi, Negri Sembilan, 2000'-2700', April 1918/ Brit. Mus. 1923-88/ *Holotrachelus comparabilis* Kln Type ♂, det. Aut. 1922. Paralectotype: 1 female specimen (NHM), Syntype/ Type/ Typus ♀/Gunong Tahan, Pahang, 2500-3500 ft., May-July 1905/ *Holotrachelus comparabilis* Kln, R. Kleine det. 1922.

devisa (Kleine, 1926) n. comb.

devisus Kleine, 1926a: 62 (*Holotrachelus*).

Type depository. NHM (HT!).

Distribution. Indonesia (Java).

Storeosomus Lacordaire, 1866

Storeosomus Lacordaire, 1866: 454.

Storeosonnus: Kleine, 1922e: 156 [lapsus].

Type species: *Brenthus decollatus* Chevrolat, 1839.

Distribution. Tropical Africa (including Madagascar); Italy (I).

decollatus (Chevrolat, 1839)

decollatus Chevrolat, 1839: 179 (*Brenthus*).

Type depository. MNHN (LT!, PLT!).

Distribution. Madagascar.

Notes. Lectotype of *B. decollatus* (MNHN) designated on a male specimen with the following labels: Madagascar/ Chevrolat T. 262/ type/ ex Musaeo G. Power/ Museum Paris 1952, coll. R. Oberthür. Paralectotype: 1 female specimen (MNHN), with the same label data as the lectotype.

riisii (Labram & Imhoff, 1841)

riisii Labram & Imhoff, 1841: fasc. 8 [page not numbered] (*Ceocephalus*).

rissii: Lacordaire, 1866: 454 (*Storeosomus*) [lapsus].

occipitalis J. Thomson, 1858: 119 (*Rhyticephalus*); syn. De Muizon, 1955b: 898.

rissi: Kolbe, 1883: 239 (*Storeosomus*) [lapsus].

sansibaricus Kolbe, 1897: 287 (*Storeosomus*); syn. Kleine, 1927a: 62.

zanzibaricus: Kleine, 1922e: 156 (*Storeosonnus*) (sic!) [lapsus].

Type depositories. *C. riisii*: unknown; *R. occipitalis*: MNHN (HT!); *S. sansibaricus*: MNHUB (LT!, PLT!).

Distribution. Tropical Africa; Italy (I).

Notes. Lectotype of *S. sansibaricus* (MNHUB) designated on a male specimen with the following labels: Usambara, Derema, 350 m, 19.X-23.XI.91, Conradt S./ *Storeosomus sansibaricus* n. sp. Paralectotype: 1 specimen (MNHUB), Bagamoio, Zanzibar/ Fischer/ 63298/ *Storeosomus sansibaricus* Kolbe.

Stroggylosternum Kleine, 1922

Stroggylosternum Kleine, 1922p: 143.

Strongylosternum: Vitzthum, 1921b: 7 [lapsus].

Type species: *Stroggylosternum opacum* Kleine, 1922.

Distribution. Madagascar.

***nitens* Kleine, 1922**

nitens Kleine, 1922p: 144.

Type depositories. DEI (LT!), MNHUB (PLT!), SMTD (PLT).

Distribution. Madagascar.

Notes. Lectotype of *S. nitens* Kleine (DEI) designated on a specimen with the following labels: S. de la baie d'Antongil/ Coll. Kraatz, v. Schönfeldt det./ Syntypus/ *Stroggylosternum nitens* Kleine, det. Auct, 1914. Paralectotype: 1 specimen (MNHUB), Tananariva, Madagascar/ Paratypus/ *Stroggylosternum nitens* Kleine, Paratypus.

***opacum* Kleine, 1922**

opacum Kleine, 1922p: 144.

Type depository. SMTD (ST).

Distribution. Madagascar.

***Temnolaimus* Chevrolat, 1839**

Temnolaimus Chevrolat, 1839: 177.

Temnolaemus Agassiz 1846b: 363 [unjustified emendation].

Temnolaemus Gemminger & Harold, 1872: 2717 (non Agassiz, 1846) [unjustified emendation].

Type species: *Temnolaimus aeneicollis* Chevrolat, 1839.

Distribution. Madagascar.

***aeneicollis* Chevrolat, 1839**

aeneicollis Chevrolat, 1839: 177.

Type depository. MNHN (HT!).

Distribution. Madagascar.

Thoracobrenthus Damoiseau, 1961

Thoracobrenthus Damoiseau, 1961a: 286.

Type species: *Thoracobrenthus semistriatus* Damoiseau, 1961.

Distribution. Democratic Republic of the Congo (ex Zaire), Ivory Coast, Republic of the Congo.

semistriatus Damoiseau, 1961

semistriatus Damoiseau, 1961a: 287.

Type depository. MRAC (HT!).

Distribution. Democratic Republic of the Congo (ex Zaire), Ivory Coast, Republic of the Congo.

Uropteroides Kleine, 1922

Uropteroides Kleine, 1922f: 217.

Type species: *Brenthus douei* Montrouzier, 1861.

Distribution. Indonesia (Moluccas: Aru, Seram [= Ceram]); Australia, New Caledonia, New Guinea (including Bismarck Archipelago: New Britain), Polynesia, Solomon Is.

douei (Montrouzier, 1861)

douei Montrouzier, 1861: 874 (*Brenthus*).

Type depository. IRSNB (?) (ST).

Distribution. Indonesia (Moluccas: Aru, Seram [= Ceram]); Australia, New Caledonia, New Guinea (including Bismarck Archipelago: New Britain), Polynesia, Solomon Is.

Notes. Damoiseau (1966a: 24) reported that holotype and allotype were held in IRSNB, but in his original description Montrouzier did not designate any of the syntypes as holotype, and we did not find any type in IRSNB.

***gestroi* (Senna, 1894)**

gestroi Senna, 1894e: 562 (*Uropterus*).

Type depositories. MSNG (LT!, PLT!), MZUF (PLT!), MNHN (PLT!), NHM (PLT!).

Distribution. Indonesia (Moluccas: Aru, Seram [= Ceram] Australia, New Caledonia, New Guinea (including Bismarck Archipelago: New Britain), Solomon Is.

Notes. Lectotype of *U. gestroi* (MSNG) designated on a male specimen with the following labels: N Guinea, Ighibirei, VII-VIII.90, Loria/ typus/ *Uropterus gestroi* n. sp. typus! Paralectotypes: 2 male and 1 female specimens (MSNG), N Guinea, Ighibirei, VII-VIII.90, Loria; 1 female specimen (MSNG), N Guinea, Dilo, VI-VII.90, Loria; 1 specimen (MNHN), N. Guinea, Ighibirei, Loria, VII-VIII.90/ Typus/ *Uropterus gestroi* Senna Typus!; 1 male specimen (NHM), Syntype/ Co-type/ N Guinea, Ighibirei, Loria, VII-VIII.90; 1 male specimen (MZUF), N Guinea, Ighibirei, VII-VIII.90, Loria/ *Uropterus gestroi* Senna, cotypi Senna/ Ann. Mus. Genova, Ser. 2, XIV, 1894, cotypi ♂ ♀/ ex-coll. Senna/ "La Specola" Firenze, 7060; 1 male and 1 female specimen (MZUF), N Guinea, Ighibirei, VII-VIII.90/ *Uropteroides gestroi* Senna, R. Damoiseau/ ex-coll. Senna/ "La Specola" Firenze, 7060.

***novaecaledoniae* Damoiseau, 1962**

novaecaledoniae Damoiseau, 1962c: 287.

Type depository. IRSNB (HT!, PT!).

Distribution. New Caledonia.

Notes. According to Damoiseau (1962c: 288) the holotype is held in IRSNB, and in his description he writes about two female specimens. We can presume that the specimen labelled as "paratype" in IRSNB is actually a true paratype even if not clearly indicated as such in the description.

Uroptera Berthold, 1827

Uroptera Berthold, 1827: 383.

Uropterus Latreille, 1828: 593; syn. Kleine, 1927a: 61.

Ceocephalus Schoenherr, 1833: 357 (non Guérin-Méneville, [1833]); syn. Dejean, 1834: 244.

Gecocephalus: Motschulsky, 1863: 524 [lapsus].

Type species: *Ceocephalus appendiculatus* Boheman, 1833.

Distribution. Mauritius Is., Reunion Is.

Notes. Damoiseau (1963e: 460) states that the type species of the genus is *Brentus caudatus* Latreille, 1817, but Latreille (1817: plate 13, fig. 7) illustrated the species without providing the Latin name, only giving the vernacular name "brente à queue". The type species for the genus *Uroptera* is *Ceocephalus appendiculatus* Boheman, 1833 as designated by Kleine (1938d: 141).

acuminata (Damoiseau, 1963)

acuminatus Damoiseau, 1963e: 463 (*Ceocephalus*).

Type depository. IRSNB (HT!).

Distribution. Mauritius Is.

appendiculata (Boheman, 1833)

appendiculatus Boheman (in Schoenherr, 1833: 358) (*Ceocephalus*).

caudatus Boheman (in Schoenherr, 1840: 511) (*Ceocephalus*); syn. Damoiseau, 1963e: 460.

codicillus Boheman (in Schoenherr, 1840: 512) (*Ceocephalus*); syn. Damoiseau, 1963e: 460.

Type depositories. *C. appendiculatus*: NHRS (HT); *C. caudatus*: IRSNB (ST!); *C. codicillus*: NHRS (HT).

Distribution. Mauritius Is. (?), Reunion Is.

Notes. According to K. E. Schedl (1961c: 203) *C. caudatus* only occurs in Reunion Is. and not in Mauritius Is.

vinsoni Kleine, 1938

vinsoni Kleine, 1938f: 291.

Type depositories. DEI (LT!, PLT!), MNHN (PLT!).

Distribution. Mauritius Is.

Notes. Lectotype and paralectotypes designated by Damoiseau (1989b: 79).

Zetophloeus Lacordaire, 1866

Zetophloeus Lacordaire, 1866: 460.

Type species: *Brenthus pugionatus* Chevrolat, 1839.

Distribution. Madagascar.

guttifer (Boheman, 1840)

guttifer Boheman (in Schoenherr, 1840: 548) (*Brenthus*).

Type depository. NHRS (HT).

Distribution. Madagascar.

pugionatus (Chevrolat, 1839)

pugionatus Chevrolat, 1839: 178 (*Brenthus*).

Type depositories. MNHN (LT!, PLT!), NHRS (PLT).

Distribution. Madagascar.

Notes. Lectotype of *B. pugionatus* (MNHN) designated on a specimen with the following labels: Madagascar/ Chevrolat 281/ type/ ex Musaeo G. Power/ Museum Paris 1952, coll. R. Oberthür. Paralectotypes: 2 specimens (MNHN), with the same label data as the lectotype.

RHYTICEPHALINI Kleine, 1922

Rhyticephalini Kleine, 1922m: 163.

Ischyromerus Labram & Imhoff, 1838

Ischyromerus Labram & Imhoff, 1838: fasc. 2 [page not numbered].

Rhyticephalus Chevrolat, 1839: 174; syn. Kleine, 1927a: 71.

Rhytitrachelus: Schoenherr, 1840: 520 [lapsus].

Rhytidocephalus Agassiz, 1846b: 327 [unjustified emendation].

Rhytidocephalus Gemminger & Harold, 1872: 2714 (non Agassiz, 1846) [unjustified emendation].

Rhyticephalue: Kleine, 1927f: 343 [lapsus].

Type species: *Brenthus madagascariensis* Labram & Imhoff, 1838.

Distribution. Comores Is., Madagascar.

dignus (Kleine, 1925)

dignus Kleine, 1925a: 195 (*Rhyticephalus*).

affinis Kleine, 1925a: 195 (*Rhyticephalus*) [original incorrect spelling].

Type depository. NHM (HT!).

Distribution. Madagascar.

Notes. This species was incorrectly described as *R. affinis*, but the legend for the drawing gives the correct name *R. dignus*. The latter name was used by Kleine in all his subsequent catalogues.

madagascariensis Labram & Imhoff, 1838

madagascariensis Labram & Imhoff, 1838: fasc. 2 [page not numbered].

brevicornis Chevrolat, 1839: 174 (*Rhyticephalus*); syn. Gemminger & Harold, 1872: 2714.

aulaconotus Chevrolat, 1839: 175 (*Rhyticephalus*); syn. Kleine, 1927a: 71.

madagascariensis Perroud, 1853: 427 (non Labram & Imhoff, [1838]) (*Rhyticephalus*); syn. Kleine, 1927a: 71.

Type depositories. *I. madagascariensis*: ETHZ (HT); *R. brevicornis*:

MNHN (LT!, PLT!); *R. aulaconotus*: MNHN (LT!, PLT!); *R. madagascariensis* Perroud: MNHN (HT!).

Distribution. Comores Is., Madagascar.

Notes. Lectotype of *R. brevicornis* (MNHN) designated on a male specimen with the following labels: Madagascar/ Chevrolat 236/ type/ ex Musaeo G. Power/ Museum Paris 1952, coll. R. Oberthür. Paralectotypes: 1 specimen (MNHN), Madagascar/ Chevrolat 236/ ex Musaeo G. Power/ Museum Paris 1952 coll. R. Oberthür; 1 female specimen (MNHN), Madagascar/ Chevrolat 236/ type/ ex Musaeo G. Power/ Museum Paris, 1952, coll. R. Oberthür; 1 female specimen (MNHN), Madagascar/ Chevrolat 236/ ex Musaeo G. Power/ Museum Paris 1952, coll. R. Oberthür.

Lectotype of *R. aulaconotus* (MNHN) designated on a male specimen with the following labels: Madagascar/ Chevrolat 238/ type/ ex Musaeo G. Power/ Museum Paris 1952, coll. R. Oberthür. Paralectotype: 1 female specimen (MNHN), with the same label data as the lectotype.

TYCHAEINI Schönfeldt, 1908

Tychaeidae Schönfeldt, 1908: 48.

Tychaeini Schönfeldt, 1910a: 31.

Nemorhinus Schoenherr, 1823

Nemorhinus Schoenherr, 1823: 1137.

Tychaeus Fischer de Waldheim, 1823: 266; syn. Alonso-Zarazaga & Lyal, 1999: 54.

Nematorhinus Agassiz, 1846b: 247 [unjustified emendation].

Type species: *Brentus myrmecophaga* Herbst, 1797.

Distribution. Belize, Costa Rica, Dominican Republic, Jamaica, Nicaragua, Panama; Bolivia, Brazil, Peru, Venezuela.

myrmecophaga (Herbst, 1797)

myrmecophaga Herbst, 1797: 185 (*Brentus*).

curvidens Lund, 1800: 59 (*Brentus*); syn. Schoenherr, 1833: 341.

nasutus Olivier, 1807: 436 (non Fabricius, 1781) (*Brentus*); syn. Schoenherr, 1833: 341.

signatus Sharp, 1895a: 63 (*Tychaeus curvidens* var.); **syn. n.**

Type depositories. *B. myrmecophaga*: MNHUB (Herbst coll.) (?) (HT); *B. curvidens*: ZMUC (ST); *T. curvidens* var. *signatus*: NHM (LT!, PLT!).

Distribution. Belize, Costa Rica, Dominican Republic, Jamaica, Nicaragua, Panama; Bolivia, Brazil, Peru, Venezuela.

Notes. According to Kleine (1928a: 87) the type for *B. myrmecophagus* could be in MNHUB, but we did not find it there.

Lectotype of *T. curvidens* var. *signatus* (NHM) designated on a male specimen with the following labels: Bugaba, 800-1500 ft., Champion/ B. C. A., Col. IV, 6. *Tychaeus curvidens* Lund/ *Tychaeus curvidens* var. *signatus*. Paralectotypes: 1 male specimen (NHM), with the same locality as the lectotype; 2 male specimens (NHM), V. de Chiriqui, 25-4000 ft., Champion/ B. C. A., Col. IV, 6. *Tychaeus curvidens* Lund; 1 female specimen (NHM), Chontales, Nicaragua, T. Belt; 1 female specimen (NHM), Chontales, Janson/ B. C. A., Col. IV, 6. *Tychaeus curvidens* Lund.

ULOCERINAE Schoenherr, 1823

Ulocerides Schoenherr, 1823: 1137.

Ulocerides: Latreille, 1828: 592 [lapsus].

Uloceroidae Agassiz, 1846b: 383.

Uloceridae Imhoff, 1856: 168.

Ulocerinae Sharp, 1895a: 79.

Ulocerini Heyne & Taschenberg, 1907a: 234.

Uloceroidea Ienistea, 1986: 32.

Ulocerides: Alonso-Zarazaga & Lyal, 1999: 56 [lapsus].

Ulocerus Schoenherr, 1823

Ulocerus Schoenherr, 1823: 1137.

Ulocerus Dalman, 1824: 25 (non Schoenherr, 1823); syn. Alonso-Zarazaga & Lyal, 1999: 56.

Cladionus Berthold, 1827: 384; syn. Alonso-Zarazaga & Lyal, 1999: 56.

Cladione Schoenherr, 1833: 373; syn. Alonso-Zarazaga & Lyal, 1999: 56.

Type species: *Ulocerus lacerosus* Schoenherr, 1823.

Distribution. Colombia, Costa Rica, Cuba, Honduras, Mexico, Nicaragua, Panama; Argentina, Bolivia, Brazil, French Guyana, Paraguay, Uruguay (?).

***albiornatus* Izecksohn, 1962**

albiornatus Izecksohn, 1962: 3.

Type depository. DZSP (HT).

Distribution. Brazil.

***amazonicus* Damoiseau, 1962**

amazonicus Damoiseau, 1962b: 20.

Type depositories. NHRS (HT), IRSNB (PT!).

Distribution. Brazil.

***aurivilliusi* Senna, 1896**

aurivilliusi Senna, 1896c: 216.

aurivillii: Schönfeldt, 1908: 80 (*Ulocerus*) [lapsus].

Type depositories. IRSNB (ST!), NHRS (?) (ST).

Distribution. Brazil.

Notes. Senna (1896c: 217) reported that a syntype of *U. aurivilliusi* was in NHRS, but it is not listed in the NHRS web site.

***bicaudatus* Suffrian, 1870**

bicaudatus Suffrian, 1870: 203.

Type depository. MNHC (HT!).

Distribution. Cuba.

***boisduvali* Senna, 1896**

boisduvali Senna, 1896c: 215.

boisduvalii: Schönfeldt, 1908: 80 (*Ulocerus*) [lapsus].

Type depository. IRSNB (HT!).

Distribution. French Guyana.

***cinereus* Senna, 1896**

cinereus Senna, 1896c: 208.

Type depositories. NHM (LT!, PLT!), IRSNB (PLT!), MZUF (PLT!), NHRS (?) (PLT).

Distribution. Brazil.

Notes. Lectotype of *U. cinereus* (NHM) designated on a specimen with the following labels: syntype/ Fry, Rio Fan/ Fry coll. 1905.100/ *Ulocerus cinereus* Senna ♂ cotypi!. Paralectotypes: 1 male specimen (NHM), syntype/ 10268/ Espirito Santo/ Fry coll. 1905.100/ *Ulocerus cinereus* Senna, Brazilia, cotypus; 1 female specimen (MZUF), Brasile, ex Mus. Tring/ *Ulocerus cinereus* Senna ♀ cotypus/ Ann. Soc. Ent. Belg., XL, 1896, cotypus ♀/ ex-coll. Senna/ "La Specola" Firenze, 7059; 1 male specimen [incorrectly labelled as holotype] (IRSNB), Brésil/ Coll. Dejean, Coll. Roelofs/ *Cinereus Latreille*/ *Ulocerus cinereus* Latreille, h. in Brasilia/ *Ulocerus cinereus* Senna ♂ cotypus/ Holotype/ Prépar. génit. No. 1206, R. Damoiseau/ *Ulocerus cinereus* Senna, R. Damoiseau det.; 1 female specimen (IRSNB), Brésil/ Coll. Roelofs/ *Ulocerus cinereus* Senna ♀ cotypus/ Allotype/ Prépar. génit. No. 1219/ *Ulocerus cinereus* Senna, R. Damoiseau det.

Senna (1896c: 209) reported that a syntype of *U. cinereus* was in NHRS but it is not listed in the NHRS web site.

***dalmani* Senna, 1896**

dalmani Senna, 1896c: 203.

dalmanni: Schönfeldt, 1908: 80 (*Ulocerus*) [lapsus].

Type depository. IRSNB (LT!, PLT!).

Distribution. Bolivia, French Guyana.

Notes. Lectotype of *U. dalmani* (IRSNB) designated on a male specimen [incorrectly labelled as holotype] with the following labels: Cayenne/ Coll.

Roelofs/ *Ulocerus Dalmani* Typus ♂ ♀ Senna/ Prépar. génit. No. 1204/ *Ulocerus dalmani* Senna ♂, R. Damoiseau rev. 1961/ Holotype. Paralectotype: 1 female specimen [head missing] (IRSNB), Cayenne/ Coll. Roelofs/ rostre manque/ Prépar. génit. No. 1205/ *Ulocerus dalmani* Senna ♀, R. Damoiseau rev. 1961/ Allotype.

***damnosus* Damoiseau, 1962**

damnosus Damoiseau, 1962b: 23.

Type depository. IRSNB (HT!).

Distribution. Brazil.

***fasciatus* Izecksohn, 1963**

fasciatus Izecksohn, 1963: 415.

Type depository. MNRJ (Campos Seabra coll.) (HT, AT, PT).

Distribution. Brazil.

***fryi* Senna, 1896**

fryi Senna, 1896c: 204.

Type depository. NHM (HT!).

Distribution. Brazil.

***immundus* Dalman, 1824**

immundus Dalman, 1824: 27.

Type depository. NHRS (HT).

Distribution. Argentina, Brazil, French Guyana.

***laceratus* Schoenherr, 1823**

laceratus Schoenherr, 1823: 1137.

laceratus Dalman, 1824: 26 (*Ulocerus*); syn. Alonso-Zarazaga & Lyal, 1999: 56.

tetraurus Labram & Imhoff, 1838: fasc. 4 [page not numbered] (*Ulocerus*); syn. Damoiseau, 1962b: 23.

pannosus Boheman (in Schoenherr, 1840: 589) (*Ulocerus*); syn. Senna, 1896c: 218.

lacertosus: Chenu, 1860: 216 (*Ulocerus*) [lapsus].

Type depositories. *U. lacerosus*: unknown; *U. laceratus*: NHRS (HT); *U. tetraurus*: unknown; *U. pannosus*: NHRS (HT).

Distribution. Argentina, Brazil.

Notes. The specimen of *U. laceratus* described by Dalman may also be the type for *laceratus* Schoenherr. Labram & Imhoff (1838) reported that the type of *U. tetraurus* was in «Mus Werthermann, Basileensis». According to E. Sprecher (NHMB) (2002, personal communication) there are no types of this species in NHMB. Part of the Imhoff collection is now in MCZ, but this species is not listed in the type catalogue on their web site.

laticornis Sharp, 1895

laticornis Sharp, 1895a: 79.

Type depository. NHM (LT!, PLT!).

Distribution. Nicaragua; Brazil, Colombia.

Notes. Lectotype of *U. laticornis* (NHM) designated on a male specimen with the following labels: *Ulocerus laticornis* ♂ Type D. S./ Chontales, Nicaragua, T. Belt/ syntype/ type/ Sharp Coll. 1905-313. Paralectotypes: 1 female specimen (NHM), *Ulocerus laticornis* Type D. S./ Chontales, Nicaragua, T. Belt/ syntype/ type/ B.C.A., Col. IV, 6. *Ulocerus laticornis* Sharp; 1 specimen (NHM), Syntype/ Chontales, Nicaragua, T. Belt/ *Ulocerus laticornis* Type D. S./ B.C.A., Col. IV, 6. *Ulocerus laticornis* Sharp.

longicornis Senna, 1896

longicornis Senna, 1896c: 197.

Type depositories. NHM (LT!, PLT!), IRSNB (PLT!).

Distribution. Brazil.

Notes. Lectotype of *U. longicornis* (NHM) designated on a male specimen

with the following labels: syntype/ 2730/ Fry, Rio Jan./ Fry coll. 1900.100/ *Ulocerus longicornis* Senna ♂ cotypi! Paralectotypes: 1 specimen (NHM), syntype/ Fry, Rio Jan./ Fry coll. 1900.100; 1 specimen (NHM), syntype/ Fry, Rio Jan./ Fry coll. 1900.100/ *Ulocerus longicornis* Senna, Brazilia, co-typus/ *Ulocerus amazonicus* Damoiseau, 1983, R. W. Donoghue det.; 1 male specimen [incorrectly labelled as holotype] (IRSNB), Brésil/ Coll. Roelofs/ *Ulocerus longicornis* (Reiche) ♀/ nov. sp. *Longicornis* Reh Brasil/ *Ulocerus longicornis* Senna cotypus! ♂/ Holotype/ Prépar. génit. 1207; 1 male specimen (IRSNB), Bahia/ Coll. Castelnau, Coll. Roelofs/ *Ulocerus longicornis* Reiche in litt. Brésil/ *Ulocerus longicornis* Senna Cotype! R. Damoiseau rev. 1961/ Paratype.

***longirostris* Izecksohn, 1963**

longirostris Izecksohn, 1963: 416.

Type depository. MNRJ (Campos Seabra coll.) (HT, AT).

Distribution. Brazil.

***luridus* Senna, 1896**

luridus Senna, 1896c: 214.

Type depositories. NHM (LT!), IRSNB (PLT!).

Distribution. Brazil.

Notes. Lectotype of *U. luridus* (NHM) designated on a male specimen [incorrectly labelled as holotype] with the following labels: Holotype/ Fry, Rio Jan./ Fry Coll. 1905.100/ *Ulocerus luridus* ♂ Typus! Senna. Paralectotype: 1 female specimen (IRSNB), Brazil/ Coll. Roelofs/ déterm. Senna/ *Ulocerus luridus* Senna ♀ typus!/ Prépar. génit. 1232/ Type.

***mexicanus* Sharp, 1895**

mexicanus Sharp, 1895a: 80.

Type depository. NHM (LT!, PLT!).

Distribution. Honduras, Mexico, Nicaragua; Colombia.

Notes. Lectotype of *U. mexicanus* (NHM) designated on a male specimen with the following labels: *Ulocerus mexicanus* ♂ Type D. S./ Cordova/ Mexico, Salle Coll./ syntype/ type/ Sharp Coll. 1905-313. Paralectotypes: 1 male specimen (NHM), *Ulocerus mexicanus* ♂ Type D. S./ Cordova/ Mexico, Salle Coll./ syntype/ type/ *mexicanus* Boh.; 1 female specimen (NHM), syntype/ type/ 2549/ Playa Vicente/ Salle Coll./ *Ulocerus mexicanus* ♀, Type D. S.

***nigrolineatus* Senna, 1896**

nigrolineatus Senna, 1896c: 212.

Type depositories. IRSNB (LT!, PLT!), NHRS (?) (PLT).

Distribution. French Guyana.

Notes. Lectotype of *U. nigrolineatus* (IRSNB) designated on a male specimen [incorrectly labelled as holotype] with the following labels: Cayenne/ Coll. Lacordaire/ Coll. Dejean, Coll. Roelofs/ *Ulocerus signaticornis* mihi D. Lacordaire h. Cayenna/ *Ulocerus nigrolineatus* Senna 2 ♂♂ 1 ♀ cotypi/ *Ulocerus nigrolineatus* Senna ♂ R. Damoiseau det. 1961/ Holotype/ Prép. génit. No. 1216 R. Damoiseau. Paralectotypes: 1 male specimen (IRSNB), Cayenne/ Coll. Lacordaire/ *Ulocerus nigrolineatus* Senna ♂ R. Damoiseau rev. 1961/ Paratype; 1 female specimen (IRSNB), Cayenne/ Coll. Lacordaire/ Prép. génit. No. 1215/ *Ulocerus nigrolineatus* Senna ♀ R. Damoiseau rev. 1961/ Allotype.

Senna (1896c: 213) reported that a syntype of *U. nigrolineatus* was in NHRS, but it is not listed in the NHRS web site.

***recticauda* Senna, 1896**

recticauda Senna, 1896c: 199.

recticauda: Haedo Rossi, 1967: 173 (*Ulocerus*) [lapsus].

Type depositories. MZUF (HT!, PT!), NHM (?) (PT), NHRS (?) (PT).

Distribution. Brazil, Paraguay.

Notes. Senna (1896c: 199) reported that paratypes of *U. recticauda* were in NHRS, but they are not listed in the NHRS web site; he also wrote that paratypes were in NHM, but we not find them there.

***rostratus* Senna, 1896**

rostratus Senna, 1896c: 205.

Type depository. NHM (HT!).

Distribution. Colombia.

***severini* Senna, 1896**

severini Senna, 1896c: 200.

severinii: Schönfeldt, 1908: 81 (*Ulocerus*) [lapsus].

Type depository. IRSNB (LT!, PLT!).

Distribution. Brazil.

Notes. Lectotype of *U. severini* (IRSNB) designated on a male specimen [incorrectly labelled as holotype] with the following labels: Bahia/ Coll. Castelnau, Coll. Roelofs/ *Ulocerus severini* Senna cotypus! ♂/ Prépar. génit. No. 1201/ *Ulocerus severini* Senna R. Damoiseau rev. 1961/ Holotype. Paralectotypes: 1 female specimen (IRSNB), Bahia/ Coll. Castelnau, Coll. Roelofs/ Prépar. génit. No. 1202/ Allotype; 2 male specimens (IRSNB), Bahia/ Coll. Castelnau, Coll. Roelofs/ Paratype; 3 male specimens (IRSNB), Bahia/ Coll. Roelofs/ Paratype.

***sharpi* Senna, 1896**

sharpi Senna, 1896c: 207.

Type depository. NHM (HT!).

Distribution. Brazil.

***sordidus* Sharp, 1895**

sordidus Sharp, 1895a: 80.

Type depository. NHM (HT!).

Distribution. Costa Rica, Mexico (?), Panama; French Guyana.

***squalidus* Dalman, 1824**

squalidus Dalman, 1824: 28.

Type depository. NHRS (HT).

Distribution. Argentina, Brazil, Paraguay, Uruguay (?).

***terrosus* Senna, 1896**

terrosus Senna, 1896c: 213.

Type depository. IRSNB (HT!).

Distribution. Brazil, Uruguay (?).

***urocerophorus* Izecksohn, 1963**

urocerophorus Izecksohn, 1963: 413.

Type depository. MNRJ (Campos Seabra coll.) (HT).

Distribution. Brazil.

INCERTAE SEDIS**Genera*****Pronomacrus* Kleine, 1929**

Pronomacrus Kleine, 1929a: 51 [published as synonym and not subsequently validated].

Notes. See Alonso-Zarazaga & Lyal (1999: 56).

***Stenorhynchus* Berthold, 1827**
(non Lamarck, 1818, nec Hemprich, 1820)

Stenorhynchus Berthold, 1827: 384 (non Lamarck, 1818, nec Hemprich, 1820).

Notes. See Alonso-Zarazaga & Lyal (1999: 56).

***Trachelizus* Gyllenhal, 1833**

Trachelizus Gyllenhal (in Schoenherr, 1833: 334).

Notes. See Alonso-Zarazaga & Lyal (1999: 56).

Species

***attenuatus* Chevrolat**

Notes. Desbrochers des Loges (1891: 4) mentioned this species as *Orychodes attenuatus* (Chevr.) from Bengala (Konbir) but we were unable to find any description for this species. Neither Schönfeldt (1910a) nor Kleine (1938d) mention this species.

***collaris* Castelnau, 1840**

collaris Castelnau, 1840: 295 (*Brenthus*).

Type depositary. Unknown.

Distribution. Philippines.

Notes. Neither Schönfeldt (1908, 1910a) nor Kleine (1938d) mention this species.

***decorus* Kleine, 1939**

decorus Kleine, 1939a: 32 (*Amerismus*).

Type depositary. Unknown.

Distribution. Madagascar.

Notes. Kleine (1939a) included this species in a key to the genus *Amerismus*, but never described it. Later, Schedl (1961c: 198) proposed including *A. decorus* in the genus *Proephebocerus*. According to Damoiseau (1966c: 139) the species must be considered *in litteris*, but as Kleine separated *A. decorus* from *A. admirabilis* by a few distinctive characters in the key, we consider the species to be valid. Nevertheless, as we were unable to examine the type for this taxon, we have included it among the "*incertae sedis*".

facilis F. Walker, 1859

facilis F. Walker, 1859: 262 (*Arrhenodes*).

Type depository. Unknown.

Distribution. Sri Lanka (?).

Notes. According to Kleine (1923f: 90) the type depository is unknown and the affinities of the species are hard to understand; neither did he mention this species in his *Genera Insectorum* (1938d). We did not find the type in NHM. Damoiseau (1971b: 268) questions if it really is a brentid.

fasciatus Schoenherr, 1833

fasciatus Schoenherr, 1833: 369 (*Brentus*) [replacement name for *cylindricornis* Duméril].
cylindricornis Duméril, 1817: 333 (non Fabricius, 1787) (*Brentus*).

Type depository. Unknown.

Distribution. Trinidad Is.

Notes. Schoenherr (1833: 369) mentions this species without any description and gives *B. cylindricornis* Duméril as synonym. Neither Schönfeldt (1908, 1910a) nor Kleine (1938d) mention this species.

filicornis Boheman, 1829

filicornis Boheman, 1829: 28 (*Brentus*).

Type depository. NHRS (HT).

Distribution. Unknown.

Notes. Schönfeldt (1910a: 33) placed this species in the genus *Baryrhynchus*, whilst Kleine (1938d) did not mention it in his *Genera Insectorum*. We did not examine the type for this taxon, so we provisionally include it among the “*incertae sedis*”.

filicornis Gyllenhal, 1833

filicornis Gyllenhal, 1833: 329 (*Arrhenodes*).

Type depository. Unknown.

Distribution. N America (?).

Notes. Schönfeldt (1910a: 33) placed this species in the genus *Baryrhynchus* as a synonym of *B. filicornis* Boheman, whilst Kleine (1938d) did not mention it in his *Genera Insectorum*. We did not examine the type for this taxon, so we provisionally include it among the “*incertae sedis*”.

monilis Olivier, 1807

monilis Olivier, 1807: 434, pl. 2, fig. 12 (non Fabricius, 1787) (*Brentus*).

monilicornis Schoenherr, 1833: 356 (*Brenthus*); syn. Gemminger & Harold, 1872: 2711.

Type depository. *B. monilis*: unknown.

Distribution. Dominican Republic.

Notes. The name *monilicornis* Schoenherr seems to be a simple replacement name for *B. monilis* Olivier, 1807 (non Fabricius, 1787). As Schoenherr (1833: 356, footnote) states that he never saw the specimen, he probably thought that the specimen illustrated in Olivier's plate (pl. 2, fig. 12) as *B. monilis* Fabricius was really a different species.

Schönfeldt (1908: 45) included *B. monilis* Olivier (with *B. monilicornis* Schoenherr as synonym) in the genus *Belopherus*. Kleine (1938d) did not mention *B. monilicornis* among the species listed in his *Genera Insectorum* and placed *B. monilis* Olivier among the “*species sedis incertae*”. Damoiseau (1962b: 32) did not mention *B. monilis* and *B. monilicornis* in the list of the species belonging to the genus *Belopherus*.

***planicollis* Schoenherr, 1826**

planicollis Schoenherr, 1826: 69 (*Brenthus*).

Type depository. Unknown.

Distribution. Unknown.

Notes. This species was quoted by Castelnau (1840: 295), but neither Schönfeldt (1908, 1910a) nor Kleine (1938d) mention this species.

***pusillus* Olivier, 1807**

pusillus Olivier, 1807: 435 (*Brentus*).

Type depository. Unknown.

Distribution. "Indes orientales".

Notes. According to Schoenherr (1840: 517) the species *Ceocephalus pusillus* occurs in "Ile de France" [= Mauritius Is.] but he also writes that the species is unknown to him. Schönfeldt (1910a: 42) dubitatively listed this species in the genus *Orychodes* whilst Kleine (1938d: 175) listed it among the species "*incertae sedis*".

***ruficollis* Herbst, 1797**

ruficollis Herbst, 1797: 193 (*Brentus*).

Type depository. "Museum Holthysen" (?) (HT).

Distribution. Unknown.

Notes. Kleine did not mention this species in his *Genera Insectorum* (1938d). We did not examine the type for this taxon, so we include it among the "*incertae sedis*".

***semilineatus* Boheman, 1840**

semilineatus Boheman (in Schoenherr, 1840: 482) (*Arrhenodes*).

Type depository. NHRS (HT).

Distribution. Brazil.

Notes. Schönfeldt (1910a: 37) listed this species in the genus *Eupsalis*; Kleine (1938d: 175) and Blackwelder (1982: 777) included this taxon among the species "*incertae sedis*". We did not examine the type for this taxon.

semipunctatus Fabricius, 1787

semipunctatus Fabricius, 1787: 96 (*Brentus*).

Type depository. Unknown.

Distribution. Unknown.

Notes. Neither Schönfeldt (1910a) nor Kleine (1938d) mentioned this species in their catalogues.

Fossil Section

inclusus Kleine, 1941

inclusus Kleine, 1941d: 41 (*Xestocoryphus*).

Type depository. Reichsstelle für Bodenforschung (Preuß. Geol. Landesanstalt) [from the original description].

nasalis Hope, 1840

nasalis Hope, 1840: 54 (*Brenthus*).

Type depository. Strong collection [from the original description].

Notes. From Hope's illustration (1840, pl. VII, fig. 8) this species seems to be *Bolbocranius csikii* or *B. distortus*, both extant species. On the other hand, Hope writes that the specimen was found in the resinous substance called "Animè", which came from Malabar (India), whilst *B. csikii* and *B. distortus* are African species.

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LITERATURE

We have attempted to list all the works on the Brentidae *sensu Auctorum* (Brentinae, Cyphagoginae, Pholidochlamydiae, Taphroderinae, Trachelizinae, Ulocerinae). No general bibliography on the brentids has been published since Kleine (1930a).

Here we list about 1,000 titles covering all the works describing genera and species, miscellaneous monographies and checklists, publications on morphology, biology, ecology, ethology, and zoo-geography as well as local faunistics papers where brentids are mentioned. We only managed to collect this vast amount of material thanks to the indispensable collaboration of colleagues and librarians from a large number of scientific institutions all over the world, whom we thank.

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APPENDIX 1

Brentid host plant species

Abbreviations of plant families:

ALO: Aloaceae; **ANA:** Anacardiaceae; **ANN:** Annonaceae; **APO:** Apocynaceae; **ARA:** Araliaceae; **ARAU:** Araucariaceae; **AST:** Asteraceae; **BAR:** Barringtonaceae; **BIG:** Bignoniaceae; **BOM:** Bombacaceae; **BOR:** Boraginaceae; **BUR:** Burseraceae; **BYT:** Byttneriaceae; **CAC:** Cactaceae; **CAE:** Caesalpiniaceae; **CEL:** Celastraceae; **CLU:** Clusiaceae; **COM:** Combretaceae; **COR:** Cornaceae; **CORY:** Corynocarpaceae; **CUC:** Cucurbitaceae; **CUP:** Cupressaceae; **CYP:** Cyperaceae; **DAT:** Datisceae; **DIL:** Dilleniaceae; **DIP:** Dipterocarpaceae; **EBE:** Ebenaceae; **ERI:** Ericaceae; **EUP:** Euphorbiaceae; **FAB:** Fabaceae; **FAG:** Fagaceae; **FLA:** Flacourtiaceae; **GRA:** Gramineae; **GUT:** Guttiferaceae; **HAM:** Hamamelidaceae; **HYP:** Hypericaceae; **IRV:** Irvingiaceae; **JUG:** Juglandaceae; **LAU:** Lauraceae; **LEC:** Lecythidaceae; **LEG:** Leguminosae; **LIL:** Liliaceae; **LIN:** Linaceae; **MAG:** Magnoliaceae; **MAL:** Malvaceae; **MEL:** Meliaceae; **MON:** Monimiaceae; **MOR:** Moraceae; **MORI:** Moringaceae; **MYRI:** Myristicaceae; **MYRS:** Myrsinaceae; **MYRT:** Myrtaceae; **OKT:** Oktoknemataceae; **OLA:** Olacaceae; **PAL:** Palmae; **PAN:** Pandaceae; **PAP:** Papilionaceae; **PAS:** Passifloraceae; **PIN:** Pinaceae; **POD:** Podocarpaceae; **PRO:** Proteaceae; **RHA:** Rhamnaceae; **RHI:** Rhizophoraceae; **ROS:** Rosaceae; **RUB:** Rubiaceae; **RUT:** Rutaceae; **SAL:** Salicaceae; **SAN:** Santalaceae; **SAM:** Samydaceae; **SAPI:** Sapindiaceae; **SAPO:** Sapotaceae; **SIM:** Simaroubaceae; **SOL:** Solenaceae; **STE:** Sterculiaceae; **STY:** Styracaceae; **THE:** Theaceae; **TIL:** Tiliaceae; **ULM:** Ulmaceae; **VER:** Verbenaceae; **VIO:** Violaceae.

Quotations are indicated by the following numbers:

White, 1846 (1); Broun, 1881 (2); Packard, 1881 (3); Harris, 1862 (4); Fleutiaux & Sallé, 1889 (5); Packard, 1890 (6); Hopkins, 1896 (7); Hopkins, 1898 (8); Felt, 1905 (9); Stebbing, 1914 (10); Denier, 1922 (11); Kleine, 1923g (12); Beeson, 1925 (13); Kleine, 1925b (14); Miller, 1925 (15); Kleine, 1927b (16); Kleine, 1927c (17); Kleine, 1927d (18); Kleine, 1927e (19); Kleine, 1931a (20); Peyerimhoff, 1931 (21); Kleine, 1933a (22); Kleine, 1933c (23); Kleine, 1933f (24); Hudson, 1934 (25); Kleine, 1934a (26); Kleine, 1935c (27); Kleine, 1936a (28); Kleine, 1936c (29); Wolcott, 1936 (30); Kleine, 1937i (31); Beeson, 1941 (32); De Muizon, 1954 (33); De Muizon, 1955b (34); Kojima, 1955a (35); Kojima, 1955b (36); Haedo Rossi, 1959b (37); De Muizon, 1960 (38); Mayné & Donis, 1960 (39); Schedl, 1961a (40); Mayné & Donis, 1962 (41); Damoiseau, 1963i (42); Damoiseau, 1963k (43); Damoiseau, 1972a (44); Schedl, 1975a (45); Meads, 1976 (46); Johnson, 1982 (47); Johnson, 1983a (48); Johnson, 1983b (49); Johnson & Hubbell, 1984 (50); Löyttyniemi & Löyttyniemi, 1987 (51); May, 1987 (52); Damoiseau, 1989a (53); Damoiseau, 1989b (54); Anderson, 1992 (55); May, 1993 (56); May, 1994 (57); Zimmerman, 1994 (58); Howden, 1995 (59); Thomas, 1996 (60); Klimaszewski & Watt, 1997 (61); Peck & Thomas, 1998 (62); Kabakov, 2001 (63); Mecke *et al.*, 2001 (64); Anderson & Kissinger, 2002 (65); Kuschel, 2003 (66).

Further information came from the following web sites:

<http://doacs.state.fl.us/~pi/enpp/98-jul-aug.htm> (ws1)
<http://forestry.about.com/library/silvics/blsilwhi.htm> (ws2)
http://oakplus.com/northern_red_oak.htm (ws3)
http://wildwnc.org/trees/Quercus_prinus.html (ws4)
http://www.acguanacaste.ac.cr/bosque_seco_virtual/bs_web_page/paginas_de_especies/bursera_simaruba.html (ws5)
http://www.forestworld.com/public/silvics/hardwoods/Quercus/michauxii/michauxii_c2.html (ws6)
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<http://www.ipecf.br/identificacao/nativas> (ws8)
http://www.na.fs.fed.us/spfo/pubs/silvics_manual/volume_2/quercus (ws9)

BRENTINAE Billberg

BRENTINI Billberg

Brentus anchorago (Linné)

ARA: *Pentapanax warmingiana* (Marchal) Harms (49); **BOM:** *Cavanillesia platanifolia* H. B. et K. (49) (50); *Pseudobombax septenatum* (Jacq.) Dugand (49); **BUR:** *Bursera simaruba* (L.) Sarg. (22) (47) (49) (55) (60) (62) (65) (ws5); *B. tomentosa* Triana et Planch. (49); *Bursera* sp. (59); **EUP:** *Sapium aucuparium* Jacq. (5); **RUT:** *Balfourodendron riedelianum* (Engl.) Engl. (ws8); **SIM:** *Simarouba* sp. (5).

Brentus approximatus Erichson

BOM: *Ceiba pentandra* (L.) Gaertn. (19) (23); **MOR:** *Castilloa costaricana* Liedm. (19) (23); *Ficus* sp. (19) (23).

Brentus armiger Herbst

BOM: *Quararibea turbinata* Poir. (19) (23).

Brentus caudatus Herbst

BOM: *Quararibea turbinata* Poir. (19) (23); **MOR:** *Ficus* sp. (19) (23).

Brentus cylindrus Lund

BOM: *Hibiscus tiliaceus* L. (12) (16) (23) (58).

Brentus guatemalensis Sharp

LAU: *Persea* sp. (19) (23).

Brentus volvulus Fabricius

ANA: *Mangifera* sp. (30); **LEG:** *Erythrina glauca* Willd. (30); **BOM:** *Gossypium* sp. (30); **RUB:** *Coffea* sp. (30).

Brentus vulneratus Gyllenhal

RUT: *Balfourodendron riedelianum* Engl. (37).

ARRHENODINI Lacordaire

Amphicordus improportionatus Heller

DIP: *Pentacme suavis* A. DC. (32).

Anepsiotes commendabilis Kleine

MYRT: *Eugenia jambolana* Lam. (32).

Anepsiotes schenklingi Kleine

HAM: *Liquidambar* sp. (63); **MOR:** *Trema* sp. (63).

Anomobrenthus kuscheli Damoiseau

ANN: *Cyathocalyx suaveolens* A. C. Smith (54).

Arrenodes minutus (Drury)

FAG: *Quercus alba* L. (3) (4) (6) (8) (ws2) (ws4); *Q. coccinea* Muenchh. (ws4) (ws7); *Q. michauxii* Nutt. (ws6); *Q. muehlenbergii* Engelm. (ws9); *Q. prinus* L. (7) (ws4) (ws7); *Q. rubra* L. (3) (6) (59) (ws3); *Q. shumardii* Buckl. (ws9); *Q. stellata* Wangenh. (3) (6); *Q. velutina* Lam. (3) (6) (ws9); *Quercus* sp. (9); **RUT:** *Citrus sinensis* Osbeck (ws1).

Baryrhynchus (Baryrhynchus) dehiscens (Gyllenhal)

DIP: *Balanocarpus heimii* King (32); *Shorea* sp. (23) (32).

Baryrhynchus (Baryrhynchus) merocephalus Kleine

FAG: *Pasania* sp. (63); *Quercus* sp. (63).

Baryrhynchus (Baryrhynchus) miles (Boheman)

ANN: *Alphonsea ventricosa* Hook f. et Th. (13) (23) (32); **COM:** *Terminalia bellerica* Roxb. (32); **EBE:** *Diospyros lancifolia* Roxb. (32); **EUP:** *Macaranga denticulata* Muell. Arg. (32); **MAG:** *Michelia champaca* L. (32); **RUT:** *Zanthoxylum budrunga* Wall. (32); **TIL:** *Elaeocarpus varunna* Buch.-Ham. (32).

Baryrhynchus (Eupsalomimus) poweri Roelofs

FAG: *Pasania* sp. (63); **PIN:** *Pinus* sp. (63); **THE:** *Sakakia ochracea* Nakai (35).

Belopherus maculatus (Olivier)

LEG: *Inga vera* Willd. (30).

Catablysmia navigator Damoiseau

HYP: *Cratoxylon arborescens* Blume (54).

Claeoderes radulirostris mexicanus Boheman

BOM: *Quararibea turbinata* Poir. (19) (23) (48).

Debora bocandei Power

CAE: *Oxystigma oxyphyllum* (Harms) J. Léonard (41); **EUP:** *Microdesmis puberula* Hook f. (41); **LIN:** *Ochthocosmus* cfr. *africana* Hook. f. (41); **RUB:** *Mitragyna* sp. (41).

Desgodinsia ambigua Kleine

STE: *Pterocymbium* sp. (23).

Ectocemus cinnamomi (Herbst)

ANA: *Mangifera zeylanica* Hook f. (32); **COM:** *Terminalia* sp. (63); **LAU:** *Cinnamomum* sp. (32); **MOR:** *Sloetia sideroxylon* Teijsm. et Binn. (32).

Ectocemus decemmaculatus (Montrouzier)**BOM:** *Ceiba pentandra* (L.) Gaertn. (32).*Estenorhinus dispar* (Linné)**RUB:** *Coffea liberica* Hiern. (32).*Estenorhinus flavolineatus* (Gyllenhal)**BOM:** *Ochroma lagopus* Sw. (19) (23) (32).*Hemiorychodes compactus* Kleine**CLU:** *Calophyllum tomentosum* Wight (20) (23) (32); **DIP:** *Hopea parviflora* Bedd. (20) (23) (32).*Orfilaia arabica* (De Muizon)**ALO:** *Aloe sabaia* Schweinf. (33).*Orfilaia reichei* (Fairmaire)**FAG:** *Quercus infectoria* Oliv. (45); *Q. ithaburensis* Decne. (45).*Orfilaia truncata* (Boheman)**BOM:** *Bombax malabaricum* DC. (20) (23) (32); **EUP:** *Euphorbia* sp. (32); **MOR:** *Artocarpus integrifolia* L. (20) (23) (24) (32); *Ficus tsiela* Roxb. (32); **MORI:** *Moringa pterygosperma* Gaertn. (32); **MYRT:** *Eugenia* sp. (32).*Orfilaia vulsellata* (Gyllenhal)**ANA:** *Lannea* sp. (41); *Sclerocarya birrea* Hochst. (41); *Spondianthus preussii* Engl. (41); **APO:** *Conopharyngia* sp. (41); *Voacanga obtusa* K. Schum. (42); **ANN:** *Annona arenaria* Thonn (= *sene-galensis* Pers.) (41); **BIG:** *Stereospermum* sp. (41); **BUR:** *Canarium schweinfurthii* Engl. (41); **CAE:** *Erythrophleum guineense* G. Don (42); **CEL:** *Maytenus senegalensis* (Lam.) Exell (41); **COM:** *Terminalia glandulosa* De Wild. (41); **EUP:** *Alchornea cordifolia* Muell. Arg. (41); *Antidesma meiocarpum* J. Leonard (41); *Bridelia ferruginea* Benth. (41); *Hymenocardia acida* Tul. (41); *Ricinodendron africanus* Muell. Arg. (41); *Uapaca guineensis* Muell. Arg. (41); **FAB:** *Erythrina indica* (41); **LEG:** *Acacia polyacantha* Willd. (41); *Acacia* sp. (32) (41); *Albizia glabrescens* Oliver (41); *Bauhinia thonningii* Schum. (41); *Daniellia alsteeniana* Duvign. (41); *Entadopsis abyssinica* (Steud. ex A. Rich.) Gilbert et Boutique (41); *Pterocarpus tinctorius* Welw. (41); **MOR:** *Chlorophora excelsa* (Welw.) Benth. et Hook f. (41); *Ficus congensis* Engl. (41); *F. mucoso* Welw. ex Ficalho (41); *F. persicifolia* Welw. ex Warb. (41); *Ficus* sp. (41); *Musanga cecropioides* R. Br. (41); *Myrianthus* sp. (41); **RHA:** *Maesopsis eminii* Engl. (41); **ROS:** *Parinarium mobola* Oliver (41); **RUB:** *Mitragyna stipulosa* Kuntze (41); **STE:** *Sterculia quinqueloba* (Gurke) K. Schum. (41); **VER:** *Vitex congolensis* De Wild. et Dur. (32).*Orychodes planicollis* (F. Walker)**ANN:** *Alphonsea ventricosa* Hook F. et Th. (13) (23) (32); **BOM:** *Bombax malabaricum* DC. (27) (32); *Bombax* sp. (63); **BUR:** *Boswellia serrata* Roxb. ex Colebr. (32); **CAE:** *Poinciana elata* L. (13) (23) (32); **DAT:** *Tetrameles nudiflora* R. Br. (32); **EBE:** *Diospyros lancifolia* Roxb. (32); **LEG:** *Albizia procera* Benth. (32); *Albizia* sp. (32); **MOR:** *Ficus religiosa* L. (32); *F. rumphii* Blume (32); *Ficus* sp. (23) (32) (63); **RUB:** *Hymenodictyon excelsum* Wall. (20) (23) (32); **SAL:** *Tre-mula* sp. (63).

Parorychodes cereus Kleine

ANN: *Alphonsea ventricosa* Hook F. et Th. (13) (23) (32).

Prophthalmus bourgeoisi Power

MOR: *Artocarpus integrifolia* L. f. (32).

Prophthalmus heikertingeri Kleine

SAM: *Casearia glomerata* Roxb. (32); *Casearia* sp. (27).

Prophthalmus longirostris (Gyllenhal)

FAG: *Castanopsis* sp. (63); *Pasania* sp. (63).

Prophthalmus potens delesserti Power

CAE: *Poinciana elata* L. (13) (23) (32).

Prophthalmus tridentatus (Lund)

BYT: *Pterospermum acerifolium* Willd. (10) (13) (23) (32).

Prophthalmus wichmanni Kleine

COM: *Terminalia* sp. (63); **MOR:** *Ficus* sp. (63); *Trema* sp. (63); **RUT:** *Citrus* sp. (63).

Pseudorychodes crassus Senna

EUP: *Macaranga denticulata* Muell. Arg. (32); **STY:** *Symplocos theaefolia* D. Don (32).

Pseudorychodes honorus Kleine

DIP: *Shorea* sp. (20) (23).

Pseudorychodes lineolatus (Kirsch)

DIP: *Shorea hypochra* Hance (29) (32).

Spatherhinus gabonicus (J. Thomson)

CAE: *Scorodophloeus zenkeri* Harms (39) (41); **LAU:** *Beilschmiedia corbisieri* (Robyns) Robyns et Wilczek (41); **LIL:** *Dracaena* sp. (41); **PAL:** *Elaeis* sp. (28); **PAN:** *Panda oleosa* Pierre (41); **VER:** *Vitex welwitschii* Guerke (41).

Spatherhinus longiceps Kolbe

ANA: *Lannea* sp. (41); *Spondianthus preussii* Engl. (41); **LEG:** *Albizia* sp. (41); **ROS:** *Parinarium mobola* Oliver (41).

Spatherhinus medioximus (J. Thomson)

ANA: *Sclerocarya birrea* Hochst. (41); **IRV:** *Oxystigma oxyphyllum* (Harms) J. Léonard (41);

LAU: *Beilschmiedia gilbertii* Robyns et Wilczek var. *glabra* (41); **MOR:** *Chlorophora excelsa* Benth. et Hook (41); **OLA:** *Ongokea gore* (Hua) Pierre (39) (41); **PAP:** *Angylocalyx pynaertii* De Wild. (41); *Pterocarpus soyauxii* Taub. (41); **RUB:** *Mitragyna stipulosa* Kuntze (41).

Spatherrhinus opacus (J. Thomson)

PAL: *Elaeis* sp. (28).

Stratiorrhina pascoei Kirsch

DIP: *Shorea* sp. (23).

Suborychodes intermedius Kleine

DIP: *Shorea assamica* Dyer. (12) (13) (23) (32); *S. laevis* Ridley (32); *S. robusta* Gaertn. f. (13) (23) (32); *S. talura* Roxb. (32).

EREMOXENINI Semenov-Tian-Shanskij

Acramorphocephalus gebieni Kleine

CAE: *Oxystigma oxyphyllum* (Harms) J. Léonard (41).

Acramorphocephalus stabilis Kleine

CAE: *Scorodophloeus zenkeri* Harms (41).

Afrocordus asper (Calabresi)

CAE: *Oxystigma oxyphyllum* (Harms) J. Léonard (41); **PAN:** *Panda oleosa* Pierre (41).

Amorphocephala mandibularis Damoiseau

LEG: *Acacia albida* Delile (21).

Symmorphocerus alluaudi Senna

SAPI: *Eriocoelum microspermum* Radlk. ex De Wild (41).

Symmorphocerus frontalis (Olivier)

LEG: *Brachystegia* sp. (51).

CYPHAGOGINAE Kolbe

CYPHAGOGINI Kolbe

Adidactus cancellatus (Lacordaire)

ANN: *Anonidium mannii* Engl. et Diels (41); *Polyalthia suaveolens* Engl. et Diels (39) (40) (41); *Xylopia gilbertii* Boutique (41); **APO:** *Alstonia boonei* De Wild. (41); *Pleiocarpa tubicina* Stapf. (41); **CAE:** *Oxystigma oxyphyllum* (Harms) J. Léonard (40) (41); *Paramacrolobium coeruleum*

(Taub.) J. Léonard (41); *Scorodophloeus zenkeri* Harms (39) (40) (41); **COM:** *Combretodendron macrocarpum* (Beauv.) Keay (41); **EUP:** *Discoglypsemna caloneura* Prain (41); *Drypetes gossweileri* S. Moore (41); *D. morocarpa* J. Léonard (41); *Hymenocardia acida* Tul. (41); *Macaranga lancifolia* Pax (41); **GUT:** *Garcinia punctata* Oliv. (40) (41); **LAU:** *Beilschmiedia corbisieri* (Robyns) Robyns et Wilczek (41); *B. gilbertii* Robyns et Wilczek var. *glabra* (40); **LEG:** *Albizia* sp. (41); *Tetrapleura tetraptera* (Thonn.) Taub. (41); **MEL:** *Guarea cedrata* Pellegr. (40) (41); *G. laurentii* De Wild. (39) (41); *G. thompsoni* Sprague et Hutch. (39) (41); **MOR:** *Chlorophora excelsa* Benth. et Hook f. (40); *Ficus recurvata* De Wild. (40); *Ficus* sp. (12) (23) (26); **OLA:** *Strombosia glaucescens* Engl. (41); *S. grandifolia* Hook. f. Benth. (41); *Strombosia* sp. (41); *Strombosiopsis tetrandra* Engl. (41); **PAN:** *Panda oleosa* Pierre (41); **PAP:** *Angylocalyx pynaertii* De Wild. (41); *Millettia* cfr. *drastica* Welw. (40); *Pterocarpus soyauxii* Taub. (41); **ROS:** *Parinarium curatellaefolium* Planch. ex Benth. (26); *Parinarium* sp. (41); **SAPI:** *Blighia welwitschii* (Hiern) Radlk. (41); *Pancovia harmsiana* Gilg. (41); *Radlkofera calodendron* Gilg. (41); **SAPO:** *Chrysophyllum africanum* DC. (41); *C. lacourtianum* De Wild. (41); *Synsepalum dulcificum* (Sch. et Th.) Daniell et Bell. (41); *S. subcordatum* De Wild. (41); **SIM:** *Hannoa klaineana* Pierre et Engl. (41); **STE:** *Cola griseiflora* De Wild. (41); **STY:** *Afrostryax lepidophyllus* (Harms) Mildbr. (41); **VIO:** *Rinorea welwitschii* Engl. (41).

Adidactus striolatus (Fairmaire)

ANN: *Anonidium mannii* Engl. et Diels (41); *Xylopia phloiadora* Mildbr. (41); **APO:** *Pleiocarpa tubicina* Stapf. (41); *Rauwolfia vomitoria* Afzel. (42); *Voacanga obtusa* K. Schum. (42); **CAE:** *Scorodophloeus zenkeri* Harms (41); **COM:** *Combretodendron macrocarpum* (Beauv.) Keay (41); **EUP:** *Drypetes gossweileri* S. Moore (41); *D. morocarpa* J. Léonard (41); *Hymenocardia ulmoides* Oliv. (41); *Ricinodendron africanus* Muell. Arg. (41); **LEG:** *Tetrapleura tetraptera* (Thonn.) Taub. (41); **MEL:** *Guarea laurentii* De Wild. (41); **PAN:** *Panda oleosa* Pierre (41); **PAP:** *Afrormosia elata* Harms (41); **RUT:** *Fagara macrophylla* (Oliv.) Engl. (41); **SAPI:** *Aphania senegalensis* Radlk. (41); *Radlkofera calodendron* Gilg. (41); **SAPO:** *Chrysophyllum lacourtianum* De Wild. (41); *Synsepalum subcordatum* De Wild. (41); **SIM:** *Hannoa klaineana* Pierre et Engl. (41); **TIL:** *Desplatsia dewevrei* (De Wild. et Th. Dur.) (41).

Allaeometrus breviceps Senna

DIP: *Shorea robusta* Gaertn. f. (13) (23) (32) (58); *Shorea* sp. (63).

Allagopus brunneus Gahan

ANA: *Antrocaryon nannanii* De Wild. (40) (41); **ANN:** *Anonidium mannii* Engl. et Diels (40) (41); *Isolona bruneelii* De Wild. (41); *Monodora myristica* (Gaertn.) Dun (41); *Monodora* cfr. *myristica* (Gaertn.) Dun (40); *Polyanthia suaveolens* Engl. et Diels (39) (40) (41); *Xylopia aethiopica* (Dun) A. Rich. (41); *X. gilbertii* Boutique (41); **APO:** *Funtumia* cfr. *latifolia* Stapf (40); *Pleiocarpa micrantha* Stapf (40); **BUR:** *Canarium schweinfurthii* Engl. (41); **CAE:** *Afzelia bella* Harms (41); *Amphimas pterocarpoides* Harms (40) (41); *Anthonotha macrophyllum* P. Beauv. (40); *Cynometra hankei* Harms (40) (41); *Dialium corbisieri* Staner (40); *D. excelsum* Louis ex Steyaert (40) (41); *Erythrophleum guineense* G. Don (39) (40) (41); *Gilbertiodendron dewevrei* (De Wild.) J. Léonard (39) (41); *Gossweilerodendron balsamiferum* (Vermoesen) (39) (40) (41); *Julbernardia sereti* (De Wild.) Troupin (40); *Pachyelasma tessmanii* (Harms) Harms (41); *Paramacrolobium coeruleum* (Taub.) J. Léonard (40) (41); *Oxystigma oxyphyllum* (Harms) J. Léonard (40) (41); *Scorodophloeus zenkeri* Harms (39) (40) (41); *Terminalia superba* Engl. et Diels (41); **COM:** *Combretodendron macrocarpum* (Beauv.) Keay (40) (41); *Combretum oblungum* K. Hoffm. (40); **EBE:** *Diospyros albiflavescens* (Gurke) F. White (41); *D. crassiflora* Hiern (40) (41); *D. undabunda* Hiern ex Greves (41); *Maba laurentii* De Wild. (40); **EUP:** *Drypetes gossweileri* S. Moore (41); *Hymenocardia ulmoides* Oliv. (40) (41); *Macaranga* cfr. *lancifolia* Pax (40) (41); *Microdesmis puberula* Hook f. (40) (41); *Phyllanthus discoideus* Muell. Arg. (40) (41); *Ricinodendron africanus* Muell. Arg. (41);

FLA: *Caloncoba glauca* Gilg (40); **GUT:** *Garcinia polyantha* Oliv. (40); *Symphonia globulifera* L. f. (40); **IRV:** *Irvingia grandifolia* Engl. (40) (41); **LAU:** *Beilschmiedia corbisieri* (Robyns) Robyns et Wilczek (41); **LEG:** *Albizia gummifera* (Gmal.) C. A. Sm. (40); *Albizia* sp. (39) (41); *Cathormion altissimum* (Oliv.) Hutch et J. E. Dandy (40); *C. leptophyllum* (Harms) Keay var. *guineense* (41); *Pentaclethra macrophylla* Benth. (40); *Tetrapleura tetraptera* (Thonn.) Taub. (40) (41); **MEL:** *Carapa procera* DC. (40); *Guarea cedrata* (A. Chev.) Pellegr. (39) (40) (41); *G. laurentii* De Wild. (39) (41); *Lovoa trichilioides* Harms. (40); *Trichilia rubescens* Oliv. (41); **MOR:** *Bosqueia angolensis* Ficalho (40) (41); *Chlorophora excelsa* (Welw.) Benth. et Hook f. (40) (41); *Myrianthus arboreus* P. Beauv. (40) (41); *Treculia africana* Decne (40) (41); **MYRI:** *Staudtia stipitata* Warb. (39) (40) (41); *Pycnanthus angolensis* (Welw.) Exell (39) (41); **MYRT:** *Eucalyptus citriodora* Hook. (40) (41); *E. saligna* Smith (40); *Eucalyptus* sp. (41); *Syzygium* sp. (40) (41); **OKT:** *Okoubaka aubrevillei* Pellegr. et Normand (41); **OLA:** *Ongokea gore* (Hua) Pierre (39) (40) (41); *Strombosia glaucescens* Engl. (40) (41); *S. grandifolia* Hook. f. Benth. (41); *S. scheffleri* Engl. (40); *Strombosiosis tetrandra* Engl. (40) (41); **PAN:** *Panda oleosa* Pierre (40) (41); **PAP:** *Afrormosia elata* Harms (39) (40) (41); *Angylocalyx pynaertii* De Wild. (40); *Millettia drastica* Welw. (41); *Pterocarpus soyauxii* Taub. (40) (41); **PAS:** *Barteria nigrifolia* Hook f. (41); **RHA:** *Maesopsis eminii* Engl. (40) (41); **ROS:** *Parinarium holstii* Engl. (40) (41); *Parinarium* sp. (28); **RUB:** *Heinsia pulchella* K. Schum. (40) (41); *Sarcocephalus* cfr. *trillesii* Pierre (40); **RUT:** *Fagara lemairei* De Wild. (40) (41); *F. macrophylla* (Oliv.) Engl. (41); **SAPI:** *Aphania senegalensis* Radlk. (41); *Blighia welwitschii* (Hiern) Radlk. (41); *Eriocoelum microspermum* Radlk. ex De Wild (41); *Lecanodiscus cupanoides* Planch. (40) (41); *Pancovia harmsiana* Gilg. (40) (41); *Radlkofera calodendron* Gilg. (40) (41); **SAPO:** *Chrysophyllum africanum* DC (40) (41) (41); *C. lacourtianum* De Wild. (40) (41); *Manilkara maleoleus* J. Louis (40) (41); *Synsepalum attenuatum* Hutchinson et Dalziel (41); *S. subcordatum* De Wild. (41); *Tridesmostemon claessensii* De Wild. (40) (41); **SIM:** *Hannoa klaineana* Pierre et Engl. (40) (41); *Klainedoxa gabonensis* Pierre var. *oblongifolia* (41); **STE:** *Cola griseiflora* De Wild. (41); *C. lateritia* K. Schum. (41); *Cola* cfr. *lateritia* K. Schum. (40); *Pterygota bequaertii* De Wild (41); **TIL:** *Desplatsia dewevrei* (De Wild. et Th. Dur.) (40) (41); **ULM:** *Celtis briei* De Wild. (40) (41); *C. mildbraedii* Engl. (41); **VIO:** *Rinorea welwitschii* Engl. (40) (41).

Allagopus yangambiensis Damoiseau

CAE: *Paramacrolobium coeruleum* (Taub.) J. Léonard (41); **COM:** *Combretodendron macrocarpum* (Beauv.) Keay (41); **PAN:** *Panda oleosa* Pierre (41).

Amphithetobrentus concolor (Kleine)

EUP: *Discoglyprena caloneura* Prain (41).

Autosebus setosellus Kolbe

ANN: *Polyanthia suaveolens* Engl. et Diels (39) (41); *Xylopia gilbertii* Boutique (41); **BUR:** *Canarium schweinfurthii* Engl. (41); **CAE:** *Scorodophloeus zenkeri* Harms (39) (41); **COM:** *Combretodendron macrocarpum* (Beauv.) Keay (41); **EBE:** *Diospyros alboflavescens* (Gurke) F. White (41); **EUP:** *Croton haumanianus* J. Léonard (41); *Discoglyprena caloneura* Prain (41); *Drypetes gossweileri* S. Moore (39) (41); **GUT:** *Garcinia smeathmanni* Oliver (41); **LEG:** *Albizia* sp. (39) (41); *Tetrapleura tetraptera* (Thonn.) Taub. (41); **MEL:** *Guarea cedrata* (A. Chev.) Pellegr. (39) (41); *G. laurentii* De Wild. (39) (41); *Trichilia lanata* A. Chev. (41); *T. rubescens* Oliv. (41); **MOR:** *Chlorophora excelsa* (Welw.) Benth. et Hook f. (39) (41); *Myrianthus arboreus* P. Beauv. (41); **MYRI:** *Pycnanthus angolensis* (Welw.) Exell (39) (41); **OLA:** *Strombosia glaucescens* Engl. (41); *S. grandifolia* Hook. f. Benth. (41); *Strombosiosis tetrandra* Engl. (41); **PAN:** *Panda oleosa* Pierre (41); **PAP:** *Angylocalyx pynaertii* De Wild. (41); **SAPI:** *Blighia welwitschii* (Hiern) Radlk. (41); **SAPO:** *Chrysophyllum pruniforme* Engl. (41); *Synsepalum subcordatum* De Wild. (41); **STY:** *Afrotyrax lepidophyllus* (Harms) Mildbr. (41).

Caenosebus fraudator Kleine

ANN: *Annona arenaria* Thonn (= *senegalensis* Pers.) (41).

Callipareius (Callipareius) bistriatus (Kleine)

FAG: *Quercus spicata* Sm. (27) (32).

Calodromus mellyi Guérin-Méneville & Gory

DIP: *Vatica lanceaefolia* Bl. (12) (13) (23); **HAM:** *Liquidambar* sp. (63); **MOR:** *Ficus* sp. (63).

Chelorhinus flexibilis (Kleine)

FAG: *Quercus serrata* Thunb. (32).

Cormopus distinctus Kleine

ANN: *Polyanthia suaveolens* Engl. et Diels (40); **APO:** *Pleiocarpa tubicina* Stapf. (41); **CAE:** *Cynometra hankei* Harms (39) (40) (41); *Dialium corbisieri* Staner (41); *Scorodophloeus zenkeri* Harms. (39) (40) (41); **COM:** *Combretodendron macrocarpum* (Beauv.) Keay (40) (41); *Terminalia superba* Engl. et Diels (39) (40) (41); **EUP:** *Drypetes leonensis* Pax var. *glabra* J. Léonard (40); *D. gossweileri* S. Moore (41); *D. morocarpa* J. Léonard (41); **GUT:** *Garcinia punctata* Oliv. (41); **IRV:** *Oxystigma oxyphyllum* (Harms) J. Léonard (41); **MEL:** *Trichilia prieureana* Juss. (40); **MYRI:** *Coelocaryon preussii* Warb. (41); **OLA:** *Strombosia grandifolia* Hook. f. Benth. (41); **PAN:** *Panda oleosa* Pierre (40) (41); **SAPO:** *Chrysophyllum africanum* A. DC. (40) (41); *C. lacourtia-num* De Wild. (41); *C. pruiniforme* Engl. (40); *Synsepalum dulcificum* (Sch. et Th.) Daniell et Bell. (41); *S. longecuneatum* De Wild. (40); *S. subcordatum* De Wild. (40) (41).

Cormopus diversus Kleine

LEG: *Brachystegia* sp. (51).

Cormopus diversitarsis Kleine

MOR: *Ficus* sp. (26); **ROS:** *Parinarium curatellaefolium* Planch. ex Benth. (26); *Parinarium* sp. (41).

Cormopus filum Kolbe

CAE: *Dialium corbisieri* Staner (40); *Gossweilerodendron balsamiferum* Harms (41); **MOR:** *Ficus* sp. (26); **ROS:** *Parinarium curatellaefolium* Planch. ex Benth. (26); **STE:** *Cola griseiflora* De Wild. (41); **TIL:** *Desplatsia dewevrei* (De Wild. et Th. Dur.) (41).

Cormopus minus Kleine

LEG: *Piptadenia* sp. (28).

Cormopus penicillifer Kolbe

ANN: *Anonidium mannii* Engl. et Diels (40); **APO:** *Alstonia boonei* De Wild. (39) (41); *Pleiocarpa tubicina* Stapf. (41); **CAE:** *Cynometra* sp. (41); *Dialium corbisieri* Staner (40); *D. pachyphyllum* Harms (41); *Gossweilerodendron balsamiferum* Harms (40); *Paramacrolobium coeruleum* (Taub.) J. Léonard (41); *Scorodophloeus zenkeri* Harms. (40) (41); **EUP:** *Drypetes gossweileri* S. Moore

(41); *D. morocarpa* J. Léonard (41); **GUT:** *Garcinia punctata* Oliv. (41); **IRV:** *Klainedoxa gabonensis* Pierre (40); *Oxystigma oxyphyllum* (Harms) J. Léonard (41); **LEG:** *Cathormion altissimum* (Oliv.) Hutch et J. E. Dandy (40); *Pentaclethra macrophylla* Benth. (40); **MEL:** *Entandrophragma* sp. (39) (41); *Trichilia heudelotii* Planch. (40); *T. prieureana* Juss. (40) (41); *T. rubescens* Oliv. (40); **MOR:** *Myrianthus arboreus* P. Beauv. (41); **OLA:** *Ongokea gore* (Hua) Pierre (40); **SAPI:** *Pancovia harmsiana* Gilg. (40) (41); **SAPO:** *Chrysophyllum africanum* A. D.C. (41); *C. lacourtia-num* De Wild. (40); *Synsepalum subcordatum* De Wild. (41); **SIM:** *Klainedoxa gabonensis* Pierre var. *oblongifolia* (41); **STE:** *Cola griseiflora* De Wild. (40) (41); **STY:** *Afrostryax lepidophyllus* (Harms) Mildbr. (41).

Cyphagogus brownei Kleine

DIP: *Shorea hypochra* Hance (28) (53); *S. robusta* Gaertn. f. (32); *Vatica lanceaefolia* Bl. (32).

Cyphagogus confertulus Kleine

CLU: *Mesua ferrea* L. (12) (13) (23) (32); **FAG:** *Castanopsis* sp. (63).

Cyphagogus confidens Kleine

DIP: *Pentacme suavis* A. DC. (32); *Shorea eximia* Scheff. (32); *S. parvifolia* Dyer (32); *S. pauciflora* King. (32); *S. platycarpa* Heim (32).

Cyphagogus corporaali Kleine

CAE: *Bauhinia purpurea* L. (13) (23) (32); **DIP:** *Pentacme suavis* A. DC. (32); **BOM:** *Bombax malabaricum* DC. (12) (13) (23) (32); **FAB:** *Butea frondosa* Roxb. (12) (13) (23) (32) (38) (58); **VER:** *Tectonia grandis* L. (13) (23) (32).

Cyphagogus cyrtotrachelus (J. Thomson)

DIP: *Shorea hypochra* Hance (32); *S. robusta* Gaertn. f. (12) (13) (23); *Shorea* sp. (23); **LAU:** *Cryptocarya wightiana* Thw. (32); **LEG:** *Albizia odoratissima* Benth. (32); **MYRI:** *Myristica andamanica* Hook. f. (20) (23) (32); *M. longifolia* Wall. (20) (23) (32); **SAPO:** *Isonandra polyantha* Kurz (32); *Mimusops littoralis* Kurz. (20) (32); **VER:** *Tectonia grandis* L. (13) (23) (32) (38).

Cyphagogus eggersi Kleine

DIP: *Shorea hypochra* Hance (32); *S. parvifolia* Dyer (29) (32).

Cyphagogus eichhorni Kirsch

BOM: *Bombax malabaricum* DC. (12) (13) (23) (32); *Bombax* sp. (63); **DIP:** *Shorea bracteolata* Dyer (29) (32); *S. hypochra* Hance (29) (32); *S. leprosula* Miq. (29) (32); *Shorea* sp. (20) (63); *Vatica lanceaefolia* Bl. (12); *Vatica* sp. (63).

Cyphagogus gemellus Kleine

MOR: *Ficus* sp. (32).

Cyphagogus incisus Senna

CLU: *Calophyllum spectabile* Willd. (20) (23) (32); **COM:** *Terminalia myriocarpa* Heurck et Muell. Arg. (27) (32); **DIP:** *Shorea* sp. (13) (23); *Vatica lanceaefolia* Bl. (13) (23) (32); **MEL:** *Chickrassia tabularis* A. Juss. (32); **MOR:** *Ficus* sp. (27) (32).

Cyphagogus longulus Senna

DIP: *Vatica lanceaefolia* Bl. (12) (13) (23) (32); **SAPO:** *Isonandra polyantha* Kurz (20) (23) (32).

Cyphagogus planifrons Kirsch

CLU: *Mesua ferrea* L. (12) (13) (23) (32); **COM:** *Terminalia myriocarpa* Heurck et Muell. Arg. (32); **DIP:** *Vatica lanceaefolia* Bl. (12) (13) (23) (32); **EUP:** *Mallotus albus* Muell. Arg. (12) (13) (32); **MYRI:** *Myristica longifolia* Wall. (20) (23) (32).

Cyphagogus signipes Lewis

JUG: *Juglans ailantifolia* Carr. (36).

Cyphagogus silvanus Senna

DIP: *Pentacme suavis* A. DC. (32).

Cyphagogus tabacicola Senna

ANA: *Melanorrhoea usitata* Wall. (12) (13) (23) (32); **DIP:** *Vatica lanceaefolia* Bl. (12) (13) (23) (32); **MOR:** *Artocarpus* sp. (63); **STE:** *Heritiera fomes* Buch. (12) (13) (23) (32).

Cyphagogus westwoodii Parry

ANA: *Lannea grandis* Engl. (27) (32); *Semecarpus gardneri* Thw. (32); **BUR:** *Canarium* sp. (23) (32); *Protium serratum* Engl. (32); **CAE:** *Poinciana elata* L. (13) (23) (32); **COM:** *Terminalia belerica* Roxb. (20) (23) (32); *Terminalia* sp. (63); **DIL:** *Wormia triquetra* Rottb. (32); **DIP:** *Hopea odorata* Roxb. (32); *Shorea* sp. (23) (32); *Vatica lanceaefolia* Bl. (12) (13) (23) (32) (58); *Vatica* sp. (63); **LAU:** *Cryptocarya wightiana* Thw. (32); **LEG:** *Intsia bakeri* Prain (23) (32); **MOR:** *Ficus* sp. (27) (32); **MYRI:** *Myristica longifolia* Wall. (20) (23) (32); **STE:** *Heritiera fomes* Buch. (12) (13) (23) (32).

Dentisebus pusio (Kolbe)

SAPO: *Chrysophyllum africanum* A. D.C. (40); *C. lacourtianum* De Wild. (40).

Diastrophocoleps femoralis (Damoiseau)

EUP: *Drypetes monocarpa* J. Léonard (41).

Eterozemus deformis Kleine

DIP: *Shorea robusta* Gaertn. f. (13) (23); **MOR:** *Ficus* sp. (63).

Eurorhinus maynei Damoiseau

MEL: *Entandrophragma* sp. (41).

Eusebus fijianus Kleine

LEG: *Albizia falcataria* (L.) Fosberg (53).

Megalosebus bidenticulatus Kolbe

ANN: *Annona arenaria* Thonn (= *senegalensis* Pers.) (41); **BUR:** *Canarium schweinfurthii* Engl. (41); **COM:** *Terminalia glandulosa* De Wild. (41); **EUP:** *Alchornea cordifolia* Muell. Arg. (41); **LEG:** *Albizia* sp. (41); *Bauhinia thonningii* Schum. (41); *Daniellia alsteeniana* Duvign. (41); *Pterocarpus tinctorius* Welw. (41); **RUB:** *Mitragyna stipulosa* Kuntze (41); *Mitragyna* sp. (42); **ROS:** *Parinarium mobola* Oliver (41); **STE:** *Sterculia quinqueloba* (Gurke) K. Schum. (41); **VER:** *Vitex* sp. (41).

Megalosebus separabilis Kolbe

MYRI: *Staudtia stipitata* Warb. (41).

Mesoderes guttatus Kleine

BOM: *Bombax malabaricum* DC. (20) (23) (32); **FAB:** *Butea frondosa* Roxb. (32).

Mesoderes sexnotatus Senna

COM: *Terminalia bialata* Steud. (20) (23) (32).

Microsebus adelphus Kolbe

FAB: *Butea frondosa* Roxb. (13) (23) (32).

Microsebus trifasciatus (Kleine)

MOR: *Ficus* sp. (63); **RUT:** *Citrus* sp. (63).

Nannobrenthus compressithorax (Senna)

ANA: *Antrocaryon nannanii* De Wild. (40); **ANN:** *Anonidium mannii* Engl. et Diels (40) (41); *Monodora myristica* (Gaertn.) Dun (41); *Polyanthia suaveolens* Engl. et Diels (39) (40) (41); *Xilopia villosa* Chipp. (40); **BUR:** *Canarium schweinfurthii* Engl. (40) (41); **CAE:** *Dialium corbisieri* Stanner (40); *Erythrophleum guineense* G. Don (41); *Oxystigma oxyphyllum* (Harms) J. Léonard (40); *Scorodophloeus zenkeri* Harms. (40) (41); **COM:** *Combretodendron macrophyllum* (P. Beauv.) Keay. (40); *Terminalia superba* Engl. et Diels (41); **EUP:** *Drypetes gossweileri* S. Moore (40); *Drypetes* cfr. *monocarpa* J. Léonard (40); *Macaranga* cfr. *lancifolia* Pax (40); **IRV:** *Klainedoxa gabonensis* Pierre (40); **LAU:** *Beilschmiedia gilbertii* Robyns et Wilczek var. *glabra* (40); **LEG:** *Albizia gummifera* (Gmal.) C. A. Sm. (40); *Tetrapleura tetraptera* (Thonn.) Taub. (40); **MEL:** *Guaerea cedrata* Pellegr. (40); *G. laurentii* De Wild. (40) (41); *G. thompsonii* Sprague et Hutch. (40); *Trichilia prieureana* Juss. (40); *T. lanata* A. Chev. (40); *T. rubescens* Oliv. (40) (41); **MOR:** *Bosqueia* sp. (41); *Chlorophora excelsa* Benth. et Hook f. (40); *Myrianthus arboreus* P. Beauv. (40); **MYRI:** *Coelocaryon preussii* Warb. (40); *Pycnanthus angolensis* (Welw.) Exell. (40); **OLA:** *Ongokea gore* (Hua) Pierre (40); *Strombosia glaucescens* Engl. (40); *S. scheffleri* Engl. (40); *Strombosiosis tetrandra* Engl. (40); **PAN:** *Panda oleosa* Pierre (40); **RUB:** *Randia octomera* Benth. (40); **RUT:** *Fagara inaequalis* Engl. (41); **SAPI:** *Blighia welwitschii* (Hiern) Radlk. (41); *Eriocoelum* cfr. *microspermum* Radlk. (40); *Pancovia harmsiana* Gilg. (40); *P. laurentii* De Wild. (40); **SAPO:** *Chrysophyllum africanum* DC. (40); *C. pruniforme* Engl. (40); *Synsepalum attenuatum* Hutchinson et Dalziel (41); *S. subcordatum* De Wild. (40); **STE:** *Cola griseiflora* De Wild. (40) (41); *C. lateritia* K. Schum. (41); *C. cfr. lateritia* K. Schum. (40); **ULM:** *Celtis brieyi* De Wild. (41); **VIO:** *Rinorea brachypetala* Kuntze (41).

Neoxybasius fatuelus Kolbe

ANN: *Anonidium mannii* Engl. et Diels (41); *Polyalthia suaveolens* Engl. et Diels (39) (41); *Xylopia gilbertii* Boutique (41); **APO:** *Funtumia* cfr. *latifolia* Stapf (41); *Pleiocarpa micrantha* Stapf (41); **BUR:** *Canarium schweinfurthii* Engl. (41); **CAE:** *Anthonothea macrophyllum* P. Beauv. (41); *Dialium pachyphyllum* Harms (41); *Erythrophleum guineense* G. Don (41); *Gossweilerodendron balsamiferum* Harms (41); *Scorodophloeus zenkeri* Harms. (39) (41); **COM:** *Combretodendron macrocarpum* (Beauv.) Keay (41); *Terminalia superba* Engl. et Diels (41); **EBE:** *Diospyros alboflavescens* (Gurke) F. White (41); *D. crassiflora* Hiern (41); **EUP:** *Croton haumanianus* J. Léonard (41); *Drypetes gossweileri* S. Moore (41); *D. morocarpa* J. Léonard (41); *Macaranga lancifolia* Pax (41); *Microdesmis puberula* Hook. f. (41); **FLA:** *Caloncoba glauca* Gilg (41); **GUT:** *Allanblackia floribunda* Oliver (41); *Garcinia punctata* Oliv. (41); *G. smeathmanni* Oliver (41); *Mammea africana* G. Don (41); **IRV:** *Irvingia gabonensis* Baill. (41); *Oxystigma oxyphyllum* (Harms) J. Léonard (41); **LAU:** *Beilschmiedia corbisieri* (Robyns) Robyns et Wilczek (41); **LEG:** *Albizia* sp. (41); **MEL:** *Carapa procera* DC. (41); *Entandrophragma candollei* Harms (41); *E. cylindricum* (41); *E. utile* (Dawe et Sprague) (41); *Guarea cedrata* Pellegr. (41); *G. laurentii* De Wild. (41); *G. thompsoni* Sprague et Hutch. (41); *Trichilia rubescens* Oliv. (41); **MOR:** *Antiaris welwitschii* Engl. (41); *Chlorophora excelsa* Benth. et Hook (41); *Myrianthus arboreus* P. Beauv. (41); **MYRI:** *Pycnanthus angolensis* (Welw.) Exell. (41); *Staudtia stipitata* Warb. (41); **MYRT:** *Syzygium* sp. (41); **OLA:** *Ongokea gore* (Hua) Pierre (41); *Strombosia glaucescens* Engl. (41); *Strombosia* sp. (41); *Strombosiopsis tetrandra* Engl. (41); **PAN:** *Panda oleosa* Pierre (41); **PAP:** *Afrormosia elata* Harms (41); *Pterocarpus soyauxii* Taub. (41); **RHA:** *Maesopsis eminii* Engl. (41); **RUB:** *Nauclea diderrichii* (De Wild.) Merrill (41); *Plectronia dewevrei* Wildem. (41); **SAPI:** *Autranella congolensis* A.Cheval. et De Wild (41); *Blighia welwitschii* (Hiern) Radlk. (41); *Pancovia harmsiana* Gilg. (41); **SAPO:** *Chrysophyllum africanum* DC. (41); *C. lacourtianum* De Wild. (41); *Manilkara malcoleus* J. Louis (41);); *Synsepalum attenuatum* Hutchinson et Dalziel (41); *S. dulcificum* (Sch. et Th.) Daniell et Bell. (41); *S. subcordatum* De Wild. (41); **SIM:** *Klainedoxa gabonensis* Pierre var. *oblongifolia* (41); **STE:** *Cola griseiflora* De Wild. (41); *C. lateritia* K. Schum. (41); **ULM:** *Celtis mildbraedii* Engl. (41).

Oncodemerus sennai Quentin

ANA: *Antrocaryon nannanii* De Wild. (41); **ANN:** *Anonidium mannii* Engl. et Diels (40) (41); *Polyalthia suaveolens* Engl. et Diels (39); *Xylopia gilbertii* Boutique (41); *X. phloioidora* Mildbr. (41); **APO:** *Alstonia boonei* De Wild. (39) (41); *Pleiocarpa micrantha* Stapf (40) (41); *P. tubicina* Stapf (40) (41); **BIG:** *Fernandoa adolfi-fridericii* (Gilg et Mildbr.) Heine (40) (41); **BOM:** *Ceiba thoningii* A. Chev. (40); **BUR:** *Canarium schweinfurthii* Engl. (41); *Dacryodes edulis* (G. Don) H. J. Lam. (41); *D. yangambiensis* J. Louis ex Troupin (41); **CAE:** *Amphimas pterocarpoides* Harms (41); *Anthonothea macrophyllum* P. Beauv. (40) (41); *Cynometra hankei* Harms (39) (41); *Dialium pachyphyllum* Harms (41); *Erythrophleum guineense* G. Don (39) (40) (41); *Gossweilerodendron balsamiferum* Harms (39) (40) (41); *Oxystigma oxyphyllum* (Harms) J. Léonard (40) (41); *Paramacrolobium coeruleum* (Taub.) J. Léonard (41); *Scorodophloeus zenkeri* Harms. (39) (40) (41); **COM:** *Combretodendron macrocarpum* (Beauv.) Keay (41); *Combretum oblungum* K. Hoffm. (41); *Terminalia superba* Engl. et Diels (41); **EBE:** *Diospyros alboflavescens* (Gurke) F. White (41); *D. crassiflora* Hiern (41); **EUP:** *Cleistanthus* sp. (40); *Croton haumanianus* J. Léonard (40); *C. megalocarpus* Hutchinson (41); *Discoglypsemna caloneura* Prain (41); *Drypetes gossweileri* S. Moore (39) (41); *D. morocarpa* J. Léonard (41); *D. cinnabarina* Pax et K. Hoffm. var. *congolensis* (41); *D. spinoso-dentata* (Pax) Hutch var. *longipeducellata* J. Léonard (40) (41); *D. cfr. louisii* J. Léonard (40) (41); *Macaranga lancifolia* Pax (41); *M. cfr. lancifolia* Pax (40) (41); *Microdesmis puberula* Hook f. (41); *Ricinodendron africanus* Muell.Arg. (41); **GUT:** *Garcinia polyantha* Oliv. (40); *G. punctata* Oliv. (41); *G. smeathmanni* Oliver (41); *Mammea africana* G. Don (40) (41); **IRV:** *Irvingia grandiflora* Engl. (41); *Irvingia* sp. (41); **LAU:** *Beilschmiedia gilbertii* Robyns et Wilczek var. *glabra* (40) (41); **LEG:** *Albizia gummifera* (Gmel.) C. A. Sm. (40); *Albizia* sp. (41); *Baphia pubescens* Hook.f. (41); *Cathormion leptophyllum* (Harms) Keay var. *guineense* (41); *Pen-*

taclethra macrophylla Benth. (41); *Piptadeniastrum africanum* (Hook. f.) Brenan (41); *Tetrapleura tetraptera* (Thonn.) Taub. (40) (41); **MEL:** *Entandrophragma* sp. (39) (41); *Guarea cedrata* (A. Chev.) Pellegr. (39) (41); *G. laurentii* De Wild. (39) (40) (41); *G. thompsonii* Sprague et Hutch. (39) (40) (41); *Trichilia laurentii* De Wild. (40); *T. thomsoni* Sprague et Hutch (40); *T. lanata* A. Chev. (40); *T. rubescens* Oliv. (40) (41); *T. welwitschii* C. DC (41); **MOR:** *Antiaris welwitschii* Engl. (41); *Bosqueia angolensis* Ficalho (40) (41); *Chlorophora excelsa* Benth. et Hook f. (39) (40) (41); *Musanga cecropioides* R. Br. (40) (41); *Myrianthus arboreus* P. Beauv. (40) (41); **MYRI:** *Coelocaryon preussii* Warb. (41); *Staudtia stipitata* Warb. (39) (40) (41); **OLA:** *Ongokea gore* (Hua) Pierre (39) (40) (41); *Strombosia glaucescens* Engl. (40) (41); *S. grandifolia* Hook. f. Benth. (41); *S. scheffleri* Engl. (40); *Strombosiaopsis tetrandra* Engl. (40) (41); **PAN:** *Panda oleosa* Pierre (40) (41); **PAP:** *Afromosia elata* Harms (39) (40) (41); *Angylocalyx pynaertii* De Wild. (40) (41); *Millettia* cfr. *drastica* Welw. (40) (41); **RUT:** *Fagara macrophylla* (Oliv.) (40) (41); *Vepris louisii* Gilbert (41); **SAPI:** *Autranella congolensis* A.Cheval. et De Wild (41); *Blighia welwitschii* (Hiern) Radlk. (41); *Pancovia harmsiana* Gilg. (40) (41); *P. laurentii* De Wild. (41); *Radlkofera calodendron* Gilg. (40) (41); **SAPO:** *Chrysophyllum africanum* DC (40) (41); *C. lacourtianum* De Wild. (40) (41); *Synsepalum attenuatum* Hutchinson et Dalziel (41); *S. dulcificum* (Sch. et Th.) Daniell et Bell. (41); *S. subcordatum* De Wild. (40) (41); *Tridesmostemon claessensii* De Wild (41); **SIM:** *Hannoa klaineana* Pierre et Engl. (40) (41); *Klainedoxa gabonensis* Pierre var. *oblongifolia* (41); **STE:** *Cola griseiflora* De Wild. (40) (41); *C. lateritia* K. Schum. (41); *Pterygota bequaertii* De Wild. (40); *P. mildbraedii* Engl. (41); *Sterculia bequaertii* De Wild. (40) (41); **TIL:** *Desplatsia dewevrei* (De Wild. et Th. Dur.) (40) (41); *Microcos coriacea* (Mast.) Burret (40) (41); **ULM:** *Celtis briei* De Wild. (40) (41); *C. mildbraedii* Engl. (39) (40) (41); **VER:** *Vitex congolensis* De Wild. et Dur. (40) (41); *V. welwitschii* Guerke (41); **ULM:** *Celtis briei* De Wild. (41); **VIO:** *Rinorea welwitschii* Engl. (41).

Opisthenoxys famulus Kleine

APO: *Dyera* sp. (17).

Opisthenoxys ochraceus Kleine

DIP: *Dipterocarpus cornutus* Dyer (23) (32); *D. pilosus* Roxb. (32); *Shorea hypochra* Hance (29) (32); *Shorea* sp. (23).

Opisthozemius appendiculatus Kolbe

COM: *Combretum splendens* Engl. (26).

Parasebasius meticulosus (Kolbe)

ANA: *Sclerocarya birrea* Hochst. (41); **ANN:** *Annona arenaria* Thonn (= *senegalensis* Pers.) (41); *Anonidium mannii* Engl. et Diels (41); *Polyanthia suaveolens* Engl. et Diels (41); **CAE:** *Paramacrolobium coeruleum* (Taub.) J. Léonard (41); *Scorodophloeus zenkeri* Harms (41); **COM:** *Combretodendron macrocarpum* (Beauv.) Keay (41); **EUP:** *Macaranga lancifolia* Pax (41); **IRV:** *Oxystigma oxyphyllum* (Harms) J. Léonard (41); **LEG:** *Albizia* sp. (41); **MEL:** *Guarea laurentii* De Wild. (41); **MOR:** *Ficus congolensis* Engl. (41); **OLA:** *Strombosiaopsis tetrandra* Engl. (41); **PAP:** *Angylocalyx pynaertii* De Wild. (41); **RUB:** *Mitragyna stipulosa* Kuntze (41); **STE:** *Sterculia quinqueloba* (Gurke) K. Schum. (41).

Podozemius comparabilis (Kolbe)

ANN: *Anonidium mannii* Engl. et Diels (41); *Polyanthia suaveolens* Engl. et Diels (41); **CAE:** *Scorodophloeus zenkeri* Harms (41); **COM:** *Combretodendron macrocarpum* (Beauv.) Keay (41); *Terminalia superba* Engl. et Diels (41); **EUP:** *Macaranga* cfr. *lancifolia* Pax (41); **IRV:** *Oxystigma*

oxyphyllum (Harms) J. Léonard (41); **LAU:** *Beilschmiedia gilbertii* Robyns et Wilczek var. *glabra* (41); **LEG:** *Albizia* sp. (39) (41); *Brachystegia* sp. (41); **MEL:** *Entandrophragma* sp. (41); **MEL:** *Guarea cedrata* Pellegr. (41); *G. laurentii* De Wild. (41); *G. thompsoni* Sprague et Hutch. (41); **MYRI:** *Pycnanthus angolensis* (Welw.) Exell. (41); **OLA:** *Strombosia glaucescens* Engl. (41); *S. grandifolia* Hook. f. Benth. (41); **PAN:** *Panda oleosa* Pierre (41); **PAP:** *Angylocalyx pynaertii* De Wild. (41); **SAPO:** *Chrysophyllum africanum* DC. (41); **SAPI:** *Pancovia harmsiana* Gilg. (41); **STY:** *Afrostrax lepidophyllum* (Harms) Mildbr. (41); **ULM:** *Celtis briei* De Wild. (41).

Podozemius mustus Kolbe

COM: *Combretodendron macrocarpum* (Beauv.) Keay (41); **EUP:** *Macaranga lancifolia* Pax (41); **IRV:** *Oxystigma oxyphyllum* (Harms) J. Léonard (41); **MOR:** *Ficus congensis* Engl. (42); **OLA:** *Strombosia glaucescens* Engl. (41); **SAPI:** *Pancovia harmsiana* Gilg. (41).

Protusambius praecursor Kolbe

ANN: *Anonidium mannii* Engl. et Diels (40) (41); *Polyanthia suaveolens* Engl. et Diels (40); *Xylopia aethiopica* (Dun) A. Rich. (40); **APO:** *Alstonia congensis* Engl. (40); **CAE:** *Afzelia bella* Harms (40); *Scorodophloeus zenkeri* Harms (40) (41); **COM:** *Terminalia superba* Engl. et Diels (40); **EUP:** *Croton haumanianus* J. Léonard (40); *Drypetes gossweileri* S. Moore (40); *Microdesmis puberula* Hook f. (40); *Tetrorchidium didmostenum* Pax. et Hoffm. (40); **LEG:** *Albizia gummifera* (Gmel.) C. A. Sm. (40); *Piptadenia africana* Hook f. (40); **MEL:** *Trichilia heudelotii* Planch. (40); *Guarea laurentii* De Wild. (40); **MOR:** *Bosqueia angolensis* Ficalho (40); *Treulia africana* Decne (40); **OLA:** *Ongokea gore* (Hua) Pierre (41); *Strombosia glaucescens* Engl. (40); **PAN:** *Panda oleosa* Pierre (40); **PAP:** *Angylocalyx pynaertii* De Wild. (40) (41); *Millettia* cfr. *drastica* Welw. (40); **SIM:** *Hannoa klaineana* Pierre et Engl. (40); **SAPI:** *Pancovia laurentii* De Wild. (40); **SAPO:** *Chrysophyllum africanum* DC. (40); *C. lacourtianum* De Wild. (40); *Synsepalum dulcificum* (Sch. et Th.) Daniell et Bell. (41); *S. longecuneatum* De Wild. (40); *S. subcordatum* De Wild. (40) (41); **STE:** *Cola griseiflora* De Wild. (40); *Sterculia bequaertii* De Wild. (40).

Pseudocyphagonus squamifer Desbrochers des Loges

COM: *Terminalia bialata* Steud. (20) (23) (32); *Terminalia* sp. (63); **DIP:** *Dipterocarpus pilosus* Roxb. (12) (13) (23) (32); *Shorea assamica* Dyer. (13) (23) (32); *Vatica* sp. (63).

Pseudoparagodus longitarsis Damoiseau

EUP: *Microdesmis puberula* Hook. f. (41); **MEL:** *Guarea cedrata* (A. Chev.) Pellegr. (41).

Pseudousambius silvanus (Kolbe)

IRV: *Oxystigma oxyphyllum* (Harms) J. Léonard (41).

Stibacephalus wagneri Kleine

ANN: *Polyanthia suaveolens* Engl. et Diels (41); **IRV:** *Oxystigma oxyphyllum* (Harms) J. Léonard (41); **SIM:** *Klainedoxa gabonensis* Pierre var. *oblongifolia* (41).

Synsebasius opacus Kolbe

OLA: *Strombosiosis tetrandra* Engl. (41).

Usambius advena (Pascoe)

CAE: *Cynometra hankei* Harms (40); *Scorodophloeus zenkeri* Harms (39) (41); **GUT:** *Garcinia*

punctata Oliv. (41); **IRV:** *Oxystigma oxyphyllum* (Harms) J. Léonard (40) (41); **LEG:** *Albizia* sp. (41); **MEL:** *Guarea cedrata* (A. Chev.) Pellegr. (41); **MOR:** *Chlorophora excelsa* Benth. et Hook (41); **OLA:** *Strombosiosis tetrandra* Engl. (41); **PAP:** *Afrormosia elata* Harms (41); **SAPI:** *Pancovia harmsiana* Gilg. (41); **SAPO:** *Chrysophyllum africanum* A. DC. (40) (41); *C. lacourtianum* De Wild. (40); *Synsepalum dulcificum* (Sch. et Th.) Daniell et Bell. (41); **SIM:** *Klainedoxa gabonensis* Pierre (40).

Usambioproctus dissimilis (Kleine)

CAE: *Amphimas pterocarpoides* Harms (41); *Julbernardia sereti* (De Wild.) Troupin (40); *Oxystigma oxyphyllum* (Harms) J. Léonard (40) (41); **EUP:** *Croton haumanianus* J. Léonard (41); **GUT:** *Garcinia punctata* Oliv. (40) (41); *G. smeathmanni* Oliver (41); **MEL:** *Guarea laurentii* De Wild. (40); *Trichilia rubescens* Oliv. (40); **MYRI:** *Staudtia stipitata* Warb. (39) (40) (41); **PAP:** *Angylocalyx pynaertii* De Wild. (40); **SAM:** *Homalium* sp. (40); **SAPO:** *Synsepalum subcordatum* De Wild. (40); *Tridesmostemon claessensi* De Wild. (41).

Usambioproctus gracilis (Kleine)

CAE: *Scorodophloeus zenkeri* Harms (41); **IRV:** *Oxystigma oxyphyllum* (Harms) J. Léonard (41); **LEG:** *Albizia* sp. (41); **MEL:** *Guarea cedrata* (A. Chev.) Pellegr. (41).

Usambioproctus planistriatus (Damoiseau)

IRV: *Oxystigma oxyphyllum* (Harms) J. Léonard (41).

Zemioses porcatus Pascoe

ANN: *Anonidium mannii* Engl. et Diels (41); *Polyanthia suaveolens* Engl. et Diels (39) (40) (41); *Xylopia gilbertii* Boutique (41); **APO:** *Alstonia boonei* De Wild. (41); **CAE:** *Dialium corbisieri* Staner (40); *Erythrophleum guineense* G. Don (40); *Gossweilerodendron balsamiferum* (Verm.) Harms. (41); *Oxystigma oxyphyllum* (Harms) J. Léonard (40) (41); *Scorodophloeus zenkeri* Harms. (40) (41); **COM:** *Combretodendron macrocarpum* (Beauv.) Keay (41); *Combretum splendens* Engl. (26); **EBE:** *Diospyros crassiflora* Hiern (40); **EUP:** *Drypetes gossweileri* S. Moore (41); *D. morocarpa* J. Léonard (41); *Macaranga lancifolia* Pax (41); **GUT:** *Allanblackia floribunda* Oliver (41); **LEG:** *Albizia gummifera* (Gmel.) C. A. Sm. (40); *Albizia* sp. (39) (41); *Newtonia buchananii* (Baker) Gilbert et Boutique (41); **MEL:** *Guarea cedrata* (A. Chev.) Pellegr. (41); *G. laurentii* De Wild. (40) (41); **MOR:** *Bosqueia angolensis* (Welw.) Fic. (40); *Ficus* sp. (12) (23); *Musanga cecropioides* R. Br. (41); **MYRI:** *Staudtia stipitata* Warb. (40); **PAP:** *Afrormosia elata* Harms (39) (41); **PAN:** *Panda oleosa* Pierre (41); **RUT:** *Fagara lemairei* De Wild. (41); **SAPI:** *Blighia welwitschii* (Hiern) Radlk. (41); *Pancovia harmsiana* Gilg. (40) (41); **SAPO:** *Chrysophyllum africanum* DC. (41); *C. pruniforme* Engl. (40); *Synsepalum longecuneatum* De Wild. (40); *S. subcordatum* De Wild. (41); **TIL:** *Microcos pinnatifida* (Mast.) Burret (40); **ULM:** *Celtis briei* De Wild. (40) (41).

Zemioses rufostriatus (Kleine)

ANN: *Anonidium mannii* Engl. et Diels (40) (41); *Isolona bruneelii* De Wild. (41); *Xylopia gilbertii* Boutique (41); *X. villosa* Chipp. (40); **BIG:** *Fernandoa adolfi-fridericii* (Gilg et Mildbr.) Heine (41); **BUR:** *Canarium schweinfurthii* Engl. (41); **CAE:** *Anthonotha macrophyllum* P. Beauv. (41); *Erythrophleum guineense* G. Don (39) (40) (41); *Gossweilerodendron balsamiferum* (Verm.) Harms. (39); *Oxystigma oxyphyllum* (Harms) J. Léonard (40) (41); *Paramacrolobium coeruleum* (Taub.) J. Léonard (41); *Scorodophloeus zenkeri* Harms (39) (41); **COM:** *Combretodendron macrocarpum* (Beauv.) Keay (41); **EUP:** *Discoglyprena caloneura* Prain (41); *Drypetes gossweileri* S. Moore (41); **LAU:** *Beilschmiedia gilbertii* Robyns et Wilczek var. *glabra* (40) (41); **LEG:** *Albizia* sp. (41); **MEL:** *Guarea cedrata* (A. Chev.) Pellegr. (39) (41); *G. laurentii* De Wild. (39) (41);

MOR: *Antiaris welwitschii* Engl. (39) (41); *Chlorophora excelsa* Benth. et Hook f. (41); *Myrianthus arboreus* P. Beauv. (41); **MYRI:** *Pycnanthus angolensis* (Welw.) Exell. (41); **OLA:** *Ongokea gore* (Hua) Pierre (40) (41); **PAN:** *Panda oleosa* Pierre (41); **PAP:** *Angylocalyx pynaertii* De Wild. (40) (41); **SAPI:** *Blighia welwitschii* (Hiern) Radlk. (40); **SAPO:** *Chrysophyllum africanum* DC. (41); *C. pruniforme* Engl. (41); *Pachystela excelsa* Baill. (41); **SIM:** *Klainedoxa gabonensis* Pierre var. *oblongifolia* (41); **STE:** *Sterculia bequaertii* De Wild. (40) (41); **STY:** *Afrostryax lepidophyllus* (Harms) Mildbr. (41); **ULM:** *Celtis briei* De Wild. (41).

ATOPOBRENTINI Damoiseau

Neoceocephalus punctatus Damoiseau

SAPO: *Synsepalum dulcificum* (Sch. et Th.) Daniell et Bell. (41).

Neoceocephalus rostralis Senna

OLA: *Ongokea gore* (Hua) Pierre (41); **SAPO:** *Chrysophyllum lacourtianum* De Wild. (41).

Neoceocephalus sculpturatus Senna

ANN: *Polyanthia suaveolens* Engl. et Diels (41); *Xylopia gilbertii* Boutique (41); **CAE:** *Scorodophloeus zenkeri* Harms (41); **COM:** *Combretodendron macrocarpum* (Beauv.) Keay (41); **IRV:** *Oxystigma oxyphyllum* (Harms) J. Léonard (41); **OLA:** *Strombosia glaucescens* Engl. (41); **SAPO:** *Chrysophyllum africanum* DC. (41); **SAPI:** *Pancovia laurentii* De Wild. (41).

HOPLOPISTHIINI Senna & Calabresi

Carcinopisthius oberthueri (Senna)

DIP: *Dipterocarpus pilosus* Roxb. (12) (13) (23) (32); **EUP:** *Sapium eugeniaefolium* Buch.-Ham. ex Wall. (27) (32); **MOR:** *Artocarpus lakoocha* Roxb. (23); *Ficus* sp. (63); **STE:** *Sterculia colorata* Roxb. (27) (32).

Hoplopisthius trichemerus Senna

FAB: *Dalbergia assamica* Benth. (12) (13) (23) (32); **STE:** *Pterocymbium* sp. (23) (32).

STEREODERMINI Sharp

Cerobates (Cerobates) aemulus Kleine

DIP: *Shorea leprosula* Miq. (29) (32).

Cerobates (Cerobates) angustipennis Senna

BOM: *Bombax malabaricum* DC. (20) (23) (32); **MOR:** *Artocarpus* sp. (63); *Ficus* sp. (32) (63).

Cerobates (Cerobates) birmanicus Senna

BUR: *Garuga pinnata* Roxb. (20) (23) (32); **MOR:** *Ficus rumphii* Blume (32); *F. tjakela* Miq. (20) (23) (32).

Cerobates (Cerobates) canaliculatus Motschulsky

MOR: *Ficus* sp. (63).

Cerobates (Cerobates) complanatus Senna

APO: *Alstonia boonei* De Wild. (41); **COM:** *Terminalia superba* Engl. et Diels (41); **EUP:** *Drypetes gossweileri* S. Moore (41); **LIL:** *Dracaena* sp. (41); **MOR:** *Chlorophora excelsa* Benth. et Hook (41); **STE:** *Pterygota* sp. (41).

Cerobates (Cerobates) conveniens Kleine

ANN: *Annona arenaria* Thonn (= *senegalensis* Pers.) (41); *Anonidium mannii* Engl. et Diels (41); **BUR:** *Canarium schweinfurthii* Engl. (41); **CAE:** *Erythrophleum guineense* G. Don (39) (40) (41); *Scorodophloeus zenkeri* Harms (39) (41); **COM:** *Combretodendron macrocarpum* (Beauv.) Keay (41); **EUP:** *Ricinodendron africanus* Muell.Arg. (41); **LEG:** *Tetrapleura tetraptera* (Thonn.) Taub. (41); **MEL:** *Entandrophragma candollei* Harms (41); *Guarea cedrata* (A. Chev.) Pellegr. (41); **OLA:** *Strombosia grandifolia* Hook. f. Benth. (41); *Strombosiaopsis tetrandra* Engl. (41); **MYRI:** *Staudtia stipitata* Warb. (41); **PAN:** *Panda oleosa* Pierre (41); **PAP:** *Afrormosia elata* Harms (39) (41); **RUB:** *Nauclea diderrichii* (De Wild.) Merrill (41); **STE:** *Sterculia tragacantha* Lindl. var. *bequaertii* De Wild. (40).

Cerobates (Cerobates) copiosus copiosus Kleine

ANN: *Polyanthia suaveolens* Engl. et Diels (40) (41); **APO:** *Clitandra* cfr. *staudtii* Stapf (40); *Conopharyngia durissima* Stapf (41); **CAE:** *Cynometra hankei* Harms (39) (41); *Scorodophloeus zenkeri* Harms (39) (41); **COM:** *Combretodendron macrocarpum* (Beauv.) Keay (40) (41); **EBE:** *Diospyros crassiflora* Hiern (41); **EUP:** *Cleistanthus* sp. (41); *Drypetes morocarpa* J. Léonard (41); *Macaranga lancifolia* Pax (41); *Microdesmis puberula* Hook. f. (41); **GUT:** *Garcinia punctata* Oliv. (41); *Symphonia globulifera* L. f. (41); **IRV:** *Irvingia gabonensis* Baill. (41); *Oxystigma oxyphyllum* (Harms) J. Léonard (41); **MEL:** *Carapa procera* DC. (41); *Guarea laurentii* De Wild. (41); *G. thompsoni* Sprague et Hutch. (39) (41); **MYRI:** *Staudtia stipitata* Warb. (39) (41); **OLA:** *Strombosia glaucescens* Engl. (41); *S. grandifolia* Hook. f. Benth. (41); **PAN:** *Panda oleosa* Pierre (41); **SAPI:** *Pancovia harmsiana* Gilg. (41); **SIM:** *Klainedoxa gabonensis* Pierre var. *oblongifolia* (41); **STE:** *Cola griseiflora* De Wild. (41); **ULM:** *Celtis briei* De Wild. (41); **VIO:** *Rinorea welwitschii* Engl. (41).

Cerobates (Cerobates) copiosus projectus Kleine

ANN: *Annona arenaria* Thonn (= *senegalensis* Pers.) (41); **COR:** *Afrocrania volkensii* (Harms) Hutchinson (41); **ERI:** *Ficalhoa laurifolia* Hiern (41); **EUP:** *Alchornea cordifolia* Muell.Arg. (41); *Neoboutonia macrocalyx* Pax (41); **FAG:** *Quercus suber* L. (41); **GUT:** *Symphonia globulifera* L. f. (41); *Symphonia* sp. (41); **LEG:** *Pterocarpus tinctorius* Welw. (41); **MOR:** *Ficus mucoso* Welw. ex Ficalho (41); *Myrianthus holstii* Engl. (41); **MYRS:** *Maesa lanceolata* Voigt (41); **MYRT:** *Eucalyptus saligna* Smith (41); **OLA:** *Strombosia grandifolia* Hook. f. Benth. (41); **POD:** *Podocarpus usambarensis* Pilg. (41); **PRO:** *Grevillea robusta* A. Cunn. (41); **RHA:** *Maesopsis eminii* Engl. (41); **ROS:** *Parinarium holstii* Engl. (41); **RUT:** *Fagara inaequalis* Engl. (41); **SAPO:** *Chrysophyllum* cfr. *fulvum* S. Moore (41); **VER:** *Tectonia* sp. (43).

Cerobates (Cerobates) corruptus Kleine

ANN: *Polyanthia suaveolens* Engl. et Diels (41).

Cerobates (Cerobates) credibilis Kleine

ANN: *Polyanthia suaveolens* Engl. et Diels (41); *Xylopia aethiopica* (Dun) A. Rich. (41); *X. gilber-*

tii Boutique (41); **APO:** *Alstonia boonei* De Wild. (39) (41); **BUR:** *Canarium schweinfurthii* Engl. (41); **CAE:** *Azelia bella* Harms (41); *Dialium excelsum* Louis ex Steyaert (41); *D. pachyphyllum* Harms (41); *Scorodophloeus zenkeri* Harms (39) (41); **COM:** *Combretodendron macrocarpum* (Beauv.) Keay (41); **EUP:** *Croton haumanianus* J. Léonard (41); *Discoglypsemna caloneura* Prain (41); *Macaranga* cfr. *lancifolia* Pax (41); *Phyllanthus discoideus* Müll. Arg. (41); **FLA:** *Caloncoba welwitschii* Gilg (41); **GUT:** *Garcinia punctata* Oliv. (41); *Mammea africana* G. Don (41); *Riciodendron africanus* Muell. Arg. (41); **IRV:** *Irvingia grandiflora* Engl. (41); **LAU:** *Beilschmiedia corbisieri* (Robyns) Robyns et Wilczek (41); **LEG:** *Albizia* sp. (41); **MEL:** *Lovoa trichilioides* Harms (41); **MOR:** *Antiaris welwitschii* Engl. (39) (41); *Bosqueia angolensis* Ficalho (41); *Chlorophora excelsa* Benth. et Hook (41); *Myrianthus arboreus* P. Beauv. (41); *Turraeanthus* sp. (41); **MYRI:** *Staudtia stipitata* Warb. (39) (41); **MYRT:** *Syzygium congolense* Vermoesen (41); **OLA:** *Strombosia glaucescens* Engl. (41); **PAN:** *Panda oleosa* Pierre (41); **RHA:** *Maesopsis eminii* Engl. (41); **ROS:** *Parinarium holstii* Engl. (41); **RUB:** *Aidia micrantha* (K. Schum.) Bullock ex F. White (41); *Canthium glabriflorum* Hiern (41); *Morinda lucida* Benth. (41); *Nauclea diderrichii* (De Wild.) Merrill (39) (41); **RUT:** *Fagara macrophylla* (Oliv.) Engl. (41); **SAPI:** *Allophyllum africanum* P. Beauv. (41); *Blighia welwitschii* (Hiern) Radlk. (41); *Pancovia harmsiana* Gilg. (41); **SAPO:** *Chrysophyllum africanum* DC. (41); *C. lacourtianum* De Wild. (41); *C. pruniforme* Engl. (41); *Synsepalum subcordatum* De Wild. (41); **SIM:** *Klainedoxa gabonensis* Pierre var. *oblongifolia* (41); **STE:** *Cola lateritia* K. Schum. (41); **TIL:** *Desplatsia dewevrei* (De Wild. et Th. Dur.) (41); *Microcos coriacea* (Mast.) Burret (41); **ULM:** *Celtis briei* De Wild. (41); *C. mildbraedii* Engl. (39) (41).

Cerobates (Cerobates) cruentatus Senna

ANN: *Anonidium mannii* Engl. et Diels (41); *Isolona bruneelii* De Wild. (41); *Polyanthia suaveolens* Engl. et Diels (39) (40) (41); *Xylopia gilbertii* Boutique (41); **APO:** *Alstonia boonei* De Wild. (39) (41); **BIG:** *Fernandoa adolfi-fridericii* (Gilg et Mildbr.) Heine (41); **BUR:** *Canarium schweinfurthii* Engl. (41); *Dacryodes edulis* (G. Don) H. J. Lam. (41); **BYT:** *Theobroma cacao* L. (41); **CAE:** *Amphimas pterocarpoides* Harms (41); *Pachyelasma tessmanii* (Harms) Harms (41); *Scorodophloeus zenkeri* Harms (39) (41); **COM:** *Combretodendron macrocarpum* (Beauv.) Keay (41); **EBE:** *Diospyros alboflavescens* (Gurke) F. White (41); *D. crassiflora* Hiern (41); **EUP:** *Drypetes gossweileri* S. Moore (39) (41); *D. morocarpa* J. Léonard (41); *Drypetes* sp. (41); *Macaranga* cfr. *lancifolia* Pax (41); *Riciodendron africanus* Muell. Arg. (41); **GUT:** *Garcinia punctata* Oliv. (41); **IRV:** *Oxystigma oxyphyllum* (Harms) J. Léonard (41); **LEG:** *Albizia* sp. (41); *Tetrapleura tetraptera* (Thonn.) Taub. (41); **MEL:** *Entandrophragma candollei* Harms (41); *E. cylindricum* (41); *Entandrophragma* sp. (39) (41); *Guarea cedrata* (A. Chev.) Pellegr. (39) (41); *G. laurentii* De Wild. (39) (41); *G. thompsoni* Sprague et Hutch. (39) (41); **MOR:** *Antiaris welwitschii* Engl. (39) (41); **OLA:** *Ongokea gore* (Hua) Pierre (39) (41); *Strombosia glaucescens* Engl. (41); *S. grandifolia* Hook. f. Benth. (41); *Strombosiopsis tetrandra* Engl. (41); **PAN:** *Panda oleosa* Pierre (40) (41); **PAP:** *Afrormosia elata* Harms (41); *Angylocalyx pynaertii* De Wild. (41); *Millettia drastica* Welw. (40); *Pterocarpus soyauxii* Taub. (41); **PAS:** *Barteria fistulosa* Mast. (41); **RUB:** *Nauclea diderrichii* (De Wild.) Merrill (39) (41); **SAPI:** *Blighia welwitschii* (Hiern) Radlk. (41); *Lecaniodiscus cupanioides* Planch. (41); *Pancovia harmsiana* Gilg. (40) (41); *P. laurentii* De Wild. (41); **SAPO:** *Chrysophyllum africanum* DC. (41); *C. lacourtianum* De Wild. (41); *Synsepalum subcordatum* De Wild. (41); *Tridesmostemon claessensii* De Wild. (41); **SIM:** *Hannoa klaineana* Pierre (41); *Klainedoxa gabonensis* Pierre var. *oblongifolia* (41); **STE:** *Cola griseiflora* De Wild. (40) (41); *C. lateritia* K. Schum. (41); *Sterculia bequaertii* De Wild. (41); **TIL:** *Desplatsia dewevrei* (De Wild. et Th. Dur.) (41); *Microcos coriacea* (Mast.) Burret (41); *M. pinnatifida* (Mast.) Burret (41); **ULM:** *Celtis mildbraedii* Engl. (39) (41).

Cerobates (Cerobates) debilis J. Thomson

ANA: *Sorindeia sparanoi* De Wild. (41); **ANN:** *Anonidium mannii* Engl. et Diels (41); *Isolona bruneelii* De Wild. (41); *Polyanthia suaveolens* Engl. et Diels (39) (40) (41); *Xylopia gilbertii* Boutique (41); *X. phloiodora* Mildbr. (41); *X. villosa* Chipp. (40); *Xylopia* sp. (41); **APO:** *Alstonia boo-*

nei De Wild. (41); *Conopharyngia durissima* Stapf (41); **BAR:** *Combretodendron macrocarpum* (P. Beauv.) Keay. (40) (41); **BIG:** *Fernandoa adolfi-fridericii* (Gilg et Mildbr.) Heine (41); **BOM:** *Ceiba pentandra* (L.) Gaertn. (41); *C. thoningii* A. Chev. (40); **BUR:** *Canarium schweinfurthii* Engl. (40) (41); *Dacryodes edulis* (G. Don) H. J. Lam. (41); *Pachylobus edulis* Don (40); **CAE:** *Azelia bella* Harms (41); *Amphimas pterocarpoides* Harms (41); *Anthonothea macrophyllum* P. Beauv. (41); *Cynometra hankei* Harms (41); *Erythrophleum guineense* G. Don (41); *Dalium excelsum* Louis ex Steyaert (40) (41); *Pachyelasma tessmanii* (Harms) Harms (41); *Paramacrolobium coeruleum* (Taub.) J. Léonard (40) (41); *Scorodophloeus zenkeri* Harms (39) (41); **COM:** *Combretodendron macrocarpum* (Beauv.) Keay (41); **EBE:** *Diospyros alboflavescens* (Gurke) F. White (41); *D. undabunda* Hiern ex Greves (41); **EUP:** *Croton haumanianus* J. Léonard (41); *Discoglypsemna caloneura* Prain (41); *Drypetes gossweileri* S. Moore (41); *D. morocarpa* J. Léonard (41); *D. cfr. louisii* J. Léonard (41); *Drypetes* sp. (41); *Hymenocardia ulmoides* Oliv. (41); *Macaranga lancifolia* Pax (41); *Phyllanthus discoideus* Muell. Arg. (41); *Ricinodendron africanus* Muell. Arg. (41); **GUT:** *Garcinia punctata* Oliv. (41); *Mammea africana* G. Don (41); **IRV:** *Iringia grandiflora* Engl. (41); *Oxystigma oxyphyllum* (Harms) J. Léonard (41); **LAU:** *Beilschmiedia corbisieri* (Robyns) Robyns et Wilczek (41); **LEG:** *Albizia gummifera* (Gmel.) C. A. Sm. (40); *Albizia* sp. (39) (41); *Parkia bicolor* A. Chevalier (41); *Piptadeniastrum africanum* (Hook. f.) Brenan (41); *Tessmannia anomala* (Micheli) Harms (41); *T. parvifolia* Harms (40); *Tetrapleura tetraptera* (Thonn.) Taub. (41); **LIL:** *Dracaena* sp. (41); **LIN:** *Ochthocosmus* cfr. *africana* Hook. f. (41); **MEL:** *Entandrophragma angolense* C. DC. (39) (41); *E. candollei* Harms (41); *E. utile* (Dawe et Sprague) (41); *Entandrophragma* sp. (41); *Guarea cedrata* (A. Chev.) Pellegr. (39) (41); *G. laurentii* De Wild. (39) (41); *G. thompsonii* Sprague et Hutch (41); *Lovoa trichilioides* Harms (41); *Melia* sp. (41); *Trichilia prieureana* Juss. (41); *T. rubescens* Oliver (41); **MOR:** *Antiaris welwitschii* Engl. (39) (41); *Bosqueia angolensis* Ficalho (40) (41); *Chlorophora excelsa* Benth. et Hook (39) (40) (41); *Musanga cecropioides* R. Br. (41); *Myrianthus arboreus* P. Beauv. (41); **MYRI:** *Coelocaryon preussii* Warb. (41); *Pycnanthus angolensis* (Welw.) Exell. (41); *Staudtia stipitata* Warb. (41); **MYRT:** *Syzygium congolense* Vermoesen (41); **OKT:** *Okoubaka* sp. (41); **OLA:** *Ongokea gore* (Hua) Pierre (41); *Strombosia glaucescens* Engl. (40) (41); *S. grandifolia* Hook. f. Benth. (41); *Strombosiaopsis tetrandra* Engl. (41); **PAN:** *Panda oleosa* Pierre (40) (41); **PAP:** *Afrormosia elata* Harms (41); *Angylocalyx pyraertii* De Wild. (41); *Millettia drastica* Welw. (41); *Pterocarpus soyauxii* Taub. (41); **PAS:** *Barteria fistulosa* Mast. (41); *B. nigrifolia* Hook. f. (41) (41); **RHA:** *Maesopsis eminii* Engl. (41); **ROS:** *Parinarium holstii* Engl. (41); **RUB:** *Canthium glabriflorum* Hiern (41); *Morinda lucida* Benth. (41); *Nauclea diderrichii* (De Wild.) Merrill (41); *Plectronia dewevrei* Wildem. (41); **RUT:** *Vepris louisii* Gilbert (41); **SAPI:** *Blighia welwitschii* (Hiern) Radlk. (41); *Lecaniodiscus cupanioides* Planch. (41); *Pancovia harmsiana* Gilg. (41); **SAPO:** *Chrysophyllum africanum* DC. (41); *C. lacourtianum* De Wild. (41); *C. pruniforme* Engl. (41); *Manilkara maleoleus* J. Louis (41); *Synsepalum subcordatum* De Wild. (41); *Tridesmostemon claessensii* De Wild (41); **SIM:** *Klainedoxa gabonensis* Pierre var. *oblongifolia* (41); **STE:** *Cola griseiflora* De Wild. (41); *Pterygota bequaertii* De Wild. (41); *P. mildbraedii* Engl. (41); *Sterculia bequaertii* De Wild. (41); **STY:** *Afrotyrax lepidophyllus* (Harms) Mildbr. (41); **TIL:** *Desplatsia dewevrei* (De Wild. et Th. Dur.) (41); *Duboscia viridiflora* Mildbr. (41); **ULM:** *Celtis briei* De Wild. (40) (41); *C. mildbraedii* Engl. (41); **VER:** *Vitex congolensis* De Wild. et Dur. (41); *Vitex* sp. (41).

Cerobates (Cerobates) dilutus Kleine

ANN: *Anonidium mannii* Engl. et Diels (40) (41); *Polyanthia suaveolens* Engl. et Diels (40) (41); **APO:** *Alstonia boonei* De Wild. (41); **BUR:** *Canarium schweinfurthii* Engl. (40) (41); **CAE:** *Scorodophloeus zenkeri* Harms. (41); **COM:** *Combretodendron macrocarpum* (P. Beauv.) Keay (40) (41); **EBE:** *Diospyros crassiflora* Hiern (41); **EUP:** *Drypetes gossweileri* S. Moore (41); *Drypetes* cfr. *louisii* J. Léonard (41); *Macaranga* cfr. *lancifolia* Pax (40) (41); *Microdesmis puberula* Hook (40) (41); *Ricinodendron africanus* Muell. Arg. (41); **FAG:** *Quercus suber* L. (41); **MEL:** *Carapa procera* DC. (40); *Entandrophragma candollei* Harms (41); *Guarea cedrata* (A. Chev.) (41); *G. laurentii* De Wild. (39) (40) (41); *G. smeathmanni* Oliver (41); *G. thompsonii* Sprague et Hutch. (41); **MYRI:** *Staudtia stipitata* Warb. (39) (40) (41); **OKT:** *Okoubaka aubrevillei* Pellegr. et Nor-

mand (41); **OLA:** *Ongokea gore* (Hua) Pierre (41); *Strombosia glaucescens* Engl. (40) (41); **PAN:** *Panda oleosa* Pierre (41); **PAP:** *Afrormosia elata* Harms (41); *Angylocalyx pynaertii* De Wild. (41); *Millettia drastica* Welw. (41); *Pterocarpus soyauxii* Taub. (41); **RUB:** *Heinsia pulchella* K. Schum. (40) (41); *Mitragyna stipulosa* Kuntze (41); *Mitragyna* sp. (40); *Morinda lucida* Benth. (41); **RUT:** *Vepris louisii* Gilbert (41); **SAPI:** *Blighia welwitschii* (Hiern) Radlk. (41); **SAPO:** *Omphalocarpum* sp. (40); *Synsepalum subcordatum* De Wild. (41); **SIM:** *Hannoa klaineana* Pierre (41); *Klainedoxa gabonensis* Pierre var. *oblongifolia* (41); **STE:** *Cola griseiflora* De Wild. (41); **ULM:** *Celtis mildbraedii* Engl. (41).

Cerobates (Cerobates) fossulatus Motschulsky

MOR: *Artocarpus lakoocha* Roxb. (13) (23) (32).

Cerobates (Cerobates) hybridus Senna

ANN: *Anonidium mannii* Engl. et Diels (40) (41); *Polyanthia suaveolens* Engl. et Diels (41); **APO:** *Alstonia boonei* De Wild. (39) (41); **BUR:** *Canarium schweinfurthii* Engl. (41); **CAE:** *Scorodophloeus zenkeri* Harms. (40) (41); **COM:** *Combretodendron macrocarpum* (Beauv.) Keay (41); **EUP:** *Macaranga* cfr. *lancifolia* Pax (41); *Ricinodendron africanus* Muell. Arg. (41); *Tetrorchidium didymostemon* Pax et Hoffm. (40) (41); **IRV:** *Oxystigma oxyphyllum* (Harms) J. Léonard (41); **LEC:** *Combretodendron macrocarpum* (Beauv.) Keay (41); **LEG:** *Tetrapleura tetraptera* (Thonn.) Taub. (40); **MEL:** *Guarea laurentii* De Wild. (39) (40) (41); *G. thompsonii* Sprague et Hutch. (41); *Trichilia prieureana* Juss. (41); **MYRI:** *Staudtia stipitata* Warb. (40); **OLA:** *Ongokea gore* (Hua) Pierre (39) (40) (41); *Strombosia glaucescens* Engl. (41); **PAN:** *Panda oleosa* Pierre (40); **PAP:** *Pterocarpus soyauxii* Taub. (41); **ROS:** *Parinarium* sp. (28); **SAPO:** *Chrysophyllum lacourtianum* De Wild. (40) (41); *Synsepalum subcordatum* De Wild. (40) (41); **STE:** *Cola griseiflora* De Wild. (41); **ULM:** *Celtis briei* De Wild. (40).

Cerobates (Cerobates) laevipennis Senna

BOM: *Bombax insigne* Wall. (20) (23) (32); **STE:** *Sterculia colorata* Roxb. (20) (23) (32).

Cerobates (Cerobates) ophthalmicus Pascoe

STE: *Brachychiton* sp. (57).

Cerobates (Cerobates) puerilis Kleine

APO: *Dyera costulata* Hook. f. (29).

Cerobates (Cerobates) punctulatus Senna

ANN: *Polyanthia suaveolens* Engl. et Diels (41); *Xylopia gilbertii* Boutique (41); *X. villosa* Chipp. (40); **APO:** *Alstonia boonei* De Wild. (41); **BUR:** *Dacryodes yangambiensis* J. Louis ex Troupin (40) (41); **COM:** *Terminalia superba* Engl. et Diels (41); **IRV:** *Irvingia gabonensis* Baill. (41); *I. grandiflora* Engl. (40) (41); **CAE:** *Cynometra alexandri* C. H. Wright (40); *C. hankei* Harms (40); *Dialium corbisieri* Staner (40) (41); *D. excelsum* Louis ex Steyaert (41); *D. pachyphyllum* Harms (41); *Erythrophleum guineense* G. Don (39) (40) (41); *Gilbertiodendron dewevrei* (De Wild.) (39) (40) (41); *Gossweilerodendron balsamiferum* (Verm.) Harms (41); *Oxystigma oxyphyllum* (Harms) J. Léonard (40); *Scorodophloeus zenkeri* Harms. (40) (41); **COM:** *Combretodendron macrocarpum* (Beauv.) Keay (41); **EBE:** *Diospyros alboflavescens* (Gurke) F. White (41); **EUP:** *Cleistanthus* sp. (40); *Drypetes gossweileri* S. Moore (41); *D. morocarpa* J. Léonard (41); *Microdesmis puberula* Hook. f. (41); **GUT:** *Garcinia punctata* Oliv. (41); **IRV:** *Oxystigma oxyphyllum* (Harms) J. Léonard (41); **LAU:** *Beilschmiedia gilbertii* Robyns et Wilczek var. *glabra* (41); **LEG:** *Piptadenia africana*

Hook. f. (40); *Piptadeniastrum africanum* (Hook. f.) Brenan (41); **MEL:** *Entandrophragma* sp. (39); *Guarea cedrata* (A. Chev.) (39) (40) (41); *G. laurentii* De Wild. (39) (40) (41); *G. thompsonii* Sprague et Hutch. (41); *Trichilia rubescens* Oliv. (40) (41); *T. welwitschii* C. DC (41); **MOR:** *Myrianthus arboreus* P. Beauv. (41); **MYRI:** *Coelocaryon preussii* Warb. (41); *Pycnanthus angolensis* (Welw.) Exell. (40) (41); *Staudtia stipitata* Warb. (39) (40) (41); **MYRT:** *Syzygium* sp. (41); **OLA:** *Strombosia glaucescens* Engl. (40) (41); *Strombosia* sp. (41); *Strombosiopsis tetrandra* Engl. (40) (41); **PAN:** *Panda oleosa* Pierre (41); **PAP:** *Afrormosia elata* Harms (39) (40) (41); *Angylocalyx pynaertii* De Wild. (40) (41); *Pterocarpus soyauxii* Taub. (40) (41); **RUB:** *Nauclea diderrichii* (De Wild.) Merrill (39) (41); *Sarcocephalus* cfr. *trillesii* Pierre (40); **RUT:** *Fagara lemairei* De Wild. (41); **SAPO:** *Chrysophyllum lacourtianum* De Wild. (40) (41); *C. pruiniforme* Engl. (40) (41); *Synsepalum subcordatum* De Wild. (41); **SAPI:** *Blighia welwitschii* (Hiern) Radlk. (40) (41); *Pancovia harmsiana* Gilg. (41); **STE:** *Cola griseiflora* De Wild. (41); *Pterygota* sp. (41); **ULM:** *Celtis mildbraedii* Engl. (39) (40) (41); *C. briei* De Wild. (41); **VIO:** *Rinorea multinervis* M. Brandt (41).

Cerobates (Cerobates) sennae Calabresi

APO: *Alstonia boonei* De Wild. (39) (41); **BUR:** *Canarium schweinfurthii* Engl. (41); **CAE:** *Scorodophloeus zenkeri* Harms. (41); **COM:** *Combretodendron macrocarpum* (Beauv.) Keay (41); **EUP:** *Croton haumanianus* J. Léonard (41); *Discoglypsemna caloneura* Prain (41); *Drypetes morocarpa* J. Léonard (41); *Microdesmis puberula* Hook (41); **IRV:** *Oxystigma oxyphyllum* (Harms) J. Léonard (41); **LEG:** *Albizia* sp. (41); *Tetrapleura tetraptera* (Thonn.) Taub. (41); **MEL:** *Guarea laurentii* De Wild. (39) (41); *G. thompsonii* Sprague et Hutch. (41); *Melia* sp. (41); **MOR:** *Myrianthus arboreus* P. Beauv. (41); **SAPI:** *Blighia welwitschii* (Hiern) Radlk. (41); **SAPO:** *Chrysophyllum fulvum* S. Moore (41); *Synsepalum subcordatum* De Wild. (41); **ULM:** *Celtis mildbraedii* Engl. (39) (41); *C. briei* De Wild. (41).

Cerobates (Cerobates) sexsulcatus Motschulsky

BOM: *Bombax malabaricum* DC. (12) (20) (32); **COM:** *Terminalia* sp. (63); **DIP:** *Shorea* sp. (32); **FAB:** *Butea frondosa* Roxb. (13) (23) (24) (32); **HAM:** *Liquidambar* sp. (63); **LEG:** *Albizia lucida* Benth. (13) (23) (24) (32); *A. procera* Benth. (32); **MEL:** *Dysoxylum binectariferum* Hook. (12) (13); *D. binectariferum* Hook. (23) (24) (32); **MOR:** *Artocarpus integrifolia* L. f. (32); *A. kunstleri* King ex Hook. f. (32); *Artocarpus* sp. (63); *Ficus* sp. (63); *Trema* sp. (63); **SAN:** *Santalum album* L. (32).

Cerobates (Cerobates) sulcatus sulcirostris J. Thomson

ANN: *Anonidium mannii* Engl. et Diels (40); *Polyanthia suaveolens* Engl. et Diels (39) (40) (41); *Xylopia villosa* Chipp. (40); **APO:** *Alstonia boonei* De Wild. (39) (41); *Alstonia* cfr. *congensis* Engl. (40); **BUR:** *Canarium schweinfurthii* Engl. (41); *Dacryodes edulis* (G. Don) H. J. Lam. (41); *Pychylobus edulis* G. Don (40); **BYT:** *Theobroma cacao* L. (41); **CAE:** *Dialium excelsum* Louis ex Steyaert (41); *Oxystigma oxyphyllum* (Harms) J. Léonard (40) (41); *Scorodophloeus zenkeri* Harms. (39) (40) (41); **COM:** *Combretodendron macrocarpum* (Beauv.) Keay (41); *Terminalia superba* Engl. et Diels (39) (41); **EBE:** *Diospyros alboflavescens* (Gurke) F. White (41); *D. undabunda* Hiern ex Greves (41); **EUP:** *Croton haumanianus* J. Léonard (41); *Discoglypsemna caloneura* Prain (40); *Drypetes gossweileri* S. Moore (40) (41); *D. ituriensis* Pax et K. Hoffm. (40); *D. morocarpa* J. Léonard (40) (41); *Ricinodendron africanus* Muell. Arg. (41); **FAG:** *Quercus suber* L. (41); **GUT:** *Mammea africana* G. Don (41); **LAU:** *Beilschmiedia corbisieri* (Robyns) Robyns et Wilczek (41); **LEG:** *Albizia gummifera* (Gmel.) C. A. Sm. (40); *Albizia* sp. (39) (41); *Tetrapleura tetraptera* (Thonn.) Taub. (41); **MEL:** *Carapa* sp. (41); *Entandrophragma angolense* DC. (40); *E. candollei* Harms (41); *Guarea cedrata* Pellegr. (39) (40) (41); *Lovoa trichilioides* Harms (40); *Melia* sp. (41); **MOR:** *Antiaris welwitschii* Engl. (40); *Macaranga* cfr. *lancifolia* Pax (41); *Myrianthus arboreus* P. Beauv. (41); **OKT:** *Okoubaka* sp. (41); **OLA:** *Ongokea gore* (Hua) Pierre (39) (40) (41); *Strombosia glaucescens* Engl. (40) (41); *S. grandifolia* Hook. f. Benth. (41); *S. scheffleri*

Engl. (40); **PAN:** *Panda oleosa* Pierre (40) (41); **PAP:** *Afrormosia elata* Harms (39) (41); *Millettia drastica* Welw. (41); *Pterocarpus soyauxii* Taub. (41); **RHI:** *Anisophyllea* sp. (40); **RUB:** *Nauclea diderrichii* (De Wild.) Merrill (39); **RUT:** *Fagara inaequalis* Engl. (41); **SAPI:** *Lecaniodiscus cupanioides* Planch. (41); *Pancovia harmsiana* Gilg. (40) (41); **SAPO:** *Chrysophyllum africanum* DC. (41); *C. pruniforme* Engl. (40) (41); *Chrysophyllum* sp.; *Synsepalum subcordatum* De Wild. (40) (41); *Tridesmostemon claessensii* De Wild. (41); **SIM:** *Klainedoxa gabonensis* Pierre var. *oblongifolia* (41); **STE:** *Cola griseiflora* De Wild. (40); *Pterygota bequaertii* De Wild. (41); *P. mildbraedii* Engl. (41); **TIL:** *Desplatsia dewevrei* (De Wild. et Th. Dur.) (41); *Duboscia viridiflora* Mildbr. (41); **ULM:** *Celtis mildbraedii* Engl. (39) (40) (41); *C. soyauxii* Engl. (40).

Cerobates (Cerobates) sumatranus Senna

MEL: *Dysoxylum binectariferum* Hook. (12) (13) (23) (32); **MOR:** *Artocarpus integrifolia* L. f. (32); *Artocarpus* sp. (63); *Ficus* sp. (63).

Cerobates (Cerobates) tristriatus (Lund)

BOM: *Bombax malabaricum* DC. (12) (13) (20) (23) (32) (38) (58); **FAB:** *Butea frondosa* Roxb. (13) (23) (32); **GRA:** "bamboo" (18); **MORI:** *Moringa pterygosperma* (32); **MOR:** *Artocarpus integrifolia* L. f. (32).

Cerobates (Ionthocerus) carinensis (Senna)

FAB: *Erythrina indica* Lam. (32); **HAM:** *Liquidambar* sp. (63); **LEG:** *Albizia procera* Benth. (32); **MOR:** *Trema* sp. (63); **ROS:** *Rubus* sp. (63); **RUB:** *Neonauclea griffithii* Merrill (27) (32).

Cerobates (Ionthocerus) conradti (Senna)

ANN: *Anonidium mannii* Engl. et Diels (41); *Xylopia gilbertii* Boutique (41); *X. phloiodora* Mildbr. (41); **BIG:** *Fernandoa adolfi-fridericii* (Gilg et Mildbr.) Heine (41); **BOM:** *Ceiba pentandra* (L.) Gaertn. (41); **BUR:** *Dacryodes edulis* (G. Don) H. J. Lam. (41); **CAE:** *Azelia bella* Harms (41); *Anthonotha macrophyllum* P. Beauv. (41); **CAE:** *Cynometra hankei* Harms (41); *Dialium excelsum* Louis ex Steyaert (41); *Erythrophleum guineense* G. Don (41); *Scorodophloeus zenkeri* Harms (41); **COM:** *Combretodendron macrocarpum* (Beauv.) Keay (41); **EBE:** *Diospyros alboflavescens* (Gurke) F. White (41); *D. undabunda* Hiern ex Greves (41); **EUP:** *Croton haumanianus* J. Léonard (41); *Discoglypsemna caloneura* Prain (41); *Hymenocardia ulmoides* Oliv. (41); **FLA:** *Caloncoba welwitschii* Gilg (41); **GUT:** *Mammea africana* G. Don (41); **IRV:** *Irvingia grandifolia* Engl. (41); **LEG:** *Albizia* sp. (41); *Tessmannia anomala* (Micheli) Harms (41); *Tetrapleura tetraptera* (Thonn.) Taub. (41); **MEL:** *Entandrophragma candollei* Harms (41); *Guarea cedrata* (A. Chev.) Pellegr. (39) (41); *Lovoa trichilioides* Harms (41); **MOR:** *Antiaris welwitschii* Engl. (41); **OKT:** *Okoubaka* sp. (41); **OLA:** *Strombosia grandifolia* Hook. f. Benth. (41); **PAN:** *Panda oleosa* Pierre (41); **PAP:** *Afrormosia elata* Harms (41); *Millettia drastica* Welw. (41); **PAS:** *Barteria fistulosa* Mast. (41); *B. nigriflora* Hook f. (41); **RUB:** *Canthium glabriflorum* Hiern (41); *Morinda lucida* Benth. (41); **SAPI:** *Blighia welwitschii* (Hiern) Radlk. (41); *Lecaniodiscus cupanioides* Planch. (41); *Pancovia harmsiana* Gilg. (41); *Plectronia dewevrei* Wildem. (41); **SAPO:** *Chrysophyllum africanum* DC. (41); *C. pruniforme* Engl. (41); *Synsepalum subcordatum* De Wild. (41); **SIM:** *Klainedoxa gabonensis* Pierre var. *oblongifolia* (41); **STE:** *Sterculia bequaertii* De Wild. (41); **TIL:** *Duboscia viridiflora* Mildbr. (41).

Cerobates (Ionthocerus) crematus Lacordaire

BUR: *Garuga pinnata* Roxb. (32); **CAE:** *Poinciana elata* L. (13) (23) (32); **EBE:** *Diospyros pyrrhocarpa* Miq. (32); **MOR:** *Artocarpus integrifolia* L. f. (32).

Cerobates (Ionthocerus) usambaricus (Senna)

ANN: *Anonidium mannii* Engl. et Diels (40) (41); *Polyanthia suaveolens* Engl. et Diels (40); *Xylopia aethiopica* (Dun) A. Rich. (40); *X. villosa* Chipp. (40); **BUR:** *Canarium schweinfurthii* Engl. (40); **CAE:** *Afzelia bella* Harms (40); *Dalium excelsum* Louis ex Steyaert (40); *D. pachyphyllum* Harms (40); *Scorodophloeus zenkeri* Harms (40); **EUP:** *Croton haumanianus* J. Léonard (40); *Discoglyprena caloneura* Prain (40); *Macaranga* cfr. *lancifolia* Pax (40); **FLA:** *Caloncoba welwitschii* Gilg (40); **GUT:** *Garcinia punctata* Oliv. (40); *Mammea africana* G. Don (41); **IRV:** *Irvingia grandiflora* Engl. (40); *Oxystigma oxyphyllum* (Harms) J. Léonard (41); **LAU:** *Beilschmiedia corbisieri* (Robyns) Robyns et Wilczek (40); *Klainedoxa gabonensis* Pierre (40); **LEG:** *Albizia gummifera* (Gmel.) C. A. Sm. (40); *Pentaclethra macrophylla* Benth. (40); **MOR:** *Antiaris welwitschii* Engl. (40); *Myrianthus arboreus* P. Beauv. (40); **MYRI:** *Staudtia stipitata* Warb. (40); **MYRT:** *Syzygium congolense* Vermoesen (40); **OLA:** *Strombosia glaucescens* Engl. (40); **PAN:** *Panda oleosa* Pierre (40); **PAS:** *Barteria fistulosa* Mast. (40); **RHA:** *Maesopsis eminii* Engl. (40); **ROS:** *Parinarium* cfr. *tenuifolium* A. Chev. (40); **RUB:** *Canthium glabriflorum* Hiern (40); *Morinda lucida* Benth. (40); *Randia congolana* De Wild et Th. Dur. (40); *Sarcocephalus* cfr. *trillesii* Pierre (40); **RUT:** *Fagara macrophylla* (Oliv.) Engl. (40); **SAPI:** *Allophyllus africanus* P. Beauv. (40); *Blighia welwitschii* (Hiern) Radlk. (40); *Eriocoelum* sp. (40); *Pancovia harmsiana* Gilg (40) (41); **SAPO:** *Chrysophyllum africanum* DC. (40); *C. lacourtianum* De Wild. (41); *C. pruiniforme* Engl. (40); **STE:** *Cola griseiflora* De Wild. (40) (41); **TIL:** *Desplatsia dewevrei* (De Wild. et Th. Dur.) (40); *Microcos coriacea* (Mast.) Burret (40); **ULM:** *Celtis briei* De Wild. (40) (41); *C. mildbraedii* Engl. (40).

Metatrachelizus abjectus Kleine

DIP: *Shorea* sp. (23) (32).

Stereodermus pygmaeus (Gyllenhal)

MOR: *Castilloa costaricana* Liedm. (19) (23).

Stereodermus infidus Senna

MOR: *Ficus* sp. (63).

TAPHRODERINAE Lacordaire

Bolbocranius anaticeps (Kolbe)

APO: *Pleiocarpa micrantha* Stapf (41); **STE:** *Cola griseiflora* De Wild. (41).

Bolbocranius bicolor Senna

ANA: *Antrocaryon nannanii* De Wild. (40) (41); *Sorindeia sparanoi* De Wild. (41); **ANN:** *Anonidium mannii* (Oliv.) Engl. et Diels (40) (41); *Isolona bruneelii* De Wild. (41); *Pleiocarpa tubicina* Stapf. (41); *Polyanthia suaveolens* Engl. et Diels (39) (40) (41); *Xylopia aethiopica* (Dun) A. Rich. (41); *X. gilbertii* Boutique (41); *X. phloioidora* Mildbr. (41); **APO:** *Alstonia boonei* De Wild. (41); **BAR:** *Combretodendron macrophyllum* (P. Beauv.) Keay. (40); **BIG:** *Fernandoa adolfi-fridericii* (Gilg et Mildbr.) Heine (41); **BUR:** *Canarium schweinfurthii* Engl. (41); *Dacryodes* sp. (41); **CAE:** *Cynometra hankei* Harms (40) (41); *Dalium corbisieri* Staner (40) (41); *D. excelsum* Louis ex Steyart (40); *D. pachyphyllum* Harms (40) (41); *Gossweilerodendron balsamiferum* (Verm.) Harms (39) (41); *Erythrophleum guineense* G. Don (41); *Oxystigma oxyphyllum* (Harms) J. Léonard (40); *Oxystigma* sp. (41); *Pachyelasma tessmanii* (Harms) Harms (40); *Scorodophloeus zenkeri* Harms

(39) (40) (41); **COM:** *Combretum oblungum* K. Hoffm. (41); *Terminalia superba* Engl. et Diels (41); **CUP:** *Cupressus lusitanica* Carr. (41); **EBE:** *Diospyros alboflavescens* (Gurke) F. White (41); *D. crassiflora* Hiern (41); **EUP:** *Cleistanthus* sp. (41); *Croton haumannianus* J. Léonard (40); *Disco-glypsemna caloneura* Prain (41); *Drypetes cinnabarina* Pax et K. Hoffm. var. *congolensis* (41); *D. leonensis* Pax var. *glabra* J. Léonard (40); *D. gossweileri* S. Moore (40) (41); *D. monocarpa* J. Léonard (40) (41); *D. undulata* Louis (40); *Drypetes* cfr. *louisii* J. Léonard (41); *D. pinoso-dentata* (Pax) Hutch. var. *longipeducellata* J. Léonard (40) (41); *Drypetes* sp. (41); *Hymenocardia acida* Tul. (41); *Macaranga lancifolia* Pax (41); *Macaranga* cfr. *lancifolia* Pax (40) (41); *Ricinodendron africanus* Muell. Arg. (41); **FLA:** *Caloncoba welwitschii* Gilg (40) (41); **GUT:** *Garcinia punctata* Oliv. (40) (41); *G. smeathmanni* Oliver (41); *Mammea africana* G. Don (41); **IRV:** *Irvingia gabonensis* Baill. (41); *I. grandifolia* Engl. (41); *Irvingia* sp. (41); *Klainedoxa gabonensis* Pierre (40); **LAU:** *Beilschmiedia gilbertii* Robyns et Wilczek var. *glabra* (41); **LEG:** *Albizia gummifera* (Gmel.) C. A. Sm. (40); *Albizia* sp. (39) (41); *Pentaclethra macrophylla* Benth. (41); *Piptadeniastrum* sp. (41); *Tetrapleura tetraptera* (Thonn.) Taub. (41); **LIL:** *Dracaena* sp. (41); **MEL:** *Entandrophragma angolense* C. DC. (41); *E. candollei* Harms (39) (40) (41); *E. utile* (Dawe et Sprague) (41); *Entandrophragma* sp. (39) (41); *Guarea cedrata* (A. Chev.) Pellegr. (39) (41); *G. laurentii* De Wild. (39) (40) (41); *G. thompsonii* Sprague et Hutch (41); *Lovoa trichilioides* Harms (39) (40) (41); *Lovoa* sp. (41); *Trichilia lanata* A. Chev. (40) (41); *T. prieureana* Juss. (41); *T. rubescens* Oliv. (41); **MOR:** *Antiaris welwitschii* Engl. (41); *Antiaris* sp. (41); *Bosqueia* sp. (41); *Chlorophora excelsa* Benth. et Hook f. (39) (40) (41); *Myrianthus arboreus* P. Beauv. (41); **MYRI:** *Coelocaryon preussii* Warb. (41); *Pycnanthus angolensis* (Welw.) Exell. (40) (41); *Staudtia stipitata* Warb. (39) (40) (41); **MYRT:** *Eucalyptus saligna* Smith (41); **OLA:** *Olea hochstetteri* Baker (41); *Ongokea gore* (Hua) Pierre (41); *Strombosia glaucescens* Engl. (40); *S. grandifolia* Hook. f. Benth. (41); *S. scheffleri* Engl. (40); *Strombosia* sp. (41); *Strombosiopsis tetrandra* Engl. (41); **PAN:** *Panda oleosa* Pierre (40) (41); **PAP:** *Afrormosia elata* Harms (41); *Angylocalyx pynaertii* De Wild. (40) (41); *Millettia* cfr. *drastica* Welw. (41); *Pterocarpus soyauxii* Taub. (40) (41); **PAS:** *Barteria nigrifolia* Hook. f. (40); **ROS:** *Parinarium* sp. (41); **RUB:** *Mitragyna stipulosa* Kuntze (41); *Nauclea diderichii* (De Wild.) Merrill (39) (41); *Pausinystalia pynaertii* De Wild. (41); *Sarcocephalus* cfr. *trillesii* Pierre (40); **RUT:** *Fagara inaequalis* Engl. (41); *F. lemairei* De Wild. (41); *Fagara* sp. (41); *Vepris louisii* Gilbert (41); **SAPI:** *Allophyllus africanus* P. Beauv. (41); *Aphania senegalensis* Radlk. (41); *Blighia welwitschii* (Hiern) Radlk. (40) (41); *Pancovia harmsiana* Gilg. (40) (41); *P. laurentii* De Wild. (41); *Radlkofera calodendron* Gilg. (41); **SAPO:** *Autranella* sp. (41); *Chrysophyllum africanum* A. DC. (40) (41); *C. fulvum* S. Moore (41); *C. lacourtianum* De Wild. (40) (41); *C. pruniforme* Engl. (40) (41); *Omphalocarpum* cfr. *mortehani* De Wild. (40) (41); *Synsepalum dulcificum* (Sch. et Th.) Daniell et Bell. (41); *S. subcordatum* De Wild. (40) (41); *Tridesmostemon claessensii* De Wild (41); **SIM:** *Hannoa klaineana* Pierre (40) (41); *Klainedoxa gabonensis* Pierre var. *oblongifolia* (41); **STE:** *Cola griseiflora* De Wild. (40) (41); *C. lateritia* K. Schum. (41); *Cola* cfr. *lateritia* K. Schum. (40); *Pterygota mildbraedii* Engl. (41); **STY:** *Afrostryx lepidophyllus* (Harms) Mildbr. (40) (41); **TIL:** *Desplatsia dewevrei* (De Wild. et Th. Dur.) (40) (41); **ULM:** *Celtis briei* De Wild. (41); **VER:** *Vitex congolensis* De Wild. et Dur. (40) (41); *V. welwitschii* Guerke (41); **VIO:** *Rinorea welwitschii* Engl. (41).

Bolbocranius csikii (Bolkay)

ANA: *Antrocaryon nannani* De Wild (41); *Sorindeia lemairei* De Wild. (40); *S. sparanoi* De Wild. (41); *Sorindeia* aff. *claessensii* (40); **ANN:** *Anonidium mannii* Engl. et Diels (40) (41); *Isolona brunneii* De Wild. (41); *Monodora myristica* (Gaertn.) Dun (41); *Polyanthia suaveolens* Engl. et Diels (39) (40) (41); *Xylopia aethiopica* (Dun) A. Rich. (40); *X. gilbertii* Boutique (41); *X. phloiadora* Mildbr. (41); *Xylopia* cfr. *ripicola* Louis (40); *Xylopia* sp.; **APO:** *Alstonia boonei* De wild. (39) (41); *Conopharyngia durissima* Stapf (40) (41); *Funtumia* cfr. *latifolia* Stapf (40) (40); *Pleiocarpa tubicina* Stapf. (41); **BAR:** *Combretodendron macrophyllum* (P. Beauv.) Keay. (40); **BIG:** *Fernandoa adolfi-fridericii* (Gilg et Mildbr.) Heine (40) (41); **BUR:** *Canarium schweinfurthii* Engl. (41); *Dacryodes edulis* (G. Don) H. J. Lam. (41); *Dacryodes* sp. (41); *Pachylobus edulis* Don (40); **BYT:** *Theobroma cacao* L. (41); **CAE:** *Afzelia bella* Harms (41); *Amphimas pterocarpoides* Harms (40)

(41); *Anthonotha macrophyllum* P. Beauv. (40) (41); *Cynometra alexandri* C. H. Wright (41); *C. hankei* Harms (39) (40) (41); *Dialium corbisieri* Staner (40) (41); *D. excelsum* Louis ex Steyaert (41); *D. pachyphyllum* Harms (40) (41); *D. tessmannii* Harms (40); *Erythrophleum guineense* G. Don (39) (40) (41); *Gossweilerodendron balsamiferum* (Vermoesen) (39) (40) (41); *Julbernardia sereti* (De Wild.) Troupin (40); *Oxystigma oxyphyllum* (Harms) J. Léonard (40) (41); *Pachyelasma tessmannii* (Harms) Harms (41); *Paramacrolobium coeruleum* (Taub.) J. Léonard (40) (41); *Scorodophloeus zenkeri* Harms (39) (40) (41); **COM:** *Combretum oblungum* K. Hoffm. (41); *Combretodendron macrocarpum* (Beauv.) Keay (41); *Combretodendron* sp. (41); *Terminalia superba* Engl. et Diels (39) (40) (41); **EBE:** *Diospyros alboflavescens* (Gurke) F. White (41); *D. crassiflora* Hiern (41); *D. undabunda* Hiern ex Greves (41); *D. undulata* Louis (40); **EUP:** *Alchornea cordifolia* Muell. Arg. (41); *Cleistanthus* sp. (40) (41); *Croton haumannianus* J. Léonard (40) (41); *C. megalocarpus* Hutchinson (41); *Discoglypemma caloneura* Prain (40) (41); *Drypetes gossweileri* S. Moore (39) (40) (41); *D. monocarpa* J. Léonard (40) (41); *Drypetes* cfr. *louisii* J. Léonard (41); *Drypetes spinoso-dentata* (Pax) Hutch. var. *longipedunculata* J. Léonard (41); *Hymenocardia acida* Tul. (41); *Macaranga lancifolia* Pax (41); *Macaranga* cfr. *lancifolia* Pax (41); *Microdesmis puberula* Hook. f. (40) (41); *Phyllanthus discoideus* Muell. Arg. (40) (41); *Ricinodendron heudelotii* (Baill.) Pierre (40); *R. africanus* Muell. Arg. (41); *Spondianthus preussii* Engl. (40); **FAB:** *Erythrina* sp. (41); **FLA:** *Caloncoba welwitschii* Gilg (40) (41); *Paropsia schiebeniana* Sleumer (40); **GUT:** *Allanblackia floribunda* Oliver (41); *Garcinia polyantha* Oliv. (40); *G. punctata* Oliv. (40) (41); *G. smeathmanni* Oliver (41); *Garcinia* sp. (26) (32); *Mammea africana* G. Don (41); *Symphonia globulifera* L. (41); **IRV:** *Irvingia gabonensis* Baill. (41); *I. grandifolia* Engl. (40) (41); *Irvingia* sp. (41); *Klainedoxa gabonensis* Pierre (40); **LAU:** *Beilschmiedia corbisieri* (Robyns) Robyns et Wilczek (40) (41); *B. gilbertii* Robyns et Wilczek var. *glabra* (41); **LEC:** *Napoleana imperialis* P. Beauv. (40); **LEG:** *Acacia polyacantha* Willd (41); *Albizia glabrescens* Oliver (41); *A. gummifera* (Gmal.) C. A. Sm. (40); *Albizia* sp. (39) (41); *Baphia pubescens* Hook. f. (41); *Bauhinia thonningii* Schum. (41); *Brachystegia* sp. (51); *Cathormion leptophyllum* (Harms) Keay var. *guineense* (41); *Daniellia alsteeniana* Duvign. (41); *Entadopsis abyssinica* (Steud. ex A. Rich.) Gilbert et Boutique (41); *Pentaclethra macrophylla* Benth. (40) (41); *Piptadeniastrum africanum* (Hook. f.) Brenan (41); *Pithecellobium dinklagei* Harms (40); *Tessmannia africana* Harms (41); *T. anomala* (Micheli) Harms (41); *T. parvifolia* Harms (40); *Tetrapleura tetraptera* (Thonn.) Taub. (41); **LIN:** *Ochthocosmus* cfr. *africana* Hook. f. (40); **MEL:** *Carapa procera* DC. (41); *Entandrophragma angolense* C. DC. (39) (41); *E. candollei* Harms (39) (41); *E. cylindricum* (Sprague) Sprague (39) (41); *E. utile* (Dawe et Sprague) (41); *Entandrophragma* sp. (39) (41); *Guarea cedrata* Pellegr. (39) (40) (41); *Guarea laurentii* De Wild. (39) (40) (41); *G. thompsonii* Sprague et Hutch. (39) (41); *Khaya anthotheca* C. DC. (41); *Melia* sp. (39); *Trichilia gilgiana* Harms (40); *T. heudelotii* Planch. (40); *T. prieureana* Juss. (41); *T. retusa* Oliver (41); *T. rubescens* Oliv. (40) (41); *T. welwitschii* C. DC (41); *T. cfr. lanata* A. Chev. (40); *Turraeanthus* sp. (41); **MOR:** *Antiaris welwitschii* Engl. (39) (41); *Bosqueia angolensis* Ficalho (40) (41); *Chlorophora excelsa* Benth. et Hook (39) (40) (41); *Ficus congensis* Engl. (41); *F. persicifolia* Welw. ex Warb. (41); *Musanga cecropioides* R. Br. (41); *Myrianthus arboreus* P. Beauv. (40) (41); *Myrianthus* sp. (41); *Treculia africana* Decne (40) (41); **MYRI:** *Coelocaryon preussii* Warb. (40) (41); *Pycnanthus angolensis* (Welw.) Exell. (39) (40) (41); *Staudtia stipitata* Warb. (39) (40) (41); **MYRT:** *Syzygium congolense* Vermoesen (40) (41); **OKT:** *Okoubaka aubrevillei* Pellegr. et Normand (41); **OLA:** *Ongokea gore* (Hua) Pierre (39) (41); *Strombosia glaucescens* Engl. (40) (41); *S. scheffleri* Engl. (40); *S. grandifolia* Hook. f. Benth. (41); *Strombosiaopsis tetrandra* Engl. (40) (41); **PAN:** *Panda oleosa* Pierre (40) (41); **PAP:** *Afrormosia elata* Harms (39) (40) (41); *Angylocalyx pynaertii* De Wild. (40) (41); *Millettia drastica* Welw. (41); *Millettia* cfr. *drastica* Welw. (41); *Pterocarpus soyauxii* Taub. (40) (41); *P. tinctorius* Welw. (40); **PAS:** *Barteria nigritiana* Hook. f. (40) (41); **RHA:** *Maesopsis eminii* Engl. (41); **ROS:** *Parinarium holstii* Engl. (40) (41); *P. mobola* Oliver (41); *Parinarium* sp. (28); **RUB:** *Canthium dewevrei* De Wild. (40); *Massularia acuminata* (G. Don) Bullock ex Hayle (41); *Mitragyna stipulosa* Kuntze (41); *Mitragyna* sp. (40); *Morinda lucida* Benth. (41); *Morinda* cfr. *lucida* Benth. (40); *Nauclea diderrichii* (De Wild.) Merrill (39) (41); *Randia oktomera* Benth. (40); *Rothmannia octomera* (Benth et Hook. f.) Fagerl. (41); *Sarcocephalus* cfr. *trillesii* Pierre (40); **RUT:** *Fagara inaequalis* Engl. (41); *F. lemairei* De Wild. (40) (41); *F. macrophylla* (Oliv.) Engl. (40); **SAM:** *Homa-*

lium sp. (40); **SAPI**: *Afania senegalensis* Radlk. (41); *Allophylus africanus* Beauv. (41); *Autranella congolensis* A. Cheval. et De Wild (41); *Blighia welwitschii* (Hiern) Radlk. (40) (41); *Deinbollia* sp. (41); *Eriocoelum microspermum* Radlk. ex De Wild (41); *Pancovia harmsiana* Gilg. (40) (41); *P. laurentii* De Wild. (40) (41); **SAPO**: *Autranella* sp. (41); *Chrysophyllum africanum* A. DC. (40) (41); *C. lacourtianum* De Wild. (40) (41); *C. pruniforme* Engl. (40) (41); *Chrysophyllum* sp. (41); *Manilkara maleoleus* J. Louis (40); *Omphalocarpum* cfr. *mortehani* De Wild. (40) (41); *Omphalocarpum* sp. (40) (41); *Pachystela excelsa* Baill. (41); *Synsepalum dulcificum* (Sch. et Th.) Daniell et Bell. (41); *S. subcordatum* De Wild. (40) (41); *Tridesmostemon claessensii* De Wild. (40) (41); **SIM**: *Hannoa klaineana* Pierre (40) (41); *Klainedoxa gabonensis* Pierre var. *oblongifolia* (41); **SOL**: *Nicotiana tabacum* L. (41); **STE**: *Cola griseiflora* De Wild. (40) (41); *C. lateritia* K. Schum. (41); *Cola* cfr. *lateritia* K. Schum. (40); *Pterygota bequaertii* De Wild. (41); *Sterculia bequaertii* De Wild. (40) (41); *S. quinqueloba* K. Schum. (40) (41); **THE**: *Nesogordonia* cfr. *leplaei* (Vermoesen) Capuron (41); **TIL**: *Desplatsia deweyrei* (De Wild. et Th. Dur.) (40) (41); *Microcos pinnatifida* (Mast.) Burret (41); **ULM**: *Holoptelea grandis* (Hutch.) Mildbr. (40); *Celtis briei* De Wild. (40) (41); *C. mildbraedii* Engl. (39) (41); **VER**: *Vitex congolensis* De Wild. et Dur. (40) (41); **VIO**: *Rinorea brachypetala* Kuntze (41); *R. multinervis* M. Brandt (41); *R. welwitschii* Engl. (41).

Bolbocranius mechowii (Kolbe)

ANN: *Polyanthia suaveolens* Engl. et Diels (39) (40) (41); *Xylopia gilbertii* Boutique (41); **CAE**: *Cynometra hankei* Harms (39); *Dialium excelsum* Louis ex Steyaert (41); *Oxystigma oxyphyllum* (Harms) J. Léonard (41); *Pachyelasma tessmanii* (Harms) Harms (41); *Scorodophloeus zenkeri* Harms (39) (40) (41); **COM**: *Combretodendron macrocarpum* (Beauv.) Keay (41); **EUP**: *Cleistanthus* sp. (41); *Croton haumannianus* J. Léonard (40); **GUT**: *Garcinia punctata* Oliv. (40) (41); **LEG**: *Albizia* sp. (41); **MEL**: *Entandrophragma candollei* Harms (39) (41); *Entandrophragma* sp. (39); *Myrianthus arboreus* P. Beauv. (41); **MYRI**: *Pycnanthus angolensis* (Welw.) Exell (41); *Staudtia stipitata* Warb. (39) (41); **OLA**: *Strombosia grandifolia* Hook. f. Benth. (41); *Strombosiosis tetrandra* Engl. (40) (41); **PAP**: *Angylocalyx pynaertii* De Wild. (41); **SAPI**: *Blighia welwitschii* (Hiern) Radlk. (40) (41); *Pancovia harmsiana* Gilg. (41); **SAPO**: *Autranella* sp. (41); *Chrysophyllum africanum* DC. (41); *C. lacourtianum* De Wild. (41); **SIM**: *Hannoa klaineana* Pierre et Engl. (40) (41); *Klainedoxa gabonensis* Pierre var. *oblongifolia* (41); **STE**: *Cola griseiflora* De Wild. (40) (41).

Bolbocranius opacus Kolbe

CAE: *Scorodophloeus zenkeri* Harms (39) (41); *Pachyelasma tessmanii* (Harms) Harms (40); **EUP**: *Drypetes gossweileri* S. Moore (39); *Sapium* sp. (26); **MEL**: *Entandrophragma candollei* Harms (39); *Guarea cedrata* (A. Chev.) Pellegr. (39); *G. laurentii* De Wild. (39); *Lovoa trichilioides* Harms (39); **RUB**: *Mitragyna stipulosa* Kuntze (41); *Nauclea diderrichii* (De Wild.) Merrill (39); **MYRI**: *Pycnanthus angolensis* (Welw.) Exell (39).

Bolbocranius rostralis (Damoiseau)

CAE: *Erythrophleum guineense* G. Don (41); **LEG**: *Tetrapleura tetraptera* (Thonn.) Taub. (41).

Bolbocranius truncatus (Damoiseau)

COM: *Combretodendron macrocarpum* (Beauv.) Keay (41).

Taphroderes nevermanni Kleine

LEG: *Inga edulis* Mart. (19) (23).

Taphroderes sahlbergi (Sharp)

ARAU: *Araucaria angustifolia* (Bertol.) Kuntze (64).

TRACHELIZINAE Lacordaire

TRACHELIZINI Lacordaire

Euschizus internatus (Pascoe)

ARAU: *Araucaria cunninghamii* A. Cunn. (58).

Hephebocerus nanus (Boheman)

LEG: *Inga* sp. (11).

Hypomiolispa compressa Kleine

HAM: *Liquidambar* sp. (63); **MOR:** *Trema* sp. (63).

Hypomiolispa incerta Kleine

RUB: *Anthocephalus cadamba* Miq. (31) (32).

Hypomiolispa tonkinensis Kleine

FAG: *Quercus* sp. (63).

Hypomiolispa trachelizoides (Senna)

APO: *Dyera* sp. (17).

Miolispoides sculpturatus Calabresi

FAG: *Castanopsis* sp. (63).

Paratrachelizus agnatus Kleine

MOR: *Castilloa costaricana* Liedm. (19) (23).

Paratrachelizus sp. prope *linearis* (Suffrian)

CYP: *Scirpus validus* Wahl. (30).

Paratrachelizus prolixus (Sharp)

TIL: *Luehea seemannii* Planch. et Triana (19) (23).

Paratrachelizus uncimanus (Boheman)

AST: *Flaveria linearis* Lag. (61); **CUP:** *Thuja* sp. (61); **SOL:** *Solanum* sp. (62).

Trachelizus bisulcatus (Lund)

ANA *Buchanania latifolia* Roxb. (32); **BOM:** *Bombax malabaricum* DC. (12) (13) (23) (32); *Kydia calycina* Roxb. (32); **CAC:** *Opuntia* sp. (27) (32); **CAE:** *Poinciana elata* L. (13) (23) (32); **COM:** *Anogeissus latifolia* Wall. (32); **CUC:** *Hodgsonia heteroclita* Hook. f. et Thoms. (32); **DAT:** *Tetrameles nudiflora* R. Br. (32); **EUP:** *Euphorbia royleana* Boiss. (32); **FAB:** *Butea frondosa*

Roxb. (12) (13) (23) (32); *Erythrina indica* Lam. (13) (23) (32); **LEG:** *Albizia* sp. (27) (32); **MOR:** *Ficus asperrima* Roxb. (12) (13) (32); *F. benghalensis* L. (32); *F. glomerata* Roxb. (13) (23) (32); *F. religiosa* L. (12) (13) (32); *F. rumphii* Blume (32); **RUB:** *Anthocephalus cadamba* Miq. (32); **STE:** *Sterculia colorata* Roxb. (20) (23) (32); **VER:** *Tectonia grandis* L. (27) (32).

Trachelizus laevigatus Senna

FAG: *Castanopsis* sp. (63); *Pasania* sp. (63).

Trachelizus politus Senna

RUB: *Anthocephalus cadamba* Miq. (32).

Tracheloschizus cameratus Lacordaire

APO: *Dyera costulata* Hook. f. (32); *Dyera* sp. (17).

Tracheloschizus multipunctatus Damoiseau

APO: *Alstonia brassii* Monachino (54).

ACRATINI Alonso-Zarazaga, Lyal, Bartolozzi & Sforzi

Acratus apicalis Sharp

MOR: *Ficus* sp. (19) (23).

Nemocephalus puncticeps (Sharp)

MOR: *Ficus* sp. (23).

ITHYSTENINI Lacordaire

Ceocephalus antennatus (Ritsema)

EBE: *Diospyros glandulosa* Lace (32).

Ceocephalus articulatus (Senna)

FAG: *Pasania* sp. (63).

Lasiorhynchus barbicornis (Fabricius)

ARAU: *Agathis australis* Steud. (1) (46) (61) (66); **AST:** *Brachyglottis repanda* Forst. (66); *Olearia rani* Druce (56); **BOM:** *Hoheria* sp. (25) (46); **CORY:** *Corynocarpus laevigata* Forst. (2) (46) (52) (56) (61) (66); **LAU:** *Beilschmiedia tarairi* Kirk (66); *B. tawa* Kirk (46) (61) (66); **MAL:** *Hoheria* sp. (66); **MEL:** *Dysoxylum spectabile* Hook f. (66); **MON:** *Hedycarya arborea* Forst. (25) (46) (61) (66); *Hedycarya* sp. (56); *Laurelia novae-zelandiae* A. Cunn. (46) (66); **POD:** *Dacrydium cupressinum* Soland. ex Forst. f. (15) (46) (66); *Prumnopitys taxifolia* (Solander ex D. Don) D. J. de Laubenfels (56); **PRO:** *Knightia excelsa* R. Br. (15) (46) (61) (66).

Mesetia amoena Blackburn

MYRT: *Leptospermum* sp. (58).

MICROTRACHELIZINI Zimmerman

Aneorhachis astricta Kleine

FAB: *Erythrina indica* Lam. (20) (23) (32).

Araiorrhinus beesoni Kleine

VER: *Tectonia grandis* L. (13) (23) (24) (32).

Entomopisthius emarginatus (K. E. Schedl)

ANN: *Anonidium mannii* Engl. et Diels (41); *Polyanthia suaveolens* Engl. et Diels (41); **COM:** *Combretodendron macrocarpum* (Beauv.) Keay (41); **EBE:** *Diospyros crassiflora* Hiern (41); **EUP:** *Drypetes gossweileri* S. Moore (41); *Macaranga denticulata* Muell. Arg. (41); *M. lancifolia* Pax (41); *Macaranga* cfr. *lancifolia* Pax (40) (41).

Entomopisthius suturalis Damoiseau

ANN: *Xylopia gilbertii* Boutique (41); **CAE:** *Scorodophloeus zenkeri* Harms. (41).

Higonius cilo Lewis

COM: *Terminalia* sp. (63); **EUP:** *Mallotus roxburghianus* Muell. Arg. (20) (32); **FAG:** *Castanopsis tribuloides* A. DC (13) (32); *Millettia auriculata* Baker. (13) (32); **MEL:** *Amoora wallichii* King. (32); **STE:** *Sterculia alata* Roxb. (20).

Higonius crux Oliff

COM: *Terminalia myriocarpa* Heurck et Muell. Arg. (32); **DAT:** *Tetrameles nudiflora* R. Br. (32); **DIP:** *Shorea robusta* Gaertn. f. (13) (23) (32); **EBE:** *Diospyros pyrrhocarpa* Miq. (32); **STE:** *Sterculia alata* Roxb. (23) (32); **VER:** *Tectonia grandis* L. (12) (13) (23) (32).

Higonius poweri Lewis

COM: *Terminalia* sp. (63); **MOR:** *Ficus* sp. (63).

Microtrachelizus accomodatus Kleine

DIP: *Shorea assamica* Dyer. (12) (13) (23) (32); *Vateria indica* L. (13) (23) (32).

Microtrachelizus aethiopicus Calabresi

ANA: *Antrocaryon nannanii* De Wild. (40) (41); *Sorindeia sparanoi* De Wild. (41); **ANN:** *Anonidium mannii* Engl. et Diels (40) (41); *Polyanthia suaveolens* Engl. et Diels (39) (40) (41); *Xylopia aethiopica* (Dun) A. Rich. (40) (41); *X. gilbertii* Boutique (41); *X. villosa* Chipp. (40); **APO:** *Alstonia boonei* De Wild (39) (41); *A. congoensis* Engl. (40); *Conopharyngia durissima* Stapf (40) (41); *Funtumia latifolia* Stapf (41); *Voacanga obtusa* K. Schum. (42); **BAR:** *Combretodendron macrophyllum* (P. Beauv.) Keay. (40); **BOM:** *Triplochiton scleroxylon* K. Schum. (44); **BUR:** *Dacryodes yangambiensis* J. Louis ex Troupin (40) (41); **CAE:** *Amphimas pterocarpoides* Harms (40) (41); *Anthonotha macrophyllum* P. Beauv. (40) (41); *Cynometra hankei* Harms (39); *Dialium corbisieri* Staner (41); *D. pachyphyllum* Harms (40) (41); *D. tessmannii* Harms (40); *Dalium* sp. (41); *Erythrophleum guineense* G. Don (39) (40) (41); *Oxystigma oxyphyllum* (Harms) J. Léonard (40) (41);

Pachyelasma tessmannii (Harms) (40) (41); *Paramacrolobium coeruleum* (Taub.) J. Léonard (40); *Scorodophloeus zenkeri* Harms. (39) (40) (41); **COM:** *Combretodendron macrocarpum* (Beauv.) Keay (41); *Combretum oblungum* K. Hoffm. (40) (41); *C. splendens* Engl. (26); *Terminalia superba* Engl. et Diels (39) (40) (41); **EBE:** *Diospyros alboflavescens* (Gurke) F. White (41); *Maba laurentii* Engl. et Diels (40); **EUP:** *Cleistanthus* sp. (40) (41); *Drypetes* cfr. *louisii* J. Léonard (40) (41); *Drypetes monocarpa* J. Léonard (40) (41); *Drypetes spinoso-dentata* (Pax) Hutch. var. *longipedunculata* J. Léonard (40) (41); *Drypetes* sp. (41); *Macaranga* cfr. *lancifolia* Pax (41); *Microdesmis puberula* Hook. f. (40) (41); **GUT:** *Garcinia polyantha* Oliv. (40); *G. punctata* Oliv. (40) (41); *G. smeathmanni* Oliver (41); **IRV:** *Irvingia* sp. (40) (41); *Klainedoxa gabonensis* Pierre (40); **LAU:** *Beilschmiedia gilbertii* Robyns et Wilczek var. *glabra* (40) (41); **LEG:** *Albizia gummifera* (Gmel.) C. A. Sm. (40); *Albizia* sp. (39) (41); *Baphia pubescens* Hook. f. (41); *Tetrapleura tetraptera* (Thonn.) Taub. (40) (41); **MEL:** *Carapa procera* DC. (40) (41); *Guarea cedrata* Pellegrin (39) (40) (41); *G. laurentii* De Wild. (39) (40) (41); *G. thompsonii* Sprague et Hutch. (39) (40) (41); *Trichilia lanata* A. Chev. (41); *T. prieureana* Juss. (40) (41); *T. welwitschii* C. DC. (41); **MOR:** *Chlorophora excelsa* Benth. et Hook f. (39) (40) (41); *Myrianthus arboreus* P. Beauv. (40); **MYRI:** *Pycnanthus angolensis* (Welw.) Exell. (39) (40) (41); *Staudtia stipitata* Warb. (39) (41); **OKT:** *Okoubaka aubrevillei* Pellegr. et Normand (41); *O. aubrevillei* var. *glabra* J. Léonard M. S. S. (40); **OLA:** *Ongokea gore* (Hua) Pierre (39) (40) (41); *Strombosia glaucescens* Engl. (40) (41); *S. grandifolia* Hook. f. Benth. (41); *S. scheffleri* Engl. (40); *Strombosiaopsis tetrandra* Engl. (40) (41); **PAN:** *Panda oleosa* Pierre (40) (41); **PAP:** *Afrormosia elata* Harms (39) (40) (41); *Angylocalyx pynaertii* De Wild. (40) (41); *Baphia soldheidii* De Wild. (40); *Millettia* cfr. *drastica* Welw. (40) (41); *Pterocarpus soyauxii* Taub. (40) (41); **PAS:** *Barteria nigrifolia* Hook f. (40) (41); **ROS:** *Pariarium curatellaefolium* Planch. ex Benth. (26); **RUB:** *Morinda lucida* Benth. (41); *Plectronia dewevrei* Wildem. (41); **RUT:** *Fagara lemairei* De Wild. (40); *Vepris louisii* Gilbert (41); *Vespris* sp. (40); **SAPI:** *Blighia welwitschii* (Hiern) Radlk. (40) (41); *Lecaniodiscus cupanioides* Planch. (41); *Pancovia harmsiana* Gilg. (40) (41); *Radlkofera calodendron* Gilg. (40) (41); **SAPO:** *Chrysophyllum africanum* A. DC. (40) (41); *C. lacourtianum* De Wild. (40) (41); *Manilkara malcoleus* J. Louis (41); *Pachystela bequaertii* De Wild. (40) (41); *Synsepalum dulcificum* (Sch. et Th.) Daniell et Bell. (40) (41); *S. subcordatum* De Wild. (40) (41); *Tridesmostemon claessensii* De Wild. (40) (41); **SIM:** *Hannoa klaineana* Pierre (40) (41); *Klainedoxa gabonensis* Pierre var. *oblongifolia* (41); **STE:** *Cola griseiflora* De Wild. (40) (41); *Cola* cfr. *lateritia* K. Schum. (40); *Nesogordonia* sp. (41); *Sterculia bequaertii* De Wild. (40) (41); **STY:** *Afrostryax lepidophyllus* (Harms) Mildbr. (40) (41); **TIL:** *Cistanthera* sp. (40); *Desplatsia dewevrei* (De Wild. et Th. Dur.) (41); *Microcos pinnatifida* (Mast.) Burret (41); **ULM:** *Celtis briei* De Wild. (40) (41); *C. mildbraedii* Engl. (39) (40) (41); **VER:** *Vitex welwitschii* Guerke (41); **VIO:** *Rinorea multinervis* M. Brandt (41); *R. welwitschii* Engl. (40) (41).

Microtrachelizus apertus Kleine

BOM: *Bombax malabaricum* DC. (32); **BOR:** *Ehretia acuminata* Br. (13) (23) (32); **CLU:** *Mesua ferrea* L. (12) (13) (23) (32); **COM:** *Terminalia* sp. (63); **EUP:** *Ostodes paniculata* Blume (32); **FAB:** *Dalbergia assamica* Benth. (12) (13) (23) (32); **MOR:** *Ficus* sp. (63); **STE:** *Sterculia villosa* Roxb. (23) (32); *Sterculia* sp. (63).

Microtrachelizus attritus Kleine

ANN: *Milusa velutina* Hook. f. et Thoms. (20) (23) (32); **FAB:** *Erythrina indica* Lam. (20) (23) (32).

Microtrachelizus beneficus Kleine

DIP: *Shorea assamica* Dyer. (12) (13) (14) (23) (32) (38) (58); *S. robusta* Gaertn. F. (20) (23) (32); **LAU:** *Litsea citrata* Blume (27) (32).

Microtrachelizus contiguus (Senna)

ANA: *Calesium grande* Kuntze (23) (32); *Odina wodier* Roxb. (20); **COM:** *Terminalia bellerica* Roxb. (32).

Microtrachelizus imbecillus Kleine

STY: *Afrostryrax lepidophyllus* (Harms) Mildbr. (41).

Microtrachelizus siamensis Kleine

EUP: *Elateriospermum* sp. (32).

Microtrachelizus tabaci Senna

DIP: *Shorea bracteolata* Dyer (23) (32).

Microtrachelizus sp.

ARAU: *Araucaria cunninghamii* A. Cunn. (58).

Parapisthius gracilis Damoiseau

EUP: *Mallotus* sp. (63).

PSEUDOCEOCEPHALINI Kleine

Eubactus sp.

PIN: *Pinus longifolia* Roxb. (10).

Eumecopodus longicornis Calabresi

ANN: *Anonidium mannii* Engl. et Diels (41); **SAPO:** *Chrysophyllum africanum* A. D.C. (41).

Gynandrorhynchus audax (Kleine)

ANN: *Polyanthia suaveolens* Engl. et Diels (41); **CAE:** *Anthonotha macrophyllum* P. Beauv. (41); *Erythrophleum guineense* G. Don (41); **OLA:** *Strombosia glaucescens* Engl. (41); **SAPO:** *Chrysophyllum lacourtianum* De Wild. (41).

Gynandrorhynchus cfr. *granulatus* (Kleine)

SAPI: *Pancovia harmsiana* Gilg. (41).

Gynandrorhynchus vittipennis (Fåhræus)

BOM: *Sida acuta* Burm.f. (42); **IRV:** *Oxystigma oxyphyllum* (Harms) J. Léonard (41); **LEG:** *Cathormion leptophyllum* (Harms) Keay var. *guineense* (41); **MEL:** *Carapa procera* DC. (41); **MOR:** *Bosqueia angolensis* (Welw.) Fic. (41); *Myrianthus arboreus* P. Beauv. (41); **RUT:** *Fagara lemairei* De Wild. (41); **SAPO:** *Omphalocarpum* sp. (41).

Heterothesis elegans Kleine

CAE: *Scorodophloeus zenkeri* Harms (39) (41); **IRV:** *Oxystigma oxyphyllum* (Harms) J. Léonard (41); **MEL:** *Guarea cedrata* (A. Chev.) Pellegr. (41).

Hormocerus reticulatus (Lund)

BOM: *Bombax malabaricum* DC. (10) (12) (13) (23) (32); **DAT:** *Tetrameles nudiflora* R. Br. (32); **FAG:** *Castanopsis tribuloides* A. DC. (10) (13) (23) (32); **MOR:** *Artocarpus integrifolia* L. f. (32); *Ficus infectoria* Roxb. (32); *Ficus* sp. (63); **RUB:** *Hymenodictyon excelsum* Wall. (32).

Isoceocephalus rufescens Quentin

ANN: *Xylopi* *gilbertii* Boutique (41); *X. villosa* Chipp. (40); **APO:** *Conopharyngia durissima* Stapf. (41); **CAE:** *Oxystigma oxyphyllum* (Harms) J. Léonard (40); *Scorodophloeus zenkeri* Harms (39); **EUP:** *Cleistanthus* sp. (41); *Drypetes* sp. (41); *Ricinodendron heudelotii* (Baill.) Pierre (40); **LEG:** *Newtonia buchananii* (Baker) Gilbert et Boutique (41); **MEL:** *Carapa procera* DC. (40); *Etandrophragma* sp. (41); *Guarea cedrata* Pellegrin (39); **MOR:** *Antiaris welwitschii* Engl. (41); *Antiaris* sp. (41); **MYRI:** *Staudtia stipitata* Warb. (40); **MYRT:** *Syzygium* sp. (41); **RUB:** *Mitragyna* sp. (40); **RUT:** *Fagara lemairei* De Wild. (40); **SAPO:** *Chrysophyllum africanum* DC. (41); *Manilkara malcoleus* J. Louis (40).

Orphanobrentus laevicollis (J. Thomson)

ANN: *Annona arenaria* Thonn (= *senegalensis* Pers.) (41); *Polyalthia suaveolens* Engl. et Diels (41); *Xylopi* *villosa* Chipp. (40); **APO:** *Conopharyngia* sp. (41); **CAE:** *Scorodophloeus zenkeri* Harms (41); **CEL:** *Maytenus senegalensis* (Lam.) Exell (41); **COM:** *Combretodendron* sp. (41); *Terminalia glandulosa* De Wild. (41); *T. superba* Engl. et Diels (41); **EUP:** *Alchornea cordifolia* Muell. Arg. (41); *Bridelia ferruginea* Benth. (41); *Hymenocardia acida* Tul. (41); **LEG:** *Acacia polyacantha* Willd. (41); *Albizia glabrescens* Oliver (41); *Albizia* sp. (41); *Pentaclethra macrophylla* Benth. (41); **LIL:** *Dracaena* sp. (41); **MEL:** *Entandrophragma utile* (Dawe et Sprague) (41); *Entandrophragma* sp. (41); *Guarea cedrata* Pellegrin (41); **MOR:** *Chlorophora excelsa* Benth. et Hook f. (39) (40) (41); *Ficus mucoso* Welw. (41); *F. persicifolia* Welw. ex Warb. (41); *Myrianthus* sp. (41); **OLA:** *Strombosia glaucescens* Engl. (41); **RUB:** *Crossopteryx febrifuga* Benth. (41); *Gaertnera* sp. (41); *Mitragyna stipulosa* Kuntze (41); *Mitragyna* sp. (40); **SOL:** *Nicotiana tabacum* L. (41); **SAPO:** *Autranella* sp. (41); **STE:** *Pterygota mildbraedii* Engl. (41); *Sterculia quinqueloba* (Gurke) K. Schum. (40) (41); **VER:** *Vitex* sp. (41).

Orphanobrentus picipes (Olivier)

ANA: *Sclerocarya birrea* Hochst. (41); **ANN:** *Annona arenaria* Thonn (= *senegalensis* Pers.) (41); *Anonidium mannii* Engl. et Diels (40) (41); *Polyalthia suaveolens* Engl. et Diels (39) (40) (41); *Xylopi* *aethiopica* (Dun) A. Rich. (41); *X. gilbertii* Boutique (41); *X. phloiodora* Mildbr. (41); *X. villosa* Chipp. (40); **ANA:** *Antrocaryon nannanii* De Wild. (41); *Lannea* sp. (40) (41); *Spondianthus preussii* Engl. (42); **APO:** *Alstonia boonei* De Wild. (41); *Conopharyngia durissima* Stapf. (40) (41); *Conopharyngia* sp. (41); *Pleiocarpa micrantha* Stapf (41); *Rauwolfia vomitoria* Afzel. (42); *Voacanga obtusa* K.Schum. (42); **BIG:** *Stereospermum* sp. (41); **BOM:** *Ceiba pentandra* (L.) Gaertn. (41); *Gossipium* sp. (28); **BUR:** *Canarium schweinfurthii* Engl. (40) (41); *Dacryodes edulis* (G. Don) H. J. Lam. (41); *D. yangambiensis* J. Louis ex Troupin (40) (41); **CAE:** *Azalia bella* Harms (40) (41); *Amphimas pterocarpoides* Harms (40) (41); *Anthonotha macrophyllum* P. Beauv. (40) (41); *Cynometra hankei* Harms (39) (40) (41); *Dialium corbisieri* Staner (41); *D. pachyphyllum* Harms (40) (41); *Erythrophleum guineense* G. Don (39) (40) (41); *Gossweilerodendron balsamiferum* (Harms) (39) (40) (41); *Oxystigma oxyphyllum* (Harms) J. Léonard (40) (41); *Paramacrolobium coeruleum* (Taub.) J. Léonard (41); *Pachylobus edulis* Don. (40); *Scorodophloeus zenkeri* Harms. (39) (40) (41); **CEL:** *Maytenus senegalensis* (Lam.) Exell (41); **COM:** *Combretodendron*

macrocarpum (Beauv.) Keay (41); *Combretum oblungum* K. Hoffm. (40) (41); *Terminalia glandulosa* De Wild. (41); *T. superba* Engl. et Diels (40) (41); *Terminalia* sp. (40); **CUP:** *Cupressus lusitanica* Carr. (41); **EBE:** *Diospyros alboflavescens* (Gurke) F. White (41); *D. undabunda* Hiern ex Greves (41); *Maba laurentii* Engl. et Diels (40); **EUP:** *Alchornea cordifolia* Muell. Arg. (41); *Antidesma meiocarpum* J. Léonard (41); *Bridelia ferruginea* Benth. (40) (41); *Cleistanthus* sp. (41); *Croton haumanianus* J. Léonard (40) (41); *C. megalocarpus* Hutchinson (41); *Discoglypsemna caloneura* Prain (41); *Drypetes gossweileri* S. Moore (39) (40) (41); *D. morocarpa* J. Léonard (41); *Drypetes* cfr. *louisii* J. Léonard (41); *Hymenocardia acida* Tul. (41); *H. ulmoides* Oliv. (41); *Macaranga lancifolia* Pax (41); *Microdesmis puberula* Hook. f. (40) (41); *Neoboutonia* sp. (42); *Phyllanthus discoideus* Muell. Arg. (40) (41); *Ricinodendron heudelotii* (Baill.) Pierre (40); *R. africanus* Muell. Arg. (41); **FLA:** *Caloncoba welwitschii* Gilg (41); **GUT:** *Garcinia punctata* Oliv. (41); **IRV:** *Irvingia grandifolia* Engl. (40); *Irvingia* sp. (41); *Klainedoxa gabonensis* Pierre (40); **LAU:** *Beilschmiedia corbisieri* (Robyns) Robyns et Wilczek (40) (41); **LEG:** *Albizia glabrescens* Oliver (41); *A. gummifera* (Gmel.) C. A. Sm. (40); *Albizia* sp. (39) (40) (41); *Bauhinia thonningii* Schum. (41); *Cathormion leptophyllum* (Harms) Keay var. *guineense* (41); *Daniellia alsteeniana* Duvign. (41); *Entadopsis abyssinica* (Steud. ex A. Rich.) Gilbert et Boutique (41); *Myroxylon pereirae* Klotzsch (28); *Newtonia buchananii* (Baker) Gilbert et Boutique (40); *Newtonia* sp. (41); *Piptadenia* sp. (28); *Pithecellobium dinklagei* Harms (40); *Tetrapleura tetraptera* (Thonn.) Taub. (40) (41); **LIN:** *Ochthocosmus* cfr. *africana* Hook. f. (41); **MEL:** *Carapa procera* DC. (41); *Etandrophragma angolense* DC. (39) (40) (41); *E. candollei* Harms. (39) (40) (41); *E. cylindricum* Sprague (39) (40) (41); *Etandrophragma* sp. (40) (41); *Guarea cedrata* Pellegrin (39) (40) (41); *G. laurentii* De Wild. (39) (40) (41); *G. thompsonii* Sprague et Hutch. (41); *Khaya anthotheca* C. DC. (41); *Khaya* sp. (40); *Lovoa trichilioides* Harms (41); *Melia* sp. (41); *Trichilia rubescens* Oliv. (40) (41); **MOR:** *Antiaris* sp. (41); *Bosqueia angolensis* (Welw.) (40) (41); *Chlorophora excelsa* Benth. et Hook f. (39) (40) (41); *Ficus congensis* Engl. (41); *F. mucoso* Welw. (40) (41); *F. persicifolia* Welw. ex Warb. (41); *Ficus* sp. (40) (41); *Musanga cecropioides* R. Br. (40) (41); *Myrianthus arboreus* P. Beauv. (40) (41); *Myrianthus* sp. (41); **MYRI:** *Coelocaryon preusii* Warb. (40) (41); *Pycnanthus angolensis* (Welw.) Exell. (39) (40) (41); *Staudtia stipitata* Warb. (39) (40) (41); **MYRT:** *Syzygium* sp. (41); **OKT:** *Okoubaka aubrevillei* Pellegr. et Normand (41); **OLA:** *Olea hochstetteri* Baker (41); *Ongokea gore* (Hua) Pierre (41); *Strombosia glaucescens* Engl. (40) (41); *S. grandifolia* Hook. f. Benth. (41); *Strombosiopsis tetrandra* Engl. (40) (41); **PAN:** *Panda oleosa* Pierre (40) (41); **PAP:** *Afrormosia elata* Harms (39) (40) (41); *Angylocalyx pynaertii* De Wild. (40) (41); *Pterocarpus soyauxii* Taub. (40) (41); **PAS:** *Barteria fistulosa* Mast. (41); **ROS:** *Parinarium curatellaefolium* Planch. ex Benth. (42); *P. holstii* Engl. (40) (41); *P. mobola* Oliver (41); *Parinarium* sp. (41); **RUB:** *Mitragyna stipulosa* Kuntze (42); *Mitragyna* sp. (40) (42); *Morinda lucida* Benth. (41); *Morinda* cfr. *lucida* Benth. (40); *Nauclea diderrichii* (De Wild.) Merrill (41); *Plectronia dewevrei* Wildem. (41); **RUT:** *Canthium dewevrei* De Wild. (40); *Fagara lemairei* De Wild. (41); *F. macrophylla* (Oliv.) Engl. (41); *Sarcocephalus* cfr. *trillesii* Pierre (40); **SAPI:** *Blighia welwitschii* (Hiern) Radlk. (40) (41); *Eriocoelum microspermum* Radlk. ex De Wild (41); *Eriocoelum* cfr. *microspermum* Radlk. (40); *Lecaniodiscus cupanioides* Planch. (41); *Pancovia harmsiana* Gilg. (41); **SAPO:** *Chrysophyllum africanum* DC. (41); *C. fulvum* S. Moore (41); *C. lacourtianum* De Wild. (41); *C. pruniforme* Engl. (41); *Ompholocarpum* sp. (40) (41); *Synsepalum dulcificum* (Sch. et Th.) Daniell et Bell. (41); *S. subcordatum* De Wild. (40) (41); *Tridesmostemon claessensii* De Wild (41); **SIM:** *Hannoa klaineana* Pierre (40) (41); *Klainedoxa gabonensis* Pierre var. *oblongifolia* (41); **STE:** *Cola griseiflora* De Wild. (40) (41); *C. lateritia* K. Schum. (41); *Cola* cfr. *lateritia* K. Schum. (40); *Pterygota bequaertii* De Wild. (40); *P. mildbraedii* Engl. (41); *Sterculia bequaertii* De Wild. (40) (41); *S. quinqueloba* (Gürke) K. Schum. (40) (41); *Theobroma cacao* L. (28); **STY:** *Afrostryax lepidophyllus* (Harms) Mildbr. (40) (41); **TIL:** *Desplatsia dewevrei* (De Wild. et Th. Dur.) Burret (40) (41); **ULM:** *Celtis briei* De Wild. (40) (41); *C. mildbraedii* Engl. (39) (40) (41); *Trema orientale* Blume (40); **VER:** *Vitex* sp. (41); **VIO:** *Rinorea welwitschii* Engl. (40) (41).

Orphanobrentus punctatissimus (Kleine)

APO: *Alstonia congoensis* Engl. (40); **MOR:** *Antiaris* sp. (41).

Peraprophthalmus applicatus Kleine

MOR: *Ficus* sp. (63).

Pithoderes gestri Calabresi

MEL: *Entandrophragma utile* (Dawe et Sprague) (41); **MYRI:** *Staudtia stipitata* Warb. (41); **RUB:** *Nauclea diderrichii* (De Wild.) Merrill (41); *Sarcocephalus diderrichii* Wildem. (34) (38); **SAPO:** *Chrysophyllum lacourtianum* De Wild. (41).

Pseudocecephalus depressus (Lund)

ANA: *Sorindeia sparanoi* De Wild. (41); **ANN:** *Xylopia aethiopica* (Dun) A. Rich. (41); *Polyanthia suaveolens* Engl. et Diels (41); **APO:** *Pleiocarpa tubicina* Stapf. (41); **BIG:** *Fernandoa adolfi-fridericii* (Gilg et Mildbr.) Heine (41); **BUR:** *Canarium schweinfurthii* Engl. (41); **CAE:** *Afzelia bella* Harms (41); *Dialium excelsum* Louis ex Steyart (40) (41); *Paramacrolobium coeruleum* (Taub.) J. Léonard (40); *Scorodophloeus zenkeri* Harms. (39) (40) (41); **COM:** *Combretum oblun-gum* K. Hoffm. (40) (41); **EUP:** *Discoglypsemna caloneura* Prain (41); *Macaranga* cfr. *lancifolia* Pax (41); *Ricinodendron africanus* Muell. Arg. (41); **IRV:** *Irvingia grandifolia* Engl. (41); *Oxy-stigma oxyphyllum* (Harms) J. Léonard (41); **LAU:** *Beilschmiedia corbisieri* (Robyns) Robyns et Wilczek (41); **LEG:** *Tetrapleura tetraptera* (Thonn.) Taub. (41); **LIN:** *Ochthocosmus* cfr. *africana* Hook. f. (40); **MEL:** *Carapa procera* DC. (41); *Entandrophragma* sp. (41); **MOR:** *Antiaris wel-witschii* Engl. (41); *Antiaris* sp. (41); *Chlorophora excelsa* Benth. et Hook f. (39) (40) (41); *Myri-anthus arboreus* P. Beauv. (41); **OKT:** *Okoubaka aubrevillei* Pellegr. et Normand (41); **OLA:** *Ongokea gore* (Hua) Pierre (39) (41); **PAN:** *Panda oleosa* Pierre (41); **PAS:** *Barteria fistulosa* Mast. (41); **RUT:** *Fagara lemairei* De Wild. (40); **SAPO:** *Chrysophyllum africanum* DC. (41); **SIM:** *Hannoa klaineana* Pierre (41); **ULM:** *Celtis briei* De Wild. (41); *C. mildbraedii* Engl. (41).

Pseudocecephalus formosus Kleine

EUP: *Macaranga* cfr. *lancifolia* Pax (41); **OLA:** *Ongokea gore* (Hua) Pierre (41); **MEL:** *Entan-drophragma candollei* Harms (41); **PAP:** *Pterocarpus soyauxii* Taub. (41); **ULM:** *Celtis mildbrae-dii* Engl. (41).

Pseudomygaleicus brunneus De Muizon

ANN: *Xylopia gilbertii* Boutique (41); **CAE:** *Scorodophloeus zenkeri* Harms (41); **EUP:** *Drypetes morocarpa* J. Léonard (41).

Pseudomygaleicus raptor (Kleine)

MOR: *Ficus* sp. (26); **ROS:** *Parinarium curatellaefolium* Planch. ex Benth. (26).

Pyrethema kuntzeni Kleine

APO: *Pleiocarpa tubicina* Stapf. (40) (41); **CAE:** *Amphimas pterocarpoides* Harms (40) (41); *Scorodophloeus zenkeri* Harms. (40); **COM:** *Combretodendron macrophyllum* (P. Beauv.) Keay (40); **FLA:** *Caloncoba welwitschii* Gilg (40); **LEG:** *Cathormion leptophyllum* (Harms) Keay var. *guineense* (41); **LIN:** *Ochthocosmus* cfr. *africanus* Hook. f. (40); **MEL:** *Guarea laurentii* De Wild. (40) (41); **MOR:** *Bosqueia angolensis* (Welw.) Fic. (40); **OLA:** *Strombosia grandifolia* Hook. f. Benth. (41); **RUB:** *Canthium dewevrei* De Wild. (40); **SAPI:** *Blighia welwitshii* (Hiern) Radlk. (41); *Plectronia dewevrei* Wildem. (41).

Rhinopteryx foveipennis (J. Thomson)

ANA: *Spondianthus preussii* Engl. (41); **ANN:** *Anonidium mannii* Engl. et Diels (40) (41); *Polyanthia suaveolens* Engl. et Diels (39) (40) (41); **APO:** *Alstonia boonei* De Wild. (39) (41); *Funtumia* cfr. *latifolia* Stapf (41); *Pleiocarpa micrantha* Stapf (40) (41); *Rauwolfia vomitoria* Afzel. (42); *Voacanga obtusa* K. Schum. (42); **BAR:** *Combretodendron macrophyllum* (P. Beauv.) Keay. (40); **BIG:** *Stereospermum* sp. (41); **BUR:** *Aucoumea klaineana* Pierre (34); **CAE:** *Cynometra hankei* Harms (39) (41); *Erythrophleum guineense* G. Don (39) (41); *Gilbertiodendron dewevrei* (De Wild.) (39) (41); *Gossweilerodendron balsamiferum* (Verm.) Harms (39) (41); *Scorodophloeus zenkeri* Harms (39) (41); **COM:** *Combretodendron macrocarpum* (Beauv.) Keay (41); *Terminalia glandulosa* De Wild. (41); **EUP:** *Croton haumanianus* J. Léonard (40) (41); *Drypetes gossweileri* S. Moore (39) (40) (41); *Hymenocardia ulmoides* Oliv. (40) (41); *Macaranga* cfr. *lancifolia* Pax (41); *Neoboutonia* sp. (42); *Ricinodendron africanus* Muell. Arg. (41); *R. heudelotii* (Baill.) Pierre (40); **FLA:** *Caloncoba welwitschii* Gilg (40); **GUT:** *Garcinia punctata* Oliv. (41); **IRV:** *Oxystigma oxyphyllum* (Harms) J. Léonard (41); **LAU:** *Beilschmiedia corbisieri* (Robyns) Robyns et Wilczek (40); **LEG:** *Acacia polyacantha* Willd. (41); *Albizia welwitschii* Olivier (26); *Cathormion leptophyllum* (Harms) Keay var. *guineense* (41); *Daniellia alsteeniana* Duvign. (41); *Pithecellobium dinklagei* Harms (40); *Tessmannia anomala* (Micheli) Harms (41); **LIL:** *Dracaena* sp. (41); **LIN:** *Ochthocosmus* cfr. *africana* Hook. f. (41); **MEL:** *Entandrophragma candollei* Harms (39) (40) (41); *Entandrophragma* sp. (41); *Guarea laurentii* De Wild. (39) (41); **MOR:** *Antiaris welwitschii* Engl. (39) (41); *Antiaris* sp. (41); *Chlorophora excelsa* Benth. et Hook (41); *Ficus mucoso* Welw. ex Ficalho (41); *Myrianthus* sp. (41); **MYRI:** *Staudtia stipitata* Warb. (39) (40) (41); **OLA:** *Ongokea gore* (Hua) Pierre (39); *Strombosiopsis tetrandra* Engl. (41); **PAN:** *Angylocalyx pynaertii* De Wild. (41); *Panda oleosa* Pierre (41); **PAP:** *Pterocarpus soyauxii* Taub. (41); **RHA:** *Maesopsis eminii* Engl. (41); **RUB:** *Canthium dewevrei* De Wild. (40); **RUB:** *Coffea* sp. (41); *Mitragyna stipulosa* Kuntze (41) (42); *Mitragyna* sp. (42); **SAPI:** *Pancovia harmsiana* Gilg. (41); *Plectronia dewevrei* Wildem. (41); **SAPO:** *Chrysophyllum africanum* DC. (41); *C. lacourtianum* De Wild. (40) (41); *Synsepalum subcordatum* De Wild. (41); *Tridesmostemon claessensii* De Wild (41); **SIM:** *Hannoa klaineana* Pierre (41); **STE:** *Cola griseiflora* De Wild. (40) (41); *Sterculia bequaertii* De Wild. (40) (41); ; *S. quinqueloba* (Gurke) K. Schum. (41); **TIL:** *Desplatsia dewevrei* (De Wild. et Th. Dur.) (41); **ULM:** *Celtis mildbraedii* Engl. (40).

Sennaiella comparabilis (Kleine)

DIP: *Dryobalanops aromatica* Gaertn. f. (32).

Schizotrachelus cavus (Walker)

ANA: *Buchaniana latifolia* Roxb. (32); *Calesium grande* Kuntze (23) (32); *Lannea grandis* Engl. (32); *Mangifera indica* L. (32); *Odina wodier* Roxb. (13); *Semecarpus anacardium* L. (32); **BOM:** *Kydia calycina* Roxb. (13) (23) (32); **COM:** *Terminalia chebula* Retz. (13) (23) (32); *T. tomentosa* W. et A. (13) (23) (32); **DIP:** *Shorea robusta* Gaertn. f. (13) (23) (32); **MOR:** *Artocarpus integrifolia* L. f. (32); **RUB:** *Anthocephalus cadamba* Miq. (32).

Schizotrachelus fasciatus (Kleine)

COM: *Terminalia bellerica* Roxb. (13) (23) (32); *T. chebula* Retz. (12) (13) (23) (32); *T. tomentosa* W. et A. (10) [see Beeson, 1925: 184]; **FAB:** *Butea frondosa* Roxb. (13) (23) (32); *Castanopsis* sp. (63).

Schizotrachelus intermedius Senna

MOR: *Ficus* sp. (63).

Schizotrachelus madens Lacordaire

MOR: *Artocarpus lakoocha* Roxb. (23) (32).

Storeosomus riisii (Labram & Imhoff)

ANA: *Antrocaryon nannanii* De Wild. (40); **ANN:** *Anonidium mannii* Engl. et Diels (41); *Polyanthia suaveolens* Engl. et Diels (40); **APO:** *Alstonia boonei* De Wild (39) (41); **BAR:** *Combretodendron macrophyllum* (P. Beauv.) Keay. (40); **BIG:** *Fernandoa adolfi-fridericii* (Gilg et Mildbr.) Heine (41); **BUR:** *Canarium schweinfurthii* Engl. (41); *Dacryodes edulis* (G. Don) H. J. Lam. (41); **BYT:** *Theobroma cacao* L. (41); **CAE:** *Amphimas pterocarpoides* Harms (40); *Dialium pachyphyllum* Harms (41); *Erythrophleum guineense* G. Don (39) (41); *Gilbertiodendron dewevrei* (De Wild) J. Léonard (39) (41); *Gossweilerodendron balsamiferum* (Verm.) Harms (41); *Oxystigma oxyphyllum* (Harms) J. Léonard (40) (41); *Paramacrolobium coeruleum* (Taub.) J. Léonard (41); *Scorodophloeus zenkeri* Harms (39) (41); **COM:** *Combretodendron macrocarpum* (Beauv.) Keay (41); *Terminalia superba* Engl. et Diels (39) (41); **EBE:** *Diospyros alboflavescens* (Gurke) F. White (41); *D. crassiflora* Hiern (41); *D. undabunda* Hiern ex Greves (41); **EUP:** *Cleistanthus* sp. (41); *Discoglypema caloneura* Prain (41); *Drypetes gossweileri* S. Moore (39) (41); *D. morocarpa* J. Léonard (41); *D. spinoso-dentata* (Pax) Hutch var. *longipedunculata* J. Léonard (41); *Macaranga lancifolia* Pax (41); *Macaranga* cfr. *lancifolia* Pax (41); *Microdesmis puberula* Hook f. (41); *Phyllanthus discoides* Muell. Arg. (41); *Ricinodendron africanus* Muell. Arg. (41); **GUT:** *Garcinia punctata* Oliv. (41); *Lebrunia busbaie* Staner (41); *Pentadesma* sp. (41); **IRV:** *Irvingia gabonensis* Baill. (41); *I. grandifolia* Engl. (40) (41); *Irvingia* sp. (40) (41); **LAU:** *Beilschmiedia gilbertii* Robyns et Wilczek var. *glabra* (41); **LEG:** *Newtonia buchananii* (Baker) Gilbert et Boutique (41); *Tetrapleura tetraptera* (Thonn.) Taub. (41); **LIN:** *Ochthocosmus* cfr. *africana* Hook. f. (41); **MEL:** *Entandrophragma angolense* C. DC. (39) (41); *E. candollei* Harms (39) (41); *E. cylindricum* (39) (41); *E. utile* (Dawe et Sprague) (39) (41); *Entandrophragma* sp. (39) (41); *Guarea cedrata* (A. Chev.) Pellegr. (39) (41); *G. laurentii* De Wild. (39) (40) (41); *G. thompsonii* Sprague et Hutch. (39) (41); *Khaya anthotheca* C. DC. (41); *Khaya* sp. (40); *Lovoa* sp. (41); *Trichilia retusa* Oliver (41); *T. rubescens* Oliv. (41); *T. welwitschii* C. DC (41); **MOR:** *Antiaris welwitschii* Engl. (39) (41); *Bosqueia angolensis* Ficalho (41); *Chlorophora excelsa* Benth. et Hook f. (39) (40) (41); *Ficus mucoso* Welw. ex Ficalho (41); *Myrianthus arboreus* P. Beauv. (41); **MYRI:** *Coelocaryon preussii* Warb. (41); *Pycnanthus angolensis* (Welw.) Exell. (39) (40) (41); *Staudtia stipitata* Warb. (39) (41); **MYRT:** *Syzygium* sp. (41); **OLA:** *Ongokea gore* (Hua) Pierre (39); *Strombosia glaucescens* Engl. (41); *S. grandifolia* Hook. f. Benth. (41); *Strombosiaopsis tetrandra* Engl. (40) (41); **PAN:** *Panda oleosa* Pierre (40) (41); **PAP:** *Afrormosia elata* Harms (39) (41); *Angylocalyx pynaertii* De Wild. (41); *Pterocarpus soyauxii* Taub. (41); **ROS:** *Parinarium holstii* Engl. (41); **RUB:** *Nauclea diderrichii* (De Wild) Merrill (39) (41); **SAPI:** *Blighia welwitschii* (Hiern) Radlk. (40) (41); *Pancovia harmsiana* Gilg. (41); **SAPO:** *Chrysophyllum africanum* DC. (41); *C. lacourtianum* De Wild. (41); *Pachystela excelsa* Baill. (41); *Synsepalum dulcificum* (Sch. et Th.) Daniell et Bell. (41); *S. subcordatum* De Wild. (41); **SIM:** *Hannoa klaineana* Pierre (40); *Klainedoxa gabonensis* Pierre var. *oblongifolia* (41); **STE:** *Cola griseiflora* De Wild. (40) (41); *Pterygota bequaertii* De Wild. (41); *Sterculia bequaertii* De Wild. (41); *S. quinqueloba* (Gürke) K. Schum. (40) (41); **TIL:** *Desplatsia dewevrei* (De Wild. et Th. Dur.) (41); **ULM:** *C. brieyi* De Wild. (41); *C. mildbraedii* Engl. (39) (41); *C. soyauxii* Engl. (40).

APPENDIX 2

Brentidae which lay eggs in wood borer beetle tunnels

Quotations are indicated by the following numbers:

Kleine, 1923g (1); Beeson, 1925 (2); Kleine, 1929b (3); Kleine, 1931e (4); Kleine, 1933f (5); Browne, 1935 (6); Kleine, 1936c (7); Kleine, 1938d (8); Beeson, 1941 (9); De Muizon, 1960 (10); Schedl, 1961a (11); Mayné & Donis, 1962 (12); Scivittaro, 1975 (13); Zimmerman, 1994 (14); Kabakov, 2001 (15); Mecke *et al.*, 2001 (16).

CYPHAGOGINAE Kolbe

CYPHAGOGINI Kolbe

Adidactus cancellatus (Lacordaire)

PLATYPODIDAE: *Platypus refertus orientalis* Strohm. (11).

Adidactus striolatus (Fairmaire)

PLATYPODIDAE: *Cylindropalpus granulosus subaffinis* Schedl (11); *C. pumilio* Schedl (11); *Cylindropalpus pusillus* Schedl (11); *Periommatius longicollis angustiformis* Schedl (11); *Periommatius* sp. (11); *Platypus hintzi* Schauf. (11); **SCOLYTIDAE:** *Doliopygus brevis medius* Schedl (11); *D. brevis minor* Schedl (11); *D. retusus* Schedl (11); *Doliopygus* sp. (11).

Allagogus brunneus Gahan

PLATYPODIDAE: *Cylindropalpus pertinax* Schedl (11); *Platypus impressus* Strohm. (11); *Triozastus banghaasi marshalli* Samps. (11); **SCOLYTIDAE:** *Doliopygus bilobatus* Schedl (11); *D. brevis brevisculus* Schedl (11); *D. brevis interstitialis* Schedl (11); *D. brevis perbrevis* Schedl (11); *D. brevis solidus* Schedl (11); *D. conjunctus* Schedl (11); *D. dubius* Samps. (11); *D. galerus* Schedl (11); *D. lecomtei* Schedl (10); *D. retusus* Schedl (11); *D. subditivus* Schedl (11); *D. tenuis* Strohm. (11); *Doliopygus* sp. (11).

Nannobrenthus compressithorax (Senna)

PLATYPODIDAE: *Triozastus banghaasi marshalli* Samps. (11); **SCOLYTIDAE:** *Doliopygus brevis solidus* Schedl (11).

Calodromus mellyi Guérin-Ménéville & Gory

SCOLYTIDAE indet. (9).

Cormopus distinctus Kleine

PLATYPODIDAE: *Cylindropalpus granulosus subaffinis* Schedl (11); *C. pertinax* Schedl (11); *C. pusillus* Schedl (11); *Periommatius excisus* Strohm. (11); *P. grandis* Schedl (11); *Platypus refertus orientalis* Strohm. (11); *P. solutus* Schedl (11); *Triozastus banghaasi propatulus* Schedl (11); **SCOLYTIDAE:** *Doliopygus brevis brevisculus* Schedl (11); *D. brevis medius* Schedl (11); *D. brevis minor* Schedl (11); *D. brevis rapax* Samps. (11); *D. serratus* Strohm. (11); *Doliopygus* sp. (11).

Cormopus filum Kolbe

SCOLYTIDAE: *Doliopygus brevis interstitialis* Schedl (11); *Doliopygus* sp. (12).

Cormopus penicillifer Kolbe

PLATYPODIDAE: *Triozastus banghaasi pertenius* Schedl (11); *T. banghaasi propatulus* Schedl (11); *Triozastus* sp. (11); SCOLYTIDAE: *Doliopygus exilis* Chap. (11); *D. lateralis* Schedl (11); *D. ghesquerei* Schedl (11).

Cyphagogus corporaali Kleine

CERAMBYCIDAE: *Xylotrechus smei* Cast. & Gory (1) (2) (10); PLATYPODIDAE: *Platypus solidus* Walker (1) (2) (9) (10).

Cyphagogus cyrtotrachelus (J. Thomson)

PLATYPODIDAE: *Crossotarsus indicus* Saup. (2) (10); SCOLYTIDAE: indet. (2) (9).

Cyphagogus eggersi Kleine

PLATYPODIDAE: *Platypus curtus* Chap. (7).

Cyphagogus eichhorni Kirsch

PLATYPODIDAE: *Crossotarsus saundersii* Chap. (1).

Cyphagogus incisus Senna

PLATYPODIDAE: *Crossotarsus saundersii* Chap. (2).

Cyphagogus planifrons Kirsch

PLATYPODIDAE: *Crossotarsus bifurcus* Schedl (6).

Cyphagogus tabacicola Senna

PLATYPODIDAE: *Crossotarsus squamulatus* Chap. (1) (2).

Cyphagogus westwoodii Parry

PLATYPODIDAE: *Crossotarsus saundersii* Chap. (1) (2) (6) (9) (10); *C. squamulatus* Chap. (1) (2) (5); *Crossotarsus* sp. (15); SCOLYTIDAE: *Xyleborus foersteri* Haged. (6) (7) (9).

Dentisebus pusio (Kolbe)

PLATYPODIDAE: *Cylindropalpus pumilio* Schedl (11); *C. pusillus* Schedl (11); *Periommatius excisus* Strohm. (11); SCOLYTIDAE: *Doliopygus brevis medius* Schedl (11).

Microsebus adelphus Kolbe

SCOLYTIDAE: *Xyleborus parvulus* Eichh. (2).

Oncodemerus costipennis (Fairmaire)

PLATYPODIDAE: *Platypus linearis* Steph. (11); **SCOLYTIDAE:** *Doliopygus ghesquerei* Schedl (11); *D. punctiventris* Schedl (11); *Doliopygus* sp. (11); *Xyleborus camerunus* Hag (12); *X. ferrugineus* F. (12).

Usambioproctus dissimilis (Kleine)

SCOLYTIDAE: *Doliopygus conradti* Strohm. (11); *D. serratus* Strohm. (11); *Doliopygus* sp. (11).

Zemioses porcatus Pascoe

PLATYPODIDAE: *Cylindropalpus granulatus fratellus* Schedl (11); **SCOLYTIDAE:** *Doliopygus serratus* Strohm. (11); *Doliopygus* sp. (11).

HOPLOPISTHIINI Senna & Calabresi*Carcinopisthius oberthueri* (Senna)

PLATYPODIDAE: *Platypus signatus* Chap. (6); Platypodidae indet. (9).

STEREODERMINI Sharp*Cerobates (Cerobates) credibilis* Kleine

SCOLYTIDAE: *Diamerus impar* Chap. (12).

MICROTRACHELIZINI Zimmerman*Microtrachelizus beneficus* Kleine

CERAMBYCIDAE: *Hoplocerambyx spinicornis* Newm. (2) (9) (10) (14).

TAPHRODERINAE Lacordaire*Bolbocranius bicolor* Senna

PLATYPODIDAE: *Periommatius excisus similis* Strohm. (11); *Platypus conjunctus* Schedl (11); *P. hintzi* Schaef. (11); *P. impressus* Strohm. (11); *P. refertus* Schedl (11); *P. schenklingsi* Strohm. (11); *P. spinulosus* Strohm. (11); **SCOLYTIDAE:** *Doliopygus brevis rapax* Schedl (11); *D. brevis solidus* Schedl (11); *D. megatoma* Schedl (11); *D. subditivus* Schedl (11); *D. ukereweensis* Schedl (11).

Bolbocranius csikii (Bolkay)

PLATYPODIDAE: *Cylindropalpus auricomans* Schaef. (11); *C. camerunus* Schedl (11); *C. granulatus fratellus* Schedl (11); *C. laudatus* Schedl (11); *Periommatius longicollis angustiformis* Schedl

(11); *Platypus hintzi* Schauf. (11); *P. impressus* Strohm. (11); *P. linearis* Steph. (11); *P. refertus orientalis* Strohm. (11); *P. solutus* Schedl (11); *Trachyostus schaufussi medius* Schedl (11); *T. schaufussi minor* Schedl (11); **SCOLYTIDAE:** *Diamerus impar* Chap. (12); *Doliopygus brevis brevisculus* Schedl (11); *D. brevis perbrevis* Schedl (11); *D. brevis solidus* Schedl (11); *D. conradti* Strohm. (11); *D. discrepans* Schedl (11); *D. dubius* Samps. (11); *D. erichsoni* Chap. (11); *D. jurioni* Schedl (11); *D. lateralis* Schedl (11); *D. lebruni* Schedl (11); *D. megatoma vexator* Schedl (11); *D. punctiventris* Schedl (11); *D. serratus* Strohm. (11); *D. subditivus* Schedl (11); *D. unicornis* Schedl (11); *Xyleborus alluaudi* Schauf. (11).

***Bolbocranius mechowii* (Kolbe)**

PLATYPODIDAE: *Trachyostus aterrimus* Schauf. (11); *T. schaufussi* Strohm. (11).

***Taphroderes nevermanni* Kleine**

SCOLYTIDAE: *Xyleborus confusus* Eichh. (3) (4) (8) (13).

***Taphroderes sahlbergi* (Sharp)**

SCOLYTIDAE: *Corthylus praealtus* Schedl (16).

***Taphroderopsis oscillator* (Sharp)**

SCOLYTIDAE: *Xyleborus confusus* Eichh. (3) (4) (8) (13).

TRACHELIZINAE Lacordaire

PSEUDOCEOCEPHALINI Kleine

***Schizotrachelus madens* Lacordaire**

PLATYPODIDAE: *Platypus signatus* Chap. (6)

APPENDIX 3

Brentid parasites (Acarina)

BRENTINAE Billberg

BRENTINI Billberg

Brentus anchorago (Linné)

Hypopus spinitarsus Hermann (Vitzthum, 1921a).

Brentus caudatus Herbst

Tyroglyphus mayaorum Vitzthum (Vitzthum, 1921a); *Uropoda diademata* Vitzthum (Vitzthum, 1921a).

Brentus sp.

Anoetus sapromyzae Dufour (Vitzthum, 1921b); *Discopoma regia* Vitzthum (Vitzthum, 1921b).

ARRHENODINI Lacordaire

Baryrhynchus (Baryrhynchus) anthracinus (Kleine)

Cillibano malayica Vitzthum (Vitzthum, 1921b).

Baryrhynchus (Baryrhynchus) latirostris (Gyllenhal)

Cillibano malayica Vitzthum (Vitzthum, 1921a); *Tyroglyphus novisimilis* Vitzthum (Vitzthum, 1921a).

Baryrhynchus (Baryrhynchus) dehiscens (Gyllenhal)

Trachytes sumatrensis Vitzthum (Vitzthum, 1921a).

Baryrhynchus (Baryrhynchus) umbraticus (Kleine)

Cillibano malayica Vitzthum (Vitzthum, 1921b); *Trachytes sumatrensis* Vitzthum (Vitzthum, 1921b); *Uropoda austroasiatica* Vitzthum (Vitzthum, 1921b).

Baryrhynchus (Eupsalomimus) poweri (Roelofs)

Cillibano malayica Vitzthum (Vitzthum, 1921b); *Uropoda austroasiatica* Vitzthum (Vitzthum, 1921b).

Claeoderes radulirostris biserrirostris Boheman

Uropoda atzeka Vitzthum (Vitzthum, 1921a).

Ectocemus cinnamomi (Herbst)

Anoetus prodectoris Vitzthum (Vitzthum, 1921a); *Histiogaster entomophagus* (Laboulbène) (Vitzthum, 1921a).

Elytracantha pogonocera (Montrouzier)

Cillibano malayica Vitzthum (Vitzthum, 1921b).

Episphales pictus (Kirsch)

Anoetus crenulatus Oudemans (Vitzthum, 1921a).

Estenorhinus dispar (Linné)

Uropoda arrhenodis Vitzthum (Vitzthum, 1921a).

Orfilaia vulsellata (Gyllenhal)

Proctophyllodes ampelidis (Buchholz) (Vitzthum, 1921b); *Uropoda transportabilis* Vitzthum (Vitzthum, 1921b).

Orychodes digramma (Boisduval)

Cillibano malayica Vitzthum (Vitzthum, 1921b); *Uropoda transportabilis* Vitzthum (Vitzthum, 1921b); *U. uvaeformis* Vitzthum (Vitzthum, 1921b).

Orychodes indus Kirsch

Uropoda caonorychodis Vitzthum (Vitzthum, 1921b).

Orychodes serrirostris Lund

Tyroglyphus macgillavryi Oudemans (Vitzthum, 1921a); *T. novisimilis* Vitzthum (Vitzthum, 1921a); *Uropoda derosa* Vitzthum (Vitzthum, 1921b); *U. caonorychodis* Vitzthum (Vitzthum, 1921b); *U. orychodis* Vitzthum (Vitzthum, 1921a).

Prophthalmus bourgeoisi Power

Cillibano applicata Vitzthum (Vitzthum, 1921a); *Tyroglyphus novisimilis* Vitzthum (Vitzthum, 1921a).

Prophthalmus mutabilis Senna

Cillibano translucida Vitzthum (Vitzthum, 1921b); *Uropoda mira* Vitzthum (Vitzthum, 1921b).

Prophthalmus potens Lacordaire

Anoetus prophthalmi Vitzthum (Vitzthum, 1921a); *Cillibano romana* (Canestrini) (Vitzthum, 1921a).

Prophthalmus ruficornis Senna

Uropoda inhaerens Vitzthum (Vitzthum, 1921b).

Prophthalmus tricolor Power

Uropoda philippinensis Vitzthum (Vitzthum, 1921a).

Pseudorychodes lineolatus (Kirsch)

Cillibano malayica Vitzthum (Vitzthum, 1921b).

Raphirhynchus longimanus (Lund)*Tyroglyphus novisimilis* Vitzthum (Vitzthum, 1921a).

CYPHAGOGINAE Kolbe

CYPHAGOGINI Kolbe

Cyphagogus eichhorni Kirsch*Anoetus puntulatus* Vitzthum (Vitzthum, 1921b).

ATOPOBRENTINI Damoiseau

Noeceocephalus sculpturatus Senna*Uropoda atlantica* Vitzthum (Vitzthum, 1921a).

STEREODERMINI Sharp

Cerobates (Cerobates) debilis J. Thomson*Uropoda adfixa* Vitzthum (Vitzthum, 1921b).*Cerobates (Ionthocerus) conradti* (Senna)*Uropoda adfixa* Vitzthum (Vitzthum, 1921b).

TRACHELIZINAE Lacordaire

TRACHELIZINI Lacordaire

Hypomiolispa exarata (Desbrochers des Loges)*Tyroglyphus novisimilis* Vitzthum (Vitzthum, 1921a); *T. novus* Oudemans (Vitzthum, 1921a).*Trachelizus bisulcatus* (Lund)*Histiogaster javensis* Oudemans (Vitzthum, 1921a); *Tyroglyphus macgillavryi* Oudemans (Vitzthum, 1921a).*Tracheloschizus bakeri* (Kleine)*Anoetus sapromyzae* Dufour (Vitzthum, 1921b).

ITHYSTENINI Lacordaire

Ithystenus bicolor Sforzi & Bartolozzi*Uropoda anthropophagorum* Vitzthum (Vitzthum, 1921b).

Ithystenus linearis Pascoe*Cillibano malayica* Vitzthum (Vitzthum, 1921b).

PSEUDOCEOCEPHALINI Kleine

Eubactrus semiaeneus Lacordaire*Anoenus polynesiacus* Vitzthum (Vitzthum, 1921a).*Hormocerus reticulatus* (Lund)*Anoenus trichophorus* Oudemans (Vitzthum, 1921a); *Cillibano applicata* Vitzthum (Vitzthum, 1921a); *Tyroglyphus macgillavryi* Oudemans (Vitzthum, 1921a).*Phacecerus olivaceus* Boheman*Uropoda admixta* Vitzthum (Vitzthum, 1921b).*Prodictor fruhstorferi* Senna*Anoetus prodictoris* Vitzthum (Vitzthum, 1921a).*Storeosomus decollatus* (Chevrolat)*Anoetus longipes* Oudemans (Vitzthum, 1921b); *Uropoda madagascariensis* Vitzthum (Vitzthum, 1921b).*Stroggylosternum nitens* Kleine*Uropoda mira* Vitzthum (Vitzthum, 1921b).

RHYTICEPHALINI Kleine

Ischyromerus madagascariensis Labram & Imhoff*Cilíbano kleinei* Vitzthum (Vitzthum, 1921b); *Uropoda admixta* Vitzthum (Vitzthum, 1921b); *U. promiscua* Vitzthum (Vitzthum, 1921b).

APPENDIX 4

Brentid host ant species

Quotations are indicated by the following numbers:

Azam C., 1884 (1); Brenske & Reitter, 1884 (2); Azam J., 1893 (3); Wasmann, 1894 (4); Mayet, 1904 (5); Lea, 1905 (6); Walker 1905 (7); Lea, 1912 (8); Sahlberg, 1913a (9); Sahlberg, 1913b (10); Kleine, 1916g (11); Kleine, 1918b (12); Gestro, 1919 (13); Kleine, 1921i (14); Kleine, 1923g (15); Menozzi, 1923 (16); Kleine, 1924d (17); Porta, 1932 (18); Mader, 1937 (19); Hoffmann, 1945 (20); Liebmann, 1945 (21); Grandi, 1951 (22); Caillol, 1954 (23); Scortecci, 1960 (24); Le Masne & Torossian, 1965 (25); Tassi, 1965 (26); Torossian, 1965 (27); Torossian, 1966 (28); Bernard, 1968 (29); Schedl K., 1970 (30); Gobbi, 1974 (31); Schedl W., 1975a (32); Español, 1981 (33); Ponel, 1981 (34); Bach & Cárdenas-Talaverón, 1984 (35); Hölldobler & Wilson, 1990 (36); García-París & Olmos, 1992 (37); Ponel, 1993 (38); Zimmerman, 1994 (39); Riti & Osella, 1996 (40).

BRENTINAE Billberg

ARRHENODINI Lacordaire

Orfilaia reichei (Fairmaire)⁽¹⁾

FORMICINAE: *Camponotus compressus* ssp. *fellah* Dalla Torre (32).

Orfilaia vulsellata (Gyllenhal)

FORMICINAE: *Plagiolepis steingroeveri* For. (14).

EREMOXENINI Semenov-Tian-Shanskij

Amorphocephala coronata (Germar)

FORMICINAE: *Camponotus aethiops* Latreille (25) (27) (28) (30); *C. cruentatus* Latreille (4) (14) (15) (23) (25) (27) (28) (29) (30) (34) (37); *C. ligniperda* Latreille (23); *C. sanguineus* L. (11) (12) (15) (19) (20); *C. sylvaticus* (Oliv.) (10) (20) (23) (25) (28) (29) (30); *C. vagus* Scopoli (= *pubescentis* F.) (2) (4) (12) (14) (15) (16) (18) (19) (20) (22) (26) (29) (30); *Camponotus* sp. (5) (20) (21) (24) (30) (31) (32) (33) (35) (36) (37) (38) (40); *Formica* sp. (32); *Lasius niger* L.⁽²⁾ (1) (3) (23); *Lasius* sp. (22) (31) (35) (37) (40); MYRMICINAE: *Crematogaster sordidula* (Nylander) (35) (40); *Crematogaster* sp. (37); *Pheidole* sp. (11) (12) (14) (30) (35) (37) (40).

Cordus ganglbaueri Senna

DOLICHODERINAE: *Iridomyrmex nitidus* Mayr (39).

Cordus hospes Schoenherr

DOLICHODERINAE: *Anonychomyma* (sub *Iridomyrmex*) *itinerans* (Lowne) (8); *Iridomyrmex nitidus* Mayr (6) (39); FORMICINAE: *Camponotus* sp. (7) (39); MYRMICINAE: *Aphaenogaster* (sub *Stenamma*) *longiceps* (Fr. Smith) (8); *Aphaenogaster longiceps* (Fr. Smith) (39); *Crematogaster laeviceps* Sm. (14) (15) (39).

Cordus kraatzii (Senna)MYRMICINAE: *Crematogaster depressa* Latreille (14) (15).*Cordus schoenherri* PowerDOLICHODERINAE: *Iridomyrmex nitidus* Mayr (14) (15) (39).*Cordus* sp.DOLICHODERINAE: *Iridomyrmex conifer* Forel (39).*Paussobrenthus bakeri* GestroDOLICHODERINAE: *Philidris* (sub *Iridomyrmex*) *myrmecodiae* (Emery) (13) (14) (15) (22).*Symmorphocerus piochardi* (Bedel)FORMICINAE: *Camponotus sylvaticus* var. *dichrous* F. (9) (10) (17) (30).

TRACHELIZINAE Lacordaire

PSEUDOCEOCEPHALINI Kleine

Orphanobrentus laevicollis (J. Thomson) ⁽¹⁾DORYLINAE: *Dorylus brevipennis* Em. (14).*Rhinopteryx foveipennis* (J. Thomson) ⁽¹⁾DORYLINAE: *Dorylus brevipennis* Em. (14); *D. wilverthi* Emery (14).

⁽¹⁾ According to Kleine (1923g: 623) this species has only occasionally been found together with ants and so may not be a true mirmecophilous species.

⁽²⁾ According to Kleine (1921i: 110) and Schedl (1970: 104) the species C. Azam (1884) and J. Azam (1893) refer to as *Lasius niger* is in fact *Camponotus cruentatus*.

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Antliarhininae Schoenherr, 1823 (Coleoptera, Curculionoidea)

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ABSTRACT

The small African brentid taxon Antliarhininae contains only two extant genera, *Antliarhinus* and *Platymerus*, with seven described species. Its discovery, classification, distribution, morphology, biology and evolutionary history are summarised. The Antliarhininae are indicated to be the adelphotaxon of Apioninae, have a fragmented distribution on the eastern side of Africa and are exclusively associated with cycads of the genus *Encephalartos*. This association is likely to be Late Cretaceous or early Tertiary in origin and to have evolved as a shift from angiosperms, possibly Proteaceae. Five new synonymies are proposed: *Antliarhinus rectirostris* Gyllenhal, 1836 **syn. n.** and *A. coriaceus* Gyllenhal, 1840 **syn. n.** of *Antliarhinus zamiae* (Thunberg, 1784); *A. dregei* Gyllenhal, 1840 **syn. n.** of *A. signatus* Gyllenhal, 1836; *Platymerus lehmanni* Gyllenhal, 1836 **syn. n.** of *P. eckloni* Gyllenhal, 1836; *P. germari* Gyllenhal, 1836 **syn. n.** of *P. winthemi* Gyllenhal, 1836. The Miocene fossil *Antliarhinites gracilis* Heer, 1864 is excluded from Antliarhininae and from Coleoptera altogether, and tentatively assigned to Heteroptera.

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INTRODUCTION

This small group of brentid weevils includes only two genera but is an extremely interesting one, both because of its exclusive association with African cycads and its generally plesiomorphic yet specialised morphology and indicated basal position in the phylogeny of the Brentidae. Its classificatory position has consequently been uncertain and is still not fully resolved. Several species remain to be described, and a comprehensive revision of the group is in progress (Oberprieler, in prep.).

HISTORICAL REVIEW

The Antliarhininae were the very first cycad-associated weevils to become known, when the Swedish explorer C. P. Thunberg discovered the first African cycads in 1772/3 in the Eastern Cape Province of South Africa (Gunn & Codd, 1981) and found their seeds infested with a long-snouted weevil, which he later described as *Curculio zamiae* (Thunberg, 1784). This remained the only antliarhinine species known to the early authors – again described by Olivier (1791) and Wesmael (1836) under different names, and transferred to a separate genus by Billberg (1820) and Schoenherr (1823) –, until Schoenherr (1836) described a second genus, *Platymerus*, and Gyllenhal (1836, 1840) additional species of *Antliarhinus* from the extensive beetle material collected from “*Zamia caffra*” in the Eastern Cape by the botanical explorers C. F. and J. F. Drège, C. F. Ecklon and C. L. P. Zeyher (Gunn & Codd, 1981). Heer (1864) described a fossil insect from Miocene deposits in Germany as *Antliarhinites gracilis* and affiliated it with *Antliarhinus*, but its true identity and relationships remained dubious.

Further material and information of Antliarhininae became available when, among the first resident botanists in South Africa, H. H. W. Pearson, A. M. Pegler, G. Rattray and H. W. R. Marloth began to investigate cycad biology in the country, although they often had difficulty distinguishing the Antliarhininae from other cycad weevils, *i.e.* the Amorphocerini (Oberprieler, 1996). Chevrolat (1878) and Champion (1914) described additional new genera from, respectively, Brazil and the Seychelles in Antliarhininae, but these were later recognised as belonging to different weevil groups. Marshall's (1955) and Verdcourt's (1957) records of *Antliarhinus* from Kenya, and de Moor's (1973) unnoticed one from Zimbabwe, were significant but isolated further contributions to the knowledge of these weevils, until Donaldson (1992, 1993a, 1993b, 1997) investigated the biology and host specialisation of *Antliarhinus* in detail and Oberprieler (1989a, 1989b, 1995a, 1995b, 1997a, 1997b, 2000) began to study the taxonomy and evolution of the group. The Antliarhininae currently comprise seven described species, but another four undescribed *Antliarhinus*

and one *Platymerus* species are known (Verdcourt, 1957, de Moor, 1973, Donaldson, 1992, Oberprieler, 1995a, 1995b, Oberprieler & Popiel, 1996).

CLASSIFICATION

Although Schoenherr (1823, 1826, 1836, 1840) classified the antliarhinines in his 'Ordo Orthoceri' and was even (1836, p. 823) very specific about this ("*haec genera indubie ad Ordinem Orthocerorum referenda, atque inter Divisiones Brenthides et Cylades inserenda esse*"), later authors, following Imhoff (1856) and the influential Lacordaire (1866), explicitly or implicitly included the group in Curculionidae Gonatoceri. Handlirsch (1925) again suspected their affinity to lie with Brentidae, but only the later thorough examinations of Kasap & Crowson (1977), Sanborne (1981) and Calder (1989, 1990) found significant character agreements with Apioninae, which these authors treated as a family. Alonso-Zarazaga (1989), Kuschel (1995), Oberprieler (1995a) and Alonso-Zarazaga & Lyal (1999) included the antliarhinines in Brentinae (rather than Apioninae), a treatment unknowingly supported by Damoiseau (1989) when he described *Platybrentus* (later recognised as a synonym of *Antliarhinus*) in Brentidae. A 'compromise' position between Brentinae and Apioninae (or between Brentidae and Apionidae when these were treated as separate families) was adopted by Sanborne (1981), Lawrence & Newton (1982, 1995), Thompson (1992), May (1993), Gønget (1997), Morrone (1998) and Lawrence *et al.* (1999), although Sanborne (1981) already identified the antliarhinines as being adelphic ('sister') to all other Apioninae. Recent phylogenetic analyses based on larval (Oberprieler, 2000), imaginal (Wanat, 2001) and molecular and combined characters (Marvaldi *et al.*, 2002) support a closer affinity to apionines than to brentines, with Wanat's (2001) analysis (spanning all primitive apionines) confirming their basal position in the group, as was suggested by Sanborne 20 years earlier. Even though Wanat's analysis requires expansion to a larger set of taxa (more brentine genera and all species of Antliarhininae) and of characters (including larval ones), a position of the antliarhinines as the most basal group of Apioninae is currently the most plausible. Their inclusion in this checklist (which excludes the Apioninae) is therefore somewhat anomalous, but as this project was commissioned before these latest analyses were published, as they are listed under Brentidae in the Alonso-Zarazaga & Lyal (1999) catalogue and as no comprehensive checklist exists of them (other than the species list in Oberprieler, 1995a), it was decided to adhere to the original plan and include them here as a subfamily of Brentidae.

NOMENCLATURE

Controversy surrounds the correct name of the type genus of the group. Billberg (1820) erected it as *Antliarhis* (gender feminine), but Schoenherr (1823) emended this to the masculine *Antliarhinus* and provided a proper description of the genus. Even though this emendation is deemed unjustified under the provisions of the modern International Code of Zoological Nomenclature (ICZN), it has been exclusively used in both the taxonomic coleopterological and the applied literature (with Billberg's original spelling only cited in synonymy), until Thompson (1992) without explanation resurrected the forgotten original spelling and May (1993), Kuschel (1995), Lawrence & Newton (1995), Alonso-Zarazaga & Lyal (1999), Lawrence *et al.* (1999) and Wanat (2001) followed him. In a footnote discussing this matter, Oberprieler (1995a) rejected this resurrection for reasons of nomenclatural instability and maintained the existing usage, as governed by Art. 23(b) of the 3rd edition of the ICZN, as then in effect.

The 4th edition of the ICZN, in effect from January 2000, in Art. 23.2. similarly intends the Principle of Priority to be used to promote stability, not "to upset a long-accepted name in its accustomed meaning ...", and, in the general recommendations of Appendix B, urges authors not to change prevailing usage of names unless this is required for scientific reasons. It furthermore includes a new article (23.9.) that allows Reversal of Precedence in cases where the senior synonym has not been used for a long time. This article applies here as follows:

- there is no scientific reason for replacing *Antliarhinus* Schoenherr, 1823 with its forgotten senior synonym *Antliarhis* Billberg, 1820, as there exists no doubt about the application of Schoenherr's name;
- nomenclatural stability is threatened by such an action, as *Antliarhinus* is widely used in the botanical and the applied horticultural literature and as its replacement with a feminine name (*Antliarhis*) would require all adjectival species names (such as *signatus*) to change their ending;
- condition 23.9.1.1. is not met as the senior synonym, *Antliarhis* Billberg, although unused as a valid name since 1823, was resurrected by Thompson (1992) and used as a valid name by a few authors since;
- condition 23.9.1.2. is met as the junior synonym, *Antliarhinus* Schoenherr, 1823, has been used as the valid name of the taxon by at least 52 authors in at least 71 works over a period of 180 years (169 years until the 1992 resurrection of *Antliarhis*).

Since condition 23.9.1.1. is not met, *Antliarhinus* Schoenherr, 1823 can unfortunately not here be treated as a *nomen protectum* in accordance with Art. 23.9.2., and *Antliarhis* Billberg, 1820 not as a *nomen oblitum*, and the matter has to be referred to the ICZN Commission for a ruling. However, Art. 23.9.3. and Art. 82 stipulate that in the meantime prevailing usage (*i.e.* the junior name) is to be maintained, and *Antliarhinus* is therefore here used as the valid name.

DISTRIBUTION

The Antliarhininae are restricted to the Afrotropical region, occurring from the Eastern Cape Province of South Africa (Figs 1, 8) northwards along the eastern side of the African continent (seemingly following the escarpment) to the Kenyan highlands. This distribution pattern largely agrees with that of their cycad hosts (see Goode, 1989, Jones, 2002), but as yet no Antliarhininae are known to occur on the central and western African cycads. In the east also, their distribution is not fully congruent with that of the cycads but considerably more fragmented, essentially restricted to three areas: the Eastern Cape in South Africa, the eastern highlands of Zimbabwe and the Kenyan highlands from near Mombasa to the Matthews Range. In the intermediate areas (the former Natal and Transvaal provinces of South Africa, and the southern part of the African Rift Valley from Malawi to eastern Congo/western Tanzania), cycads are much sparser both in species numbers and in population sizes, and antliarhinines are likely to have disappeared from these areas as their cycad diversity and density declined.

MORPHOLOGY

The structural characters of the Antliarhininae have not yet been comprehensively studied. This is largely due to the scarcity of specimens in collections, and nearly all investigations have been based only on *Antliarhinus zamiae*, the most common but also the most specialised species of the group. Kasap & Crowson (1977) and Calder (1989, 1990) studied aspects of its internal anatomy, while Sanborne (1981) gave an overview of its morphology and Thompson (1992) investigated its abdominal structure in greater detail, in a comparative study of higher curculionoid taxa. Howden (1995) investigated its ovipositor structure in relation to oviposition behaviour in weevils. Sanborne (1981) and May (1993) examined the larval characters of the group, again based only on *A. zamiae*, and Oberprieler (2000) analysed phylogenetically relevant larval characters of several species of *Antliarhinus* as well as of *Platymerus*. Recently, Wanat (2001) undertook an extensive comparative study of the external imaginal morphological characters of the group, based on several species of both genera, but since the important northern *Antliarhinus verdcourti* species group is largely undescribed and was not included, even this treatment is not comprehensive. The pending revision of the Antliarhininae (Oberprieler, in prep.) will address this issue.

Among the Brentidae, the Antliarhininae exhibit numerous primitive features in the antennal scrobes, elytra, legs, ventrites and male genitalia (Wanat, 2001: 362-363) but are also highly specialised morphologically, with some unique apomorphies related to their life style in tight spaces in cycad cones.



Fig. 1 - Habitat of *Antliarhinus zamiae* (Thunberg) with host plant *Encephalartos princeps* (Zamia-ceae): Kubusi Valley, Eastern Cape Province, South Africa (photo by R. Oberprieler).



Fig. 2 - Host of *Antliarhinus zamiae* (Thunberg): *Encephalartos villosus* female plant with ripe cone; Komga, Eastern Cape Province, South Africa (photo by R. Oberprieler).

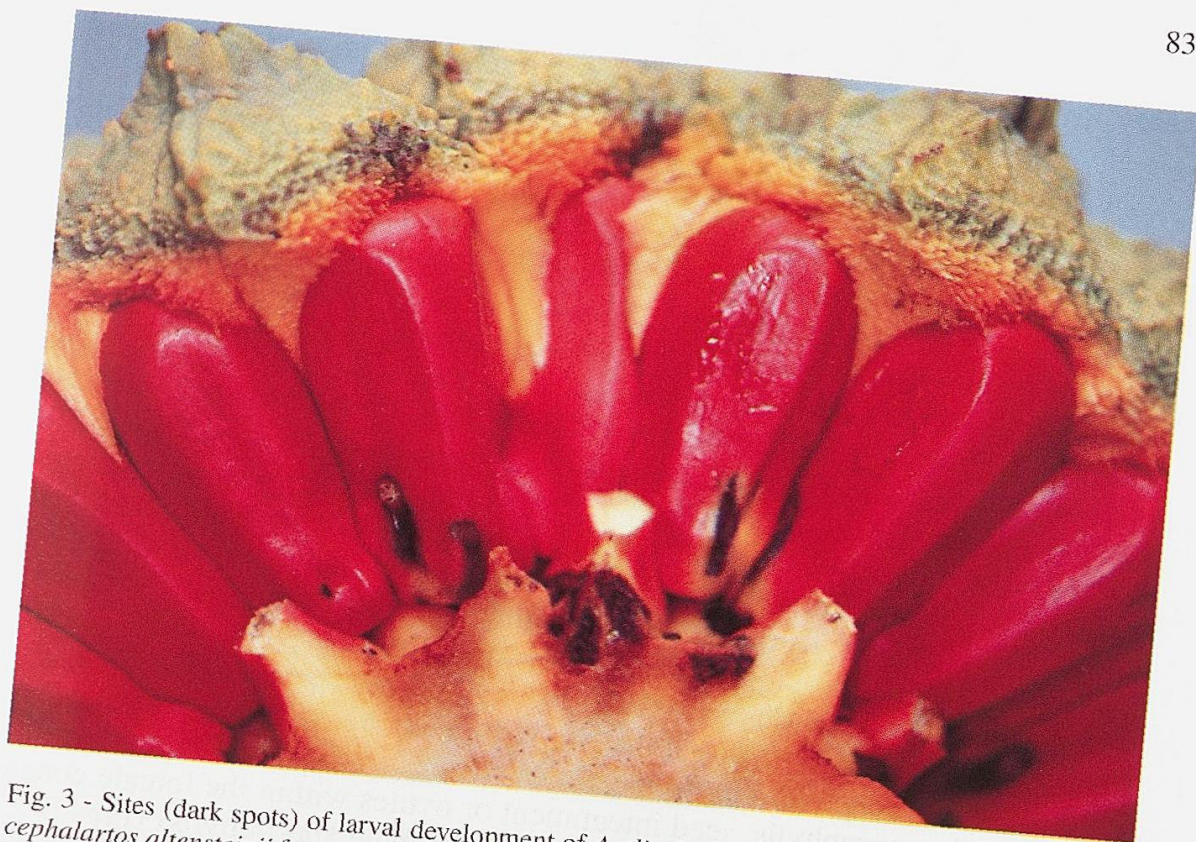


Fig. 3 - Sites (dark spots) of larval development of *Antliarhinus* sp. n. in seed flesh and axis of *Encephalartos altensteinii* female cone; Port Alfred, Eastern Cape Province, South Africa (photo by R. Oberprieler).



Fig. 4 - *Antliarhinus signatus* Gyllenhal male (bottom) and female on cut seed of *Encephalartos altensteinii*; Port Alfred, Eastern Cape Province, South Africa (photo by R. Oberprieler).

BIOLOGY

Early records of antliarhinine biology were limited to vague indications of hostplants (given as *Zamia caffra*; Thunberg, 1781, Linnaeus, 1790, Gyllenhal, 1836), and even though *A. zamiae* was later well known to develop in cycad seeds (Dyer, 1965, Giddy, 1974, 1984, Crowson, 1981, Annecke & Moran, 1982), its biology and that of the group as a whole group remained largely uncertain due to a general confusion pertaining to the identities of the various weevils that inhabit *Encephalartos* cones in the Eastern Cape Province (Rattray, 1913, Marloth, 1914, Oberprieler, 1996). Marshall's (1955) record of the eastern African *A. verdcourti* developing not in cycad seeds but in the rhachis of female cones added to the conundrum surrounding antliarhinine biology, and only Donaldson's (1992, 1993a, 1993b) comprehensive studies of four species of *Antliarhinus* established details of mating and oviposition behaviour, larval development and host relationships in this genus. Oberprieler (1989a) provided the only account of the biology of the enigmatic *Platymerus* and later summarised the biology of both genera (Oberprieler, 1995a). It is now established that the Antliarhininae are exclusively associated with cycads of the genus *Encephalartos* in Africa, where their larvae develop in the rhachis, sporophylls, seed integument or ovules within the female cones (only *A. peglerae* sometimes also developing in male sporophylls) (Figs 3-7, 9-10). Except for the seed-feeding *A. zamiae* and *A. signatus*, all species appear to be monophagous on single cycad species, but none of them are indicated to be significantly involved in the pollination of their hosts. *Antliarhinus zamiae* and *A. signatus* are widely regarded as serious "pests" of cycads both in the wild and in horticulture (Giddy, 1974, 1984, Goode, 1989, Jones, 1993, 2002, Whitelock, 2002).

EVOLUTIONARY HISTORY

The age and origin of the cycad association of the Antliarhininae remain unclear. Crowson (1991) surmised the group to be of Tertiary age, and Oberprieler (1995b) suggested a Late Cretaceous origin based on their high morphological specialisation and intimacy of host association, the discovery of primitive brentids in the early Late Cretaceous (Cenomanian) of southern Africa (Kuschel *et al.*, 1994), and the apparent origin of modern cycad genera in the early Tertiary. The recently confirmed position of the antliarhinines as the most primitive Apioninae (Wanat, 2001) is broadly in accord with such an age, with derived Apioninae appearing in the fossil record in the Late Eocene. In contrast, recent allozyme and molecular analyses of cycad phylogeny (Van der Bank *et al.*, 2001, Treutlein & Wink, 2002, Schneider *et al.*, 2002) propose a much younger age for the diversification of the extant cycads, dating the divergence between the African *Encephalartos* and the Australian

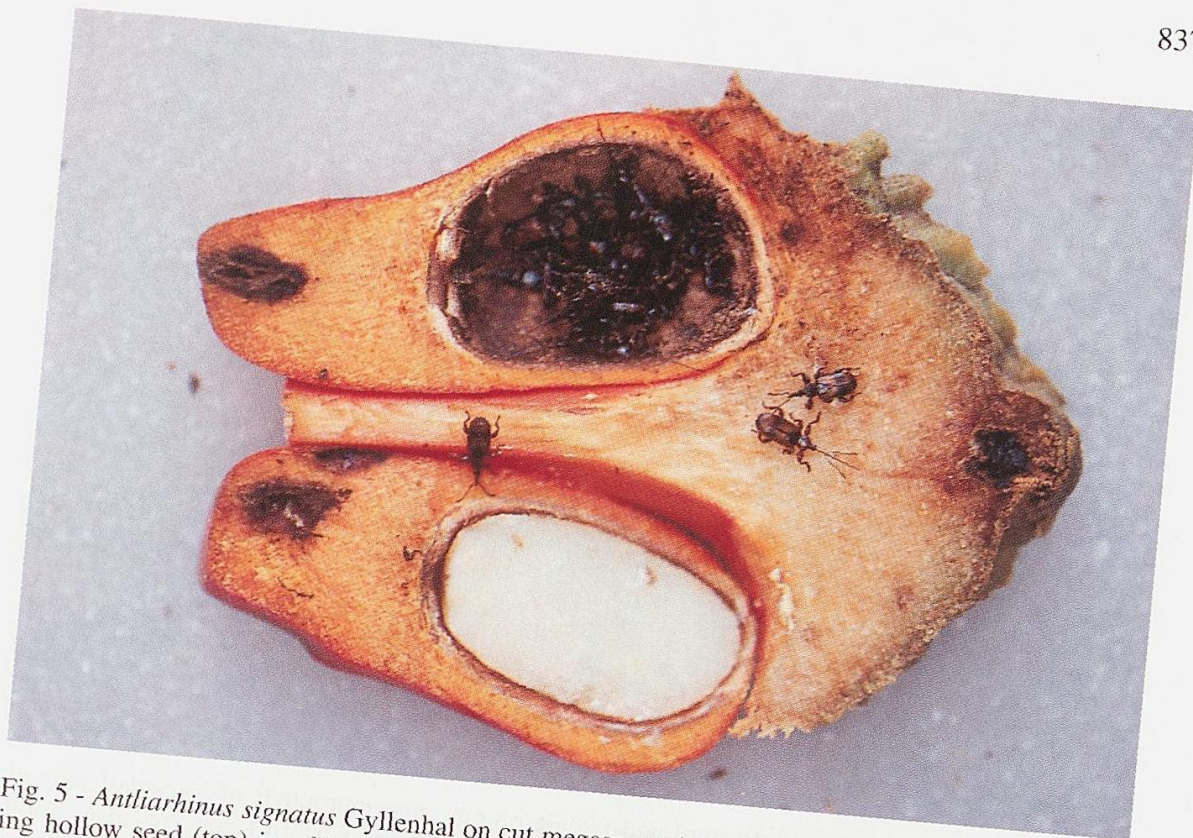


Fig. 5 - *Antliarhinus signatus* Gyllenhal on cut megasporophyll of *Encephalartos altensteinii* showing hollow seed (top) in which they developed; Port Alfred, Eastern Cape Province, South Africa (photo by R. Oberprieler).

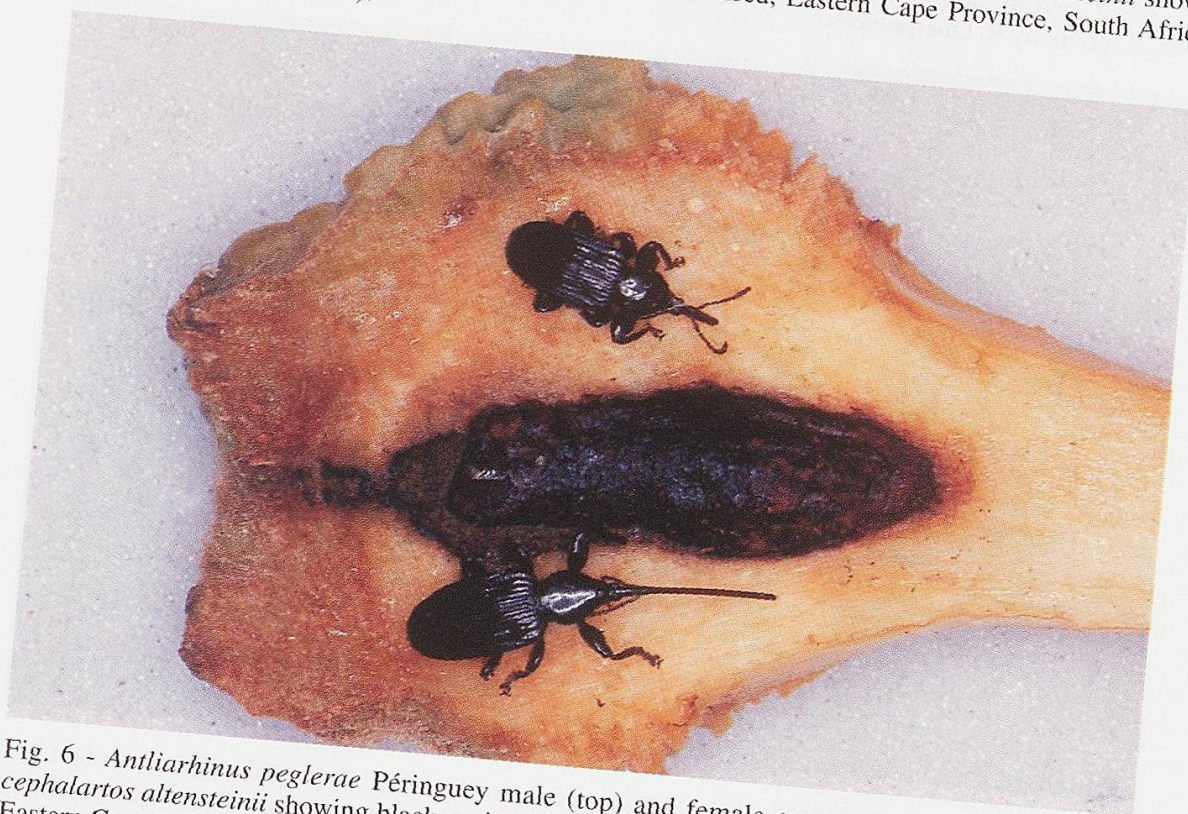


Fig. 6 - *Antliarhinus peglerae* Péringuey male (top) and female on cut megasporophyll of *Encephalartos altensteinii* showing black cavity in which the female developed; King William's Town, Eastern Cape Province, South Africa (photo by R. Oberprieler).



Fig. 7 - *Antliarhinus* sp. n. male (top) and female on cut megasporophyll of *Encephalartos al-tensteinii* next to cavities in which they developed; Port Alfred, Eastern Cape Province, South Africa (photo by R. Oberprieler).



Fig. 8 - Habitat of *Platymerus zeyheri* Gyllenhal and *P. eckloni* Gyllenhal, with host plant *Encephalartos friderici-guilielmi* growing along hill top; Queenstown, Eastern Cape Province, South Africa (photo by R. Oberprieler).



Fig. 9 - *Platymerus zeyheri* Gyllenhal male (left) and female on woolly female cone of *Encephalartos friderici-guilielmi* in which they developed; Queenstown, Eastern Cape Province, South Africa (photo by R. Oberprieler).

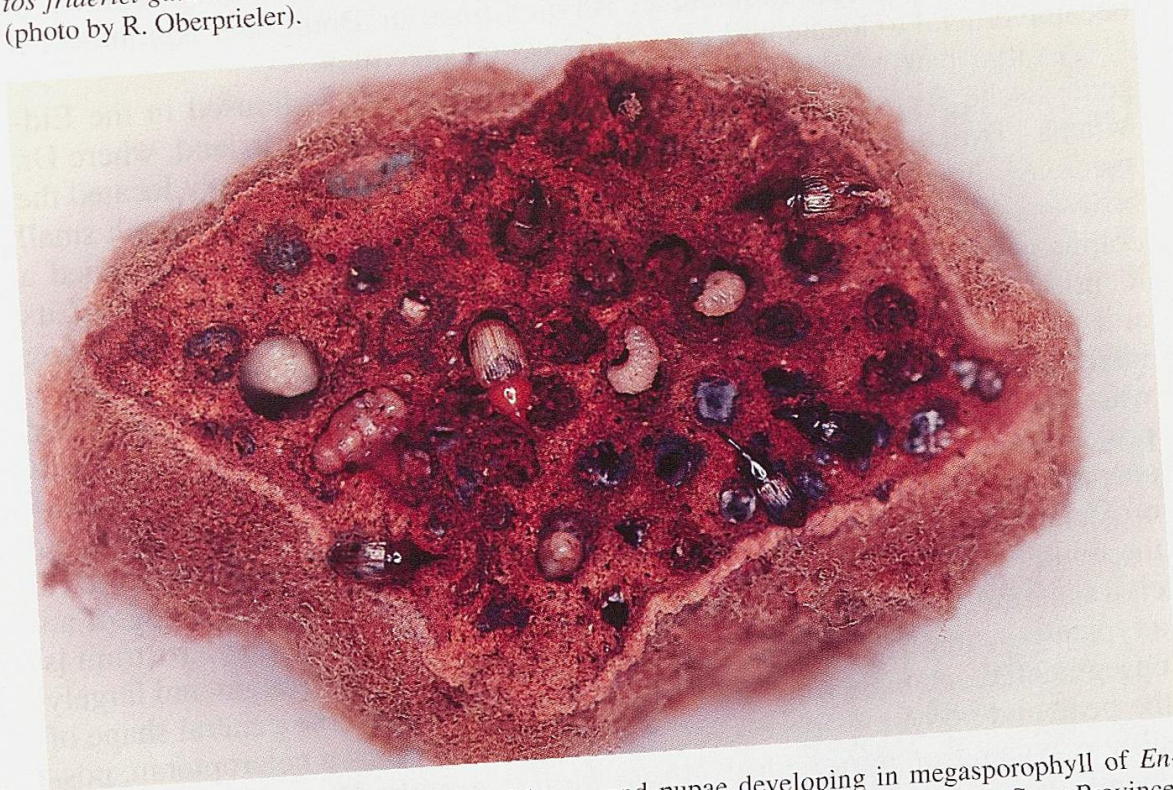


Fig. 10 - *Platymerus zeyheri* Gyllenhal larvae and pupae developing in megasporophyll of *Encephalartos friderici-guilielmi*, top (outer) section cut away; Queenstown, Eastern Cape Province, South Africa (photo by R. Oberprieler).

Macrozamia/Lepidozamia as Early Miocene and the diversification within *Encephalartos* as Middle Miocene (ca 10 mya) in age. These results are in need of corroboration, and a Late Cretaceous to Early Tertiary age for the Antliarhininae currently remains the most plausible. Their association with cycads is almost certainly derived from an original angiosperm host, as is the rule in Brentidae, probably as a shift from Proteaceae, which are the hosts of other primitive African Apioninae (Tanaini, Mecolenini) (Wanat, 2001).

THE IDENTITY OF *ANTLIARHINITES GRACILIS* HEER, 1864

Heer (1864: 371, 374) described and illustrated a fossil insect specimen from the Late Miocene deposits at Öhningen in southern Germany as *Antliarhinites gracilis* and related it to *Antliarhinus*, on account of "its thin, long rostrum and the basally inserted antennae". Scudder (1891) later listed it under Attelabidae: Rhynchitinae and Handlirsch (1925) and Carpenter (1992) under Curculionidae, but Alonso-Zarazaga & Lyal (1999) placed it back in Antliarhininae. None of these authors had, however, inspected Heer's specimen, and even the late V. V. Zherikhin, who had begun to systematically re-examine many described weevil fossils in recent years, had not seen it (Zherikhin, 2002, pers. comm. 2000). Since Heer's description and small, poor line drawing of *A. gracilis* (fig. 246) do not allow a reliable placement in either Antliarhininae, Attelabidae: Rhynchitinae or Brentidae: Apioninae, it became crucial to locate and inspect the actual specimen.

A large part of Heer's collection of insect fossils is housed in the Eidgenössische Technische Hochschule (ETH) in Zürich, Switzerland, where Dr. Milena Pika-Biolzi searched through the collection and eventually located the specimen. It appears that Heer's fossils are still packed in their original, small boxes and have never been studied again. Dr. Pika-Biolzi kindly supplied a colour photograph of *A. gracilis*, which finally allows a reassessment of its identity. The photograph (Fig. 12) shows a mirror image of Heer's drawing and probably depicts the counterpart of Heer's specimen, but it is readily identifiable as the same specimen. From the photograph it is clear that Heer was grossly mistaken in assigning this fossil to the weevils. The appendage projecting from the front of the head is evidently not a rostrum but the second antenna, and the abdomen shows the venter rather than two closed elytra, as on Heer's drawing. The distinct, curved, longitudinal, median line in front of the middle legs, which Heer seems to have interpreted as the beginning of an elytral suture, evidently is a thin rostrum folded backwards. Such a rostrum is a typically hemipteran feature and, together with the long, straight and largely unsegmented antennae, the slender femora and tibiae and the general shape of the head and body, most likely identifies the specimen as a heteropteran, possibly a lygaeid or pyrrhocorid. *Antliarhinites gracilis* Heer, 1864 is consequently here excluded from Curculionoidea and tentatively assigned to Heteroptera, pending a detailed examination of the fossil itself.

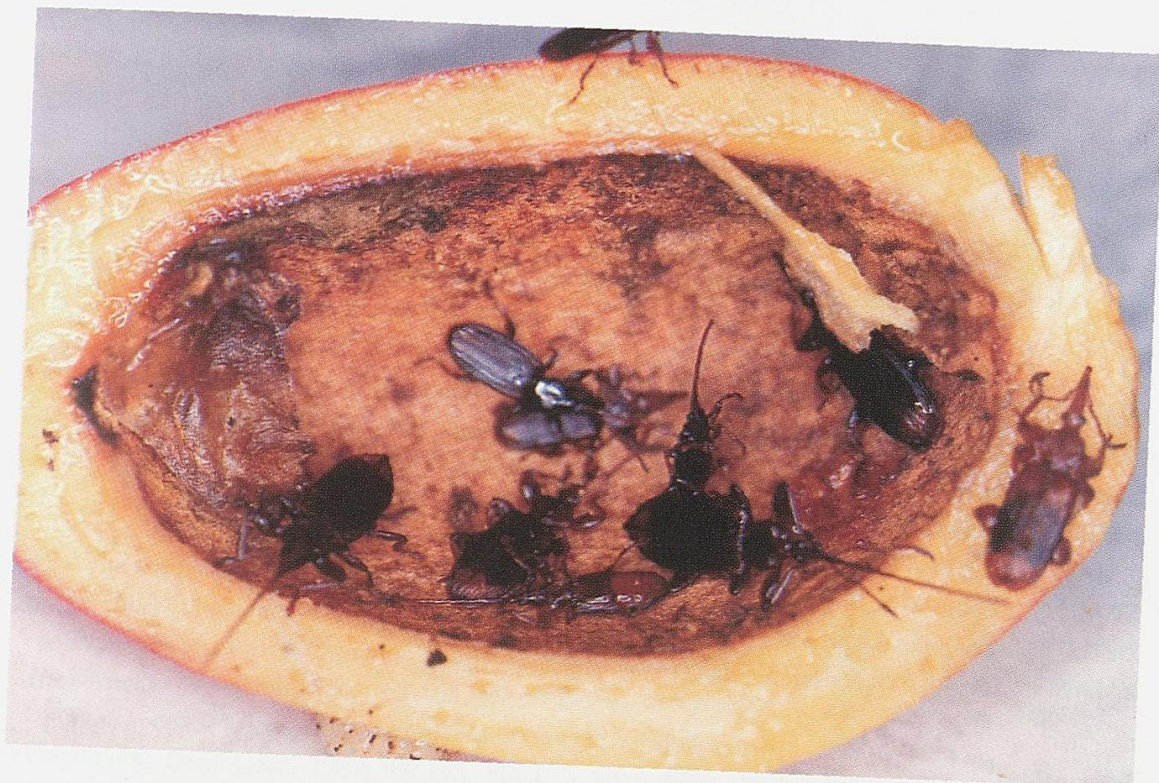


Fig. 11 - *Antliarhinus zamiae* (Thunberg) in opened seed of *Encephalartos villosus* hollowed out by their larvae; East London, Eastern Cape Province, South Africa (photo by R. Oberprieler).



Fig. 12 - Fossil *Antliarhinites gracilis* Heer (Heteroptera), incorrectly related to Antliarhininae but not a weevil; Miocene of Öhningen, Germany (photo by M. Pika-Biolzi).

ABBREVIATIONS

HT	Holotypus
ST	Syntypus

The codens of the institutions holding type material of Antliarhininae follow Samuelson *et al.* (2002):

BMNH	The Natural History Museum, London, United Kingdom
ISNB	Institut Royal des Sciences Naturelles de Belgique, Brussels, Belgium
NHRS	Naturhistoriska Riksmuseet, Stockholm, Sweden
NMBZ	Natural History Museum of Zimbabwe, Bulawayo, Zimbabwe
NMKE	National Museum of Kenya, Nairobi, Kenya
SAMC	South African Museum, Cape Town, South Africa
UZIU	Zoological Museum, Uppsala University, Uppsala, Sweden
ZMAN	Zoologisch Museum, Amsterdam, The Netherlands

CHECKLIST

ANTLIARHININAE Schoenherr, 1823

Antliarhinides Schoenherr, 1823: c. 1137.

Antliarhininae Schoenherr: Handlisch, 1925: 678.

Antliarhinus Schoenherr, 1823

Antliarhis Billberg, 1820: 40.

Type species: *Curculio haustellatus* Olivier, by monotypy.

Antliarhinus Schoenherr, 1823: c. 1137 [emend. for *Antliarhis* Billberg].

Type species: *Curculio zamiae* Thunberg, by original designation

Mitorhynchus Wesmael, 1836: 163-165 (*Mitorhinchus*, *Mithorinchus*); syn. Lacordaire, 1866: 182.

Type species: *Mitorhynchus brunneus* Wesmael, by monotypy.

Platybrentus Damoiseau, 1989: 94; syn. Sforzi & Oberprieler (in Alonso-Zarazaga & Lyal, 1999: 50).

Type species: *Platybrentus paradoxus* Damoiseau, by original designation.

Type species: *Curculio zamiae* Thunberg, 1784.

Distribution. Southern and eastern Africa.

zamiae (Thunberg, 1784)

zamiae Thunberg, 1784: 29, pl. 1 fig. 7 (*Curculio*).

haustellatus Olivier, 1791: 517 (*Curculio*); syn. Schoenherr, 1826: 68.

brunneus Wesmael, 1836: 165, pl. 6 fig. 2 (*Mitorhynchus*); syn. Gyllenhal, 1840: 583.

rectirostris Gyllenhal, 1836: 825 **n. syn.**

coriaceus Gyllenhal, 1840: 584 **n. syn.**

paradoxus Damoiseau, 1989: 94 (*Platybrentus*); syn. Sforzi & Oberprieler (in Alonso-Zarazaga & Lyal, 1999: 50).

Type depositories. *C. zamiae*: UZIU (ST); *C. haustellatus*: not traced; *M. brunneus*: ISNB (2 ST); *A. rectirostris*: NHRS (2 ST!); *A. coriaceus*: NHRS (2 ST!); *P. paradoxus*: ZMAN (HT! ♂).

Distribution. Southern Africa, Eastern Cape Province, introduced into KwaZulu-Natal, Gauteng and Mpumalanga (type locality only given as “*Caput bona spei*”, but likely to be at the mouth of the Seekoei River near the

present-day Humansdorp in the Eastern Cape Province, after details of Thunberg's travels in Gunn & Codd, 1981) (Figs 1, 2).

Hosts. Larvae communally in seeds of several *Encephalartos* species (see Oberprieler, 1995a) (Fig. 11).

Notes. Lectotypes for all relevant species names will be designated in the revision of the group in preparation. Synonymy of *rectirostris* Gyllenhal and *coriaceus* Gyllenhal with *zamiae* Thunberg is, however, effected here, based on a comparative study of the relevant type specimens. Both ST of *rectirostris* (labelled "Caffraria / in Zamia sp. / Ecklon & Zeyh.") as received from the NHRS are at the larger end of the size variation of *A. zamiae* (male 7 mm, female 22 mm total body length), and the female has the typically large rostrum:body length ratio (1.98) of such specimens; the straightness of its rostrum is a non-diagnostic feature dependent mainly on the preservation technique employed. The two ST of *coriaceus* (labelled "Cap. b. sp. / Drège") are near the smaller end of the size variation of *A. zamiae* (male 6.4 mm, female 12 mm total body length) and with a correspondingly small rostrum:body length ratio (1.5) in the female. Rostrum length in the female varies allometrically in relation to body size, the latter being determined by the food resources available to the developing larva and dependent on the size of the cycad seed and the number of larvae developing in it. In all other critical characters the types of *rectirostris* and *coriaceus* agree fully with typical specimens of *A. zamiae*.

signatus Gyllenhal, 1836

signatus Gyllenhal, 1836: 825.

dregei Gyllenhal, 1840: 585 n. syn.

Type depositories. *A. signatus*: NHRS (2 ST!); *A. dregei*: NHRS (2 ST!).

Distribution. Southern Africa, Eastern Cape Province (precise type locality not deducible from either the specimen labels or the literature).

Hosts. Larvae communally in seeds of several *Encephalartos* species (see Oberprieler 1995a) (Figs 4, 5).

Notes. The synonymy of *dregei* Gyllenhal with *signatus* Gyllenhal is based on a comparison of their respective type specimens. The male ST of *signatus* (labelled "Cap. b. sp. / Drège" and bearing a dark-red "Typus" label, but not being validly designated as lectotype) is a specimen of *Platymerus eckloni* (already so labelled by Kuschel in 1992). The female ST (with same

locality label but a dark-red "Allotypus" label - again not a valid paralectotype designation) is relatively large (7 mm in length). Female type(s) with a black prothorax, identified as "variety β " by Gyllenhal (1836), were not examined. The STs of *dregei* are indistinguishable from the female ST of *signatus*, representing only small specimens of the same species, with the male (labelled "Africa Austr. / Drège" and bearing a dark-red "Typus" label but also not validly designated as lectotype) measuring 4 mm in length and the female (labelled "Caffrafria / in Zamia Caffra / Eckl. & Zeyh." and with a dark-red "Allotypus" label) 6 mm. Size variation in this species is again determined by the number of larvae developing on the limited food resource of a cycad seed.

peglerae Péringuey, 1908

peglerae Péringuey, 1908: 325.

Type depository. SAMC (3 ST!).

Distribution. Southern Africa, Eastern Cape Province to southern KwaZulu-Natal (type locality: Kentani in the former Transkei).

Hosts. Larvae in megasporophylls (occasionally also microsporophylls) of several *Encephalartos* species (see Oberprieler, 1995a) (Fig. 6).

Notes. One ST carries the SAMC type number 4066 and is labelled as type; however Péringuey (1908) did not identify a holotype and no lectotype has been validly designated thus far.

verdcourti Marshall, 1955

verdcourti Marshall, 1955: 300.

Type depositories. BMNH (37 ST!), SAMC (2 ST!).

Distribution. Kenya (type locality: Lolokwe in the Matthews Range).

Hosts. Larvae in rhachis of female cones of *E. tegulaneus* (see Oberprieler, 1995a), similar to those of undescribed South African species on *E. altensteinii* (Figs 3, 7).

Notes. The type series was bred in 1954 by B. Verdcourt from the rhachis of a female cone of *E. tegulaneus*. Another 27 specimens from the same locality, collected in July 1954, are in NMBZ.

Platymerus Schoenherr, 1836

Platymerus Schoenherr, 1836: 826.

Type species: *Platymerus eckloni* Gyllenhal, by original designation.

Type species: *Platymerus eckloni* Gyllenhal, 1836.

Distribution. Southern Africa.

eckloni Gyllenhal, 1836

eckloni Gyllenhal, 1836: 827.

lehmanni Gyllenhal, 1836: 829 **n. syn.**

Type depositories. *P. eckloni*: NHRS (2 ST!); *P. lehmanni*: NHRS (2 ST!).

Distribution. Southern Africa, Eastern Cape Province (type locality not specified, but from Ecklon's and Zeyher's collecting localities in the 'Tambukiland' (after Gunn & Codd, 1981) it is between Queenstown and Cathcart, probably the 'Friedrich-Wilhelmsberg' near Silo (Shiloh, at present-day Whittelee)) (Fig. 8).

Hosts. Larvae in megasporophyll peduncles and seed integument (occasionally also boring into ovules) of *E. friderici-guilielmi* (see Oberprieler, 1989b, 1995a).

Notes. All types are labelled "Caffraria / in Zamia Caffra / Eckl. & Zeyh.", with the males additionally bearing dark-red "Typus" labels and the females similar "Allotypus" labels (these not representing valid lectotype designations). The ST of *lehmanni* are indistinguishable from those of *eckloni* on all critical taxonomic characters, representing only somewhat lighter-coloured specimens of the same species. Both these names are established in the same publication; *eckloni* is here given preference as the valid name of the species because it is already in use to denote the species as the type species of *Platymerus*.

winthemi Gyllenhal, 1836

winthemi Gyllenhal, 1836: 829.

germari Gyllenhal, 1836: 830 **n. syn.**

Type depositories. *P. winthemi*: NHRS (2 ST!); *P. germari*: NHRS (2 ST!).

Distribution. Southern Africa, Eastern Cape Province (type locality as for *P. eckloni*).

Hosts. Larvae in outer layer of megasporophyll bullae of *E. friderici-guilielmi* (see Oberprieler, 1989b, 1995a).

Notes. All ST are labelled in the same way as those of *P. eckloni*. They differ from the latter only in their smaller size, narrower shape and paler coloration with larger yellow maculae on the elytra. Clear species-diagnostic differences are absent, and it is possible that *winthemi* represents only minor males and females of *P. eckloni*. The types form a transition series (*eckloni* – *lehmanni* – *germari* – *winthemi*) from large, robust and dark specimens to smaller, more elongate, paler ones, with the gap between *eckloni/lehmanni* and *germari/winthemi* being somewhat more distinct and, in the absence of a thorough study of long series of specimens, for the moment precluding the synonymy also of *germari* and *winthemi* with *eckloni*. The status of *P. winthemi* as a distinct species, as here preserved, is, however, in need of review, taking into consideration also the apparent differences in sites of larval development (see Oberprieler 1989b). The ST of *germari* are indistinguishable from those of *winthemi* except for minor differences in coloration. Both these names are again established in the same publication; *winthemi* is here given preference as the valid name of the species in line with its previous use to denote this species (Oberprieler, 1989b, 1995a,b).

zeyheri Gyllenhal, 1836

zeyheri Gyllenhal, 1836: 828.

Type depository. NHRS (4 ST!).

Distribution. Southern Africa, Eastern Cape Province (type locality as for *P. eckloni*).

Hosts. Larvae in megasporophyll bullae of *E. friderici-guilielmi* (see Oberprieler, 1989b, 1995a) (Figs 9, 10).

Notes. All STs are labelled in the same way as those of *P. eckloni* and *P. winthemi*. One pair represents Gyllenhal's "variety β ", more lightly coloured specimens in which particularly the pronotum is of a more reddish colour than in darker-coloured specimens.

ACKNOWLEDGEMENTS

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Cyladinae Schoenherr, 1823 (Coleoptera, Curculionoidea)

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ABSTRACT

A review of the systematic and nomenclatural state of the Cyladinae is presented. The single included genus, *Cylas* Latreille, 1802, has 24 species. These are catalogued, their synonymy and distribution summarized, and type depositories given. A general introduction to history, classification, distribution and biology opens this treatment.

The following new synonymy is proposed: *Cylas longicollis* Chevrolat, 1844 **syn. n.** of *Cylas longicollis* Guérin-Ménéville, 1833.

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INTRODUCTION

This is a group of 24 species (although it seems that many more are extant in collections and awaiting description), associated with tubers and fruits of Convolvulaceae, mainly in the tropical genus *Ipomoea*. All these species are currently included in the single genus *Cylas*.

HISTORICAL REVIEW

The first member of the genus *Cylas* to be described was the type species as *Brentus brunneus* by Olivier (1790) from Senegal (although Fabricius, 1792 is wrongly quoted in literature as the author). A few years later, Fabricius (1798) described the commonest species twice, the males as *Brentus formicarius* and the females as *Attelabus formicarius*. This particular species has deserved much attention from the applied entomologists since it is a red-outable enemy of the sweet potato crops.

In 1802, Latreille erected the genus *Cylas* for the inclusion of *B. brunneus* and placed it between *Brentus* and *Attelabus*.

In 1941, Pierce described a new genus, *Protocylas*, to include the species having globose elytral shapes and short rostra. Both features intergrade in some species and the only phylogenetic analysis available for the moment (Wolfe, 1991) shows that the included species belong to different non-monophyletic groupings in the cladogram.

CLASSIFICATION

The genus *Cylas* was isolated in its own group (as Divisio 11, Cylades) by Schoenherr (1823), who recognised its peculiar features. Cylades was placed between Divisio 10, Brenthides, and 12, Ulocerides, that is, more or less where they stand now.

The relationships of this group have been always a debatable question, and the authors placed *Cylas* either as a member of its own family (e.g. Imhoff, 1856) or as a member of Curculionidae Apioninae (e.g. Wagner, 1910 in the tribe Eurhynchini; Kissinger, 1968) or Brentidae (e.g. LeConte, 1876; LeConte & Horn, 1883) depending upon the importance given to the considered characters. In general, it was always clear that the genus belonged somewhere in the "brentoid complex" but the affinities were uncertain.

It was not until Morimoto (1976) showed unmistakably the presence of a particular apomorphy of the Brentidae (namely, the labial palpi borne in deep sockets of the labium) that the position of *Cylas* as a peculiar member of

Brentidae became clear. The last world catalogue of weevil family and genus group names (Alonso-Zarazaga & Lyal, 1999) recognised this situation by placing Cyladinae as one of eight subfamilies of Brentidae Billberg, 1820. A recent phylogenetic analysis (Wanat, 2001) showed that the Cyladinae and the rest of the Brentidae (Brentinae *s. l.*) are sister groups, Antliarhinae being placed as the most basal clade within Apionidae. However, this situation may require a reappraisal when the Microcerinae are included.

Inside Brentidae, Cyladinae show several autapomorphic features, which depart from the basic body structure of the remainder of Brentidae, like the united antennal club, the sulciform scrobes, the fused claws, the pedunculate prothorax, the tangent procoxae, the large mesothoracic pit, the confusely punctate elytra, the reduced tibial spurs, the anal cell of wing absent and the specialized (probably sensory) pits inside the antennal scrobe, among others.

Within *Cylas*, several species groups can be recognised, some of them monospecific (Wolfe, 1991). At least four African species await description. Wolfe (*l. c.*) recognises three species groups among pest species (although he did not include non-pest species in groups): the *C. formicarius* group, including *C. formicarius* alone; the *C. brunneus* group, including *C. brunneus* and *C. femoralis*; and the *C. puncticollis* group, divided into two subgroups: the *C. puncticollis* subgroup, including *C. puncticollis* (and its rather undefinable subspecies *opacus*) and *C. nigrocoeruleans*, and the *C. compressus* subgroup, including *C. compressus* and *C. hovanus*. The other species have not been assigned to any group.

NOMENCLATURE

From the most modern species list (Wolfe, 1991), one species must be dismissed: *Cylas rufescens* Fairmaire, 1900 is a member of genus *Lispotherium* in Apionidae.

DISTRIBUTION

The Cyladinae were restricted originally to the Old World Tropics, namely to the Afrotropical and Oriental Realms. According to Wolfe (1991) the common ancestor of the genus evolved in mid-Jurassic times (ca. 150 Mybp) in the Gondwanian area that gave later West Africa, Madagascar and India. With the subsequent spread of their host plants as human food resources, a few of the species have become introduced outside of their original range since the XVIth century, mainly in the Americas. *C. formicarius* is native from the Oriental Region, and has successfully reached the Americas and less so Africa, where it has to compete against local pests of sweet potato and other *Ipomoea* spp.

BIOLOGY

In general, *Cylas* spp. attack only *Ipomoea* spp., although they can be found on a variety of other plants (Chalfant *et al.*, 1990). Eleven species have been identified as pests of sweet potato, although there are real records only for nine of these (Wolfe, 1991). Little is known about the non-pest species biology. Larvae are non-typical brentid larvae, lacking legs (Kemner, 1924), although this absence is not recognised by Wibmer and O'Brien (1986), who state that larvae do have legs. Larvae and pupae are killed by parasite fungi and adults are hosts of parasite nematodes (Kemner, 1924; Chalfant *et al.*, 1990).

In *C. formicarius*, dispersal ability of males by flight is moderate to low (Miyatake *et al.*, 1997) and they only fly at night (Sakuratani *et al.*, 1994), females tend to disperse less, usually by crawling. Dispersal is influenced by several factors, among which climatic factors, seasons and availability of hosts (Miyatake *et al.*, 1995), as well as age, mating status and degree of starvation (Moriya & Hiroyoshi, 1998).

Females use their rostrum for chewing holes in the tuber necks, each hole is provided with a single egg, which is covered with a faecal plug. Larvae develop internally by boring the plant tissues, and they undergo 3-5 moults within a mean of 3 weeks, although the larval life span (10-36 days) is strongly influenced by outer temperature (Risbec, 1947; Chalfant *et al.*, 1990). The pupal stage takes a mean of 15 days (range: 7-28 days). Adults feed on leaves, stems and tubercles. In favourable cases, there are up to 9 generations a year, the mean adult female longevity is 76-110 days, the males tend to have a shorter life (data for *C. formicarius*) (Chalfant *et al.*, 1990).

Several braconid, pteromalid and encyrtid wasps have been described as natural enemies (Risbec, 1947; Chalfant *et al.*, 1990).

ABBREVIATIONS

AT	Allotype
HT	Holotype
PT	Paratype
ST	Syntype
MNHN	Museum National d'Histoire Naturelle, Paris, France
MRAC	Musée Royal de l'Afrique Centrale, Tervuren, Belgium
NHM	The Natural History Museum, London, United Kingdom
NHMB	Naturhistorisches Museum, Basel, Switzerland
NHRS	Naturhistoriska Riksmuseet, Stockholm, Sweden
NZSI	National Zoological Collection, Zoological Survey of India, Calcutta, India
SMTD	Staatliches Museum für Tierkunde, Dresden, Germany
ZMUC	Zoological Museum, University of Copenhagen, Denmark

CHECKLIST

CYLADINAE Schoenherr, 1823

Cylades Schoenherr, 1823: 1137.
 Cyladae; Imhoff, 1856: XVIII.
 Cylinae; Pascoe, 1870: 436.
 Cyladidae; LeConte, 1876: 327.
 Cyladinae; LeConte & Horn, 1883: 503.
 Cyladini; Faust, 1894: 233.
 Cyladina; Champion, 1914: 450.
 Cyladinini; Hustache, 1933: XI.

Cylas Latreille, 1802

Cylas Latreille, 1802: 196.
Cylanus Rafinesque, 1815: 115 [unjustified emendation].
Cyclus: Leach, 1815: 107 [lapsus].
Protocylas Pierce, 1941: 219 [described as valid genus; type species: *Cylas laevicollis* Boheman, 1833 by original designation; synonymy by Kissinger (1968: 12)].

Type species: *Brentus brunneus* Olivier, 1790 by monotypy.

Distribution. Circumtropical; originally Old World tropics; New World tropics through introduction.

aeneus Hustache, 1922

aeneus Hustache, 1922: 498.

Type depository. Depository not stated, probably MNHN (ST).

Distribution. Cameroon, Chad, Democratic Republic of the Congo (ex Zaire), Ethiopia, Mali, Republic of the Congo, Senegal.

brunneus (Olivier, 1790)

brunneus Olivier, 1790: 190 (*Brentus*).
angustatus Labram & Imhoff, 1838: nr. 25 (*Cylas*); syn. Pierce, 1940: 217.

Type depositories. *B. brunneus*: MNHN (ST?); *C. angustatus*: NHMB (ST?).

Distribution. Angola, Burundi, Ghana, Ivory Coast, Kenya, Nigeria, Rwanda, Senegal, Sierra Leone, Togo, Uganda.

Notes. Species attacking sweet potato crops (Risbec, 1947; Wolfe, 1991; Smit *et al.*, 2001).

coimbatorensis (Subramanian, 1958)

coimbatorensis Subramanian, 1958: 204 (*Protocylas*).

Type depository. Agricultural College and Research Institute, Coimbatore, India (HT, AT, PT).

Distribution. India.

compressus Hartmann, 1899

compressus Hartmann, 1899: 22.

Type depository. SMTD (ST).

Distribution. Kenya, Senegal, Tanzania, Uganda.

Notes. Species attacking sweet potato crops (Wolfe, 1991).

curtipennis Fairmaire, 1884

curtipennis Fairmaire, 1884: CXLVII.

Type depository. MNHN (ST?).

Distribution. Cameroon, Chad, Gabon, Kenya, Senegal, Tanzania, Zambia.

cyanescens Boheman, 1833

cyanescens Boheman, 1833: 371.

Type depository. NHRS (ST).

Distribution. Angola, Ethiopia, Mali, Republic of the Congo, Senegal, South Africa.

Notes. Species attacking sweet potato crops (Risbec, 1947); considered uncertain by Wolfe (1991).

femoralis Faust, 1898

femoralis Faust, 1898: 24.

Type depository. SMTD (ST).

Distribution. Cameroon, Chad, Democratic Republic of the Congo (ex Zaire), Gabon, Guinea, Ivory Coast, Liberia, Mali, Niger, Republic of the Congo.

Notes. A pest of sweet potato (Risbec, 1947; Hoffmann, 1963; Wolfe, 1991).

formicarius (Fabricius, 1798)
(Figs 1, 2)

formicarius Fabricius, 1798: 174 (*Brentus*).

formicarius Fabricius, 1798: 163 (non Linnaeus, 1758) (*Attelabus*); syn. Fabricius, 1801: 549.

turcipennis Boheman, 1833: 369 (*Cylas*); syn. Kemner, 1924: 401.

elegantulus Summers, 1875: 401 (*Otidocephalus*) (see notes); syn. LeConte, 1876: 327.

Type depositories. *A. formicarius*: ZMUC (ST); *B. formicarius*: ZMUC (ST); *C. turcipennis*: NHRS (ST); *O. elegantulus*: lost.

Distribution. Circumtropical through introduction; not native in Western Hemisphere, doubtfully native in Africa. Angola, Cameroon, Chad, Comoros, Democratic Republic of the Congo (ex Zaire), Ghana, Kenya, Liberia, Madagascar, Mauritius, Mozambique, Reunion, Rodrigues Is., Senegal, Seychelles, Somalia, South Africa, Sudan, Swaziland, Tanzania, Uganda, Zimbabwe; Bangladesh, Burma, Cambodia, Chagos Arch., China (Fujian, Guangdong, Guangxi, Guizhou, Hunan, Jiangsu, Jiangxi, Shandong, Sichuan, Yunnan, Zhejiang), Christmas I., Cocos-Keeling I., India (Andaman Is., Andhra Pradesh, Assam, Bihar, Gujarat, Karnataka, Kerala, Madhya Pradesh, Maharashtra, Manipur, Nagaland, Orissa, Rajasthan, Tamil Nadu, Uttar Pradesh),



Fig. 1 - *Cylas formicarius*: male, dorsal view.

Indonesia (Aru, Java, Kalimantan, Madura, Maluku, Riau, Sangir, Sulawesi, Sumatra, Tanimbar, Timor), Japan (Kyushu, Ryu-Kyu Is., Ogasawara Is.), Korea, Laos, Malaysia (Brunei, Labuan, Sabah, Sarawak, West Malaysia), Pakistan, Philippines, Singapore, Sri Lanka, Taiwan, Thailand, Vietnam; American Samoa, Australia (New South Wales, Queensland), Caroline Is., Cook Is., Fiji, Hawaii, Kiribati, Marquesas Is., Marianas Is., New Guinea (Irian Jaya, Papua New Guinea), New Zealand, Society Is., Solomon Is., Tonga, Vanuatu, Wallis Is.; Antilles (Anguilla, Antigua, Barbados, St. Eustatius, St. Kitts-Nevis, Virgin Is.), Bahamas, Belize, Cayman Is., Cuba, Dominican Republic, Guatemala, Haiti, Jamaica, Mexico, Puerto Rico, Trinidad; SE, SW USA, Bermuda; Guyana, Peru, Venezuela.

Notes. This is the “sweet potato weevil”, a dreadful pest of sweet potato, *Ipomoea batatas* (L.) Lam. and other related wild species, such as *I. pes-caprae* (L.) R. Br. (seashore convolvulus), *I. acuminata* (Vahl) Roemer & Schultes (morning glory) or *I. digitata* L. (vidari-kanda). *Manihot palmata* Müller. Arg. (sweet cassava) (Euphorbiaceae) has also been reported as attacked (Risbec, 1947) and some other plants may act as refuges. The literature on its bionomics and ecology is overwhelming, and will not be reported here.

Summer’s description of *Otidocephalus elegantulus* was not published in a scientific journal, therefore it must be deemed as unpublished, not satisfying the criteria of Art. 8.1.1 of the International Code on Zoological Nomenclature (1999).



Fig. 2 - *Cylas formicarius*: details of head and antennae, showing difference in size of antennal club and eyes: male on the left, female on the right.

freyi Voss, 1966

freyi Voss, 1966: 319.

Type depository. NHMB (HT, PT?).

Distribution. Mali, Nigeria.

glabripennis Hartmann, 1897

glabripennis Hartmann, 1897: 282.

Type depository. SMTD (ST).

Distribution. Cameroon, Chad, Gabon, Nigeria, Tanzania.

hovanus Hustache, 1933

hovanus Hustache, 1933: 49.

Type depository. MNHN (ST).

Distribution. Madagascar.

Notes. Species attacking sweet potato crops (Wolfe, 1991).

impunctatus Faust, 1891

impunctatus Faust, 1891: 282.

Type depository. SMTD (HT?).

Distribution. India.

laevicollis Boheman, 1833

laevicollis Boheman, 1833: 372.

Type depository. NHRS (ST).

Distribution. Indonesia (Java).

***laevigatus* Fähræus, 1871**

laevigatus Fähræus, 1871: 237.

Type depository. NHRS (HT).

Distribution. South Africa.

***longicollis* Guérin-Méneville, 1833**

longicollis Guérin-Méneville, 1833: pl. 36, f. 10.

longicollis Chevrolat, 1844: 139 (non Guérin-Méneville, 1833) (*Cylas*); **syn. n.**

Type depository. Unknown, either NHRS or MNHN.

Distribution. Ivory Coast, Republic of the Congo, Senegal.

Notes. Both nominal species refer to the same taxon and originated from the same specimens.

***nigrocoerulans* Fairmaire, 1902**

nigrocoerulans Fairmaire, 1902: 384.

Type depository. MNHN (HT?).

Distribution. Madagascar.

Notes. Species attacking sweet potato crops (Wolfe, 1991).

***nitens* Hustache, 1929**

nitens Hustache, 1929: 485.

Type depository. MNHN (ST).

Distribution. Kenya.

***pumilus* Marshall, 1953**

pumilus Marshall, 1953: 108.

Type depository. NHM (ST).

Distribution. Angola, Namibia.

***puncticollis puncticollis* Boheman, 1833**

puncticollis Boheman, 1833: 372.

Type depository. NHRS (ST).

Distribution. Burundi, Cameroon, Cape Verde, Central African Republic, Chad, Democratic Republic of the Congo (ex Zaire), Ethiopia, Guinea, Ivory Coast, Kenya, Madagascar, Malawi, Mali, Mozambique, Nigeria, Republic of the Congo, Rwanda, Senegal, Sierra Leone, Somalia, Sudan, Tanzania, Uganda, South Africa, Zambia.

Notes: Species attacking sweet potato crops (Risbec, 1947; Wolfe, 1991; Smit *et al*, 2001).

***puncticollis opacus* Voss, 1966**

puncticollis opacus Voss, 1966: 319.

Type depository. NHMB (HT, PT?).

Distribution. Guinea, Nigeria.

Notes. Doubtfully different from the nominotypical subspecies.

***robustus* Faust, 1895**

robustus Faust, 1895: 149.

Type depository. SMTD (ST).

Distribution. Eritrea, Ethiopia.

***rufipes* Faust, 1893**

rufipes Faust, 1893: 513.

Type depository. Unknown, probably in coll. Fleutiaux (MNHN) (ST) and in coll. Faust (SMTD) (ST).

Distribution. Vietnam.

***semipunctatus* Fåhraeus, 1871**

semipunctatus Fåhraeus, 1871: 237.

Type depository. NHRS (HT).

Distribution. South Africa.

***submetallicus* Desbrochers, 1890**

submetallicus Desbrochers, 1890: 214.

Type depository. MNHN (ST), NZSI (ST).

Distribution. India.

***vandenplasi* Burgeon, 1936**

vandenplasi Burgeon, 1936: 508.

Type depository. MRAC (ST).

Distribution. Chad, Democratic Republic of the Congo (ex Zaire), Ethiopia, Mali, Republic of the Congo, Tanzania.

Notes. Species uncertainly attacking sweet potato crops (Wolfe, 1991).

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Eurhynchinae Lacordaire, 1863 (Coleoptera, Curculionoidea)

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ABSTRACT

The small Australo-New-Guinean brentid taxon Eurhynchinae contains three extant genera (*Eurhynchus*, *Ctenaphides* and *Aporhina*) and one extinct one (*Orapaeus*), with together 30 currently recognised species. Its discovery, classification, distribution, morphology, biology and evolutionary history are summarised, and all its described species are catalogued. The Eurhynchinae are indicated to represent a primitive and ancient brentid lineage that may have survived in temperate and tropical Australia and in New Guinea since the mid-Cretaceous. The precise relationships and classificatory position of the group remain uncertain. Its biology and host associations are poorly known, and some further biological observations are here reported. A replacement name, *Aporhina pulchra* Oberprieler, **nom. n.**, is proposed for *Aporhina splendida* (Blackburn, 1894) because of secondary homonymy with *Aporhina splendida* (Blanchard, 1849). Following the earlier restoration of *Aporhina* as senior synonym of *Chalcocybeus*, the following new combinations are effected: *Aporhina alboguttata* (Snellen van Vollenhoven, 1866) **comb. n.**, *A. aspericollis* (Heller, 1910) **comb. n.**, *A. assimilis* (Györfyi, 1917) **comb. n.**, *A. bennigseni* (Wagner, 1912) **comb. n.**, *A. biroi* (Györfyi, 1917) **comb. n.**, *A. bispinosa aruensis* (Heller, 1896) **comb. n.**, *A. bispinosa guttifera* (Heller, 1896) **comb. n.**, *A. bispinosa intermedia* (Heller, 1896) **comb. n.**, *A. bispinosa macrospilota* (Heller, 1896) **comb. n.**, *A. exarmata* (Heller, 1905) **comb. n.**, *A. granosispina* (Heller, 1925) **comb. n.**, *A. helleri* (Wagner, 1912) **comb. n.**, *A. inermis* (Heller, 1897) **comb. n.**, *A. insignis* (Heller, 1905) **comb. n.**, *A. levigata* (Heller, 1925) **comb. n.**, *A. magdalenae* (Györfyi, 1917) **comb. n.**, *A. massutei* (Heller, 1901) **comb. n.**, *A. massutei interrupta* (Heller, 1901) **comb. n.**, *A. mesospila* (Györfyi, 1917) **comb. n.**, *A. mutica* (Heller, 1925) **comb. n.**, *A. nitens* (Snellen van Vollenhoven, 1866) **comb. n.**, *A. nitens papuana* (Heller) **comb. n.**, *A. obtusispina* (Heller, 1925) **comb. n.**, *A. richteri* (Faust, 1892) **comb. n.**

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INTRODUCTION

The Eurhynchinae (Fig. 1) are a small and peculiar group of brentid weevils extantly restricted to Australia and New Guinea but with fossil representatives in southern Africa and possibly South America. The group appears to represent the remnants of an old brentid stock that may have originated in the mid-Cretaceous, and it may also preserve an ancient type of brentid host associations and larval habits. Its classification and taxonomic status have varied considerably over the last century and remain somewhat uncertain and controversial, despite a number of recent phylogenetic analyses that have included eurhynchine taxa and characters. The Australian fauna was comprehensively revised by Zimmerman (1994), but the New Guinean fauna has received virtually no attention for about 80 years and remains very incompletely sampled and studied. No comprehensive catalogue exists of the group as a whole since Wagner (1910, 1912a). Such an updated catalogue is presented here, together with a summary of the classification, distribution, morphology, biology and evolutionary history of the group.

HISTORICAL REVIEW

The seemingly very first eurhynchines to become known were collected by Robert Brown in "New Holland" and deposited in the Macleay collection, from where Kirby described *Eurhynchus scabrior*, *E. laevior* (Fig. 1) and *E. muricatus* in 1819. Further specimens were taken in the new colony in ensuing years by British, French and German collectors and described in their respective homelands, by Boisduval (1835), Boheman (1839, 1859), Erichson (1842), Blanchard (1849) and Pascoe (1870a, b, 1873). By 1873, all species



Fig. 1 - *Eurhynchus laevior* (Kirby): male on leaf of hostplant *Persoonia levis* (Proteaceae); Ter-meil, N.S.W., Australia (photo by D. Rentz).

of *Eurhynchus* and *Ctenaphides* were described, except for Lea's (1907) later description of *E. gymnostictus* (a synonym of *Ctenaphides maculatus*) and a new *Ctenaphides* recorded by Zimmerman (1994). A wealth of information on these authors and their collections is contained in Zimmerman's (1993) superb annotated bibliography.

The first *Aporhina* was collected on the *Astrolabe* expedition on Waigeo, the large north-western island of New Guinea (Boisduval, 1835), and two further species from the same area were described by Snellen van Vollenhoven (1866). Blackburn (1894) first recorded *Aporhina* from Australia, and Faust (1892) started the description of numerous species of the genus from the former German colony of "Kaiser-Wilhelms-Land", in north-eastern New Guinea, which was continued by Heller (1895, 1896, 1897, 1901, 1905, 1910, 1925), Wagner (1912b) and Györfyi (1917). Sixteen of the 27 species-group taxa of New Guinean *Aporhina* were described from this territory, and thirteen of these from the Huon Peninsula between Astrolabe Bay (Bongu) and the Sattelberg (Finschhafen), thus creating the impression of an unusually large species diversity existing in this region. However, 19 of these 27 taxa are based on single specimens, and often on mere differences in the development of the elytral spines and setal patches, and the species diversity of this area appears as much overemphasised as that of the rest of New Guinea is underrepresented among the described species. The exploration of the New Guinean eurhynchine fauna is still very much in its infancy.

Later workers focussed on the classification (Wagner 1909, 1912a, Crowson, 1955, Voss, 1961, Kissinger, 1968, Morimoto, 1976), morphology (Calder, 1898, 1990, Thompson, 1992, Wanat, 2001) and phylogenetic position (Kuschel, 1995, Oberprieler, 2000, Wanat, 2001, Marvaldi *et al.*, 2002) of the Eurhynchinae. Froggatt (1896) and Oberprieler (2000) provided the only accounts of their biology, and Kuschel *et al.* (1994) described a fossil form (Fig. 2). The most comprehensive modern treatment of the Eurhynchinae is Zimmerman's (1994) detailed and copiously illustrated review of the Australian fauna, which covers nomenclature and taxonomy as well as morphology and classification and represents the first major advance in our knowledge of the actual fauna in more than 100 years. By contrast, knowledge of the New Guinean fauna has not progressed at all and remains at the haphazard and incomplete descriptive level it had reached by 1925, only Voss (1961) attempting a cursory and very superficial grouping of the New Guinean *Aporhina* species.

CLASSIFICATION

The checkered classificatory position of the Eurhynchinae has been discussed by Zimmerman (1994), Oberprieler (2000) and Wanat (2001). Essentially it has swung between inclusion in Apioninae or Brentinae or a position between these two groups, and, depending on the status afforded to the latter, its rank varied from tribe to family. The affinity of the eurhynchines to the Brentidae *sensu lato* has been demonstrated on the basis on numerous characters, including molecular ones (Marvaldi *et al.*, 2002); however, Wanat's (2001) recent cladistic analysis produced a position adelphic to (Curculionidae + Brentidae) + Apionidae. Further studies, and combinations of data sets in phylogenetic analyses, are required to elucidate their proper classificatory position and relationships, but currently a subfamilial placement as one of the primitive, relictual groups of Brentidae *sensu lato* - alongside Microcerinae and probably Ithycerinae (Oberprieler, 2000, Marvaldi *et al.*, 2002) - seems the most satisfactory position.

The early occasional inclusion of *Ctenaphides* in Belidae has been dismissed (Zimmerman, 1994), and most recent classifications also exclude *Cylas* Latreille from Eurhynchinae, as had frequently been proposed in the past. Alonso-Zarazaga & Lyal (1999) included in Eurhynchinae also the Madagascan *Lispothorium* Faust, which was mostly placed in Cyladinae but moved to Myrmacicelinae by Wanat (2001). The inclusion in Eurhynchinae of the southern African fossil *Orapaeus* Kuschel & Oberprieler has been questioned by Zimmerman (1994), but no convincing alternative classification has thus far been proposed for this fossil form.

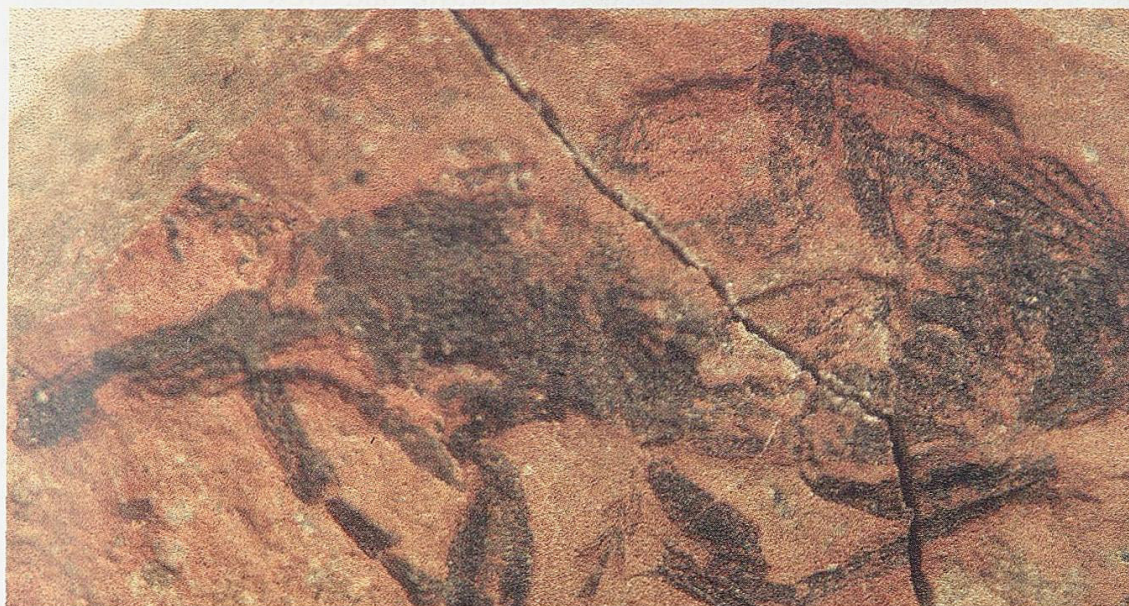


Fig. 2 - *Orapaeus cretaceus* Kuschel & Oberprieler, holotype: Cenomanian of Orapa, Botswana (photo by R. Rayner).

NOMENCLATURE

The nomenclature of both the family-group name and the type genus has been embroiled in a nomenclatural tangle dating back to the beginning of the nineteenth century, and although the use of the names *Eurhynchus* and *Eurhynchinae* has been quite stable and consistent since Schoenherr (1833) and Lacordaire (1863), the controversy was reopened when Kissinger (1968) reverted to Kirby's (1819a) original spelling *Eurhinus*. E. C. Zimmerman and R. T. Thompson researched the full complexities of the situation and in 1983 presented the case to the International Commission on Zoological Nomenclature, which resolved the conundrum by a ruling published in Opinion 1352 of 1985. Relevant details and references are given by Zimmerman (1994: 257-258) and Alonso-Zarazaga & Lyal (1999: 56-57).

Aporhina has also been plagued by nomenclatural difficulties because Boisduval (1835) did not furnish a proper description for it. The early authors consequently regarded it as an unavailable name and used its junior synonym, *Chalcocybebus*, as the valid name for the genus. Kissinger (1968) and Zimmerman (1994) reestablished the validity of *Aporhina* under Art. 12.2.2.6. of the ICZN (combined generic and specific description before 1931), but did not formally transfer all described species names to *Aporhina*. This is rectified here.

DISTRIBUTION

The *Eurhynchinae* are restricted to Australia and New Guinea. In Australia, *Ctenaphides* is endemic to the south-western part of the country, but because of the paucity of records its proper distribution remains very poorly known. *Eurhynchus* occurs on the eastern side of the continent from Tasmania and South Australia into northern Queensland, in forested habitats from sea level to higher elevations in the mountains of the Great Dividing Range (Fig. 3). *Aporhina* is in Australia restricted to coastal Queensland and northern New South Wales, occurring also at altitudes around 1000 m (Fig. 4), but in New Guinea it is widespread and diverse. Most of the New Guinean records are from coastal areas (Fig. 5), but this may reflect collecting bias rather than an actual distribution pattern. The New Guinean fauna is very poorly explored and its taxonomy inadequately studied, featuring species allegedly distributed across the entire island in numerous subspecies (*A. bispinosa*) next to others known from single localities. It appears that the genus does not extend westwards into the Moluccas.



Fig. 3 - Habitat of *Eurhynchus scabrior* (Kirby): Brindabella Mts., Canberra, A.C.T., Australia (photo by K. Pullen).

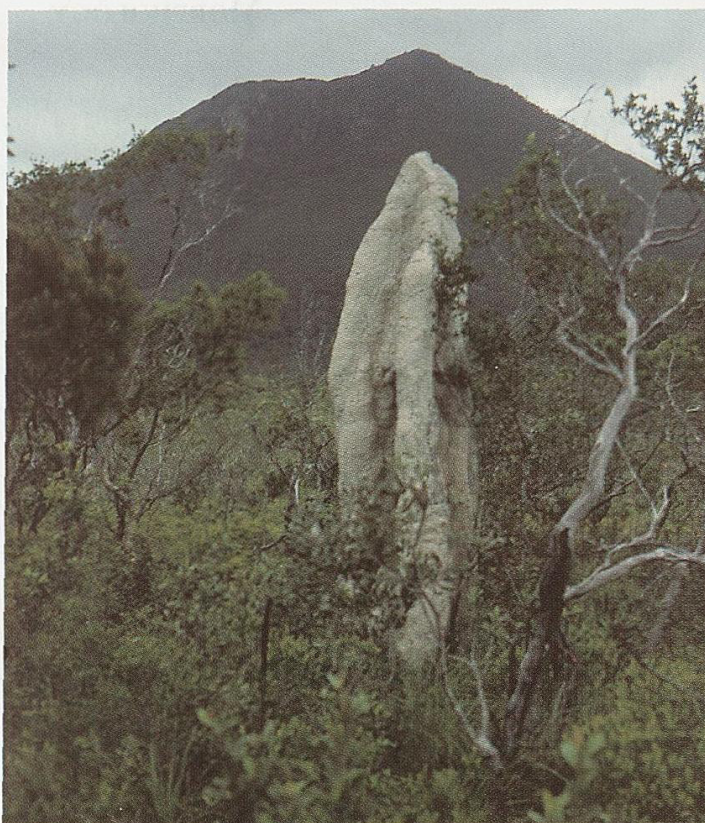


Fig. 4 - Habitat of *Aporhina australis* (Heller): Mount Tozer, northern Queensland, Australia (photo by R. Oberprieler).



Fig. 5 - Habitat of several *Aporhina* spp. described from Astrolabe Bay/Bongu: Ohu village, Madang, Papua New Guinea (photo by G. Setliff).

MORPHOLOGY

The structural characters of the Eurhynchinae have been studied by various authors, although rarely comprehensively. Wagner (1912a) discussed the main taxonomic characters of the group and its three extant genera, Crowson (1955) and Morimoto (1976) illustrated the mouthparts and metendosternite, Kissinger (1968) the tibial spurs and external male and female genitalia, Calder (1989, 1990) the alimentary canal, nervous system and internal genitalia, and Thompson (1992) the abdominal venter and male sternite 8. Zimmerman (1994) gave the most comprehensive account of the eurhynchine morphology, and Wanat (2001) compared an extensive range of detailed imaginal characters with those of other higher weevils, mainly Apioninae. Oberprieler (2000) described the larvae and assessed their salient features in relation to other phylogenetically critical brentid taxa. The main imaginal characters of the group were included in phylogenetic analyses of weevils by Kuschel (1995), Wanat (2001) and Marvaldi *et al.* (2002).

Within the Brentidae, the Eurhynchinae exhibit a large set of primitive characters (such as tibial spurs, short trochanters, loosely segmented antennal clubs, complete suture between ventrites 1 and 2) but are characterised by some unique apomorphies (such as emarginated protibial apex, submedially inserted labial palps, bar-like sclerite in membrane of ovipositor), see Zimmerman (1994: 238-239) and Wanat (2001: 360).

BIOLOGY

The biology of the Eurhynchinae remains poorly known. For more than a century, Froggatt's (1896) report of the larva of *Eurhynchus laevior* tunnelling in stems of *Persoonia lanceolata* (Proteaceae) remained the only record of the immature stages and larval hosts for the entire group, until Oberprieler (2000) rediscovered the *E. laevior* larva (Fig. 6) and fully described it and its biology. In the same paper he also described the larva of *Aporhina australis*, which was then known from a single specimen and remains the only *Aporhina* larva discovered thus far. In 2002, Jack Hasenpusch (Innisfail, Qld.) obtained more larvae of *A. australis* (Figs 8, 9) by sleeving females on suitable stems of its host, *Litsea leefiana* (Lauraceae), and following the development of the larval galls (Fig. 7) over several months. Gregory Setliff (Madang, Papua New Guinea) recently collected *Aporhina* nr. *nitens* in Papua New Guinea also on a *Litsea* sp. and reports the adults feeding of leaves of this plant and of *L. timoriana* in captivity. He also found empty galls in stems of *L. timoriana* that are very similar to the galls induced by *A. australis* on *L. leefiana*. Alexander Riedel (Germany) collected *Aporhina* nr. *australis* in Papua New Guinea on *Cryptocarya* (Oberprieler, 2000), and thus it appears that Lauraceae, and *Litsea* in particular, may be the main larval hosts of *Aporhina*.

Eurhynchus other than *E. laevior* is frequently collected on *Eucalyptus* (Oberprieler, 2000), and it appears that this genus and perhaps related Myr-



Fig. 6 - *Eurhynchus laevior* (Kirby): submature larva outside tunnel it excavated in stem of *Persoonia levis*; Termeil, N.S.W., Australia (photo by D. Rentz).

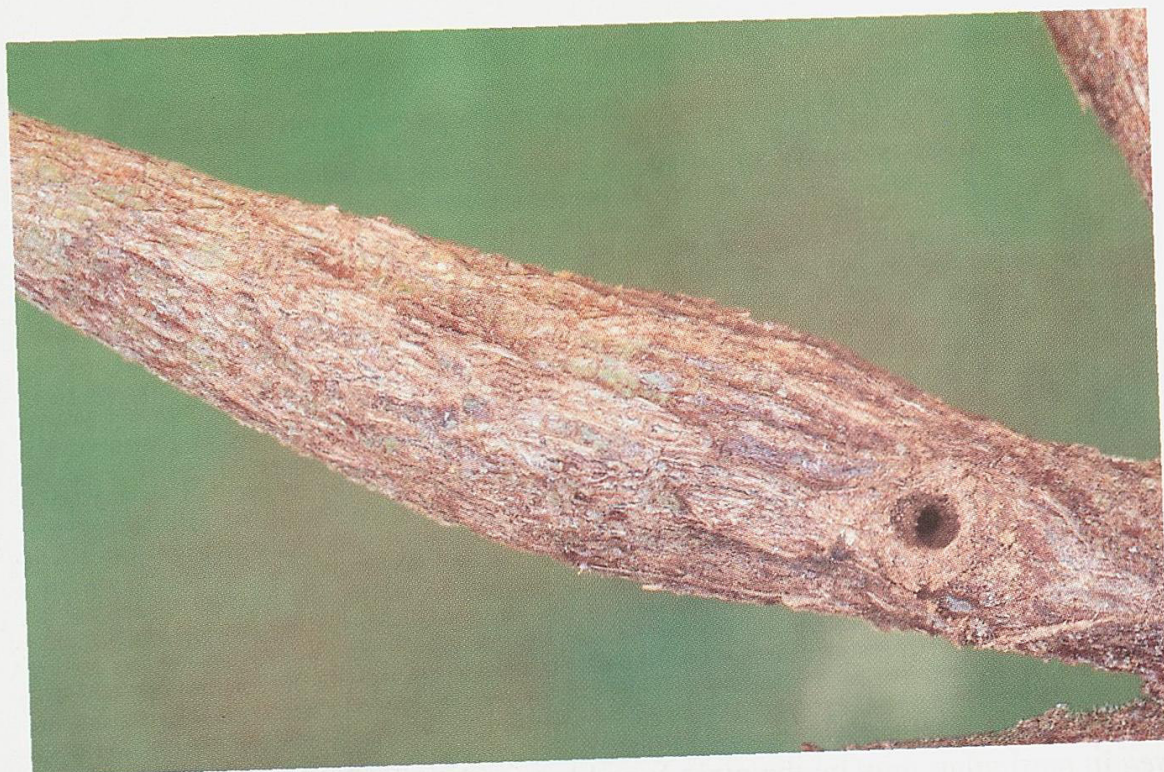


Fig. 7 - *Aporhina australis* (Heller): larval gall in stem of *Litsea leefiana* (Lauraceae); Innisfail, Qld, Australia (photo by R. Oberprieler).

taceae may serve as larval hosts for these species. All available (but putative) host records for *Ctenaphides* are of Proteaceae (*Banksia*, *Dryandra*). The discovery of the larvae of this genus and of other species of *Eurhynchus* and *Aporhina* will add significant information about the biology of the group and ascertain whether all larvae are stem borers characteristically expelling frass through a ventral opening in the larval tunnel.

EVOLUTIONARY HISTORY

The large suite of primitive characters of the Eurhynchinae and their restricted distribution in Australia and New Guinea indicate that the group is a relictual one and of considerable age. No fossils closely similar to the extant genera are known, but Kuschel *et al.* (1994) assigned the fossil genus *Orapaeus* (Fig. 2) from the early Late Cretaceous of Orapa, Botswana, to Eurhynchinae because of character similarities of the antennae and legs. A similar, undescribed fossil from the late Early Cretaceous (Aptian) Santana Formation in Ceara, Brazil, has been tentatively placed in the group as well (V.V. Zherikhin, pers. comm. 1999). Both the age and geographical location of these fossils is reconcilable with the indicated early origin of the Eurhynchinae, and it appears that they represent remnants of an ancestral brentid stock that either never existed in the northern hemisphere or became extinct there.



Fig. 8 - *Aporhina australis* (Heller): larva in opened gall; Innisfail, Qld, Australia (photo by R. Oberprieler).



Fig. 9 - *Aporhina australis* (Heller): larva in opened gall, enlarged; Innisfail, Qld, Australia (photo by R. Oberprieler).

Both recorded host plant families of the Eurhynchinae, Lauraceae and Proteaceae, are relatively primitive families in the angiosperms, the former grouping in the Magnoliidae (related to the monocots) and the latter at the base of the eudicots (Angiosperm Phylogeny Group, 2003). Neither Lauraceae nor Proteaceae are recorded in the fossil flora of Orapa (Rayner *et al.*, 1997), but Magnoliidae and some eudicot subclasses were identified from this unique, heavily angiosperm-dominated flora (Kuschel *et al.*, 1994). It seems that the extant eurhynchines may also preserve an ancient type of brentid host association and life style.

ABBREVIATIONS

HT	Holotypus
LT	Lectotypus
PT	Paratypus
ST	Syntypus

The codens of the institutions holding type material of Eurhynchinae follow Samuelson *et al.* (2002):

ANIC	Australian National Insect Collection, Canberra, Australia
BMNH	The Natural History Museum, London, United Kingdom
BPIPR	Bernard Price Institute for Palaeontological Research, University of the Witwatersrand, Johannesburg, South Africa
DEI	Deutsches Entomologisches Institut, Eberswalde Finow (formerly Berlin-Dahlem), Germany
HNHM	Hungarian Natural History Museum, , Budapest, Hungary
ISNB	Institut Royal des Sciences Naturelles de Belgique, Brussels, Belgium
MNHN	Museum National d'Histoire Naturelle, Paris, France
MSNM	Museo Civico di Storia Naturale, Milano, Italy
NHRS	Naturhistoriska Riksmuseet, Stockholm, Sweden
RMNH	Nationaal Natuurhistorische Museum ("Naturalis"), Leiden, Netherlands
SAM	South Australian Museum, Adelaide, Australia
SMTD	Staatliches Museum für Tierkunde, Dresden, Germany
ZMHB	Museum für Naturkunde, Humboldt-Universität, Berlin, Germany

CHECKLIST

EURHYNCHINAE Lacordaire, 1863

Eurhynchides Lacordaire, 1863: 527.

Eurhynchinae Lacordaire: Pascoe, 1870a: 204.

Eurhinini Kissinger, 1968: 10; syn. by validation of Eurhynchinae Lacordaire, 1863 by ICZN Opinion 1352 (1985)

Eurhynchus Kirby, 1828

Eurhinus Kirby, 1819a: 427.

Eurhynchus Kirby (in Kirby & Spence, 1828: 324) (replacement name for *Eurhinus* Kirby, 1819 nec Illiger, 1807, a junior homonym as ruled by ICZN Opinion 1352, September 1985).

Type species: *Eurhinus scabrior* Kirby, by subsequent designation (Schoenherr, 1833: 248).

Type species: *Eurhinus scabrior* Kirby, 1819.

Distribution. Eastern Australia.

acanthopterus Boisduval, 1835

acanthopterus Boisduval, 1835: 308, pl. 7 fig. 17.

tetracanthus Boheman, 1839: 363; syn. Pascoe, 1873: 281.

fulvofasciatus Blanchard, 1849: 143 (*Eurhinus*); syn. Zimmerman, 1994: 259.

bellicosus Boheman, 1859: 119; syn. Zimmerman, 1994: 259.

scapularis Pascoe, 1870a: 204; syn. Zimmerman, 1994: 259.

Type depositories. *E. acanthopterus*: ISNB (HT); *E. tetracanthus*: NHRS (HT); *E. fulvofasciatus*: MNHN ('LT'); *E. bellicosus*: NHRS (HT ♀); *E. scapularis*: BMNH (HT ♂).

Distribution. South-eastern Australia, from Tasmania through Victoria and New South Wales into Queensland (type locality of *acanthopterus* given only as Nouvelle Hollande by Boisduval but interpreted as Hobart by Zimmerman (1994: 259), of *bellicosus*: Sydney, of *tetracanthus* and *fulvofasciatus*: prob. Sydney, of *scapularis*: Queensland).

Hosts. *Eucalyptus* (Myrtaceae) (Oberprieler, 2000: 758).

Notes. For details of the types see Zimmerman (1994: 259, 261). He examined a type of *fulvofasciatus* labelled as lectotype by D. G. Kissinger, but Kissinger did not publish such a designation and the lectotype designation is referable to Zimmerman (1994). However, it is not evident from Blanchard's description that he had more than one specimen before him, and the specimen may actually be the holotype.

***laevior* (Kirby, 1819)**

laevior Kirby, 1819a: 429, pl. 22 fig. 8 (*Eurhinus*).

laevior (Kirby, 1819) (*Eurhynchus*): Gyllenhal, 1833: 248.

muricatus Kirby, 1819b: 468 (*Eurhinus*).

muricatus (Kirby, 1819) (*Eurhynchus*): Schoenherr, 1833: 249; syn. Zimmerman, 1994: 263.

Type depositories. Types of both *laevior* and *muricatus* apparently lost (Zimmerman, 1994: 263).

Distribution. South-eastern Australia, seemingly only coastal New South Wales (type localities: prob. Sydney).

Hosts. *Persoonia lanceolata* (Proteaceae) (Froggatt, 1896), *P. levis*, *P. myrtilloides* (Oberprieler, 2000: 757).

Notes. For details of the types see Zimmerman (1994: 263). The life history and larva were described by Oberprieler (2000).

***quadridens* Erichson, 1842**

quadridens Erichson, 1842: 186.

Type depository. ZMHB (HT ♂).

Distribution. South-eastern Australia, from Tasmania (type locality: Woolnorth area) through Victoria and South Australia to New South Wales and Queensland (Zimmerman, 1994: 268).

Hosts. *Eucalyptus* (Myrtaceae) (Oberprieler, 2000: 758).

Notes. For details of the type and other old specimens see Zimmerman (1994: 268).

***quadrituberculatus* Boheman, 1839**

quadrituberculatus Boheman, 1839: 361.

quadrinodosus Erichson, 1842: 186; syn. Zimmerman, 1994: 268.

Type depositories. *E. quadrituberculatus*: NHRS (HT ♂, PT); *E. quadrinodosus*: ZMHB (HT ♂).

Distribution. South-eastern Australia, from Tasmania (type locality of *quadrinodosus*: Woolnorth area) through South Australia and New South Wales (type locality of *quadrituberculatus*: prob. Sydney) to Queensland (Zimmerman, 1994: 268).

Hosts. *Eucalyptus* (Myrtaceae) (Tepper, 1887: 34).

Notes. For details of the type and other old specimens see Zimmerman (1994: 268, 270). He reports examining the holotype of *quadrituberculatus* and seeing "other specimens [that] have the squamules yellow". Boheman described such (a) specimen(s) as "var. β ", so must have had more than one specimen before him. Although he did not specifically designate a holotype, the specimen with white squamules, as referred to in the diagnosis and description, should be regarded as the holotype (regardless of the "Typus" label attached to its pin).

***scabrior* (Kirby, 1819)**

scabrior Kirby, 1819a: 428 (*Eurhinus*).

scabrior (Kirby, 1819) (*Eurhynchus*): Schoenherr, 1833: 248.

Type depository. BMNH (ST).

Distribution. South-eastern Australia, New South Wales, Australian Capital Territory and Victoria (type locality: prob. Sydney).

Hosts. *Eucalyptus* (Myrtaceae) (Oberprieler, 2000: 758).

Notes. Zimmerman (1994: 263) reports that the type material is in the BMNH but does not give the number of specimens or indicate whether a lectotype has been designated.

Ctenaphides Pascoe, 1870

Ctenaphides Pascoe, 1870b: 476 (not p. 428, as cited by Zimmerman, 1994: 246).

Type species: *Ctenaphides porcellus* Pascoe, by monotypy.

Type species: *Ctenaphides porcellus* Pascoe, 1870.

Distribution. Western Australia.

Notes. Lea (1907: 165) expressed doubts about the distinctness of *Ctenaphides* from *Eurhynchus* and Kissinger (1968: 10) synonymised the two generic names, but Zimmerman (1994: 240) restored *Ctenaphides* as a valid genus. Apart from the two described species, Zimmerman (1994: 255) records another undescribed one known from a single, damaged male.

maculatus (Pascoe, 1873)

maculatus Pascoe, 1873: 281 (*Eurhynchus*).

maculatus (Pascoe, 1873) (*Ctenaphides*): Lea, 1918: 270.

gymnostictus Lea, 1907: 164, syn. Lea, 1918: 270.

Type depositories. *E. maculatus*: BMNH (HT ♀); *E. gymnostictus*: HT ♀ apparently lost (Zimmerman, 1994: 254).

Distribution. South-western Australia (type localities - *maculatus*: Swan River, *gymnostictus*: King George's Sound, = Albany).

Hosts. Probably *Dryandra* and other Proteaceae.

Notes. For details of the types and other old specimens see Zimmerman (1994: 254). It is a rare species, known from only about a dozen specimens. A female was collected on *Dryandra* in the Fitzgerald River National Park in 1985 by C. Reid and P. Gullan (ANIC).

porcellus Pascoe, 1870

porcellus Pascoe, 1870b: 477, pl. 18 fig. 10.

porcellus (Pascoe, 1870) (*Eurhinus*): Kissinger, 1968: 10.

porcellus (Pascoe, 1870) (*Eurhynchus*): Calder, 1989: 1211.

Type depository. BMNH (ST).

Distribution. South-western Australia (type locality: Champion Bay, = Geraldton).

Hosts. Seemingly *Banksia speciosa* (Proteaceae) (Calder, 1989) and perhaps other Proteaceae.

Notes. Pascoe described the species from three males obtained from F. H. DuBoulay, one remaining in his collection (in BMNH). The locations of the other two are not recorded, and the one in Pascoe's collection has not been designated as lectotype. It is a rare species, known only from a handful of specimens.

Aporhina Boisduval, 1835

Aporhina Boisduval, 1835: 310.

Type species: *Aporhina bispinosa* Boisduval, by monotypy.

Chalcocybebus Snellen van Vollenhoven, 1866: 224; syn. Faust, 1892: 205, footnote 11.

Type species: *Chalcocybebus nitens* Snellen van Vollenhoven, by subsequent designation (Zimmerman, 1994: 241).

Type species: *Aporhina bispinosa* Boisduval, 1835.

Distribution. Australia, New Guinea.

Notes. *Aporhina* was not explicitly described by Boisduval and therefore treated as an unavailable name by Lacordaire (1863: 527) and subsequent workers, until Kissinger (1968) a century later restored it as an available and valid name, though without explanation. Only Zimmerman (1994) later stated the reason (combined generic and specific description). In resurrecting *Aporhina*, Kissinger (1968: 11) also newly synonymised *Chalcocybebus* with it, but overlooked that Faust (1892) had already proposed this synonymy much earlier, except that he and later workers used *Chalcocybebus* as the valid name for the genus. Lacordaire (1863) had treated *Aporhina* as a synonym of *Eurhynchus*, and Gemminger & von Harold (1871: 2457) also listed *Chalcocybebus* as a (new) synonym of *Eurhynchus*, but Faust (1892) restored *Chalcocybebus* as a distinct genus, with *Aporhina* in synonymy.

Voss (1961: 432) stated that there are specimens in the HNHM labelled as types of *Chalcocybebus nigripes* Györffy, *C. politus* Györffy, *C. biroi* Györffy, *C. magdalenae* Györffy and *C. similis* Györffy, but only the last three species names have been validated (Györffy, 1917 - the last as *C. assimilis*).

***alboguttata* (Snellen van Vollenhoven, 1866) comb. n.**

alboguttatus Snellen van Vollenhoven, 1866: 226 (*Chalcocybebus*).

alboguttatus (Snellen van Vollenhoven, 1866) (*Eurhynchus*): Gemminger & von Harold, 1871: 2457, syn. of *bispinosa* Boisduval.

Type depository. RMNH (HT ♂).

Distribution. New Guinea: West Papua (type locality: Salawati Isl.).

Hosts. Unknown.

Notes. Faust (1892: 205) restored *A. alboguttata* as a valid species from synonymy with *A. bispinosa* (Gemminger & von Harold 1871). Heller (1896) examined the single type and recorded additional specimens from north-western New Guinea, incl. Waigeo Isl.

***aspericollis* (Heller, 1910) comb. n.**

aspericollis Heller, 1910: 27 (*Chalcocybebus*).

Type depository. SMTD (HT ♀).

Distribution. New Guinea: Papua New Guinea (type locality: Torricelli Mts, 640 m).

Hosts. Unknown.

Notes. The single type is from the Schlaginhaufen collection, in the SMTD.

***assimilis* (Györfi, 1917) comb. n.**

assimilis Györfi, 1917: 276, fig. 1 (*Chalcocybebus*).

Type depository. SMTD (HT ♀).

Distribution. New Guinea: Papua New Guinea (type locality: Camp Walsh R., = Kemp Welch River, at present-day Kalo, south-east of Port Moresby).

Hosts. Unknown.

Notes. Seemingly described from a single female collected by Emil Weiske (see Heller, 1901: 1-2, for information on the collector), but Voss (1961: 432) indicates having seen a specimen from the HNHM labelled as type of "*similis*" Györfy.

***australis* (Heller, 1896)**

bispinosus australis Heller, 1896: 19 (*Chalcocybebus*).

australis (Heller, 1896) (*Eurhynchus*): Lea, 1908: 158.

australis (Heller, 1896) (*Aporhina*): Zimmerman, 1994: 246.

Type depository. BMNH (4 ST)

Distribution. North-eastern Australia (coastal Queensland and Torres Strait Islands, type locality: "Geraldton", = Innisfail).

Hosts. *Litsea leefiana* (Lauraceae) (Oberprieler, 2000).

Notes. Zimmerman (1994: 246) states that Lea (1908), in raising *australis* from a subspecies of *A. bispinosus* to species level, had examined the type of *australis*, but Lea (1908: 158, footnote 1) only reports having received a specimen from Heller, rather than any of the four syntypes which are, according to Heller (1896: 19), in the BMNH (ex Tring Museum). In Lea's collection (SAM) there is a male labelled 'N. Queensland / E. Weiske' and bearing a determination label by Heller; this is evidently the specimen sent to Lea by Heller but, not being labelled "Geraldton", it is unlikely to be a type. Also, Heller's paper is dated April 1896 and Weiske only collected in Queensland from late 1896 to early 1897 (Heller, 1901: 1). A lectotype remains to be designated for the species. It was first recorded from Australia by Blackburn (1894) as *Eurhynchus bispinosus*. The life history and larva were described by Oberprieler (2000).

***bennigseni* (Wagner, 1912) comb. n.**

bennigseni Wagner, 1912b: 310 (*Chalcocybebus*).

Type depository. DEI (HT ♀).

Distribution. New Guinea: Papua New Guinea (type locality: Kani Mts.).

Hosts. Unknown.

Notes. Described from a single female collected by R. von Bennigsen, erstwhile governor of Kaiser-Wilhelms-Land.

***biroi* (Györffy, 1917) comb. n.**

biroi Györffy, 1917: 277, fig. 2 (*Chalcocybebus*).

Type depository. HNHM (HT ♀).

Distribution. New Guinea: Papua New Guinea (type locality: Sattelberg).

Hosts. Unknown.

Notes. Seemingly described from a single female, collected by L. Biró in 1899.

***bispinosa* Boisduval, 1835**

bispinosa Boisduval, 1835: 310.

spinosa: Lacordaire, 1863: 528, footnote 3 (misspelling).

bispinosus (Boisduval) (*Eurhynchus*): Gemminger & von Harold, 1871: 2457.

splendidus Blanchard, 1849: 144 (*Eurhinus*).

splendidus (Blanchard, 1849) (*Eurhynchus*): Lacordaire, 1863: 528, footnote 3, syn.

superbus Heller, 1895: 12 (*Eurhynchus*).

superbus (Heller, 1895) (*Chalcocybebus*): Heller, 1896: 19, syn.

***aruensis* (Heller, 1896) comb. n.**

bispinosus aruensis Heller, 1896: 19 (*Chalcocybebus*).

***guttifera* (Heller, 1896) comb. n.**

bispinosus guttifer Heller, 1896: 19 (*Chalcocybebus*).

***intermedia* (Heller, 1896) comb. n.**

bispinosus intermedius Heller, 1896: 20 (*Chalcocybebus*).

***macrospilota* (Heller, 1925) comb. n.**

bispinosus ab. *macrospilotus* Heller, 1925: 287 (*Chalcocybebus*).

Type depositories. *E. bispinosus*: ISNB (HT ♀), *E. splendidus*: MNHN (HT), *E. superbus*: SMTD (HT ♂), *E. bispinosus aruensis*: RMNH (ST ♀), MSNM? (ST ♂), *E. bispinosus guttifer*: BMNH (HT ♀), *E. bispinosus intermedius*: MSNM? (HT ♀), *E. bispinosus macrospilotus*: SMTD (2 ST), DEI (18 ST).

Distribution. New Guinea: West Papua and Papua New Guinea (type lo-

calities - *bispinosa*: Waigeo Isl., *superba*: Bongu, *aruensis*: Aru, *guttifera*: Andai, *intermedia*: Port Moresby, *macrospilota*: Bongu/Sattelberg).

Hosts. Unknown.

Notes. The single female type of *bispinosa* was redescribed by Heller (1896: 22). It was collected by d'Urville on the *Astrolabe* expedition and originally in Dejean's collection. *Eurhinus splendidus* Blanchard, apparently described from a single male collected by L. Rousseau, was synonymised with *bispinosus* (as *Aporhina* "*spinosa*") by Lacordaire (1863). Its origin ("une des îles de l'océan Pacifique") is very imprecise but probably refers to one of the New Guinean islands. Unfortunately the name is a senior homonym of *splendida* Blackburn, 1894 in *Aporhina*, necessitating a replacement name for the latter (see below). The HT of *intermedius* and one ST of *aruensis* were received by Heller from J. R. H. Neervoort van de Poll's collection (Beukenstein Museum), the Curculionidae of which are now in the MSNM. However, the presence of these types in the MSNM has not been verified.

***exarmata* (Heller, 1905) comb. n.**

exarmatus Heller, 1905: 73 (*Chalcocybebus*).

Type depository. ZMHB (HT ♀).

Distribution. New Guinea: Papua New Guinea (type locality: Sattelberg).

Hosts. Unknown.

Notes. The single type is from the collection of G. Hauser, Erlangen, now in the ZMHB. It is a female without elytral spines.

***granosispina* (Heller, 1925) comb. n.**

granosispina Heller, 1925: 289 (*Chalcocybebus*).

Type depository. DEI (HT ♀).

Distribution. New Guinea: Papua New Guinea (type locality: Bongu/Sattelberg).

Hosts. Unknown.

Notes. Described from a single female collected by Carl Wahnes.

***helleri* (Wagner, 1912) comb. n.**

helleri Wagner, 1912b: 312, fig. 2a, b (*Chalcocybebus*).

Type depository. DEI (HT ♀).

Distribution. New Guinea: Papua New Guinea (type locality: Kani Mts.).

Hosts. Unknown.

Notes. Wagner's (1912b: 313) figs. 2a and 2b are labelled as *C. bennigseni* but included in the description of *C. helleri* and compared with figs. 1a, b (*C. insignis* Heller, which is very similar to *C. helleri*) and refer to *C. helleri*, not *C. bennigseni*. The single type is from the Bennigsen collection, in the DEI.

***inermis* (Heller, 1897) comb. n.**

inermis Heller, 1897: 3 (*Chalcocybebus*).

Type depository. BMNH (HT ♀).

Distribution. New Guinea: Papua New Guinea (type locality: between Alexander and Nisbet Mts., in the Owen Stanley Range, north-west of Port Moresby).

Hosts. Unknown.

Notes. This species is also described from a single female without elytral spines, collected by Anthony in 1896.

***insignis* (Heller, 1905) comb. n.**

insignis Heller, 1905: 72 (*Chalcocybebus*).

Type depository. ZMHB (HT ♂).

Distribution. New Guinea: Papua New Guinea (type locality: Sattelberg).

Hosts. Unknown.

Notes. The single type is from the collection of G. Hauser, Erlangen (see *A. exarmata*). Heller did not give a type locality for the species, but the holotype bears a "Sattelberg" label (Frisch, pers. comm.) and Wagner (1910, 1912a) also lists this as the (type) locality.

***levigata* (Heller, 1925) comb. n.**

levigatus Heller, 1925: 288 (*Chalcocybebus*).

Type depository. SMTD (2 ST), DEI (5 ST).

Distribution. New Guinea: Papua New Guinea (type locality: Bongu/Sattelberg).

Hosts. Unknown.

Notes. Described from seemingly 7 specimens of both sexes (leg. C. Wahnes) but no holotype designated.

***magdalenae* (Györffy, 1917) comb. n.**

magdalenae Györffy, 1917: 280, fig. 4 (*Chalcocybebus*).

Type depository. HNHM (ST).

Distribution. New Guinea: Papua New Guinea (type locality: Sattelberg).

Hosts. Unknown.

Notes. Described from both sexes but number of specimens not indicated and no holotype designated. The types were collected by L. Biró in 1899.

***massutei* (Heller, 1901) comb. n.**

massutei Heller, 1901: 16 (*Chalcocybebus*).

***interrupta* (Heller, 1901) comb. n.**

massutei var. *interrupta* Heller, 1901: 16 (*Chalcocybebus*).

Type depositories. *C. massutei*: SMTD (HT ♂); *C. massutei* var. *interrupta*: SMTD (HT ♀).

Distribution. New Guinea: Papua New Guinea (type localities – *massutei*: Aroa River (near Hisiu, north-west of Port Moresby), *interrupta*: Astrolabe Mts (near Birribi, east of Port Moresby)).

Hosts. Unknown.

Notes. Heller (1901: 1-2) gives a useful, brief account of the expeditions in New Guinea and Australia of the collector of the types, Emil Weiske.

***mesospila* (Györffy, 1917) comb. n.**

mesospilus Györffy, 1917: 279, fig. 3 (*Chalcocybebus*).

Type depository. HNHM (ST), DEI (1 ST).

Distribution. New Guinea: Papua New Guinea (type locality: Sattelberg).

Hosts. Unknown.

Notes. Described from both sexes but no holotype designated. The types were collected by L. Biró in 1899.

***mutica* (Heller, 1925) comb. n.**

muticus Heller, 1925: 288 (*Chalcocybebus*).

Type depository. SMTD (ST ♂), DEI (ST ♀).

Distribution. New Guinea: Papua New Guinea (type locality: Bongu/Sattelberg).

Hosts. Unknown.

Notes. Described from a single pair collected by C. Wahnes, but no holotype designated.

***nitens* (Snellen van Vollenhoven, 1866) comb. n.**

nitens Snellen van Vollenhoven, 1866: 225, pl. 12 fig. 2 (*Chalcocybebus*).

***papuana* (Heller, 1896) comb. n.**

nitens papuanus Heller, 1896: 20 (*Chalcocybebus*).

Type depositories. *C. nitens*: RMNH (HT ♀); *C. nitens papuana*: SMTD (3 ST), MSNM? (5 ST).

Distribution. New Guinea: West Papua and Papua New Guinea (type localities - *nitens*: Waigeo Isl., *papuana*: Bongu).

Hosts. Probably *Litsea* (Lauraceae) (Setliff, pers. comm. 2003).

Notes. *Chalcocybebus nitens papuanus* var. *auratus* Heller, 1896: 20, named for 2 females from Humboldt Bay, is an infrasubspecific name and unavailable. Two of the ST of *papuana* in the SMTD are from Faust's collection, and the ones in the MSNM were collected by Rhode. For the location of the latter, see under *A. bispinosa* above.

***obtusispina* (Heller, 1925) comb. n.**

obtusispina Heller, 1925: 287 (*Chalcocybebus*).

Type depository. SMTD (2 ST), DEI (4 ST).

Distribution. New Guinea: Papua New Guinea (type locality: Bongu/Sattelberg).

Hosts. unknown.

Notes. Described from six specimens comprising both sexes (leg. C. Wahnes), but no holotype designated. One male has a malformed left antenna (Heller, 1925: 288).

***pulchra* Oberprieler, h. o. nomen novum**

splendidus Blackburn, 1894: 163 (*Eurhynchus*).

splendidus (Blackburn, 1894) (*Chalcocybebus*): Heller, 1896: 20.

splendida (Blackburn, 1894) (*Aporhina*): Zimmerman, 1994: 246.

pulchra Oberprieler, replacement name for *Aporhina splendida* Blackburn, 1894 (non Blanchard, 1849).

Type depository. BMNH (HT ♂).

Distribution. North-eastern Australia, mostly Queensland but also northern New South Wales (type locality: N. Queensland).

Hosts. Unknown.

Notes. *Eurhynchus splendidus* Blackburn, 1894 is a junior secondary homonym of *Eurhinus splendidus* Blanchard, 1849 when both placed in the same genus, *Aporhina*. Blackburn did not specify a precise type locality, but Heller (1896: 20) records having examined a male in the BMNH from Geraldton (= Innisfail, see under *A. australis*), which may be the holotype. A rare and pretty species known only from few specimens, which stronger sculpture and vestiture than other *Aporhina* species (noted by Heller 1896: 24).

richteri (Faust, 1892) **comb. n.**

richteri Faust, 1892: 205 (*Chalcocybebus*).

Type depository. SMTD (HT ♂).

Distribution. New Guinea (type locality: not specified).

Hosts. Unknown.

Notes. Heller (1896: 20) reports a pair collected by Rhode at Astrolabe Bay, which may also be the locality of Faust's type. The single type is from the collection Paul Richter.

Fossil Section

Orapaeus Kuschel & Oberprieler, 1994

Orapaeus Kuschel & Oberprieler (in Kuschel G., Oberprieler R. G., Rayner R. J., 1994: 139).

Type species: *Orapaeus cretaceus* Kuschel & Oberprieler, by original designation.

Distribution: Southern Africa.

***cretaceus* Kuschel & Oberprieler, 1994**

cretaceus Kuschel & Oberprieler (in Kuschel G., Oberprieler R. G., Rayner R. J., 1994: 140, figs. 1-11, 18-26).

Type depository. BPIPR (HT, PT).

Distribution. Botswana: Orapa, early Late Cretaceous (Cenomanian).

Hosts. Unknown.

Notes. The description is based on 10 specimens, all illustrated by Kuschel *et al.* (1994).

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LITERATURE

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Microcerini Lacordaire, 1863 (Coleoptera, Curculionoidea)

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ABSTRACT

The present taxonomic state of the Microcerini (Brachycerinae: Curculionidae: Coleoptera) is fully catalogued. A checklist provides the microcerine genera and species in alphabetical order, for each of which the original description reference, list of synonymies, depositories of primary and secondary types, and *distribution by country* is presented. All taxon names are alphabetically indexed and a full microcerine related bibliography is provided.

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INTRODUCTION

The Microcerini are comprised of three genera, i. e. *Microcerus* (with 23 extant species), *Episus* (with 42 extant species) and *Gyllenhalia* (with 2 extant species) and exhibit an exclusive subsaharan Africa distribution. The present taxonomic state of affairs is regarded to depict a fairly accurate nomenclatorial scenario within the Microcerini and, at most, only a few species in obscure collections still await description.

A historical review of microcerinae taxonomy, as well as the ecology, phylogeny and zoogeography of the group is discussed in Louw (1986). However, since the seminal publication by Kuschel (1995) on the higher classification of the Curculionoidea, the traditional 'Microcerinae' have become a topic of debate regarding their phylogenetic classification. Kuschel (1995) places the genera *Microcerus*, *Episus* and *Gyllenhalia* (as the tribe Microcerini) within Brachycerinae of the Curculionidae, mentioning the peculiarity of a short-scaped, non-geniculate antenna in *Microcerus*. He also goes on to regard the Brachycerinae as one of six lineages, and the most primitive, within the Curculionidae. Oberprieler (2000), however, questions this placement, arguing that closer examination of the larvae and adults of Microcerini in fact point to a close relationship to traditional Brentidae. In the larva this affinity is demonstrated in open frontal sutures (i. e. absence of a fronto-epicranial brachon and the antennal placement being fully on the frons), two-segmented legs, only two dorsal lobes on the abdominal segments and four Malpighian tubules. On the adults, in turn, this affinity comes to the fore mainly by the labial palps which are sunken into grooves, the eyes which are covered by a corneal layer, the orthocerous antennae (already pointed out by Kuschel (1995)), the abdominal ventrites which are stepped between 2 and 3 and the metendosternite which is a non-curculionid type, but resembles that of *Cylas*. Oberprieler (2000), however, goes on to stress the point that overall this placement of the microcerinines within the brentid clade is rather weakly supported and that alternatives closer to the apionines do also exist. This lack of confidence is demonstrated by the fact that the male microcerinine terminalia are not distinctly brentine, even though, as mentioned above, other adult features again favour the placement within the brentines, possibly in close proximity to the cyladinines. Therefore, the unquestionable phylogenetic classification of the Microcerini still awaits critical scrutiny across a full, broad suit of characters.

ABBREVIATIONS

HT	Holotypus
LT	Lectotypus
NT	Neotypus
PLT	Paralectotypus
PT	Paratypus
IRSNB	Institut Royal des Sciences Naturelles de Belgique, Brussels, Belgium
MNHN	Museum National d'Histoire Naturelle, Paris, France
MNHUB	Museum für Naturkunde der Humboldt-Universität, Berlin, Germany
MRAC	Musée Royal de l'Afrique Centrale, Tervuren, Belgium
MSNG	Museo Civico di Storia Naturale "Giacomo Doria", Genova, Italy
NCIP	National Collection of Insects, Pretoria, South Africa
NHM	The Natural History Museum, London, United Kingdom
NHMB	Naturhistorisches Museum, Basel, Switzerland
NHRS	Naturhistoriska Riksmuseet, Stockholm, Sweden
NMBS	National Museum, Bloemfontein, South Africa
NMBZ	National Museum, Bulawayo, Zimbabwe
NMNH	National Museum of Natural History, Washington D. C., USA
OXUM	Oxford University Museum of Natural History, Oxford, Great Britain
SAMC	South African Museum, Cape Town, South Africa
SMWN	State Museum, Windhoek, Namibia
TMSA	Transvaal Museum, Pretoria, South Africa
ZMUC	Zoological Museum, University of Copenhagen, Denmark
ZSM	Zoologische Staatssammlung, Munich, Germany



Fig. 1 - *Episus aculeatus* Labram & Imhoff: De Koei, Richtersveld, South Africa (photo courtesy of R. G. Oberprieler).

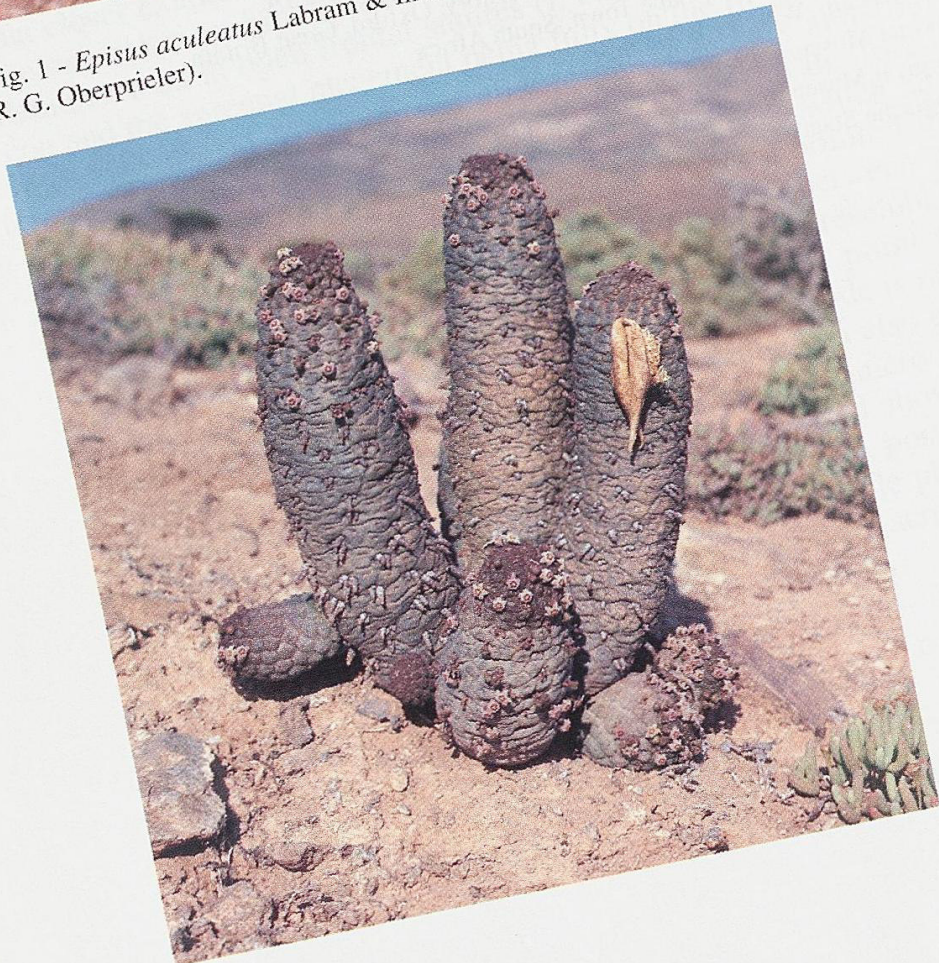


Fig. 2 - Habitat of *Episus aculeatus* Labram & Imhoff: De Koei, Richtersveld, South Africa (photo courtesy of R. G. Oberprieler).

CHECKLIST

MICROCERINAE Lacordaire, 1863

Microcerides Lacordaire, 1863: 20.

Microcerinae Jekel, 1875: 104.

Microcerini Bovie, 1909: 2.

Type genus: *Microcerus* Schoenherr, 1833.

Episus Billberg, 1820

Episus Billberg, 1820: 40 (nec Schoenherr, 1826).

Lagenisus Jekel, 1875: 105.

Type species: *Curculio rostratus* Fabricius, 1781 (by subsequent designation: Schoenherr, 1826).

Distribution. Africa, south of the Sahara.

aculeatus Labram & Imhoff, 1838

(Figs 1, 2, 4, 5)

aculeatus Labram & Imhoff, 1838: 26.

stricticollis Jekel, 1875: 119 (*Episus*); syn. Aurivillius, 1885: 12.

stricticollis: Aurivillius, 1887: 35 (*Episus aculeatus* ssp.); syn. Louw, 1986: 117.

Type depositories. *E. aculeatus*: NHMB (HT); *E. aculeatus stricticollis*: MSNG (HT).

Distribution. Namibia, South Africa.

angulicollis Aurivillius, 1885

angulicollis Aurivillius, 1885: 16.

Type depository. MNHUB (HT).

Distribution. Central and eastern South Africa.

***angusticollis* Fåhraeus, 1871**

(Fig. 3)

angusticollis Fåhraeus, 1871: 4.**Type depositories.** NHRS (HT); MRAC (PT).**Distribution.** Botswana, South Africa, Zimbabwe.***cognatus* Fåhraeus, 1871***cognatus* Fåhraeus, 1871: 5.**Type depository.** NHRS (HT).**Distribution.** Central and northern South Africa, southern Zimbabwe.***contractus* Fåhraeus, 1871***contractus* Fåhraeus, 1871: 6.*bohemani* Aurivillius, 1885: 12 (*Episus*); syn. Louw, 1986: 109.**Type depositories.** *E. contractus*: NHRS (HT); *E. bohemani*: unknown.**Distribution.** Angola, Botswana, Namibia, South Africa, Zimbabwe.***cyathiformis cyathiformis* Gyllenhal, 1833***cyathiformis* Gyllenhal, 1833: 376.*coenosus* Jekel, 1875: 107 (*Lagenisus*); syn. Aurivillius, 1885: 7.*caenosus*: Giachino, 1982: 407 (*Episus*) [lapsus].**Type depositories.** *E. cyathiformis*: unknown; *L. coenosus*: NHRS (HT).**Distribution.** Chad, northern Democratic Republic of the Congo (ex Zaire), Gambia, Nigeria, Senegal, Sudan.



Fig. 3 - *Episus angusticollis* Fåhraeus: Boshoff, Free State Province, South Africa (photo courtesy of R. G. Oberprieler).



Fig. 4 - Habitat of *Episus inermicollis* Gyllenhal, *E. occidentalis* Louw, *E. mendosus* Gyllenhal, *E. aculeatus* Labram & Imhoff and *Microcerus grisescens* Gyllenhal: Steinkopf, Namaqualand, South Africa (photo courtesy of R. G. Oberprieler).

cyathiformis dorsalis Fåhræus, 1871

dorsalis Fåhræus, 1871: 4.

wahlbergi Jekel, 1875: 109 (*Lagenisus*); syn. Louw, 1986: 49.

wahlbergi: Aurivillius, 1887: 14 (*Episus cyathiformis* ssp.).

parallelus Aurivillius, 1887: 14 (*Episus cyathiformis* ssp.); syn. Louw, 1986: 49.

paucidentatus Aurivillius, 1887: 15 (*Episus cyathiformis* ssp.); syn. Louw, 1986: 49.

Type depositories. *E. dorsalis*: NHRS (HT); *L. wahlbergi*: NHRS (HT); *E. cyathiformis parallelus*: NHRS (HT); *E. cyathiformis paucidentatus*: NHRS (HT).

Distribution. Southern Angola, Botswana, Namibia, central and northern South Africa, Zimbabwe.

cyathiformis tuberosus Gerstäcker, 1871

tuberosus Gerstäcker, 1871: 68.

Type depository. MNHUB (HT).

Distribution. Botswana, eastern Democratic Republic of the Congo (ex Zaire), Kenya, Malawi, north-eastern Mozambique, northern South Africa, Tanzania, Uganda, Zambia, Zimbabwe.

cylindrus Louw, 1986

cylindrus Louw, 1986: 78.

Type depositories. NHM (HT, PT); NMBS (PT), MNHN (PT); SAMC (PT); MNHUB (PT).

Distribution. Malawi, Mozambique, Tanzania, Zambia, Zimbabwe.

devylderi Aurivillius, 1885

devylderi Aurivillius, 1885: 10.

Type depository. NHRS (HT).

Distribution. Southern Angola, Botswana, Democratic Republic of the Congo (ex Zaire), Kenya, Malawi, Mozambique, northern Namibia, northern South Africa, Tanzania, Zimbabwe.

dolus Louw, 1986

dolus Louw, 1986: 147.

Type depository. NHM (HT, AT).

Distribution. South-eastern South Africa.

dregei Boheman, 1840

dregei Boheman, 1840: 595.

rostratus Olivier, 1790: 186 (*Brachycerus*) [nomen preoccupatum].

obliquus, Jekel, 1875: 113 (*Episus*); syn. Pape, 1910: 4.

punctatus Aurivillius, 1885: 15 (*Episus*); syn. Louw, 1986: 153.

Type depositories. *E. dregei*: NHM (HT, PT); *B. punctatus*: MNHN (HT); *E. obliquus*: unknown.

Distribution. Eastern and southern South Africa.

echinatus Boheman, 1840

echinatus Boheman, 1840: 594.

muricatus Boheman, 1840: 598 (*Episus*); syn. Louw, 1986: 156.

dubius Aurivillius, 1887: 50 (*Episus muricatus* ssp.); syn. Louw, 1986: 156.

Type depositories. *E. echinatus*: NHRS (HT); *E. muricatus*: NHRS (HT); *E. muricatus dubius*: MNHUB (HT, PT).

Distribution. Botswana, South Africa.

elongatus Aurivillius, 1885

elongatus Aurivillius, 1885: 10.

Type depository. IRSNB (HT).

Distribution. South-western South Africa.

endroedyi Louw, 1986

endroedyi Louw, 1986: 126.

Type depositories. TMSA (HT, PT); NMBS (AT, PT); NCIP (PT).

Distribution. Western South Africa.

exilis Louw, 1986

exilis Louw, 1986: 81.

Type depositories. TMSA (HT); SMWN (PT); MRAC (PT); ZSM (PT); NHM (PT); NCIP (PT); NMBS (PT); NMBZ (PT).

Distribution. Southern Angola, south-eastern Democratic Republic of the Congo (ex Zaire), northern Namibia, Zimbabwe.

fahraei Aurivillius, 1885

fahraei Aurivillius, 1885: 10.

funicularis Voss, 1974: 438 (*Episus fahraei* ssp.); syn. Louw, 1986: 75.

fahreusi: Hustache, 1935: 167 [lapsus].

Type depositories. *E. fahraei*: IRSNB (HT); *E. fahraei funicularis*: unknown.

Distribution. Southern Angola, southern Democratic Republic of the Congo (ex Zaire), northern Mozambique, northern Namibia, Tanzania, Zimbabwe.

fictus Gyllenhal, 1840

fictus Gyllenhal, 1840: 597.

t-album Jekel, 1875: 11 (*Episus*); syn. Louw, 1986: 151.

Type depositories. *E. fictus*: NHRS (HT); *E. t-album*: MSNG (HT).

Distribution. Central and eastern South Africa.

flexuosus Boheman, 1840

flexuosus Boheman, 1840: 599.

Type depository. NHRS (HT).

Distribution. Lesotho, central South Africa.

gibbosus Aurivillius, 1885

gibbosus Aurivillius, 1885: 16.

krebsi Aurivillius, 1887: 42 (*Episus gibbosus* ssp.); syn. Louw, 1986: 123.

Type depositories. *E. gibbosus*: MNHUB (HT); *E. gibbosus krebsi*: MNHUB (HT).

Distribution. Central South Africa.

hieroglyphicus Fåhræus, 1871

hieroglyphicus Fåhræus, 1871: 3.

Type depository. NHRS (HT, PT).

Distribution. Southern Mozambique, central and northern South Africa.

holmi Louw, 1986

holmi Louw, 1986: 112.

Type depositories. MNHUB (HT, PT); MNHN (PT); NHM (PT); SAMC (PT).

Distribution. Kenya, Sudan.

hopei Boheman, 1840

hopei Boheman, 1840: 600.

dohrni Aurivillius, 1885: 11 (*Episus*); syn. Louw, 1986: 131.

Type depositories. *E. hopei*: OXUM (HT); *E. dohrni*: MNHUB (HT).

Distribution. Southern Botswana, southern Mozambique, central South Africa.

hypocritus Gyllenhal, 1833

hypocrita Gyllenhal, 1833: 375.

cristatus Gyllenhal, 1840: 596 (*Episus*); syn. Aurivillius, 1885: 15.

Type depositories. *E. hypocrita*: NHRS (HT); *E. cristatus*: unknown.

Distribution. Central and eastern South Africa.

impressicollis Aurivillius, 1885

impressicollis Aurivillius, 1885: 11.

brevicollis nigrovittatus Aurivillius, 1887: 30 (*Episus*); syn. Louw, 1986: 94.

Type depositories. *E. impressicollis*: SAMC (NT); *E. brevicollis nigrovittatus*: NHRS (HT).

Distribution. South-western Angola, Botswana, southern Mozambique, Namibia, northern & central South Africa, western Zimbabwe.

inermicollis Gyllenhal, 1840

(Figs 4, 5)

inermicollis Gyllenhal, 1840: 590.

dentatus Boheman, 1840: 592 (*Episus*); syn. Aurivillius, 1885: 11.

gravidus Aurivillius, 1885: 11 (*Episus*); syn. Louw, 1986: 97.

brevicollis Jekel, 1875: 117 (*Episus*); syn. Louw, 1986: 97.

Type depositories. *E. inermicollis*: NHRS (HT); *E. dentatus*: NHRS (HT); *E. gravidus*: MNHUB (NT); *E. brevicollis*: MSNG (HT).

Distribution. Southern Angola, Botswana, Namibia, South Africa.

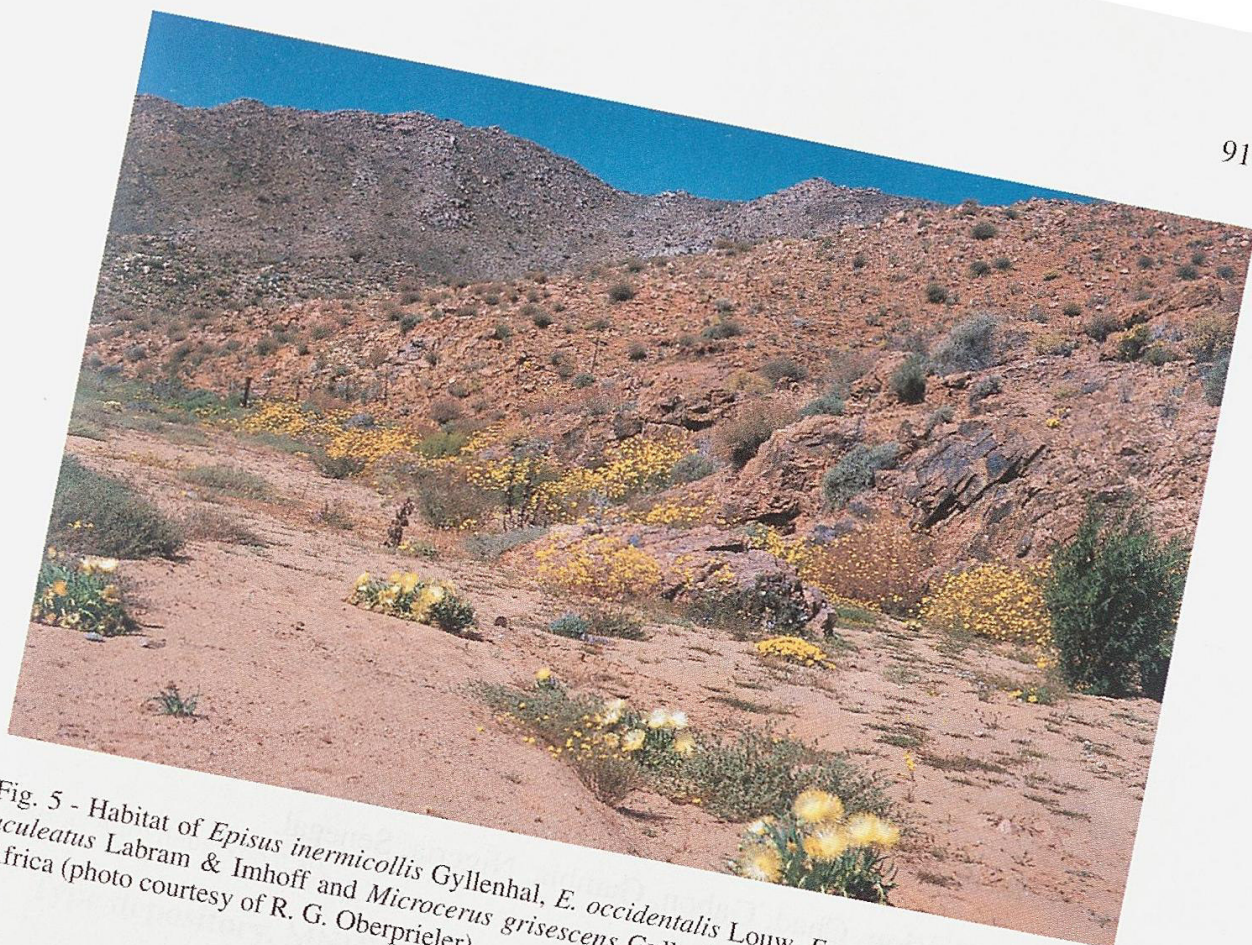


Fig. 5 - Habitat of *Episus inermicollis* Gyllenhal, *E. occidentalis* Louw, *E. mendosus* Gyllenhal, *E. aculeatus* Labram & Imhoff and *Microcerus grisescens* Gyllenhal: Gamoep, Namaqualand, South Africa (photo courtesy of R. G. Oberprieler).

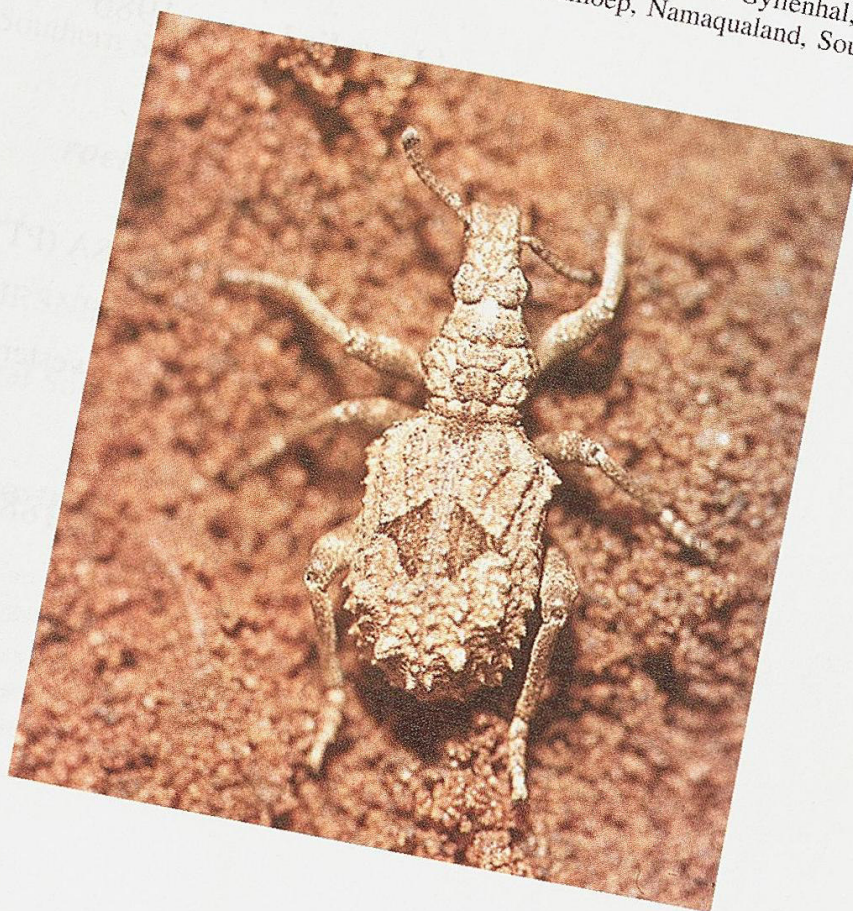


Fig. 6 - *Microcerus latipennis* Fåhræus: Pretoria, Gauteng Province, South Africa (photo courtesy of R. G. Oberprieler).

***mendosus* Gyllenhal, 1833**
(Figs 4, 5)

mendosus Gyllenhal, 1833: 377.

mandosus: Giachino, 1982: 407 (*Episus*) [lapsus].

Type depository. NHRS (HT).

Distribution. Western and southern South Africa.

***oberthuri* Aurivillius, 1885**

oberthuri Aurivillius, 1885: 12.

Type depository. MNHN (HT)

Distribution. Chad, Gabon, Gambia, Nigeria, Senegal.

***occidentalis* Louw, 1986**
(Figs 4, 5)

occidentalis Louw, 1986: 53.

Type depositories. NMBS (HT, AT, PT); TMSA (PT); NCIP (PT); SMWN (PT).

Distribution. South-western Namibia, north-western South Africa.

***opalinus* Aurivillius, 1885**

opalinus Aurivillius, 1885: 10.

Type depository. SAMC (HT).

Distribution. Southern South Africa.

penrithae Louw, 1986

penrithae Louw, 1986: 92.

Type depository. SMWN (HT).

Distribution. Angola.

planus Louw, 1986

planus Louw, 1986: 59.

Type depository. TMSA (HT).

Distribution. Western South Africa.

robustus Aurivillius, 1885

robustus Aurivillius, 1885: 10.

Type depository. MNHUB (HT, PT).

Distribution. Southern South Africa.

roelofsi Aurivillius, 1885

roelofsi Aurivillius, 1885: 14.

Type depository. IRSNB (HT).

Distribution. Central South Africa.

rostratus (Fabricius, 1781)

rostratus Fabricius, 1781: 194 (*Curculio*).

rostratus: Thunberg, 1799: 32 (*Brachycerus*); syn. Aurivillius 1887: 43.

thunbergi Boheman, 1840: 591 (*Episus*); syn. Aurivillius 1885: 13.

albulus Lacordaire, 1863: 21 (*Episus*); syn. Aurivillius 1885: 13.

quadrulifer Jekel, 1875: 115 (*Episus*); syn. Louw 1986: 140.

thunbergi: Giachino, 1982: 407 (*Episus*) [lapsus].

Type depositories. *C. rostratus*: NHM (HT); *E. quadrulifer*: MSNG (HT); *E. thunbergi*: unknown; *E. albulus*: unknown.

Distribution. Southern South Africa.

rotundicollis Aurivillius, 1885

rotundicollis Aurivillius, 1885: 13.

Type depository. SAMC (HT).

Distribution. South-western South Africa.

saritae Louw, 1986

saritae Louw, 1986: 120.

Type depositories. SMWN (HT, PT); TMSA (PT); SAMC (PT); NHM (PT); NMBS (PT).

Distribution. Southern Namibia, north-western South Africa.

simulator Dalman, 1833

simulator Dalman, 1833: 377.

peringueyi Aurivillius, 1885: 9 (*Episus*); syn. Louw, 1986: 66.

Type depositories. *E. simulator*: NHRS (HT); *E. peringueyi*: SAMC (HT).

Distribution. Southern South Africa.

spinosus ganglionicus Boheman, 1840

ganglionicus Boheman 1840: 603.

ganglianicus: Giachino, 1982: 407 (*Episus*) [lapsus].

Type depository. NHRS (HT).

Distribution. Central South Africa.

***spinosus nodicollis* Aurivillius, 1885**

nodicollis Aurivillius, 1885: 14.

Type depository. MNHUB (LT, PLT).

Distribution. Southern and eastern South Africa.

Notes. Lectotypes designated by Louw (1986).

***spinosus spinosus* Boheman, 1840**

spinosus Boheman, 1840: 602.

Type depository. MNHN (HT).

Distribution. Central South Africa.

***sputatilius* Gyllenhal, 1833**

sputatilius Gyllenhal, 1833: 378.

Type depository. NHRS (HT).

Distribution. South-eastern South Africa.

***tenuis* Fairmaire 1887**

tenuis Fairmaire, 1887: 310.

Type depository. MNHN (HT).

Distribution. Eastern Democratic Republic of the Congo (ex Zaire), Ethiopia, Kenya, Somalia, Tanzania, northern Zambia.

***truncatus* Boheman, 1845**

truncatus Boheman, 1845: 374.

Type depository. NHRS (HT).

Distribution. South-western South Africa.

tubervultus Louw, 1986

tubervultus Louw, 1986: 105.

Type depository. SAMC (HT).

Distribution. Southern South Africa.

westermanni Aurivillius, 1885

westermanni Aurivillius, 1885: 9.

Type depository. ZMUC (HT).

Distribution. Angola, Botswana, Namibia, South Africa.

whiteheadi Louw, 1986

whiteheadi Louw, 1986: 100.

Type depository. SAMC (HT).

Distribution. Southern South Africa.

Gyllenhalia Aurivillius, 1885

Gyllenhalia Aurivillius, 1885: 20.

Type species: *Curculio spectrum* Fabricius, 1781.

Distribution. Southern Africa.

crinita Marshall, 1959

crinita Marshall, 1959b: 392.

Type depositories. NHM (HT, PT); TMSA (PT).

Distribution. Southern Namibia, central-northern South Africa.

spectrum (Fabricius, 1781)

spectrum Fabricius, 1781: 194 (*Curculio*).

spectrum: Fabricius, 1792: 383 (*Brachycerus*).

Type depository. NHM (HT).

Distribution. Central and southern South Africa.

Microcerus Schoenherr, 1833

Microcerus Schoenherr, 1833: 8.

Type species: *Curculio retusus* Fabricius, 1781.

Distribution. Africa, south of the Sahara.

annuliger Harold, 1880

annuliger Harold, 1880: 265.

Type depository. ZMHUB (LT, PLT).

Distribution. Kenya, Mozambique, Somalia, Tanzania, Zimbabwe.

Notes. Lectotypes designated by Louw (1986).

attenuarus Louw, 1986

attenuarus Louw, 1986: 209.

Type depositories. MRAC (HT, PT); NHM (AT); IRSNB (PT); NMBS (PT); MNHN (PT); MNHUB (PT).

Distribution. Kenya, Malawi, Zambia, Zimbabwe.

besckei Fåhraeus, 1840

besckei Fåhraeus, 1840: 729.

albiventer Gerstäcker, 1855: 84 (*Microcerus*); syn. Louw, 1986: 198.

Type depositories. *M. besckei*: NHRS (HT); *M. albiventer*: MNHUB (HT).

Distribution. Central Africa.

bicolor Louw, 1986

bicolor Louw, 1986: 217.

Type depositories. MNHM (HT, PT); NHM (AT, PT); NMBS (PT); SAMC (PT); MNHUB (PT).

Distribution. Kenya, Sudan, Tanzania.

borrei Aurivillius, 1885

borrei Aurivillius, 1885: 18.

Type depository. IRSNB (HT).

Distribution. Angola, Democratic Republic of the Congo (ex Zaire), Kenya, Mozambique, Namibia, South Africa, Tanzania, Zambia, Zimbabwe.

costalis Fåhraeus, 1871

costalis Fåhraeus, 1871: 7.

fahraei Jekel, 1875: 121 (*Microcerus*); syn. Louw, 1986: 224.

costalis fahrei: Péringuey, 1892: 71 (*Microcerus*) [lapsus].

fahraei sulcifrons Aurivillius, 1887: 73 (*Microcerus*); syn. Louw, 1986: 224.

Type depositories. *M. costalis*: NHRS (HT); *M. fahraei*: MSNG (HT); *M. fahraei sulcifrons*: unknown; *M. planifrons*: NHRS (PT).

Distribution. Botswana, Mozambique, Namibia, South Africa, Swaziland, Tanzania, Zimbabwe.

cretaceus Gyllenhal, 1840

cretaceus Gyllenhal, 1840: 727.

Type depository. SAMC (NT).

Distribution. Central South Africa.

***cribellatus* Fairmaire, 1887**

cribellatus Fairmaire, 1887: 312.

abyssinicus Fairmaire, 1893: 33 (*Microcerus*); syn. Louw, 1986: 213.

Type depositories. *M. cribellatus*: MNHN (HT); *M. abyssinicus*: MSNG (HT).

Distribution. Democratic Republic of the Congo (ex Zaire), Ethiopia, Kenya, Malawi, Mozambique, Somalia, Sudan, Tanzania, Uganda, Zambia.

***dorsofumatus* Fairmaire, 1885**

dorsofumatus Fairmaire, 1885: 147.

vermiculatus Fairmaire, 1887: 310 (*Microcerus*); syn. Louw, 1986: 196.

fissirostris Fairmaire, 1887: 313 (*Microcerus*); syn. Louw, 1986: 196.

Type depositories. *M. dorsofumatus*: MNHN (HT); *M. vermiculatus*: MNHN (HT), *M. fissirostris*: MNHN (HT).

Distribution. Kenya, Somalia, Tanzania.

***fallax* Fåhræus, 1871**

fallax Fåhræus, 1871: 8.

Type depository. NHRS (HT, PT).

Distribution. Botswana, Mozambique, South Africa, Zimbabwe.

***gracilis* Louw, 1986**

gracilis Louw, 1986: 220.

Type depositories. TMSA (HT); NHM (PT); NMBS (PT); SMWN (PT); SAMC (PT); MNHUB (PT).

Distribution. Angola, Botswana, Namibia.

griseus Gyllenhal, 1840

(Figs 4, 5)

griseus Gyllenhal, 1840: 728.*tutanus* Jekel, 1875: 127 (*Microcerus*); syn. Louw, 1986: 232.*titanus*: Giachino, 1982: 174 (*Microcerus*) [lapsus].*depressus* Aurivillius, 1885: 19 (*Microcerus*); syn. Louw, 1986: 232.**Type depositories.** *M. griseus*: NHRS (HT); *M. tutanus*: MSNG (HT); *M. depressus*: MNHUB (HT).**Distribution.** Botswana, Namibia, South Africa.***inaequalis*** Gyllenhal, 1840*inaequalis* Gyllenhal, 1840: 730.**Type depository.** NHRS (HT).**Distribution.** Central South Africa.***interstitiopunctatus*** Aurivillius, 1885*interstitiopunctatus* Aurivillius, 1885: 19.**Type depository.** NHRS (HT).**Distribution.** Botswana, South Africa, Mozambique, Zimbabwe.***latipennis*** Fåhraeus, 1871

(Figs 6, 7, 8, 9, 10)

latipennis Fåhraeus, 1871: 6.*kirschi* Jekel, 1875: 125 (*Microcerus*); syn. Louw, 1986: 180.*kirschii*: Aurivillius, 1885: 18 (*Microcerus*) [lapsus].*fossilis* Aurivillius, 1887: 61 (*Microcerus*); syn. Louw, 1986: 180.**Type depositories.** *M. latipennis*: NHRS (HT); *M. kirschi*: MSNG (HT); *M. fossilis*: IRSNB (HT).**Distribution.** Angola, Botswana, Namibia, South Africa, Zambia, Zimbabwe.

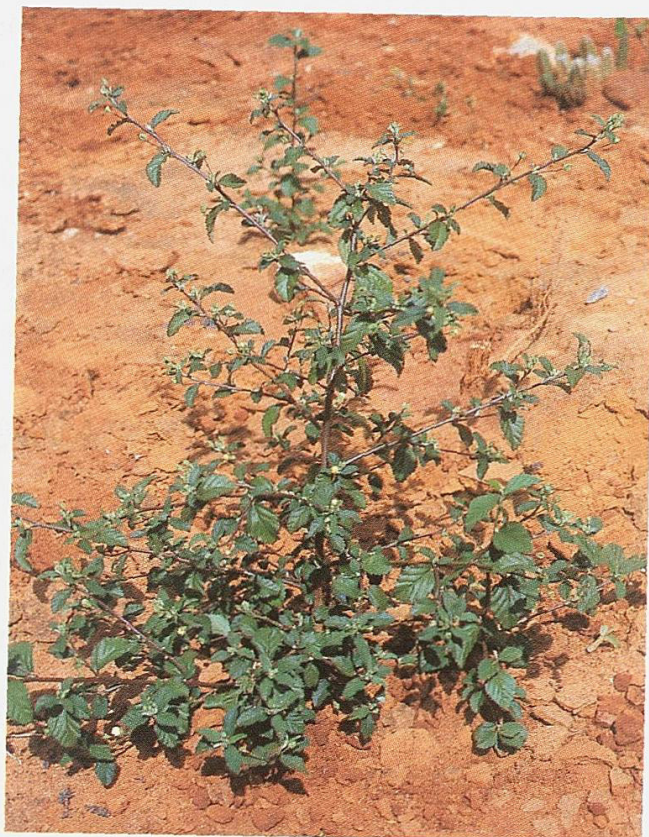


Fig. 7 - Larval hostplant of *Microcerus latipennis* Fähræus: *Sida rhombifolia* (Malvaceae); Pretoria, Gauteng Province, South Africa (photo courtesy of R. G. Oberprieler).



Fig. 8 - Late instar larva of *Microcerus latipennis* Fähræus at root of *Sida rhombifolia* (Malvaceae); Pretoria, Gauteng Province, South Africa (photo courtesy of R. G. Oberprieler).

marshalli Louw, 1986*marshalli* Louw, 1986: 204.**Type depositories.** MRAC (HT, AT, PT); NMBS (PT); IRSNB (PT); MNHN (PT); NHM (PT); NMNH (PT).**Distribution.** Angola, Democratic Republic of the Congo (ex Zaire), Zambia.*melancholicus* Gyllenhal, 1840*melancholicus* Gyllenhal, 1840: 726.**Type depository.** NMBS (NT).**Distribution.** Mozambique, South Africa.**Notes.** Neotype designated by Louw (1986).*oblongus* Aurivillius, 1885*oblongus* Aurivillius, 1885: 20.**Type depository.** NHRS (HT).**Distribution.** Southern Namibia.*pseudoannuliger* Louw, 1986*pseudoannuliger* Louw, 1986: 195.**Type depository.** NHM (HT).**Distribution.** North-eastern South Africa.*retusus* (Fabricius, 1781)*retusus* Fabricius, 1781: 195 (*Curculio*).*retusus*: Olivier, 1790: 186 (*Brachycerus*).*ephippiatus* Sparrman, 1785: 52 (*Curculio*); syn. Pape, 1910: 8.*bimaculatus* Thunberg, 1799: 33 (*Brachycerus*); syn. Aurivillius, 1887: 59.*spissus* Aurivillius, 1887: 59 (*Microcerus retusus* ssp.); syn. Louw, 1986: 186.

rotundatus Aurivillius, 1887: 60 (*Microcerus retusus* ssp.); syn. Louw, 1986: 186.

dregei Gyllenhal, 1840: 725 (*Microcerus*); syn. Pape, 1910: 7.

idolum Gyllenhal, 1833: 443 (*Microcerus*); syn. Louw, 1986: 186.

Type depositories. *C. resutus*: NHM (HT); *M. retusus spissus*: MNHUB (HT); *M. retusus rotundatus*: MNHUB (HT); *M. dregei*: unknown; *M. idolum*: NHRS (HT).

Distribution. Central and southern South Africa.

spiniger cavirostris Fairmaire, 1887

cavirostris Fairmaire, 1887: 312.

tuberifrons Fairmaire, 1893: 154 (*Microcerus*); syn. Louw, 1986: 180.

Type depositories. *M. cavirostris*: MNHN (HT); *M. tuberifrons*: unknown.

Distribution. Eastern Africa.

spiniger spiniger Gerstäcker, 1855

spiniger Gerstäcker, 1855: 83.

Type depository. MNHUB (HT).

Distribution. Angola, Botswana, western Democratic Republic of the Congo (ex Zaire), southern Malawi, northern Mozambique, north-eastern Namibia, northern South Africa, southern Zambia, Zimbabwe.

subcaudatus Gerstäcker, 1855

subcaudatus Gerstäcker, 1855: 38.

pascoei Jekel, 1875: 121 (*Microcerus*); syn. Louw, 1986: 228.

planifrons Aurivillius, 1887: 74 (*Microcerus*); syn. Louw, 1986: 228.

Type depositories. *M. subcaudatus*: MNHUB (HT); *M. pascoei*: MSNG (HT); *M. planifrons*: NHRS (HT).

Distribution. Angola, Botswana, Democratic Republic of the Congo (ex Zaire), Malawi, Mozambique, Namibia, South Africa, Swaziland, Tanzania, Zambia, Zimbabwe.



Fig. 9 - Larval feeding scar (root girdling) of *Microcerus latipennis* Fähræus on taproot of *Sida rhombifolia* (Malvaceae); Pretoria, Gauteng Province, South Africa (photo courtesy of R. G. Oberprieler).



Fig. 10 - Habitat of *Microcerus latipennis* Fähræus: Lapalala Wilderness Reserve, Waterberg, Limpopo Province, South Africa (photo courtesy of R. G. Oberprieler).

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In bold small caps: valid subfamilies and tribes; in bold italics: valid genera; in italics: valid species; in roman: subfamily, tribe, genus, and species synonym names; the two dots sign indicates a lapsus. The italic page numbers indicate a quotation of the taxon in the text; the roman page numbers refer to the taxon in the checklist.

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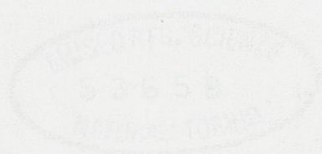
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La famiglia Brentidae trattata in questo volume comprende le seguenti sottofamiglie e tribù: Brentinae, Cyphagoginae, Pholidochlamyinae, Taphroderinae, Trachelizinae, Ulocerinae (queste 6 sottofamiglie corrispondono ai Brentidae *sensu Auctorum*), più Antliarhininae, Cyladinae, Eurhynchinae e Microcerini.

In questo libro, che aggiorna l'ultima checklist della famiglia, pubblicata da Richard Kleine nel 1938, vi sono anche capitoli sulla morfologia, la biologia, l'ecologia, l'etologia e la zoogeografia, così come una parte storica sugli specialisti che si sono occupati dei brentidi e una bibliografia completa sulla famiglia. Tutti i taxa descritti sono elencati nella checklist.

Attualmente i Brentidae *sensu Auctorum* (Brentinae, Cyphagoginae, Pholidochlamyinae, Taphroderinae, Trachelizinae, Ulocerinae) includono 293 generi con 1690 specie valide, per la maggior parte a distribuzione circumtropicale. Gli Antliarhininae sono tutti africani, con 2 generi e 7 specie, e sono associati esclusivamente alle cicadee del genere *Encephalartos*. I Cyladinae consistono del solo genere circumtropicale *Cylas*, con 24 specie. Gli Eurhynchinae comprendono 3 generi con 30 specie di Australia e Nuova Guinea. Infine, la tribù Microcerini raggruppa 3 generi con 67 specie a distribuzione sub-Sahariana.

Alessandra Sforzi, nata a Pistoia il 1 Ottobre 1966, ha iniziato a studiare i Brentidae nel 1991, dopo aver conseguito la Laurea in Scienze Biologiche presso l'Università degli Studi di Firenze. Attualmente ha un contratto come Ricercatrice presso la Sezione di Zoologia del Museo di Storia Naturale dell'Università di Firenze. Ha partecipato a numerose spedizioni scientifiche in vari paesi dell'Africa, Asia e Sud America e ha studiato le collezioni di brentidi dei principali musei europei. Ha pubblicato numerosi lavori sulla tassonomia e faunistica della famiglia, descrivendo diversi nuovi taxa.

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I capitoli su Antliarhininae e Eurhynchinae sono opera di Rolf Oberprieler, Ricercatore presso la Sezione di Entomologia dello CSIRO di Canberra, Australia, e specialista della sistematica dei Curculionoidea.

Il capitolo sui Cyladinae è stato scritto da Miguel Angel Alonso-Zarazaga, Entomologo presso il Museo Nazionale di Scienze Naturali di Madrid, Spagna, e specialista della sistematica dei Curculionoidea e della tassonomia di Apionidae e Naphyidae.

Il capitolo sui Microcerini è opera di Schalk vdM Louw, Professore presso il Dipartimento di Zoologia e Entomologia della Università del Free State, Bloemfontein, Sud Africa, specialista della sistematica e tassonomia dei Curculionoidea, e delle interazioni insetto-pianta.

In copertina:

Estenorhynus sp., Ecuador (disegno di Cristina Girard)

Retro di copertina:

Nemorhynus myrmecophagus (Herbst) (tavola da Labram & Imhoff, 1838)

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